



NIH PUBLIC ACCESS

Author Manuscript

J Asian Nat Prod Res. Author manuscript; available in PMC 2011 March 1.

Published in final edited form as:

J Asian Nat Prod Res. 2010 March ; 12(3): 227–232. doi:10.1080/10286021003591617.

Cancer preventive agents 10. Prenylated dehydrozingerone analogs as potent chemopreventive agents

Jin Tatsuzaki^a, Kyoko Nakagawa-Goto^a, Harukuni Tokuda^b, and Kuo-Hsiung Lee^{a,*}^a Natural Products Research Laboratories, Eshelman School of Pharmacy, University of North Carolina, Chapel Hill, NC 27599-7360, USA^b Department of Biochemistry, Kyoto Prefectural University of Medicine, Kyoto 602-0841, Japan

Abstract

Dehydrozingerone analogs and related compounds were screened as potential antitumor promoters by using the *in vitro* short-term 12-*O*-tetradecanphorbol-13-acetate (TPA)-induced Epstein-Barr virus early antigen (EBV-EA) activation assay. Among 40 synthesized compounds, the prenylated analogs **16** and **34–36** showed the most significant and promising activity (100% inhibition of activation at 1×10^{-3} mol ratio/TPA, and 82–80%, 37–35%, 13–11% inhibition at 5×10^{-2} , 1×10^{-2} , 1×10^{-1} mol ratio/TPA, respectively) in this screening. Their activity profiles were comparable to that of the reference standard curcumin. While a prenyl moiety conferred potent chemopreventive activity, an extended prenyl unit such as a farnesyl moiety did not improve activity. Because *in vitro* inhibitory effects in this assay generally correlate well with *in vivo* inhibitory effects on tumor promotion, our results strongly suggested that prenylated **16** and **34–36** are likely to be promising antitumor promoters.

Keywords

dehydrozingerone; antitumor-promoting effect; Epstein-Barr virus; two-stage carcinogenesis

1. Introduction

The natural product dehydrozingerone (DZ, **1**) is the “half analog” of curcumin (**2**) (Figure 1), which is known to have potent anti-oxidant, anti-inflammatory, and antitumor promoting (chemopreventive) activities [1,2]. Curcumin inhibits epidermal inflammation in mice and 12-*O*-tetradecanoylphorbol-13-acetate (TPA)-induced tumor promotion in mouse skin [3]. Structure-activity relationship (SAR) correlations of curcumin analogs as antitumor promoting agents have also been investigated [4,5]. Dehydrozingerone (**1**) and isoeugenol (**3**), which have similar catechol skeletons, but different alkenyl side chains, had stronger antitumor promotion effects than curcumin [6]. In the search for antitumor promoters from natural sources, the antitumor promoting properties of phenylpropanoids have also been reported [7–9]. The above evidence strongly supports the use of **1** as a lead to develop novel antitumor promoters and to further explore the structural features necessary for chemopreventive activity. Because certain natural products and synthetic compounds containing a prenyl moiety showed strong activity against the Epstein-Barr virus early antigen (EBV-EA) activation induced by TPA in Raji cells [10–14], prenyl derivatives of **1** were thus synthesized and evaluated for *in vitro* inhibitory activity against EBV-EA. In this

*Corresponding author. khlee@unc.edu.

paper, we report the synthesis and SAR study of dehydrozingerone analogs as chemopreventive agents.

2. Results and discussion

All compounds were previously synthesized [15]. Figure 2 shows the structures of analogs **4–11**, which are derivatives of **1**, and of the related chalcones **12–15**, in which the terminal methyl group is replaced by phenyl. Figure 3 shows the structures of analogs **16–33**, in which various types of alkyl and alkenyl groups were added to the C-4' alcohol of **1** and **3**, as well as four additional prenyloxy derivatives **34–37**. The structures of methylated and prenylated curcumins (**38** and **39**, respectively) are shown in Figure 4. All analogs were evaluated *in vitro* as inhibitors of EBV-EA activation induced by TPA in Raji cells [16–18], and the inhibitory data are shown in Tables 1 and 2. Figure 5 indicates the typical fluorescent finding of EBV-EA activation in Raji cell through the fluorescence microscope.

Compounds with 3,4-disubstituted benzalacetone structures (**1**, **5**, and **8**) showed slightly better activity than compounds with other disubstitution patterns (e.g. 2,3- or 2,4-). The data for **12–15** indicated that a phenyl group at C4 decreased the activity. No remarkable difference was observed between methoxy and ethoxy groups (**1** vs. **8** and **4** vs. **9**). However, by comparing **10** and **11** with **1** and **4–9**, the presence of fluorine on the benzene ring might decrease the activity.

Compounds **16–24** (dehydrozingerone analogs), **25–33** (isoeugenol analogs), **34–37** (prenylated analogs), and **38** and **39** (curcumin analogs) were also tested using an *in vitro* synergistic assay on EBV-EA activation induced by TPA. The inhibitory effects of tested compounds and the associated viability of Raji cells are shown in Table 2. Curcumin and **3** were used as positive controls. In this assay, all compounds showed inhibitory effects on EBV-EA activation without high cytotoxicity on Raji cells. At high concentrations (1×10^3 mol ratio), dehydrozingerone (**16–24** and **34–37**), isoeugenol (**25–33**), and curcumin (**38**, **39**) derivatives showed 100% inhibition, and at lower concentrations, were as or more potent than the parent compounds. The prenylated analogs, **16** and **25**, showed significant potency compared with other alkylated analogs in the respective series (see **16–24** for dehydrozingerone analogs, and **25–33** for isoeugenol analogs). Prenylated dehydrozingerone analogs **34–37** showed comparable activity with **16**, which showed the best activity in the alkylated series. These findings support the reported conclusions that a prenyl moiety is important for optimal inhibitory effects on EBV-EA activation [7–9]. Compound **37** was less active than **16** and **34–36**, indicating that fluorine does not affect the activity. Although analog **19** containing a geranyl group (two prenyl units), was more active than **20** and **29** with farnesyl groups (three prenyl units), it was less active compared with other analogs in the dehydrozingerone series. Compounds **17**, **18**, **22–24** and **26**, **27**, **31**, **32**, which contain allyl, 2-butenyl, ethyl, propyl, and isopentyl substituents, respectively, showed similar activity, while methylated compounds **21** and **30** showed slightly lower activity. When analogs with structurally similar alkyl and alkenyl groups were compared, (**16** vs **24**, **17** and **18** vs **23**), the presence of a double bond did not seem to affect the activity.

In summary, prenylated dehydrozingerone **16** and its analogs **34–36** showed the most significant and promising activity in this screening (100% inhibition of activation at 1×10^3 mol ratio/TPA, and 82–80%, 37–35%, 13–11% inhibition at 5×10^2 , 1×10^2 , 1×10^1 mol ratio/TPA, respectively). While a prenyl moiety conferred potent chemopreventive activity, an extended prenyl unit such a farnesyl moiety did not improve activity. Hydrophobicity might be important for inhibition of TPA-induced EBV-EA activation. Because *in vitro* inhibitory effects in this assay generally correlate well with *in vivo* inhibitory effects on tumor

promotion [4,5,19,20], our results suggested that **16** and **34–36** are promising antitumor promoters and further in vivo investigations are now in progress.

3. Experimental

3.1 In vitro EBV-EA activation experiments

EBV-EA positive serum from a patient with nasopharyngeal carcinoma (NPC) was a gift from Professor H. Hattori, Department of Otorhinolaryngology, Kobe University. The EBV genome carrying lymphoblastoid cells (Raji cells derived from Burkitt's lymphoma) were cultured in 10% fetal bovine serum (FBS) in RPMI-1640 medium (Sigma R8758, USA). Spontaneous activation of EBV-EA in our subline of Raji cells was less than 0.1%. The inhibition of EBV-EA activation was assayed using Raji cells (virus non-producer type) as described below. The cells were incubated at 37°C for 48 h in 1 mL of medium containing *n*-butyric acid (4 mM), TPA [32 pM = 20 ng in 2 µL dimethyl sulfoxide (DMSO)] and various amounts of the test compounds dissolved in 2 µL of DMSO. Smears were made from the cell suspension. The EBV-EA inducing cells were stained by the means of an indirect immunofluorescence technique. In each assay, at least 500 cells were counted, and the number of stained cells (positive cells) was recorded. Triplicate assays were performed for each compound. The average EBV-EA induction of the test compound was expressed as a ratio relative to the control experiment (100%), which was carried out with *n*-butyric acid (4 mM) plus TPA (32 pM). EBV-EA induction was ordinarily around 35%. The viability of treated Raji cells was assayed by the Trypan blue staining method. The cell viability of the TPA positive control was greater than 80%. Therefore, only these compounds that induced less than 80% (% of control) of the EBV-active cells (those with a cell viability of more than 60%) were considered able to inhibit the activation caused by promoter substances.

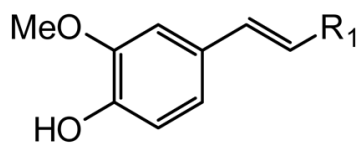
Acknowledgments

This investigation was supported in part by a grant CA 17625 from the National Cancer Institute awarded to K. H. Lee. This study was also supported in part by a grant from the Ministry of Education, Sciences, Sports and Cultures, and Ministry of Health and Welfare, Japan (Kyoto).

References

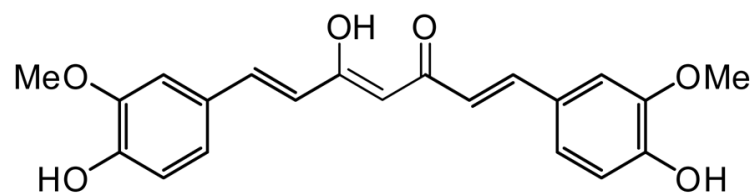
1. Sugiyama Y, Kawakishi S, Osawa T. *Biochem Pharmacol* 1996;52:519. [PubMed: 8759023]
2. Rudy AJ, Kuttan G, Babu KD, Rajasekharan KN, Kuttan R. *Cancer Lett* 1995;94:79. [PubMed: 7621448]
3. Huang MT, Smart RC, Wong CQ, Conney AH. *Cancer Res* 1988;48:5941. [PubMed: 3139287]
4. Ishida J, Kozuka M, Wang H, Konoshima T, Tokuda H, Okuda M, Mou XY, Nishino H, Sakurai N, Lee KH, Nagai M. *Cancer Lett* 2000;159:135. [PubMed: 10996724]
5. Ishida J, Kozuka M, Tokuda H, Nishino H, Nagumo S, Lee KH. *Bioorg Med Chem* 2002;10:3361. [PubMed: 12150883]
6. Motohashi N, Yamagami C, Tokuda H, Konoshima T, Okuda Y, Mukainaka T, Nishino H, Saito Y. *Cancer Lett* 1998;134:37. [PubMed: 10381128]
7. Ito C, Itoigawa M, Furukawa H, Ichiishi E, Mukainaka T, Okuda M, Ogata M, Tokuda H, Nishino H. *Cancer Lett* 1999;142:49. [PubMed: 10424780]
8. Ito C, Itoigawa M, Otsuka T, Tokuda H, Nishino H, Furukawa H. *J Nat Prod* 2000;63:1344. [PubMed: 11076549]
9. Itoigawa M, Ito C, Tokuda H, Enjo F, Nishino H, Furukawa H. *Cancer Lett* 2004;214:165. [PubMed: 15363542]
10. Ito C, Itoigawa M, Takakura T, Ruangrunsi N, Enjo F, Tokuda H, Nishino H, Furukawa H. *J Nat Prod* 2003;66:200. [PubMed: 12608849]

11. Ito C, Itoigawa M, Kojima N, Tokuda H, Hirata T, Nishino H, Furukawa H. *J Nat Prod* 2004;67:1125. [PubMed: 15270565]
12. Akihisa T, Tokuda H, Ukiya M, Iizuka M, Schneider S, Ogasawara K, Mukainaka T, Iwatsuki K, Suzuki T, Nishino H. *Cancer Lett* 2003;201:133. [PubMed: 14607326]
13. Ito C, Itoigawa M, Miyamoto Y, Onoda S, Rao KS, Mykainaka T, Tokuda H, Nishino H, Furukawa H. *J Nat Prod* 2003;66:206. [PubMed: 12608850]
14. Itoigawa M, Ito C, Wu TS, Enjo F, Tokuda H, Nishino H, Furukawa H. *Cancer Lett* 2003;193:133. [PubMed: 12706869]
15. Tatsuzaki J, Bastow KF, Nakagawa-Goto K, Nakamura S, Itokawa H, Lee KH. *J Nat Prod* 2006;69:1445. [PubMed: 17067159]
16. Takasaki M, Konoshima T, Fujitani K, Yoshida S, Nishimura H, Tokuda H, Nishino H, Iwashima A, Kozuka M. *Chem Pharm Bull (Tokyo)* 1990;38:2737. [PubMed: 1963812]
17. Tokuda H, Ohigashi H, Koshimizu K, Ito Y. *Cancer Lett* 1986;33:279. [PubMed: 3802058]
18. Henle G, Henle W. *J Bacteriol* 1966;91:1248. [PubMed: 4160230]
19. Konoshima T, Takasaki M, Tatsumoto T, Kozuka M, Kasai R, Tanaka O, Nie R, Tokuda H, Nishino H, Iwashima A. *Biol Pharm Bull* 1994;17:668–671. [PubMed: 7920430]
20. Sakurai N, Kozuka M, Tokuda H, Nobukuni Y, Takayasu J, Nishino H, Kusano A, Kusano G, Nagai M, Sakurai Y, Lee KH. *Bioorg Med Chem* 2003;11:1137. [PubMed: 12614901]



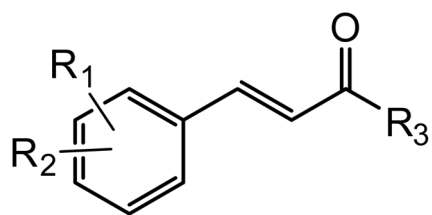
1: Dehydrozingerone R₁ = COCH₃

3: Isoeugenol R₁ = CH₃



2: Curcumin

Figure 1.
Structures of Dehydrozingerone (1), Curcumin (2), and Isoeugenol (3)

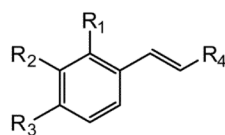


1, 4-11: R₃ = Me

12-15: R₃ = Ph

	R ₁	R ₂	R ₃
1	3-OMe	4-OH	Me
4	2-OH	3-OMe	Me
5	3-OH	4-OMe	Me
6	2-OH	4-OMe	Me
7	H	H	Me
8	3-OEt	4-OH	Me
9	2-OH	3-OEt	Me
10	2-OH	3-F	Me
11	3-F	4-OMe	Me
12	3-OMe	4-OH	Ph
13	2-OH	3-OMe	Ph
14	3-OH	4-OMe	Ph
15	2-OH	4-OMe	Ph

Figure 2.
Structures of Deydrozingerone Analogs



Dehydrozingerone R ₄ = COMe	R ₁	R ₂	R ₃	Isoeugenol R ₄ = Me
16	H	OMe		25
17	H	OMe		26
18	H	OMe		27
19	H	OMe		28
20	H	OMe		29
21	H	OMe	OMe	30
22	H	OMe	OEt	31
23	H	OMe	OPr	32
24	H	OMe		33
34		OMe	H	-
35	H		OMe	-
36		H	OMe	-
37		F	H	-

Figure 3. Structures of Dehydrozingerone (**16–24, 34–37**) and Isoeugenol (**25–33**) Analogs

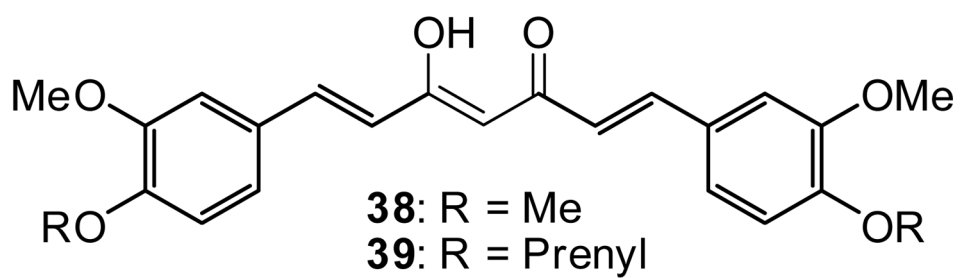


Figure 4.
Curcumin Analogs **38** and **39**

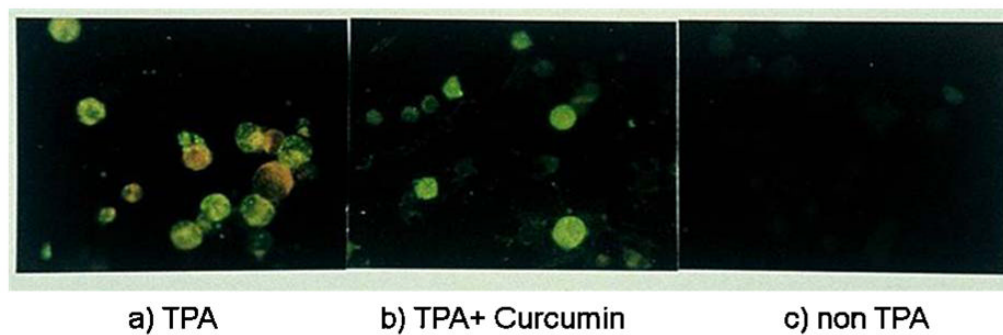


Figure 5. Typical fluorescent findings of EBV-EA activation

a) EBV-EA expression by treatment with TPA; b) EBV-EA expression was reduced by Curcumin; c) No EBV-EA expression was observed without TPA.

Table 1

Relative ratio^a of EBV-EA activation with respect to positive control in presence of dehydrozingerone analogs.

Compound	Percentage EBV-EA positive cells				IC ₅₀
	1000	500	100	10	
Dehydrozingerone (1) ^c	0 (70)	44.2	75.5	95.4	372
4	0 (70)	46.7	77.7	97.6	380
5	0 (70)	44.1	74.6	94.3	369
6	0 (70)	45.8	76.9	96.3	378
7	0 (70)	48.6	78.1	98.9	378
8	0 (70)	42.2	73.7	93.1	368
9	0 (70)	43.9	74.8	95.3	370
10	0 (60)	51.3	79.4	98.0	385
11	0 (60)	53.4	81.0	99.0	389
12	0 (60)	37.2	79.4	100	379
13	0 (60)	38.6	81.5	100	381
14	0 (60)	37.9	80.7	100	380
15	0 (60)	37.2	80.3	100	380

^aValues represent percentages relative to the positive control value (100%).

^bTPA concentration is 20 ng/mL (32 pmol/mL).

^cValues in parentheses are viability percentages of Raji cells.

Table 2

Relative ratio^a of EBV-EA activation with respect to positive control in presence of dehydrozingerone analogs and related compounds.

Compound	Percentage EBV-EA positive cells					IC ₅₀
	1000	500	100	10	1	
Dehydrozingerone (1) ^c	0±0.5 (70) ^d	44.2±1.9	75.5±2.6	95.4±0.2	372	
16	0±0.2 (60)	19.0±1.3	65.2±2.0	88.7±0.4	216	
17	0±0.4 (60)	21.9±1.5	67.8±2.1	90.0±0.2	234	
18	0±0.3 (60)	20.7±1.5	66.9±2.1	89.3±0.3	220	
19	0±0.4 (60)	23.5±1.5	71.0±2.3	95.1±0.1	242	
20	0±0.4 (60)	24.9±1.3	73.9±2.3	97.7±0.2	250	
21	0±0.4 (60)	22.8±1.3	70.9±2.0	93.7±0.3	239	
22	0±0.6 (60)	38.5±1.7	72.0±2.3	92.1±0.2	368	
23	0±0.2 (60)	21.5±1.2	68.9±2.1	91.7±0.3	238	
24	0±0.3 (60)	20.1±1.3	66.2±2.0	89.0±0.3	220	
Isoetgenol (3) ^c	16.7±1.5 (70)	53.5±1.9	81.0±2.7	100±0.1	490	
25	0±0.2 (60)	19.9±1.1	66.0±1.9	89.5±0.4	219	
26	0±0.4 (60)	22.8±1.4	68.0±2.2	92.7±0.2	236	
27	0±0.3 (60)	21.8±1.3	67.3±2.1	90.3±0.3	234	
28	NA ^e	-	-	-	-	
29	0±0.5 (60)	26.3±1.4	75.3±2.3	98.8±0.1	261	
30	0±0.4 (60)	23.6±1.5	72.0±2.3	95.6±0.2	246	
31	NA	-	-	-	-	
32	0±0.4 (60)	22.7±1.3	69.3±2.3	92.8±0.1	240	
33	0±0.4 (60)	21.2±1.4	67.3±2.1	91.5±0.2	237	
34	0±0.2 (60)	19.1±1.1	64.8±2.0	88.0±0.4	222	
35	0±0.2 (60)	18.0±1.0	63.2±2.0	86.8±0.3	207	
36	0±0.2 (60)	19.7±1.1	63.2±2.0	86.8±0.3	219	
37	0±0.2 (60)	22.8±1.3	67.7±2.1	90.5±0.3	231	

Compound	Percentage EBV-EA positive cells				
	Compound concentration (mol ratio/TPA ^b)				
	1000	500	100	10	IC ₅₀
Curcumin (2) ^c	0±0.4 (60)	21.1±1.1	80.1±2.4	100±0.1	379
38	0±0.2 (60)	19.6±1.1	76.5±2.4	98.5±0.1	253
39	0±0.1 (60)	17.3±0.9	72.6±2.1	92.4±0.3	240

^aValues represent percentages relative to the positive control value (100%).

^bTPA concentration is 20 ng/mL (32 pmol/mL).

^cPositive control

^dValues in parentheses are viability percentages of Raji cells.

^eNot applicable