

Published in final edited form as:

Nat Genet. 2010 October ; 42(10): 885–892. doi:10.1038/ng.669.

A locus on 19p13 modifies risk of breast cancer in *BRCA1* mutation carriers and is associated with hormone receptor–negative breast cancer in the general population

Antonis C Antoniou¹, Xianshu Wang², Zachary S Fredericksen³, Lesley McGuffog¹, Robert Tarrell³, Olga M Sinilnikova^{4,5}, Sue Healey⁶, Jonathan Morrison¹, Christiana Kartsonaki¹, Timothy Lesnick³, Maya Ghoussaini⁶, Daniel Barrowdale¹, EMBRACE^{1,133}, Susan Peock¹, Margaret Cook¹, Clare Oliver¹, Debra Frost¹, Diana Eccles⁷, D Gareth Evans⁸, Ros Eeles⁹, Louise Izatt¹⁰, Carol Chu¹¹, Fiona Douglas¹², Joan Paterson¹³, Dominique Stoppa-Lyonnet¹⁴, Claude Houdayer¹⁴, Sylvie Mazoyer⁵, Sophie Giraud⁴, Christine Lasset^{15,16}, Audrey Remenieras¹⁷, Olivier Caron¹⁸, Agnès Hardouin¹⁹, Pascaline Berthet¹⁹, GEMO Study Collaborators^{20,133}, Frans B L Hogervorst²¹, Matti A Rookus²², Agnes Jager²³, Ans van den Ouweland²⁴, Nicoline Hoogerbrugge²⁵, Rob B van der Luijt²⁶, Hanne Meijers-Heijboer²⁷, Encarna B Gómez García²⁸, HEBON^{29,133}, Peter Devilee^{30,31}, Maaïke P G Vreeswijk³², Jan Lubinski³³, Anna Jakubowska³³, Jacek Gronwald³³, Tomasz Huzarski³³, Tomasz Byrski³³, Bohdan Górski³³, Cezary Cybulski³³, Amanda B Spurdle⁶, Helene Holland⁶, kConFab^{34,133}, David E Goldgar³⁵, Esther M John³⁶, John L Hopper³⁷, Melissa Southey³⁷, Sandra S Buys³⁸, Mary B Daly³⁹, Mary-Beth Terry⁴⁰, Rita K Schmutzler^{41,42}, Barbara Wappenschmidt^{41,42}, Christoph Engel⁴³, Alfons Meindl⁴⁴, Sabine Preisler-Adams⁴⁵, Norbert Arnold⁴⁶, Dieter Niederacher⁴⁷, Christian Sutter⁴⁸, Susan M Domchek⁴⁹, Katherine L Nathanson⁴⁹, Timothy Rebbeck⁴⁹, Joanne L Blum⁵⁰, Marion Piedmonte⁵¹, Gustavo C Rodriguez⁵², Katie Wakeley⁵³, John F Boggess⁵⁴, Jack Basil⁵⁵, Stephanie V

© 2010 Nature America, Inc. All rights reserved.

Correspondence should be addressed to F.J.C. (couch.fergus@mayo.edu).

¹³³A full list of members is provided in the supplementary Note.

Note: Supplementary information is available on the Nature Genetics website.

AUTHOR CONTRIBUTIONS

F.J.C., A.C.A. and D.F.E. designed the study and obtained financial support. G.C.-T. founded CIMBA in order to provide the infrastructure for the *BRCA1* GWAS. F.J.C. and X.W. coordinated collection of samples. A.C.A. directed the statistical analysis. D.F.E. advised on the statistical analysis. C.K., Z.S.F. and T.L. carried out analyses. Z.S.F., R.T., J.M., L.M. and D.B. provided bioinformatics and database support. F.J.C., H. Hakonarson and X.W. directed the genotyping of the *BRCA1* carrier and triple-negative samples. M.G. directed the genotyping of the UK case-control samples. A.C.A., F.J.C. and D.F.E. drafted the manuscript. F.J.C. was the overall project leader.

O.M.S. and S.H. coordinated the *BRCA1* mutation classification. T.K., J.V., M.M.G., D.A. and C.G. were involved in the *BRCA2* GWAS genotyping and coordination. K.O. led the *BRCA2* GWAS.

S.P., M.C., C.O., D.F., D.E., D.G.E., R.E., L.I., C.C., F.D., J.P., O.M.S., D.S.-L., C.H., S.M., S.G., C.L., A.R., O.C., A.H., P.B., F.B.L.H., M.A.R., A.J., A.v.d.O., N.H., R.B.v.d.L., H.M.-H., E.B.G.G., P.D., M.P.G.V., J.L., A.J., J.G., T.H., T.B., B.G., C.C., A.B.S., H.H., D.E., E.M.J., J.L.H., M.S., S.S.B., M.B.D., M.-B.T., R.K.S., B.W., C.E., A.M., S.P.-A., N.A., D.N., C.S., S.M.D., K.L.N., T.R., J.L.B., M.P., G.C.R., K.W., J.F.B., J.B., S.V.B., E.F., B.K., Y.L., R.M., I.L.A., G.G., H.O., N.L., K.H., J.R., H.E., A.-M.G., M.T., L.S., P.P., S.M., B.B., A.V., P.R., T.C., M.d.I.H., C.F.S., A.F.-R., M.H.G., P.L.M., J.T.L., L.G., N.M.L., T.V.O.H., F.C.N., I.B., C.L., J.G., S.J.R., S.A.G., C.P., S.N., C.I.S., J.B., A.O., H.N., T.H., M.A.C., M.S.B., U.H., A.K.G., M.M., C.C., S.L.N., B.Y.K., N.T., A.E.T., J.W., O.O., J.S., P.S., W.S.R., A.A. and G.R. collected data and samples on *BRCA1* and/or *BRCA2* mutation carriers.

N.G.M., G.W.M., J.C.-C., D.F.-J., H.B., G.S., L.B., A.C., S.S.C., P.M., S.M.G., W.T., D.Y., G.F., P.A.F., M.W.B., I.d.S.S., J.P., D.L., R.P., T.R., A.F., R.W., K.P., R.B.D., A.M.L., J.E.-P., C.V., F.B., K.D., A.D. and P.P.D.P. collected data and samples for the TNBCC case-control and/or the SEARCH studies.

All authors provided critical review of the manuscript.

COMPETING FINANCIAL INTERESTS

The authors declare no competing financial interests.

Reprints and permissions information is available online at <http://npg.nature.com/reprintsandpermissions/>.

Blank⁵⁶, Eitan Friedman⁵⁷, Bella Kaufman⁵⁸, Yael Laitman⁵⁷, Roni Milgrom⁵⁷, Irene L Andrulis^{59,60,61}, Gord Glendon⁵⁹, Hilmi Ozcelik⁶⁰, Tomas Kirchhoff^{62,63}, Joseph Vijai^{62,63}, Mia M Gaudet^{64,65}, David Altshuler⁶⁶, Candace Guiducci⁶⁶, SWE-BRCA^{67,133}, Niklas Loman⁶⁸, Katja Harbst⁶⁸, Johanna Rantala⁶⁹, Hans Ehrencrona⁷⁰, Anne-Marie Gerdes⁷¹, Mads Thomassen⁷², Lone Sunde⁷³, Paolo Peterlongo^{74,75}, Siranoush Manoukian⁷⁶, Bernardo Bonanni⁷⁷, Alessandra Viel⁷⁸, Paolo Radice^{74,75}, Trinidad Caldes⁷⁹, Miguel de la Hoya⁷⁷, Christian F Singer⁸⁰, Anneliese Fink-Retter⁸⁰, Mark H Greene⁸¹, Phuong L Mai⁸¹, Jennifer T Loud⁸¹, Lucia Guidugli², Noralane M Lindor⁸², Thomas V O Hansen⁸³, Finn C Nielsen⁸³, Ignacio Blanco⁸⁴, Conxi Lazaro⁸⁴, Judy Garber⁸⁵, Susan J Ramus⁸⁶, Simon A Gayther⁸⁶, Catherine Phelan⁸⁷, Stephen Narod⁸⁸, Csilla I Szabo², MOD SQUAD^{89,133}, Javier Benitez⁹⁰, Ana Osorio⁹⁰, Heli Nevanlinna⁹¹, Tuomas Heikkinen⁹¹, Maria A Caligo⁹², Mary S Beattie^{93,94,95}, Ute Hamann⁹⁶, Andrew K Godwin³⁹, Marco Montagna⁹⁷, Cinzia Casella⁹⁷, Susan L Neuhausen⁹⁸, Beth Y Karlan^{99,100}, Nadine Tung¹⁰¹, Amanda E Toland¹⁰², Jeffrey Weitzel¹⁰³, Olofunmilayo Olopade¹⁰⁴, Jacques Simard^{105,106}, Penny Soucy¹⁰⁵, Wendy S Rubinstein¹⁰⁷, Adalgeir Arason¹⁰⁸, Gad Rennert¹⁰⁹, Nicholas G Martin¹¹⁰, Grant W Montgomery¹¹⁰, Jenny Chang-Claude¹¹¹, Dieter Flesch-Janys¹¹², Hiltrud Brauch^{113,114}, GENICA^{115,133}, Gianluca Severi¹¹⁶, Laura Baglietto¹¹⁶, Angela Cox¹¹⁷, Simon S Cross¹¹⁸, Penelope Miron¹¹⁹, Sue M Gerty⁷, William Tapper⁷, Drakoulis Yannoukakos¹²⁰, George Fountzilas^{121,122}, Peter A Fasching¹²³, Matthias W Beckmann¹²⁴, Isabel dos Santos Silva¹²⁵, Julian Peto¹²⁵, Diether Lambrechts¹²⁶, Robert Paridaens¹²⁷, Thomas Rüdiger¹²⁸, Asta Försti^{96,129}, Robert Winqvist¹³⁰, Katri Pylkäs¹³⁰, Robert B Diasio¹³¹, Adam M Lee¹³¹, Jeanette Eckel-Passow³, Celine Vachon³, Fiona Blows¹, Kristy Driver¹, Alison Dunning¹, Paul P D Pharoah¹, Kenneth Offit⁶¹, V Shane Pankratz³, Hakon Hakonarson¹³², Georgia Chenevix-Trench⁶, Douglas F Easton¹, and Fergus J Couch²

¹ Centre for Cancer Genetic Epidemiology, Department of Public Health and Primary Care, University of Cambridge, Cambridge, UK ² Department of Laboratory Medicine and Pathology, Mayo Clinic, Rochester, Minnesota, USA ³ Department of Health Sciences Research, Mayo Clinic, Rochester, Minnesota, USA ⁴ Unité Mixte de Génétique Constitutionnelle des Cancers Fréquents, Centre Hospitalier Universitaire de Lyon/Centre Léon Bérard, Lyon, France ⁵ Equipe labellisée LIGUE 2008, UMR5201 CNRS, Centre Léon Bérard, Université de Lyon, Lyon, France ⁶ Queensland Institute of Medical Research, Brisbane, Australia ⁷ Wessex Clinical Genetics Service, Princess Anne Hospital, Southampton, UK ⁸ Genetic Medicine, Manchester Academic Health Sciences Centre, Central Manchester University Hospitals National Health Service (NHSFT) Foundation Trust, Manchester, UK ⁹ Oncogenetics Team, The Institute of Cancer Research and Royal Marsden NHS Foundation Trust, Sutton, UK ¹⁰ Clinical Genetics, Guy's and St. Thomas' NHS Foundation Trust, London, UK ¹¹ Yorkshire Regional Genetics Service, Leeds, UK ¹² Institute of Human Genetics, Centre for Life, Newcastle Upon Tyne Hospitals NHS Trust, Newcastle upon Tyne, UK ¹³ Department of Clinical Genetics, East Anglian Regional Genetics Service, Addenbrookes Hospital, Cambridge, UK ¹⁴ INSERM U509, Service de Génétique Oncologique, Institut Curie, Université Paris-Descartes, Paris, France ¹⁵ CNRS UMR5558, Université Lyon 1, Lyon, France ¹⁶ Unité de Prévention et d'Epidémiologie Génétique, Centre Léon Bérard, Lyon, France ¹⁷ Genetics Department, Institut de Cancérologie Gustave Roussy, Villejuif, France ¹⁸ Consultation de Génétique, Département de Médecine, Institut de Cancérologie Gustave Roussy, Villejuif, France ¹⁹ Centre François Baclesse, Caen, France ²⁰ GEMO study, Cancer Genetics Network 'Groupe Génétique et Cancer', Fédération Nationale des Centres de Lutte Contre le Cancer, Paris, France ²¹ Family Cancer Clinic, Netherlands Cancer Institute, Amsterdam, The Netherlands ²² Department of Epidemiology, Netherlands Cancer Institute, Amsterdam, The Netherlands ²³ Department of Medical Oncology, Rotterdam Family Cancer Clinic, Erasmus University Medical Center, Rotterdam, The Netherlands ²⁴ Department of Clinical Genetics, Rotterdam Family Cancer Clinic, Erasmus University Medical Center, Rotterdam, The Netherlands ²⁵ Department of Human Genetics, Radboud University Nijmegen

Medical Centre, Nijmegen, The Netherlands²⁶ Department of Medical Genetics, University Medical Center Utrecht, Utrecht, The Netherlands²⁷ Department of Clinical Genetics, Vrije Universiteit (VU) Medical Center, Amsterdam, The Netherlands²⁸ Department of Clinical Genetics, School for Oncology and Developmental Biology, Maastricht University Medical Center, Maastricht, The Netherlands²⁹ HEBOON, Hereditary Breast and Ovarian Cancer Research Group, The Netherlands³⁰ Department of Human Genetics, Leiden University Medical Center, Leiden, The Netherlands³¹ Department of Pathology, Leiden University Medical Center, Leiden, The Netherlands³² Department of Toxicogenetics, Leiden University Medical Center, Leiden, The Netherlands³³ International Hereditary Cancer Center, Department of Genetics and Pathology, Pomeranian Medical University, Szczecin, Poland³⁴ kConFab, Kathleen Cuninghame Foundation Consortium for Research into Familial Breast Cancer, Peter MacCallum Cancer Centre, Melbourne, Australia³⁵ Department of Dermatology, University of Utah School of Medicine, Salt Lake City, Utah, USA³⁶ Cancer Prevention Institute of California, Stanford University School of Medicine, Stanford, California, USA³⁷ The University of Melbourne, Melbourne, Australia³⁸ Huntsman Cancer Institute, University of Utah Health Sciences Centre, Salt Lake City, Utah, USA³⁹ Fox Chase Cancer Center, Philadelphia, Pennsylvania, USA⁴⁰ Columbia University, New York, New York, USA⁴¹ Centre of Familial Breast and Ovarian Cancer, Department of Obstetrics and Gynaecology, University of Cologne, Cologne, Germany⁴² Centre for Integrated Oncology (CIO), University of Cologne, Cologne, Germany⁴³ Institute for Medical Informatics, Statistics and Epidemiology, University of Leipzig, Leipzig, Germany⁴⁴ Department of Obstetrics and Gynaecology, Division of Tumor Genetics, Klinikum rechts der Isar, Technical University Munich, Munich, Germany⁴⁵ Institute of Human Genetics, University of Muenster, Muenster, Germany⁴⁶ Department of Obstetrics and Gynaecology, University Hospital of Schleswig-Holstein, Campus Kiel, Christian-Albrechts University, Kiel, Germany⁴⁷ Department of Obstetrics and Gynaecology, Division of Molecular Genetics, University Hospital Düsseldorf, Heinrich-Heine University Düsseldorf, Dusseldorf, Germany⁴⁸ Institute of Human Genetics, Division of Molecular Diagnostics, University Heidelberg, Heidelberg, Germany⁴⁹ University of Pennsylvania, Philadelphia, Pennsylvania, USA⁵⁰ Baylor-Charles A. Sammons Cancer Center, Dallas, Texas, USA⁵¹ Gynecology Oncology Group Statistical and Data Center, Roswell Park Cancer Institute, Buffalo, New York, USA⁵² Northshore University Health System, Evanston Northwestern Healthcare, Evanston, Illinois, USA⁵³ Tufts University, New England Medical Center, Boston, Massachusetts, USA⁵⁴ University of North Carolina, Chapel Hill, North Carolina, USA⁵⁵ St. Elizabeth Medical Center, Edgewood, Kentucky, USA⁵⁶ New York University School of Medicine, New York, New York, USA⁵⁷ The Susanne Levy Gertner Oncogenetics Unit, Sheba Medical Center, Tel-Hashomer, Israel⁵⁸ Oncology Institute, Sheba Medical Center, Tel-Hashomer, Israel⁵⁹ Ontario Cancer Genetics Network, Cancer Care Ontario, Toronto, Ontario, Canada⁶⁰ Fred A. Litwin Center for Cancer Genetics, Samuel Lunenfeld Research Institute, Mount Sinai Hospital, Toronto, Ontario, Canada⁶¹ Department of Molecular Genetics, University of Toronto, Toronto, Ontario, Canada⁶² Clinical Genetics Service, Memorial Hospital, New York, New York, USA⁶³ Cancer Biology and Genetics Program, Sloan-Kettering Institute, Memorial Sloan-Kettering Cancer Center, New York, New York, USA⁶⁴ Department of Epidemiology and Population Health, Albert Einstein College of Medicine, New York, New York, USA⁶⁵ Department of Obstetrics and Gynecology and Women's Health, Albert Einstein College of Medicine, New York, New York, USA⁶⁶ Broad Institute of Harvard and Massachusetts Institute of Technology (MIT), Cambridge, Massachusetts, USA⁶⁷ SWE-BRCA, Swedish Breast Cancer Study, Sweden⁶⁸ Department of Oncology, Lund University, Lund, Sweden⁶⁹ Department of Clinical Genetics, Karolinska University Hospital, Stockholm, Sweden⁷⁰ Department of Genetics and Pathology Rudbeck Laboratory, Uppsala University, Uppsala, Sweden⁷¹ Department of Clinical Genetics, Rigshospitalet, Copenhagen, Denmark⁷² Department of Clinical Genetics, Odense University Hospital, Odense, Denmark⁷³ Department of Clinical Genetics, Aalborg and Aarhus University Hospital, Aarhus, Denmark⁷⁴ Unit of Genetic Susceptibility to Cancer, Department of

Experimental Oncology and Molecular Medicine, Fondazione Istituto di Ricovero e Cura a Carattere Scientifico (IRCCS), Istituto Nazionale Tumori (INT), Milan, Italy ⁷⁵ IFOM, Fondazione Istituto Fondazione Italiana per la Ricerca sul Cancro (FIRC) di Oncologia Molecolare, Milan, Italy ⁷⁶ Unit of Medical Genetics, Department of Preventive and Predictive Medicine, Fondazione IRCCS Istituto Nazionale dei Tumori (INT), Milan, Italy ⁷⁷ Division of Cancer Prevention and Genetics, Istituto Europeo di Oncologia (IEO), Milan, Italy ⁷⁸ Division of Experimental Oncology, Centro di Riferimento Oncologico (CRO), IRCCS, Aviano (PN), Italy ⁷⁹ Hospital Clinico San Carlos, Madrid, Spain ⁸⁰ Department of Obstetrics/Gynecology, Medical University of Vienna, Vienna, Austria ⁸¹ Clinical Genetics Branch, Division of Cancer Epidemiology and Genetics, National Cancer Institute, Bethesda, Maryland, USA ⁸² Medical Genetics, Mayo Clinic Rochester, Rochester, Minnesota, USA ⁸³ Department of Clinical Biochemistry, Copenhagen University Hospital, Copenhagen, Denmark ⁸⁴ Hereditary Cancer Program, Catalan Institute of Oncology, Gran Via de l'Hospitalet, Barcelona, Spain ⁸⁵ Dana-Farber Cancer Institute, Boston, Massachusetts, USA ⁸⁶ Department of Gynaecological Oncology, University College London, Elizabeth Garrett Anderson (EGA) Institute for Women's Health, London, UK ⁸⁷ Women's College Research Institute, Toronto, Ontario, Canada ⁸⁸ Department of Epidemiology and Genetics, H. Lee Moffitt Cancer Center, Tampa, Florida, USA ⁸⁹ MOD SQUAD, Modifier Study of Quantitative Effects on Disease ⁹⁰ Human Cancer Genetics Programme, Spanish National Cancer Research Centre, Madrid, Spain ⁹¹ Helsinki University Central Hospital, Department of Obstetrics and Gynecology, Helsinki, Finland ⁹² Division of Surgical, Molecular and Ultrastructural Pathology, Department of Oncology, University of Pisa, Pisa University Hospital, Pisa, Italy ⁹³ Department of Medicine, University of California, San Francisco, San Francisco, California, USA ⁹⁴ Department of Epidemiology and Biostatistics, University of California, San Francisco, San Francisco, California, USA ⁹⁵ Cancer Risk Program, Helen Diller Family Cancer Center, University of California, San Francisco, San Francisco, California, USA ⁹⁶ German Cancer Research Center (DKFZ), Heidelberg, Germany ⁹⁷ Istituto Oncologico Veneto-IRCCS, Immunology and Molecular Oncology Unit, Padua, Italy ⁹⁸ Department of Population Sciences, The Beckman Research Institute of the City of Hope, Duarte, California, USA ⁹⁹ Women's Cancer Research Institute at the Samuel Oschin Cancer Institute, Division of Gynecologic Oncology, Cedars-Sinai Medical Center, Los Angeles, California, USA ¹⁰⁰ Department of Obstetrics and Gynecology, David Geffen School of Medicine at the University of California Los Angeles, Los Angeles, California, USA ¹⁰¹ Beth Israel Deaconess Medical Center, Boston, Massachusetts, USA ¹⁰² Comprehensive Cancer Center, Department of Molecular Virology, Immunology and Medical Genetics, Division of Human Genetics, Department of Internal Medicine, Ohio State University, Columbus, Ohio, USA ¹⁰³ Division of Clinical Cancer Genetics, City of Hope Cancer Center, Duarte, California, USA ¹⁰⁴ University of Chicago Medical Center, Chicago, Illinois, USA ¹⁰⁵ Cancer Genomics Laboratory, Centre Hospitalier Universitaire de Quebec, Québec City, Quebec, Canada ¹⁰⁶ Université Laval, Centre de recherche du Centre Hospitalier Universitaire de Québec (CHUQ), Québec City, Quebec, Canada ¹⁰⁷ Northshore University Health System, Evanston, Illinois, USA ¹⁰⁸ Department of Pathology, Landspítali University Hospital, Reykjavik, Iceland ¹⁰⁹ QIMR GWAS Collective, Queensland Institute of Medical Research, Brisbane, Australia ¹¹⁰ Clalit Health Services National Cancer Control Center, Department of Community Medicine and Epidemiology, Carmel Medical Center and B. Rappaport Faculty of Medicine, Technion, Haifa, Israel ¹¹¹ Division of Cancer Epidemiology, German Cancer Research Center, Heidelberg, Germany ¹¹² Institute for Medical Biometrics and Epidemiology, University Clinic Hamburg-Eppendorf, Hamburg, Germany ¹¹³ Dr. Margarete Fischer-Bosch-Institute of Clinical Pharmacology, Stuttgart, Germany ¹¹⁴ University Tübingen, Tübingen, Germany ¹¹⁵ GENICA, Gene Environment Interaction and Breast Cancer in Germany ¹¹⁶ Melbourne Collaborative Cohort Study (MCCS), Cancer Epidemiology Centre, The Cancer Council Victoria, Melbourne, Australia ¹¹⁷ Institute for Cancer Studies, Department of Oncology, Faculty of Medicine, Dentistry and Health, University of Sheffield, Sheffield, UK ¹¹⁸ Academic Unit of Pathology, Department of Neuroscience, Faculty of Medicine,

Dentistry and Health, University of Sheffield, Sheffield, UK ¹¹⁹ Department of Cancer Biology, Dana-Farber Cancer Institute, Boston, Massachusetts, USA ¹²⁰ Molecular Diagnostics Laboratory Institute of Radioisotopes and Radiodiagnostic Products, National Centre for Scientific Research 'Demokritos', Athens, Greece ¹²¹ Department of Medical Oncology, Papageorgiou Hospital, Aristotle University of Thessaloniki School of Medicine, Thessaloniki, Greece ¹²² Hellenic Cooperative Oncology Group, Greece ¹²³ University of California at Los Angeles, David Geffen School of Medicine, Department of Medicine, Division of Hematology and Oncology, Los Angeles, California, USA ¹²⁴ University Hospital Erlangen, Department of Gynecology and Obstetrics, Erlangen, Germany ¹²⁵ Cancer Research UK Epidemiology and Genetics Group, Department of Epidemiology and Population Health, London School of Hygiene and Tropical Medicine, London, UK ¹²⁶ Vesalius Research Center (VRC), VIB, Leuven, Belgium ¹²⁷ Multidisciplinary Breast Center, University Hospitals Leuven, Leuven, Belgium ¹²⁸ Institute of Pathology, Städtisches Klinikum Karlsruhe, Karlsruhe, Germany ¹²⁹ Center for Primary Health Care Research, University of Lund, Lund, Sweden ¹³⁰ Laboratory of Cancer Genetics, Department of Clinical Genetics and Biocenter Oulu, University of Oulu, Oulu University Hospital, Oulu, Finland ¹³¹ Department of Pharmacology, Mayo Clinic, Rochester, Minnesota, USA ¹³² Center for Applied Genomics, Children's Hospital of Philadelphia, Philadelphia, Pennsylvania, USA

Abstract

Germline *BRCA1* mutations predispose to breast cancer. To identify genetic modifiers of this risk, we performed a genome-wide association study in 1,193 individuals with *BRCA1* mutations who were diagnosed with invasive breast cancer under age 40 and 1,190 *BRCA1* carriers without breast cancer diagnosis over age 35. We took forward 96 SNPs for replication in another 5,986 *BRCA1* carriers (2,974 individuals with breast cancer and 3,012 unaffected individuals). Five SNPs on 19p13 were associated with breast cancer risk ($P_{\text{trend}} = 2.3 \times 10^{-9}$ to $P_{\text{trend}} = 3.9 \times 10^{-7}$), two of which showed independent associations (rs8170, hazard ratio (HR) = 1.26, 95% CI 1.17–1.35; rs2363956 HR = 0.84, 95% CI 0.80–0.89). Genotyping these SNPs in 6,800 population-based breast cancer cases and 6,613 controls identified a similar association with estrogen receptor–negative breast cancer (rs2363956 per-allele odds ratio (OR) = 0.83, 95% CI 0.75–0.92, $P_{\text{trend}} = 0.0003$) and an association with estrogen receptor–positive disease in the opposite direction (OR = 1.07, 95% CI 1.01–1.14, $P_{\text{trend}} = 0.016$). The five SNPs were also associated with triple-negative breast cancer in a separate study of 2,301 triple-negative cases and 3,949 controls ($P_{\text{trend}} = 1 \times 10^{-7}$ to $P_{\text{trend}} = 8 \times 10^{-5}$; rs2363956 per-allele OR = 0.80, 95% CI 0.74–0.87, $P_{\text{trend}} = 1.1 \times 10^{-7}$).

Pathogenic *BRCA1* and *BRCA2* mutations confer high risks of breast and ovarian cancer. Variation in risk estimates by degree of family history suggests that these risks are modified by other genetic variants^{1–5}. Recent studies from the Consortium of Investigators of Modifiers of *BRCA1/2* (CIMBA) have demonstrated that common breast cancer susceptibility alleles, identified through genome-wide association studies (GWAS) in the general population^{6–9}, are also associated with the risk of developing breast cancer in *BRCA1* or *BRCA2* mutation carriers^{10,11}. However, although five of six alleles were associated with risk of breast cancer for *BRCA2* mutation carriers, only two polymorphisms (in the *TOX3* and 2q35 regions) were associated with risk for *BRCA1* carriers. These findings are consistent with the distinct pathology of breast cancer in *BRCA1* tumors^{12,13} and suggest that the genetic variants that modify breast cancer risk for *BRCA1* mutation carriers may differ from the modifiers of risk for *BRCA2* carriers or for non-carriers.

To search for genetic loci associated with breast cancer in *BRCA1* carriers, we conducted a two-stage GWAS. In stage 1, we genotyped 2,500 *BRCA1* carriers using the Illumina

Infinium 610K array, which included 620,901 SNPs. Mutation carriers were selected on the basis of an invasive breast cancer diagnosis at under 40 years of age ($n = 1,250$) or the absence of breast cancer when 35 years of age or older ($n = 1,250$). After quality control exclusions, 2,383 carriers (1,193 unaffected and 1,190 affected) from 20 centers in 11 different countries and 555,616 SNPs were available for analysis (Supplementary Tables 1 and 2). Genotype associations were evaluated using a 1 degree-of-freedom (d.f.) score test for trend, based on modeling the retrospective likelihood of the observed genotypes conditional on the disease phenotypes, stratified by country of residence. A kinship-adjusted version of the score test statistic was used to allow for the dependence between related individuals.

There was little evidence for inflation in the test statistic of association (inflation factor (λ) = 1.036; Supplementary Fig. 1). Ninety-six SNPs were significant at the $P < 10^{-4}$ level compared with 55.6 SNPs which were expected by chance. In stage 2, we genotyped 86 of these SNPs, seven surrogate SNPs (within 10 kb of the significant SNPs and pair-wise $r^2 > 0.90$) and three additional SNPs in 6,332 *BRCA1* carriers. After quality control exclusions, 89 SNPs and 5,986 *BRCA1* mutation carriers (3,012 unaffected and 2,974 affected) were used in the stage 2 analysis. The most significant associations were for five SNPs on 19p13 ($P < 0.002$), which had hazard ratios in the same direction as in stage 1 (Table 1 and Supplementary Table 3). In the combined analysis of stage 1 and 2, there was strong evidence of association¹⁴ with breast cancer for these SNPs ($P = 2.3 \times 10^{-9}$ to $P = 3.9 \times 10^{-7}$).

The minor alleles of rs8170 and rs4808611 were associated with an increased breast cancer risk for *BRCA1* carriers (per allele HR = 1.26, 95% CI 1.17–1.35 for both SNPs). In contrast, SNPs rs8100241, rs2363956 and rs3745185 were associated with decreased breast cancer risk (HR = 0.84, 95% CI 0.80–0.89 for rs8100241 and rs2363956; HR = 0.86, 95% CI 0.81–0.91 for rs3745185) (Table 1). The HR estimates for rs8170 and rs4808611 were similar in stages 1 and 2, but for rs8100241, rs2363956 and rs3745185, the HRs were stronger in stage 1; this may be due to the sample selection criteria for stage 1 or a ‘winner’s curse’ effect¹⁵. There was no evidence of heterogeneity in the HR estimates among the countries of residence in stages 1 and 2 combined (Fig. 1; rs8170, $P = 0.10$; rs4808611, $P = 0.14$; rs8100241, $P = 0.18$; rs2363956, $P = 0.17$; and rs3745185, $P = 0.48$).

The strength of the association with breast cancer could also be affected by the inclusion of prevalent cases if these SNPs were associated with breast cancer survival. To address this possibility, we excluded breast cancer cases diagnosed with the disease >5 years before study entry. The HR estimates were similar to the overall analysis after this exclusion (Supplementary Table 4). This indicates that the inclusion of prevalent breast cancer cases was unlikely to have influenced the overall results.

To investigate whether any of these SNPs were associated with ovarian cancer risk for *BRCA1* carriers, we analyzed the data within a competing risks framework and estimated HR simultaneously for breast and ovarian cancer. There was no evidence of association with ovarian cancer risk for any of the SNPs, and the breast cancer associations were virtually identical to the primary analysis both in terms of significance and in the HR estimates (Table 2). We repeated the breast cancer association analysis after excluding all individuals who developed ovarian cancer either before or after a breast cancer diagnosis. Despite the sample size reduction, the top four SNPs remained significant at $P < 10^{-7}$ and the HR estimates were identical to the analysis which included individuals with ovarian cancer as unaffected individuals (Supplementary Table 4). We also evaluated ovarian cancer associations after excluding individuals with ovarian cancer who were recruited >3 years after their cancer diagnosis in order to account for a potential survival bias. No significant associations were

observed after this exclusion ($P_{\text{trend}} = 0.44$ to $P_{\text{trend}} = 0.96$ using competing risk analysis). We conclude that the associations with breast cancer were not confounded by the competing risk of ovarian cancer.

We evaluated the SNP associations by the predicted functional consequences of *BRCA1* mutation type^{16–18}. Class 1 mutations correspond to loss-of-function mutations and are expected to result in a reduced transcript or protein level due to nonsense-mediated RNA decay, whereas class 2 mutations are likely to generate stable proteins with potential residual or dominant negative function^{18–20}. Among class 1 mutation carriers (combined stage 1 and 2, $n = 5,732$), the five most significant associated SNPs included rs6994019, an intronic SNP in *MMP16* on chromosome 8 ($P_{\text{trend}} = 2.9 \times 10^{-6}$) and four SNPs in the 19p13 region ($P_{\text{trend}} = 7.6 \times 10^{-6}$ to $P_{\text{trend}} = 1.6 \times 10^{-4}$). The *MMP16* SNP rs6994019 was the ninth most significant SNP in the primary analysis of all mutations combined ($P_{\text{trend}} = 2.7 \times 10^{-4}$ in stage 1 and 2 combined; Supplementary Table 3). The strongest association with breast cancer risk for carriers of class 2 mutations was at the five SNPs in the 19p13 region ($P_{\text{trend}} = 1.8 \times 10^{-6}$ to $P_{\text{trend}} = 1.2 \times 10^{-4}$; Supplementary Table 3). The HR estimates for the five SNPs in 19p13 were larger for class 2 mutations, but the differences between class 1 and class 2 mutations were significant for only rs8170 and rs3745185 ($P = 0.03$ and $P = 0.004$, respectively). These differences might reflect a stronger modifying effect on breast cancer risk for tumors retaining residual or dominant negative *BRCA1* function.

Tumor estrogen or progesterone receptor status was available for 1,197 breast cancer cases in stage 1 and 2 combined. A case-only analysis revealed significant differences in the associations for the 19p13 SNPs between estrogen receptor–positive and estrogen receptor–negative disease and between estrogen receptor– or progesterone receptor–positive and estrogen receptor– and progesterone receptor–negative disease, particularly for SNPs rs8100241, rs2363956 and rs3745185 ($P = 0.002$ to $P = 0.04$; Supplementary Table 5). The OR estimates suggest that these SNPs are more strongly associated with estrogen receptor–negative disease.

The two most significant SNPs (rs8170 and rs4808611) were strongly correlated ($r^2 = 0.87$) in the *BRCA1* samples but displayed a low correlation with the other associated SNPs ($r^2 < 0.23$). rs8100241 and rs2363956 were perfectly correlated ($r^2 = 1$), whereas the least significant SNP, rs3745185, had weaker correlations with both sets of SNPs ($r^2 = 0.17$ and $r^2 = 0.74$ with rs8170 and rs8100241, respectively).

To evaluate the contribution of the 19p13 locus to breast cancer risk in the general population, we genotyped rs8170 and rs2363956 in 6,800 breast cancer cases and 6,613 controls from the SEARCH (Studies of Epidemiology and Risk Factors in Cancer Heredity) study in the UK. Neither SNP was associated with overall breast cancer risk ($P = 0.65$ and $P = 0.79$; Table 3). However, stratification of tumors by estrogen receptor status indicated that both SNPs were associated with estrogen receptor–negative breast cancer (rs8170, per-allele OR = 1.21, 95% CI 1.07–1.37, $P = 0.0029$ and rs2363956, OR = 0.83, 95% CI 0.75–0.92, $P = 0.0003$; Table 3). These effect sizes were similar to the estimated HRs for *BRCA1* carriers, consistent with the observation that *BRCA1* mutations predispose predominately to estrogen receptor–negative disease. Weaker associations were observed in the opposite direction for estrogen receptor–positive disease (rs8170, per-allele OR = 0.91, 95% CI 0.84–0.98, $P = 0.011$ and rs2363956, OR = 1.07, 95% CI 1.01–1.14, $P = 0.016$). Similar patterns were observed when tumors were stratified by progesterone receptor status or estrogen receptor and progesterone receptor status combined (Table 3).

The majority of breast tumors in *BRCA1* carriers exhibit a triple-negative (estrogen receptor, progesterone receptor and HER2 negative) phenotype. To evaluate the association of the

19p13 locus with triple-negative disease in the general population, we obtained genotype data for the five SNPs from up to 2,301 cases from 15 centers in six countries involved in the triple-negative breast cancer consortium (TNBCC). Genotype data from up to 3,949 geographically matched controls were also available (Supplementary Table 5). All SNPs were associated with triple-negative breast cancer, and the ORs were comparable to the HRs seen in the *BRCA1* carriers and the ORs for estrogen receptor–negative breast cancer seen in the SEARCH population-based study (rs2363956, per-allele OR = 0.80, 95% CI 0.74–0.87, $P = 1.1 \times 10^{-7}$ and rs8170, OR = 1.28, 95% CI 1.16–1.41, $P = 1.2 \times 10^{-6}$; Table 3 and Supplementary Table 5).

Two of the SNPs (rs8170 and rs2366956) were genotyped in 2,486 *BRCA2* mutation carriers as part of an ongoing GWAS. Neither SNP was associated with breast cancer risk for *BRCA2* carriers ($P_{\text{trend}} = 0.17$ and $P_{\text{trend}} = 0.07$), but the HR estimates were in line with the ORs estimated for estrogen receptor–positive disease in the SEARCH study (Table 3).

All five SNPs were located in a region that spans 39 kb on 19p13 (Fig. 2). In an analysis for the joint effect of these SNPs on breast cancer risk for *BRCA1* carriers, it was not possible to distinguish between rs8170 and rs4808611, as neither SNP improved the model fit significantly when the other was included ($P = 0.11$ and $P = 0.22$ for rs8170 and rs4808611, respectively). rs8100241 was retained in preference to rs3745185 (P for inclusion of rs3745185 in model = 0.79). Thus, the most parsimonious model included SNPs rs8170 and either rs8100241 or rs2363956 (P for inclusion = 7.7×10^{-5} and $P = 6.7 \times 10^{-5}$ for rs8170 and rs8100241, respectively) and had a 2 d.f. $P = 6.3 \times 10^{-13}$ for inclusion of both SNPs. This suggests that these associations may be driven by a single causative variant that is partially correlated with all five SNPs. To investigate this further, we evaluated the associations for SNPs identified through the 1000 Genomes Project using imputation. 1,055 SNPs in a 300-kb interval with a minor allele frequency >0.01 in samples of European ancestry, were evaluated. Thirty-one SNPs, none of which were genotyped in stage 1, displayed $P < 1.76 \times 10^{-9}$ (Fig. 2 and Supplementary Table 3). The most significant associations with the imputed genotypes in stage 1 and 2 combined were for eight perfectly correlated SNPs within a 13-kb region (the most significant SNP was rs4808075, $P = 9.4 \times 10^{-12}$; Supplementary Table 3). These SNPs were correlated with the four genotyped SNPs ($r^2 = 0.37$ to $r^2 = 0.58$ based on the 1000 Genomes Project data; Supplementary Fig. 2). This suggests that one or more of these imputed SNPs may be causally associated with breast cancer risk. However, some rare SNPs may have been missed because the 1000 Genome Project data used were based on the resequencing of only 56 individuals. Therefore, the possibility that the association is driven by a rarer variant, or a more cryptic common variant not detected in the resequencing, cannot yet be ruled out.

Of the five genotyped SNPs in the region and the eight most significant imputed SNPs, only rs8170 and rs2363956 were located in coding regions. The smaller 13-kb region, defined by the most strongly associated SNPs, contains three genes: *ABHD8* (encoding abhydrolase domain containing 8), *ANKLE1* (encoding ankyrin repeat and LEM domain containing 1) and *C19orf62*. The eight most significant imputed SNPs were clustered in and around *ANKLE1*, which encodes a protein of undefined function. However, *C19orf62*, which encodes MERIT40 (Mediator of Rap80 Interactions and Targeting 40 kD), is a more plausible genetic modifier of breast cancer in *BRCA1* carriers because MERIT40 interacts with BRCA1 in a protein complex. MERIT40 is a component of the BRCA1 A complex containing BRCA1-BARD1, Abraxas1, RAP80, BRCC36 and BRCC45 that is required for recruitment and retention of the BRCA1-BARD1 ubiquitin ligase and the BRCC36 deubiquitination enzyme at sites of DNA damage^{21–23}. Thus, a variant that modifies MERIT40 function or expression might influence BRCA1-dependent DNA repair and checkpoint activity in mammary epithelial cells of *BRCA1* carriers sufficiently, before loss

of the wildtype *BRCA1* allele, to increase the risk of breast cancer. However, until the SNPs that increase risk of cancer have been definitively linked to MERIT40, it remains possible that the other genes in the region or genes influenced by long range chromatin remodeling or by transcriptional events account for the breast cancer association.

Genetic variation at this locus, in combination with other risk modifiers, may prove useful in individual cancer risk assessment for breast cancer in *BRCA1* carriers. In addition, understanding the functional basis of this association may provide important insights into the etiology of *BRCA1*-associated breast cancer and hormone receptor–negative breast cancer in the general population. Our results suggest that GWAS in *BRCA1* mutation carriers or GWAS restricted to specific breast cancer subtypes may identify further breast cancer susceptibility variants.

METHODS

Methods and any associated references are available in the online version of the paper at <http://www.nature.com/naturegenetics/>.

URLs

1000 Genomes Project, <http://www.1000genomes.org/>; MACH software, <http://www.sph.umich.edu/csg/yli/mach/index.html/>.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

Financial support for this study was provided by the Breast Cancer Research Foundation (BCRF), Susan G. Komen for the Cure and US National Institutes of Health grant CA128978 to F.J.C. and by Cancer Research UK to D.F.E. and A.C.A. A.C.A. is a Cancer Research UK Senior Cancer Research Fellow and D.F.E. is a Cancer Research UK Principal Research Fellow. The authors thank Cancer Genetic Markers of Susceptibility (CGEMS) and Wellcome Trust Case Control Consortium (WTCCC) for provision of genotype data from controls. Study specific acknowledgments listed in Supplementary Note.

References

1. Antoniou A, et al. Average risks of breast and ovarian cancer associated with *BRCA1* or *BRCA2* mutations detected in case series unselected for family history: a combined analysis of 22 studies. *Am J Hum Genet.* 2003; 72:1117–1130. [PubMed: 12677558]
2. Antoniou AC, et al. The BOADICEA model of genetic susceptibility to breast and ovarian cancers: updates and extensions. *Br J Cancer.* 2008; 98:1457–1466. [PubMed: 18349832]
3. Begg CB, et al. Variation of breast cancer risk among *BRCA1/2* carriers. *J Am Med Assoc.* 2008; 299:194–201.
4. Milne RL, et al. The average cumulative risks of breast and ovarian cancer for carriers of mutations in *BRCA1* and *BRCA2* attending genetic counseling units in Spain. *Clin Cancer Res.* 2008; 14:2861–2869. [PubMed: 18451254]
5. Simchoni S, et al. Familial clustering of site-specific cancer risks associated with *BRCA1* and *BRCA2* mutations in the Ashkenazi Jewish population. *Proc Natl Acad Sci USA.* 2006; 103:3770–3774. [PubMed: 16537453]
6. Easton DF, et al. Genome-wide association study identifies novel breast cancer susceptibility loci. *Nature.* 2007; 447:1087–1093. [PubMed: 17529967]
7. Hunter DJ, et al. A genome-wide association study identifies alleles in *FGFR2* associated with risk of sporadic postmenopausal breast cancer. *Nat Genet.* 2007; 39:870–874. [PubMed: 17529973]

8. Stacey SN, et al. Common variants on chromosomes 2q35 and 16q12 confer susceptibility to estrogen receptor-positive breast cancer. *Nat Genet.* 2007; 39:865–869. [PubMed: 17529974]
9. Stacey SN, et al. Common variants on chromosome 5p12 confer susceptibility to estrogen receptor-positive breast cancer. *Nat Genet.* 2008; 40:703–706. [PubMed: 18438407]
10. Antoniou AC, et al. Common breast cancer-predisposition alleles are associated with breast cancer risk in *BRCA1* and *BRCA2* mutation carriers. *Am J Hum Genet.* 2008; 82:937–948. [PubMed: 18355772]
11. Antoniou AC, et al. Common variants in *LSP1*, 2q35 and 8q24 and breast cancer risk for *BRCA1* and *BRCA2* mutation carriers. *Hum Mol Genet.* 2009; 18:4442–4456. [PubMed: 19656774]
12. Lakhani SR, et al. The pathology of familial breast cancer: predictive value of immunohistochemical markers estrogen receptor, progesterone receptor, HER-2, and p53 in patients with mutations in *BRCA1* and *BRCA2*. *J Clin Oncol.* 2002; 20:2310–2318. [PubMed: 11981002]
13. Lakhani SR, et al. Prediction of *BRCA1* status in patients with breast cancer using estrogen receptor and basal phenotype. *Clin Cancer Res.* 2005; 11:5175–5180. [PubMed: 16033833]
14. Wellcome Trust Case Control Consortium. Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared controls. *Nature.* 2007; 447:661–678. [PubMed: 17554300]
15. Zollner S, Pritchard JK. Overcoming the winner's curse: estimating penetrance parameters from case-control data. *Am J Hum Genet.* 2007; 80:605–615. [PubMed: 17357068]
16. Buisson M, Anczukow O, Zetoune AB, Ware MD, Mazoyer S. The 185delAG mutation (c. 68_69delAG) in the *BRCA1* gene triggers translation reinitiation at a downstream AUG codon. *Hum Mutat.* 2006; 27:1024–1029. [PubMed: 16941470]
17. Mazoyer S, et al. A *BRCA1* nonsense mutation causes exon skipping. *Am J Hum Genet.* 1998; 62:713–715. [PubMed: 9497265]
18. Perrin-Vidoz L, Sinilnikova OM, Stoppa-Lyonnet D, Lenoir GM, Mazoyer S. The nonsense-mediated mRNA decay pathway triggers degradation of most *BRCA1* mRNAs bearing premature termination codons. *Hum Mol Genet.* 2002; 11:2805–2814. [PubMed: 12393792]
19. Antoniou AC, et al. *RAD51* 135G→C modifies breast cancer risk among *BRCA2* mutation carriers: results from a combined analysis of 19 studies. *Am J Hum Genet.* 2007; 81:1186–1200. [PubMed: 17999359]
20. Liu HX, Cartegni L, Zhang MQ, Krainer AR. A mechanism for exon skipping caused by nonsense or missense mutations in *BRCA1* and other genes. *Nat Genet.* 2001; 27:55–58. [PubMed: 11137998]
21. Feng L, Huang J, Chen J. *MERIT40* facilitates *BRCA1* localization and DNA damage repair. *Genes Dev.* 2009; 23:719–728. [PubMed: 19261748]
22. Shao G, et al. *MERIT40* controls *BRCA1*-Rap80 complex integrity and recruitment to DNA double-strand breaks. *Genes Dev.* 2009; 23:740–754. [PubMed: 19261746]
23. Wang B, Hurov K, Hofmann K, Elledge SJ. *NBA1*, a new player in the *Brca1* A complex, is required for DNA damage resistance and checkpoint control. *Genes Dev.* 2009; 23:729–739. [PubMed: 19261749]

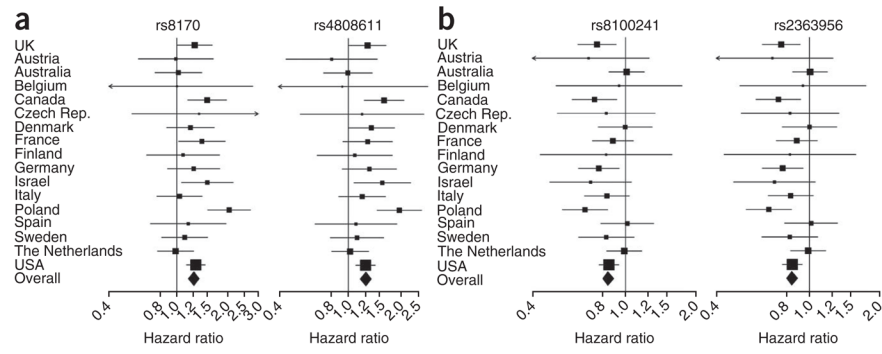


Figure 1. Forest plots of the associations by country of residence of *BRCA1* mutation carriers in the combined stage 1 and stage 2 samples. **(a,b)** Squares indicate the country specific per-allele HR estimates for SNPs rs8170, rs4808611 **(a)** and rs8100241, rs2363956 **(b)**. The area of the square is proportional to the inverse of the variance of the estimate. Horizontal lines indicate 95% CIs.

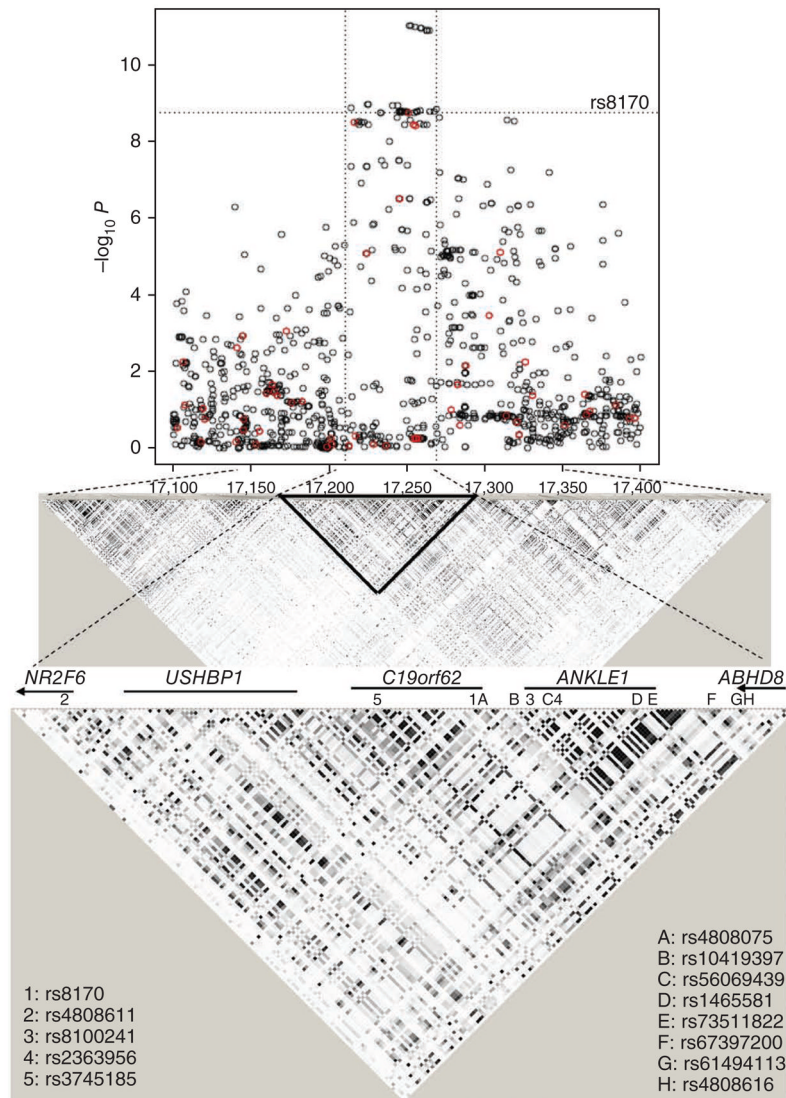


Figure 2.

Above, results of the kinship-adjusted score test statistic (1 d.f.) by position (kb) in stage 1 and 2 samples combined for genotyped and imputed SNPs in the associated region (chromosome 19, positions 17,100–17,400 kb). Genotyped SNPs in stages 1 or 2 are shown in red and imputed SNPs are shown in black. The horizontal dotted line indicates the P values for the strongest association among genotyped SNPs (rs8170). At middle, the linkage disequilibrium (LD) blocks around the top five associated SNPs (chromosome 19, positions 17,150–17,350 kb) in the combined analysis of stage 1 and stage 2 samples based on the 1000 Genomes Project data for the samples of European ancestry. Squares in the LD blocks indicate pairwise correlations between the SNPs (r^2) by grayscale (darker symbols indicate correlations closer to 1). Below, details of the region containing the most significantly associated genotyped and imputed SNPs (chromosome 19, positions 17,210–17,268 kb). Location of genotyped SNPs shown by numbers 1–5 and the eight most significantly associated imputed SNPs are shown in letters A–H ($P = 9.0 \times 10^{-12}$ to $P = 1.0 \times 10^{-11}$).

Table 1
Associations with breast cancer risk in *BRCA1* mutation carriers for the five most significant sNPs on 19p13

SNP, position, allele 1/allele 2	Stage	Number		Allele 2 frequency		HR (95% CI) ^b			<i>P</i> _{trend} ^e
		Unaffected ^a	Affected ^a	Unaffected	Affected	Per allele ^c	Heterozygote	Homozygote ^d	
rs8170	Stage 1	1,193	1,190	0.16	0.20	1.25 (1.12–1.39)	1.23 (1.08–1.41)	1.61 (1.13–2.30)	1.1 × 10 ⁻⁴
17,250,704	Stage 2	3,010	2,970	0.17	0.20	1.26 (1.15–1.38)	1.28 (1.14–1.43)	1.54 (1.17–2.03)	4.1 × 10 ⁻⁶
G/A	Combined	4,203	4,160	0.17	0.20	1.26 (1.17–1.35)	1.26 (1.16–1.37)	1.57 (1.26–1.95)	2.3 × 10 ⁻⁹
rs4808611	Stage 1	1,191	1,190	0.16	0.19	1.26 (1.13–1.41)	1.23 (1.08–1.41)	1.72 (1.21–2.45)	7.9 × 10 ⁻⁵
17,215,825	Stage 2	3,000	2,964	0.16	0.19	1.26 (1.15–1.39)	1.30 (1.16–1.46)	1.43 (1.06–1.92)	6.4 × 10 ⁻⁶
G/A	Combined	4,191	4,154	0.16	0.19	1.26 (1.17–1.35)	1.27 (1.17–1.39)	1.53 (1.22–1.93)	2.7 × 10 ⁻⁹
rs8100241	Stage 1	1,191	1,189	0.53	0.47	0.81 (0.74–0.88)	0.82 (0.71–0.95)	0.65 (0.55–0.77)	1.8 × 10 ⁻⁶
17,253,894	Stage 2	3,008	2,972	0.51	0.49	0.86 (0.80–0.92)	0.93 (0.82–1.05)	0.74 (0.65–0.85)	1.1 × 10 ⁻⁴
G/A	Combined	4,199	4,161	0.52	0.48	0.84 (0.80–0.89)	0.88 (0.81–0.97)	0.71 (0.63–0.79)	3.9 × 10 ⁻⁹
rs2363956	Stage 1	1,193	1,190	0.53	0.47	0.81 (0.74–0.88)	0.82 (0.71–0.95)	0.65 (0.55–0.77)	1.5 × 10 ⁻⁶
17,255,124	Stage 2	3,006	2,970	0.51	0.49	0.87 (0.81–0.93)	0.92 (0.82–1.04)	0.75 (0.65–0.86)	1.7 × 10 ⁻⁴
A/C	Combined	4,199	4,160	0.52	0.48	0.84 (0.80–0.89)	0.88 (0.80–0.97)	0.71 (0.64–0.79)	5.5 × 10 ⁻⁹
rs3745185	Stage 1	1,193	1,190	0.46	0.40	0.83 (0.76–0.90)	0.81 (0.71–0.93)	0.69 (0.57–0.82)	2.3 × 10 ⁻⁵
17,245,267	Stage 2	3,009	2,972	0.44	0.41	0.88 (0.82–0.95)	0.89 (0.80–1.00)	0.77 (0.67–0.89)	1.2 × 10 ⁻³
G/A	Combined	4,202	4,162	0.44	0.41	0.86 (0.81–0.91)	0.86 (0.81–0.91)	0.74 (0.66–0.83)	3.9 × 10 ⁻⁷

^a Affected, unaffected with breast cancer.

^b Estimated hazard ratio and 95% CI.

^c Per copy of allele 2.

^d Two copies of allele 2.

^e Kinship-adjusted score test.

Table 2

Competing risk analysis; associations with breast and ovarian cancer risk for *BRCA1* mutation carriers in the combined stage 1 and 2 samples

SNP	Genotype	Unaffected (%)	Breast cancer (%)	Ovarian cancer (%)	Ovarian cancer			Breast cancer		
					HR	95% CI	<i>P</i> ^a	HR	95% CI	<i>P</i> ^a
rs8170	GG	2,306 (68.4)	2,631 (63.4)	584 (69.3)	1.00			1.00		
	GA	973 (28.9)	1,360 (32.8)	238 (28.2)	1.10	0.92–1.31		1.27	1.17–1.39	
	AA	91 (2.7)	159 (3.8)	21 (2.5)	1.06	0.68–1.66		1.58	1.27–1.97	
	Per allele				1.07	0.93–1.24	0.33	1.27	1.18–1.36	1.5 × 10 ⁻¹⁰
rs4808611	GG	2,353 (70.0)	2,696 (65.1)	593 (70.7)	1.00			1.00		
	GA	923 (27.5)	1,307 (31.5)	229 (27.3)	1.14	0.96–1.36		1.29	1.18–1.41	
	AA	86 (2.6)	141 (3.4)	17 (2.0)	0.99	0.58–1.69		1.54	1.22–1.94	
	Per allele				1.10	0.94–1.27	0.34	1.27	1.18–1.37	1.6 × 10 ⁻¹⁰
rs8100241	GG	793 (23.6)	1,100 (26.5)	188 (22.4)	1.00			1.00		
	GA	1,676 (49.8)	2,118 (51.0)	428 (50.9)	1.01	0.83–1.23		0.89	0.81–0.98	
	AA	899 (26.7)	933 (22.5)	225 (26.8)	0.89	0.71–1.11		0.70	0.62–0.78	
	Per allele				0.94	0.84–1.05	0.28	0.84	0.79–0.88	1.6 × 10 ⁻¹⁰
rs2363956	AA	793 (23.6)	1,100 (26.5)	188 (22.3)	1.00			1.00		
	AC	1,678 (49.8)	2,116 (51.0)	429 (51.0)	1.01	0.83–1.23		0.89	0.80–0.97	
	CC	896 (26.6)	934 (22.5)	225 (26.7)	0.89	0.71–1.12		0.70	0.63–0.78	
	Per allele				0.94	0.85–1.05	0.30	0.84	0.79–0.88	2.4 × 10 ⁻¹⁰
rs3745185	GG	1,051 (31.2)	1,423 (34.3)	245 (29.1)	1.00			1.00		
	GA	1,675 (49.7)	2,048 (49.3)	437 (51.8)	1.03	0.85–1.23		0.86	0.79–0.94	
	AA	643 (19.1)	681 (16.4)	161 (19.1)	0.92	0.73–1.15		0.73	0.65–0.82	
	Per allele				0.97	0.86–1.08	0.54	0.86	0.81–0.91	7.1 × 10 ⁻⁸

^aRobust Wald statistic.

Table 3

Associations with breast cancer risk in the seArCH study overall and by tumor subtype, associations with triple negative breast cancer in the tNBCC study and associations with overall breast cancer risk for *BRCA2* mutation carriers

rs8170		rs2363956						
Study/subtype	Controls (%)	Cases (%)	OR/HR ^a (95% CI)	P	Controls (%)	Cases (%)	OR/HR ^a (95% CI)	P
SEARCH								
All cases								
GG	4,288 (65.8)	4,227 (66.5)	1.00		1,628 (24.7)	1,556 (24.3)	1.00	
GA	1,999 (30.7)	1,885 (29.7)	0.96 (0.89–1.03)		3,261 (49.4)	3,174 (49.7)	1.02 (0.93–1.11)	
AA	229 (3.5)	241 (3.8)	1.07 (0.89–1.29)		1,714 (26.0)	1,660 (26.0)	1.01 (0.92–1.12)	
Per allele			0.99 (0.93–1.05)	0.65	Per allele		1.01 (0.96–1.06)	0.79
Estrogen receptor status								
Estrogen receptor positive								
GG	4,288 (65.8)	2,437 (68.7)	1.00		1,628 (24.7)	817 (22.7)	1.00	
GA	1,999 (30.7)	988 (27.9)	0.87 (0.79–0.95)		3,261 (49.4)	1,791 (49.8)	1.09 (0.99–1.21)	
AA	229 (3.5)	123 (3.5)	0.95 (0.75–1.18)		1,714 (26.0)	992 (27.6)	1.15 (1.03–1.29)	
Per allele			0.91 (0.84–0.98)	0.011	Per allele		1.07 (1.01–1.14)	0.016
Estrogen receptor negative								
GG	4,288 (65.8)	503 (61.4)	1.00		1,628 (24.7)	240 (28.8)	1.00	
GA	1,999 (30.7)	272 (33.2)	1.16 (0.99–1.36)		3,261 (49.4)	421 (50.5)	0.88 (0.74–1.04)	
AA	229 (3.5)	44 (5.4)	1.64 (1.17–2.29)		1,714 (26.0)	172 (20.7)	0.68 (0.55–0.84)	
Per allele			1.21 (1.07–1.37)	0.0029	Per allele		0.83 (0.75–0.92)	0.0003
Heterogeneity ^b								
2.9 × 10 ⁻⁵								
Progesterone receptor status								
Progesterone receptor positive								
GG	4,288 (65.8)	1,087 (68.1)	1.00		1,628 (24.7)	368 (23.3)	1.00	
GA	1,999 (30.7)	447 (28.0)	0.88 (0.78–1.00)		3,261 (49.4)	759 (48.0)	1.03 (0.90–1.18)	
AA	229 (3.5)	62 (3.9)	1.07 (0.80–1.43)		1,714 (26.0)	454 (28.7)	1.17 (1.01–1.37)	
Per allele			0.94 (0.85–1.04)	0.21	Per allele		1.08 (1.00–1.17)	0.038
Progesterone receptor negative								
GG	4,288 (65.8)	451 (62.4)	1.00		1,628 (24.7)	199 (27.5)	1.00	

Study/subtype	rs8170						rs2363956					
	Controls (%)	Cases (%)	OR/HR ^a (95% CI)	P	Controls (%)	Cases (%)	OR/HR ^a (95% CI)	P	Controls (%)	Cases (%)	OR/HR ^a (95% CI)	P
GA	1,999 (30.7)	237 (32.8)	1.13 (0.95–1.33)		AC	3,261 (49.4)	375 (51.7)	0.94 (0.78–1.13)				
AA	229 (3.5)	35 (4.8)	1.45 (1.01–2.10)		CC	1,714 (26.0)	151 (20.8)	0.72 (0.58–0.90)				
Per allele			1.16 (1.01–1.33)	0.031	Per allele			0.85 (0.77–0.95)	0.004			
Heterogeneity ^b				0.0088					0.0002			
Estrogen receptor and progesterone receptor status												
Estrogen receptor or progesterone receptor positive												
GG	4,288 (65.8)	2,515 (68.6)	1.00		AA	1,628 (24.7)	848 (22.8)	1.00				
GA	1,999 (30.7)	1,019 (27.8)	0.87 (0.79–0.95)		AC	3,261 (49.4)	1,838 (49.5)	1.08 (0.98–1.20)				
AA	229 (3.5)	130 (3.6)	0.97 (0.78–1.21)		CC	1,714 (26.0)	1,026 (27.6)	1.15 (1.03–1.29)				
Per allele			0.91 (0.85–0.98)	0.014	Per allele			1.07 (1.01–1.13)	0.017			
Estrogen receptor and progesterone receptor negative												
GG	4,288 (65.8)	280 (59.5)	1.00		AA	1,628 (24.7)	134 (28.3)	1.00				
GA	1,999 (30.7)	169 (35.9)	1.29 (1.07–1.58)		AC	3,261 (49.4)	256 (54.0)	0.95 (0.77–1.19)				
AA	229 (3.5)	22 (4.7)	1.47 (0.93–2.32)		CC	1,714 (26.0)	84 (17.7)	0.60 (0.45–0.79)				
Per allele			1.26 (1.07–1.48)	0.0054	Per allele			0.79 (0.69–0.90)	0.0004			
Heterogeneity ^b				0.0002					8.3 × 10 ⁻⁶			
TNBC												
Estrogen receptor, progesterone receptor and HER2 negative												
GG	2,610 (66.2)	1,388 (60.7)	1.00		AA	890 (22.6)	614 (26.9)	1.00				
GA	1,200 (30.5)	791 (34.6)	1.30 (1.15–1.47)		AC	1,938 (49.3)	1,115 (48.9)	0.83 (0.72–0.95)				
AA	131 (3.3)	106 (4.6)	1.55 (1.16–2.07)		CC	1,103 (28.1)	550 (24.1)	0.65 (0.55–0.76)				
Per allele			1.28 (1.16–1.41)	1.2 × 10 ⁻⁶	Per allele			0.80 (0.74–0.87)	1.1 × 10 ⁻⁷			
BRCA2												
GG	784 (65.1)	864 (69.5)	1.00		AA	302 (24.9)	297 (23.8)	1.00				
GA	373 (31.0)	337 (27.1)	0.86 (0.71–1.04)		AC	608 (50.2)	599 (47.9)	1.03 (0.82–1.28)				
AA	47 (3.9)	43 (3.5)	0.92 (0.58–1.46)		CC	301 (24.9)	354 (28.3)	1.25 (0.98–1.61)				
Per allele			0.90 (0.77–1.05)	0.17 ^c	Per allele			1.12 (0.99–1.27)	0.07 ^c			

^aOR estimates for the SEARCH and TNBCC studies and HR estimates for the BRCA2 associations.

^bDifference in OR between hormone receptor–positive and hormone receptor–negative breast cancer tumors.

^cBased on the kinship-adjusted score test statistic.

NIH-PA Author Manuscript

NIH-PA Author Manuscript

NIH-PA Author Manuscript