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# Autoimmunity in the Pathogenesis and Treatment of Keratoconjunctivitis Sicca

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# Abstract

Dry eye is a chronic corneal disease that impacts the quality of life of many older adults. keratoconjunctivitis sicca (KCS), a form of aqueous-deficient dry eye, is frequently associated with Sjögren's syndrome and mechanisms of autoimmunity. For KCS and other forms of dry eye, current treatments are limited, with many medications providing only symptomatic relief rather than targeting the pathophysiology of disease. Here, we review proposed mechanisms in the pathogenesis of autoimmune-based KCS: genetic susceptibility and disruptions in antigen recognition, immune response, and immune regulation. By understanding the mechanisms of immune dysfunction through basic science and translational research, potential drug targets can be identified. Finally, we discuss current dry eye therapies as well as promising new treatment options and drug therapy targets.

# Keywords

Dry eye; Autoimmunity; Keratoconjunctivitis sicca; Sjögren's disease; Pathogenesis; Treatment; Immune regulation; Humoral immunity

# INTRODUCTION

Dry eye is a significant ocular disease that affects up to 35 % of the population aged 65 years and over [1]. Dry eye is a dysfunction of the nasolacrimal unit consisting of the nasolacrimal glands, corneal surface, and eyelids that results in an insufficient tear film. Patients experience ocular irritation often described as burning, gritty sensation, or dryness. The symptoms generally vary during the day and are often worse at night. Other symptoms

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**Compliance with Ethics Guidelines** 

**Conflict of Interest** 

Katy C. Liu, Kyle Huynh, Joseph Grubbs Jr., and Richard M. Davis declare that they have no conflict of interest.

Human and Animal Rights and Informed Consent

This article does not contain any studies with human or animal subjects performed by any of the authors.

include photophobia, itching, mucous accumulation, and tearing. Dry eye poses a significant problem, as it can lead to complications such as visual impairment, corneal ulceration, infection, anxiety, depression, and decreased quality of life.

Dysfunction in dry eye can be classified by mechanism: aqueous-deficient dry eye, evaporative dry eye, or mixed mechanism. In aqueous-deficient dry eye, the lacrimal duct produces an insufficient volume of tears, either due to dysfunction or destruction of the lacrimal glands; the latter group is mostly associated with autoimmune diseases such as Sjögren's disease. In evaporative dry eye, poor tear quality and tear film hyperosmolarity stem from defects such as meibomian (sebaceous) gland dysfunction, lagophthalmos (inability to close the eyelids completely), or decreased blink function [2].

Aqueous-deficient dry eye is also referred to as keratoconjunctivitis sicca (KCS). Henrik Sjögren first described KCS in 1933 as ocular findings in patients with primary Sjögren's disease. The prevalence of KCS is 4 % in adults over age 65. KCS is generally insidious in onset, presenting more commonly in females and Caucasians. In addition to Sjögren's disease, other causes of KCS include age-related atrophy of secreting glands and drug-induced KCS. Specifically, KCS has been associated with the use of beta-blockers, diuretics, antihistamines, and antidepressant drugs [1]. In this review, we focus on Sjögren's-associated KCS, and the autoimmune-based mechanisms and treatments for keratoconjunctivitis sicca.

### MECHANISMS OF PATHOGENESIS IN AUTOIMMUNE-MEDIATED KCS

Although precise mechanisms underlying autoimmune-mediated keratoconjunctivitis sicca are not well understood, the pathogenesis of keratoconjunctivitis sicca is likely multi-factorial with genetic and environmental components contributing to autoimmunity. Research has revealed potential mechanisms of dysfunction and dysregulation in the physiologic immune response resulting in the pathogenesis of KCS. In this review, we emphasize genetic susceptibility to the disease as well as disruptions in antigen recognition, immune response, and immune regulation, in the context of autoimmune-mediated KCS.

#### **Genetic Susceptibility**

Major histocompatibility complex (MHC) class II molecules have long been implicated in autoimmune diseases such as Sjögren's disease. On a transcriptional level, certain human leukocyte antigen (HLA) genes, such as HLA-DR $\beta$ 1, encode specific MHC class II molecules and are upregulated in patients with Sjögren's disease [3]. The upregulation of such HLA alleles is thought to genetically predispose individuals to Sjögren's disease and thus has utility for clinical diagnosis. To our knowledge, there are no specific HLA genes that predispose individuals to non-Sjögren's-associated KCS.

#### Antigen Recognition

**Autoantibodies**—Antibodies against self-antigens are a well-established mechanism for antigen recognition and autoimmunity. Autoantibodies have long been used as diagnostic markers for Sjögren's disease. In particular, anti-Ro/SSA, anti-La/SSB, and anti-nuclear antibodies (ANA) are often detected at high levels in patients with Sjögren's disease.

Interestingly, autoantibodies can potentially be used to discriminate between Sjögren'sassociated KCS versus other causes of aqueous-deficient dry eye. Compared to dry eye patients without Sjögren's disease, anti-Ro and anti-La antibodies have only been detected in Sjögren's-associated KCS [4]. New antibody markers for Sjögren's disease continue to be discovered and are directed against a variety of antigens, including nuclear, cytoplasmic, membrane-bound, and secreted proteins [5]. For example, NuMA (nuclear mitotic apparatus) and MCAs (mitotic chromosomal autoantigens) have recently been reported [6]. Nonetheless, only anti-Ro/SSA and anti-La/SSB antibodies are routinely used in diagnostic testing of Sjögren's disease. Notably, autoantibodies alone are unable to elicit or predict the development of autoimmune disease, since autoantibodies can be detected in healthy patients due to tolerance to self-antigens [5, 6].

**Molecular Mimicry**—Pathogens such as viruses can also elicit an autoimmune response in a process called molecular mimicry, whereby antigen epitopes on microbial proteins cross-react with mammalian self-antigens. It is hypothesized that molecular mimicry mediates post-viral KCS. The herpes simplex virus has been shown to react with La/SSB antigen in Sjögren's patients [7], and Coxsackie virus 2B can cross-react with the Rh60 autoantigen [8]. Interestingly, the two aforementioned viruses have also been shown to cross-react with autoantigens in systemic lupus erythematosus (SLE) patients [7, 8].

**Apoptosis and Antigen Presentation**—Apoptosis is a process of controlled cell death that can be used to eliminate injured or damaged cells. Apoptosis has been linked with autoimmune diseases such as Sjögren's disease [9] and SLE [10]. In the context of autoimmune disease, apoptosis-produced cell debrism such as DNA and/or RNA fragmentsm likely induce autoantigens and subsequent inflammation. Interestingly, an accelerated rate of apoptosis has been observed in Sjögren's disease [9, 11, 12]. Within the eye, increased corneal epithelial cell apoptosis has been observed in both Sjögren's- and non-Sjögren's-associated KCS in a mouse model of environmentally-induced KCS [13]. These results suggest that, in addition to autoimmune-mediated KCS, apoptosis likely plays an important role in other forms of KCS such as evaporative dry eye. At present, it is unclear how autoantigens are elicited from apoptotic cell debris. In Sjögren's disease, it is hypothesized that the process of autoantigen generation involves an initial subcellular redistribution and clustering of molecules into blebs on the surface of apoptotic cells [14].

DNA and/or RNA fragments from apoptotic cells also have the ability to activate toll-like receptors (TLRs). TLRs are found on immune cells such as macrophages and dendritic cells, as well as in the gut, where they respond to pathogen-associated molecular patterns to elicit an immune response. In one of the major TLR signaling pathways, MyD88 recruits IRAK followed by downstream NF-κB-associated nuclear translocation and transcription that ultimately induces a pro-inflammatory state. Specific toll-like receptors such as TLR3, TLR7/8m and TLR9 are activated in systemic autoimmune diseases like Sjögren's disease [15]. On the corneal surface of KCS patients, increased apoptosis is observed with chromatic and small ribonuclear particles (snRNPs) likely activating TLRs [13].

In autoimmune diseases, apoptosis is regulated by key molecules, many of which have been elucidated in recent studies. I $\kappa$ B is a transcription factor associated with NF- $\kappa$ B [16], and

has recently been shown to be important in autoimmunity and inflammation in a mouse model with I $\kappa$ B-deficient epithelial cells, Okuma et al. demonstrated accelerated apoptosis and development of a Sjögren's-like autoimmune syndrome [17]. This suggests I $\kappa$ B is required for the normal time course of apoptosis. Furthermore, I $\kappa$ B has been shown to be regulated by STAT3, a transcription factor needed for Th17 differentiation. Th17 induces cytokines and inflammation and thus may play a critical role in autoimmune diseases such as KCS [18]. In addition, the cytokine IFN- $\gamma$  has been identified as a modulator of increased apoptosis in the conjunctival epithelium [19]. In this study, Zhang et al. found that goblet cells were strongly targeted for IFN- $\gamma$ -dependent apoptosis in a desiccating stress-induced mouse model of KCS (induced by scopolamine, air draft exposure, and low humidity).

Another apoptotic pathway likely involved in Sjögren's disease and KCS is death receptor Fas/Fas ligand (FasL) binding. In apoptosis, the Fas/FasL interaction activates caspases and proteinases that result in DNA fragmentation and cell death. Higher levels of Fas-positive cells and soluble FasL were detected in the serum of Sjögren's-associated KCS than in non-Sjögren's KCS patients [20]. Although this study suggests a role for Fas/FasL and apoptosis in Sjögren's-associated KCS, a direct correlation has not been established between increased serum Fas/FasL and increased rate of apoptosis. Future research is needed to further elucidate the role of Fas/FasL binding and its mechanism of action in autoimmune-mediated apoptosis.

Antigen Presenting Cells—In KCS, antigens derive from multiple sources autoantigens, molecular mimicry, and apoptosis. These antigens are presented to immature antigen-presenting cells (APCs), which are then activated by major histocompatibility complex (MHC) class II cell surface receptors. Mature APCs then present antigen to T cells to generate an inflammatory and immune response.

The process of antigen presentation in KCS appears to be altered, at least in some cases, at the step involving MHC class II and other costimulatory molecules. Within the conjunctival epithelium of patients with Sjögren's disease, specific MHC II molecules such as HLA-DR are upregulated [20]. Interestingly, this HLA-DR upregulation has been observed in Sjögren's- and non-Sjögren's-associated KCS patients [21]; however, significantly higher levels of HLA-DR are expressed in Sjögren's-associated versus non-Sjögren's-associated KCS. These data suggest that HLA-DR upregulation has a role in the pathophysiology of aqueous-deficient dry eye and, in particular, Sjögren's-associated KCS. Because of its upregulation, HLA-DR has been used as a marker for active disease in KCS, as reduced HLA-DR expression has been observed in experimental testing of the efficacy of pharmacologic treatments such as pranoprofen and cyclosporine A for dry eye [22, 23].

Also, in the cornea, MHC class II molecules show distinct localization patterns that are disrupted in KCS. Endogenous MHC class II-expressing and non-expressing cells localize to the corneal periphery, while, in the central corneal epithelium, only MHC II non-expressing cells are detected [24]. However, in the conjunctival epithelium of patients with KCS, MHC II molecules are overexpressed [25, 26] ,and APCs, notably dendritic cells, are significantly upregulated in the central corneal epithelium [27]. Similarly, MHC class II molecules are upregulated in the salivary glands of Sjögren's patients [28].

In addition, costimulatory molecules such as CD40 and B7 are upregulated in corneal inflammation as well as Sjögren's-associated KCS [20, 29]. With ongoing injury to the ocular surface in KCS, MHC class II and costimulatory molecules are repeatedly triggered, further perpetuating the cycle of antigen recognition, immune response, and inflammation [29]. How exactly MHC II and other costimulatory molecules are upregulated on the ocular surface remains unknown. A recent study has revealed several molecular pathways for control of MHC class II presentation via regulation of transcription and protein translocation to the cell membrane. These pathways involve TGF-B, the actomyosin cytoskeleton, and regulation by GTPases [30]. The role of TGF- $\beta$  in MHC II presentation is particularly intriguing given the presence of endogenous TGF- $\beta$  in lacrimal tears [31], increased activity of TGF- $\beta$  in dry eye, and even higher TGF- $\beta$  activity in Sjögren's-associated KCS [32]. In a mouse model of dry eye, disruption of TGF- $\beta$  was found to suppress APC maturation in the cornea, leading to an improvement in dry eye phenotype and in conjunctival inflammation [33, 34]. Furthermore, factors that regulate TGF- $\beta$  have been shown to be involved in the pathogenesis of Sjögren's-like KCS in mice [35]. Thus, the TGF- $\beta$  signaling pathway poses an intriguing drug therapy target for the treatment of KCS.

#### **Immune Response**

**Cell-mediated Immunity**—It is clear that a disruption in cell-mediated immunity is important in the pathogenesis of autoimmune diseases such as Sjögren's disease, as mice with defective T cell development acquire symptoms of Sjögren's disease as well as anti-Ro/SSA and anti-La/SSB antibodies [36]. Moreover, the introduction of desiccating stress in mice has been shown to induce T cell-mediated inflammation, specifically in the cornea, conjunctiva, and lacrimal gland. In this study, primed T cells were adoptively transferred to T cell-deficient nude mice and produced a similar KCS-type inflammatory response [37]. T cells and cell-mediated immunity are important in the inflammatory pathway in KCS following antigen presentation.

**Humoral Immunity**—The humoral immune response is mediated by B cells, which produce antibodies targeted against pathogenic antigens. B cell dysfunction and the resultant disruption of host immune tolerance have been observed in autoimmune diseases such as SLE, rheumatoid arthritis, type I diabetes, and multiple sclerosis [38]. B cell dysfunction has also been implicated in the pathogenesis of Sjögren's disease [39]. In the NOD mouse model of Sjögren's disease, mice lacking mature B cells did not develop lacrimal and salivary gland dysfunction [40]. This suggests that B cells are necessary for the development of Sjögren's disease. B cell hyper-reactivity followed by hypergammaglobulinemia have been observed in Sjögren's disease [41]. Furthermore, upregulation of B cell activating factors, BAFF and APRIL, has been detected in the lacrimal glands of Sjögren's patients [42, 43]. BAFF (BLyS or B-lymphocyte stimulator) is a B cell activating factor of the tumor necrosis factor family found to be significantly overexpressed in salivary glands of Sjögren's disease patients [44] with the effect of preventing the apoptosis of autoreactive B cells [45]. As for APRIL (a proliferation-inducing ligand), it is yet to be determined whether its upregulation induces B cell hyperactivity and hypergammaglobulinemia. Interestingly, immunoglobulin is sufficient to induce KCS, as immunoglobulin serum derived from a KCS mouse injected into a nude-recipient mouse produced KCS and inflammation [21]. Furthermore, goblet cell

apoptosis was observed in the KCS-immunoglobulin recipient mice [21], thus illustrating the complexity of the pathogenesis of immune dysfunction in autoimmune disease.

#### Immune Regulation

Helper T cells—Helper or effector T cells are activated after APC stimulation and facilitate cytokine secretion. In KCS, research has focused on the effector T cells,  $T_{H}$  and  $T_H 17$  [32].  $T_H 17$  induces the secretion of IL-17 in response to TGF- $\beta$  or IL-6, which then stimulates the production of pro-inflammatory cytokines and matrix metalloproteinases [46]. Matrix metalloproteinases are upregulated in the lacrimal and salivary glands in Sjögren's disease [47], and significant tissue injury and destruction can result from their activity [48]. The  $T_H 17$ -centric cytokine IL-17 has been analyzed in the tear film, revealing highest levels in Sjögren's-associated KCS as well as increased levels in non-Sjögren's-associated KCS versus control subjects [49]. Interestingly, the induction of IL-17A into murine salivary glands was sufficient to induce a Sjögren's-like phenotype of reduced salivary function, inflammation, and positive antibody markers such as anti-ANA [50]. T<sub>H</sub>17 has been implicated in other autoimmune diseases such as Crohn's disease [51] and collagen-induced arthritis [52]. Thus, the regulation of pro-inflammatory cytokines and matrix metalloproteinases has a significant role in KCS that results in tissue injury and reduced function of exocrine glands. From this work, drug targets such as localized anti-IL17 have been proposed [50].

**Regulatory T cells**—Regulatory T cells (Tregs) are thought to maintain immune tolerance to autoimmune antigens. Correspondingly, dysfunction of regulatory T cells is observed in autoimmune diseases such as Sjögren's disease, SLE, and rheumatoid arthritis [53]. Notably, Tregs are important for the development of Sjögren's-associated KCS: in a mouse model of KCS using Treg-deficient nude mice, T cells from KCS-primed mice were adoptively transferred, resulting in the development of Sjögren's-like disease even without a desiccating stress trigger [37]. This study suggests that Treg deficiency could predispose to Sjögren's disease and KCS. Interestingly, T<sub>H</sub>17 cells implicated in the pathogenesis of KCS have been shown to be resistant to Treg-mediated suppression [54]. Correspondingly, a demonstrated imbalance between Treg cells and IL-17, with a disproportionate increase in IL-17, in mild to moderate inflammation in Sjögren's disease has been hypothesized to lead to tissue injury and pathology [55]. The imbalance between IL-17 and Treg cells provides another focus for potential therapies to alter the immune response in Sjögren's-associated KCS.

# TREATMENTS FOR KCS

Our understanding of the mechanisms of dysfunction of the immune system has guided the development of drug therapy for KCS. However, treatments for dry eye, particularly in the context of autoimmune dysfunction, remain limited. Ongoing research in autoimmune-mediated KCS is critical, with the hope of elucidating novel drug targets and therapies for this chronic and debilitating eye disease.

Thus far, treatments for autoimmune-based KCS have been targeted toward stimulation of tear secretion, anti-inflammatory agents, artificial tear replacement, and tear film

maintenance. As a first-line treatment option, artificial tears can provide temporary, symptomatic relief. Frequent topical application hinders its use for long-term treatment, and, furthermore, artificial tear replacement does not address the pathophysiology of dry eye disease. Tear secretion stimulators, such as cyclosporine A and cevimeline, are available and will be discussed below. Other drugs stimulating tear secretion include pilocarpine and diquafosol (approved for treatment in Japan). Also, anti-inflammatory agents have been assessed for the treatment of dry eye. Topical corticosteroids, although likely effective in alleviating the symptoms of dry eye [56, 57], are not recommended for long-term therapy due to complications such as increased intraocular pressure and cataract formation. Finally, topical NSAIDs have also been evaluated for dry eye treatment, but, unfortunately, topical NSAIDs are not as effective at providing symptomatic improvement versus topical corticosteroids or artificial tear replacement alone [57].

**Cyclosporine A**—Cyclosporine A is a generally well-tolerated treatment strategy for dry eye that has been FDA-approved for over a decade. Although cyclosporine is an immunosuppressive agent when administered systemically, topical cyclosporine acts as a partial immunomodulator in the eye and has been used for a wide range of ocular diseases with an inflammatory component, such as dry eye, posterior blepharitis, post-LASIK dry eye, atopic keratoconjunctivitis, and graft-versus-host disease [58]. In two mouse models of Sjögren's disease, cyclosporine has been shown to increase tear production [59]. In agreement with mouse models, randomized controlled trials in patients with moderate to severe dry eye showed improved ocular discomfort, less blurred vision, and improved Schirmer test scores with cyclosporine [60].

Another mechanism of action of cyclosporine in the treatment of dry eye is the inhibition of apoptosis. In a dry eye mouse model and in human conjunctival epithelial cells, cyclosporine reduced apoptosis as well as the number of infiltrating lymphocytes [61, 62]. In NOD mice, cyclosporine reduced Fas ligand expression in infiltrating lymphocytes [59]. In addition to cyclosporine, other means to inhibit apoptosis have been proposed as potential treatment modalities; for example, local Fas ligand transfer via adenoviral vectors [63] or the use of the anti-apoptotic agent D-beta-hydroxybutyrate [64] have been suggested as potential drug therapies.

**Cevimeline**—Cevimeline activates muscarinic M3 receptors and has been used to treat xerostoma (dry mouth) in patients with Sjögren's disease. In randomized prospective double-blinded trials to evaluate the efficacy of cevimeline in KCS, the cevimeline treatment arm had improved tear biometrics, as measured by Schirmer testing, rose Bengal and fluorescein staining, and tear break-up time, as well as subjective patient symptom reporting [65–67]. Although promising in terms of efficacy, systemic cevimeline has been associated with side effects of nausea, abdominal pain, sweating, headache, dizziness and cardiac arrhythmias, resulting in a withdrawal rate of 14–19 % from these clinical trials [66, 67].

#### Other potential drug targets

**Rituximab**—Rituximab is a monoclonal antibody directed against CD20 that targets B cells. Although rituximab has been studied in the treatment of systemic manifestations of

Sjögren's disease, less is known about the role of rituximab specifically in Sjögren'sassociated KCS. Other B cell-depleting therapies, such as interferon-alpha and anti-B-cell activating factor (BAFF), have been investigated, and short-term benefit has been demonstrated with both [68]. Success with rituximab and other B cell depleting therapies suggests that B cells play an important role in Sjögren's disease, and, as a general strategy, B cell-targeted therapies have shown potential for treatment of KCS.

**Gene therapy**—Gene therapy is a promising approach for targeted and long-term treatment with unique advantages of avoiding multiple daily applications of eye drops and side effects of chronic medication use. In gene therapy, a gene of interest is introduced into the organ of choice by a viral vector. In the case of dry eye, the tissues such as the lacrimal gland [69] and the corneal epithelial surface have been targeted in mouse models with some success. In a mouse model for radiation-induced dry eye, pretreatment with a viral vector to express erythropoietin improved the quality of the ocular surface as well as the exocrine function of lacrimal and salivary glands [70]. In another mouse model for dry eye, a plasmid with the MUC5AC gene, encoding glycoprotein mucin 5AC, was introduced onto the ocular surface by cationized gelatin-based nanoparticles. Results showed that the treatment was well tolerated with a favorable reduction in inflammation and improved tear production [71]. While currently in the preliminary stages, gene transfer has shown exciting potential for the treatment of dry eye.

# CONCLUSIONS

In keratoconjunctivitis sicca, autoimmunity plays a large role, particularly in Sjögren'sassociated KCS. While the complexity of the pathogenesis of disease has hindered progress in treatments for autoimmune-mediated KCS, research has identified key steps in the immune response that produce dysregulation, tissue injury, and symptomatic disease. Immune dysregulation is likely multi-factorial, with contributing factors including genetic susceptibility and dysfunction of antigen recognition, immune response, and/or immune regulation. The combination of basic science and translational and clinical research has elucidated numerous potential drug targets for KCS, a disease that currently has limited treatment options. As a chronic disease, KCS and its prevalence are expected to increase with the aging population. Thus, the development of new drug therapies for KCS will yield great benefit in the coming years.

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# References

- Schein OD, Hochberg MC, Munoz B, Tielsch JM, Bandeen-Roche K, Provost T, et al. Dry eye and dry mouth in the elderly: a population-based assessment. Archives of internal medicine. 1999 Jun 28; 159(12):1359–63. [PubMed: 10386512]
- The definition and classification of dry eye disease: report of the Definition and Classification Subcommittee of the International Dry Eye WorkShop. The ocular surface. 2007 Apr; 5(2):75–92. [PubMed: 17508116]

- Gottenberg JE, Busson M, Loiseau P, Cohen-Solal J, Lepage V, Charron D, et al. In primary Sjogren's syndrome, HLA class II is associated exclusively with autoantibody production and spreading of the autoimmune response. Arthritis and rheumatism. 2003 Aug; 48(8):2240–5. [PubMed: 12905478]
- Chen KS, Jiang MC, Li CJ, Liu OK, Tsai CS. Discrimination between Sjogren's and non-Sjogren's sicca syndrome by sialoscintigraphy and antibodies against alpha-fodrin and Ro/La autoantigens. The Journal of international medical research. 2009 Jul-Aug;37(4):1088–96. [PubMed: 19761691]
- 5. Mimori T. Autoantibodies in connective tissue diseases: clinical significance and analysis of target autoantigens. Internal medicine. 1999 Jul; 38(7):523–32. [PubMed: 10435356]
- Muller S, Briand JP, Barakat S, Lagueux J, Poirier GG, De Murcia G, et al. Autoantibodies reacting with poly(ADP-ribose) and with a zinc-finger functional domain of poly(ADP-ribose) polymerase involved in the recognition of damaged DNA. Clinical immunology and immunopathology. 1994 Nov; 73(2):187–96. [PubMed: 7923925]
- Haaheim LR, Halse AK, Kvakestad R, Stern B, Normann O, Jonsson R. Serum antibodies from patients with primary Sjogren's syndrome and systemic lupus erythematosus recognize multiple epitopes on the La(SS-B) autoantigen resembling viral protein sequences. Scandinavian journal of immunology. 1996 Jan; 43(1):115–21. [PubMed: 8560190]
- Stathopoulou EA, Routsias JG, Stea EA, Moutsopoulos HM, Tzioufas AG. Cross-reaction between antibodies to the major epitope of Ro60 kD autoantigen and a homologous peptide of Coxsackie virus 2B protein. Clinical and experimental immunology. 2005 Jul; 141(1):148–54. [PubMed: 15958081]
- Manganelli P, Fietta P. Apoptosis and Sjogren syndrome. Seminars in arthritis and rheumatism. 2003 Aug; 33(1):49–65. [PubMed: 12920696]
- Franz S, Gaipl US, Munoz LE, Sheriff A, Beer A, Kalden JR, et al. Apoptosis and autoimmunity: when apoptotic cells break their silence. Current rheumatology reports. 2006 Aug; 8(4):245–7. [PubMed: 16839503]
- Matsumura R, Umemiya K, Kagami M, Tomioka H, Tanabe E, Sugiyama T, et al. Glandular and extraglandular expression of the Fas-Fas ligand and apoptosis in patients with Sjogren's syndrome. Clinical and experimental rheumatology. 1998 Sep-Oct;16(5):561–8. [PubMed: 9779303]
- Polihronis M, Tapinos NI, Theocharis SE, Economou A, Kittas C, Moutsopoulos HM. Modes of epithelial cell death and repair in Sjogren's syndrome (SS). Clinical and experimental immunology. 1998 Dec; 114(3):485–90. [PubMed: 9844061]
- Yeh S, Song XJ, Farley W, Li DQ, Stern ME, Pflugfelder SC. Apoptosis of ocular surface cells in experimentally induced dry eye. Investigative ophthalmology & visual science. 2003 Jan; 44(1): 124–9. [PubMed: 12506064]
- 14. Rosen A, Casciola-Rosen L. Altered autoantigen structure in Sjogren's syndrome: implications for the pathogenesis of autoimmune tissue damage. Critical reviews in oral biology and medicine : an official publication of the American Association of Oral Biologists. 2004; 15(3):156–64.
- 15. Theofilopoulos AN, Kono DH, Beutler B, Baccala R. Intracellular nucleic acid sensors and autoimmunity. Journal of interferon & cytokine research : the official journal of the International Society for Interferon and Cytokine Research. 2011 Dec; 31(12):867–86.
- Yamazaki S, Muta T, Takeshige K. A novel IkappaB protein, IkappaB-zeta, induced by proinflammatory stimuli, negatively regulates nuclear factor-kappaB in the nuclei. The Journal of biological chemistry. 2001 Jul 20; 276(29):27657–62. [PubMed: 11356851]
- 17\*. Okuma A, Hoshino K, Ohba T, Fukushi S, Aiba S, Akira S, et al. Enhanced apoptosis by disruption of the STAT3-IkappaB-zeta signaling pathway in epithelial cells induces Sjogren's syndrome-like autoimmune disease. Immunity. 2013 Mar 21; 38(3):450–60. This recent study finds Sjögren's-like disease develops with disruption of the STAT3-IκB apoptotic pathway. [PubMed: 23453632]
- Yang XO, Panopoulos AD, Nurieva R, Chang SH, Wang D, Watowich SS, et al. STAT3 regulates cytokine-mediated generation of inflammatory helper T cells. The Journal of biological chemistry. 2007 Mar 30; 282(13):9358–63. [PubMed: 17277312]

- Zhang X, Chen W, De Paiva CS, Corrales RM, Volpe EA, McClellan AJ, et al. Interferon-gamma exacerbates dry eye-induced apoptosis in conjunctiva through dual apoptotic pathways. Investigative ophthalmology & visual science. 2011 Aug; 52(9):6279–85. [PubMed: 21474767]
- Brignole F, Pisella PJ, Goldschild M, De Saint Jean M, Goguel A, Baudouin C. Flow cytometric analysis of inflammatory markers in conjunctival epithelial cells of patients with dry eyes. Investigative ophthalmology & visual science. 2000 May; 41(6):1356–63. [PubMed: 10798650]
- Stern ME, Schaumburg CS, Siemasko KF, Gao J, Wheeler LA, Grupe DA, et al. Autoantibodies contribute to the immunopathogenesis of experimental dry eye disease. Investigative ophthalmology & visual science. 2012 Apr; 53(4):2062–75. [PubMed: 22395876]
- 22. Liu X, Wang S, Kao AA, Long Q. The effect of topical pranoprofen 0.1% on the clinical evaluation and conjunctival HLA-DR expression in dry eyes. Cornea. 2012 Nov; 31(11):1235–9. [PubMed: 22677643]
- Baudouin C, Brignole F, Pisella PJ, De Jean MS, Goguel A. Flow cytometric analysis of the inflammatory marker HLA DR in dry eye syndrome: results from 12 months of randomized treatment with topical cyclosporin A. Advances in experimental medicine and biology. 2002; 506(Pt B):761–9. [PubMed: 12613989]
- 24. Hamrah P, Liu Y, Zhang Q, Dana MR. The corneal stroma is endowed with a significant number of resident dendritic cells. Investigative ophthalmology & visual science. 2003 Feb; 44(2):581–9. [PubMed: 12556386]
- Pisella PJ, Brignole F, Debbasch C, Lozato PA, Creuzot-Garcher C, Bara J, et al. Flow cytometric analysis of conjunctival epithelium in ocular rosacea and keratoconjunctivitis sicca. Ophthalmology. 2000 Oct; 107(10):1841–9. [PubMed: 11013183]
- 26. Barabino S, Chen Y, Chauhan S, Dana R. Ocular surface immunity: homeostatic mechanisms and their disruption in dry eye disease. Progress in retinal and eye research. 2012 May; 31(3):271–85. [PubMed: 22426080]
- 27. Lin H, Li W, Dong N, Chen W, Liu J, Chen L, et al. Changes in corneal epithelial layer inflammatory cells in aqueous tear-deficient dry eye. Investigative ophthalmology & visual science. 2010 Jan; 51(1):122–8. [PubMed: 19628746]
- Moutsopoulos HM, Hooks JJ, Chan CC, Dalavanga YA, Skopouli FN, Detrick B. HLA-DR expression by labial minor salivary gland tissues in Sjogren's syndrome. Annals of the rheumatic diseases. 1986 Aug; 45(8):677–83. [PubMed: 3527087]
- Hamrah P, Liu Y, Zhang Q, Dana MR. Alterations in corneal stromal dendritic cell phenotype and distribution in inflammation. Archives of ophthalmology. 2003 Aug; 121(8):1132–40. [PubMed: 12912691]
- Paul P, van den Hoorn T, Jongsma ML, Bakker MJ, Hengeveld R, Janssen L, et al. A Genomewide multidimensional RNAi screen reveals pathways controlling MHC class II antigen presentation. Cell. 2011 Apr 15; 145(2):268–83. [PubMed: 21458045]
- Gupta A, Monroy D, Ji Z, Yoshino K, Huang A, Pflugfelder SC. Transforming growth factor beta-1 and beta-2 in human tear fluid. Current eye research. 1996 Jun; 15(6):605–14. [PubMed: 8670763]
- Zheng X, De Paiva CS, Rao K, Li DQ, Farley WJ, Stern M, et al. Evaluation of the transforming growth factor-beta activity in normal and dry eye human tears by CCL-185 cell bioassay. Cornea. 2010 Sep; 29(9):1048–54. [PubMed: 20539212]
- Shen L, Barabino S, Taylor AW, Dana MR. Effect of the ocular microenvironment in regulating corneal dendritic cell maturation. Archives of ophthalmology. 2007 Jul; 125(7):908–15. [PubMed: 17620569]
- 34. De Paiva CS, Volpe EA, Gandhi NB, Zhang X, Zheng X, Pitcher JD 3rd, et al. Disruption of TGFbeta signaling improves ocular surface epithelial disease in experimental autoimmune keratoconjunctivitis sicca. PloS one. 2011; 6(12):e29017. [PubMed: 22194977]
- Turpie B, Yoshimura T, Gulati A, Rios JD, Dartt DA, Masli S. Sjogren's syndrome-like ocular surface disease in thrombospondin-1 deficient mice. The American journal of pathology. 2009 Sep; 175(3):1136–47. [PubMed: 19700744]
- 36. Li H, Dai M, Zhuang Y. A T cell intrinsic role of Id3 in a mouse model for primary Sjogren's syndrome. Immunity. 2004 Oct; 21(4):551–60. [PubMed: 15485632]

- Niederkorn JY, Stern ME, Pflugfelder SC, De Paiva CS, Corrales RM, Gao J, et al. Desiccating stress induces T cell-mediated Sjogren's Syndrome-like lacrimal keratoconjunctivitis. Journal of immunology. 2006 Apr 1; 176(7):3950–7.
- 38. Salinas GF, Braza F, Brouard S, Tak PP, Baeten D. The role of B lymphocytes in the progression from autoimmunity to autoimmune disease. Clinical immunology. 2013 Jan; 146(1):34–45. [PubMed: 23202542]
- Cornec D, Devauchelle-Pensec V, Tobon GJ, Pers JO, Jousse-Joulin S, Saraux A. B cells in Sjogren's syndrome: from pathophysiology to diagnosis and treatment. Journal of autoimmunity. 2012 Sep; 39(3):161–7. [PubMed: 22749831]
- 40. Robinson CP, Brayer J, Yamachika S, Esch TR, Peck AB, Stewart CA, et al. Transfer of human serum IgG to nonobese diabetic Igmu null mice reveals a role for autoantibodies in the loss of secretory function of exocrine tissues in Sjogren's syndrome. Proceedings of the National Academy of Sciences of the United States of America. 1998 Jun 23; 95(13):7538–43. [PubMed: 9636185]
- 41. Voulgarelis M, Moutsopoulos HM. Lymphoproliferation in autoimmunity and Sjogren's syndrome. Current rheumatology reports. 2003 Aug; 5(4):317–23. [PubMed: 14531960]
- Salomonsson S, Jonsson MV, Skarstein K, Brokstad KA, Hjelmstrom P, Wahren-Herlenius M, et al. Cellular basis of ectopic germinal center formation and autoantibody production in the target organ of patients with Sjogren's syndrome. Arthritis and rheumatism. 2003 Nov; 48(11):3187–201. [PubMed: 14613282]
- 43. Hjelmervik TO, Petersen K, Jonassen I, Jonsson R, Bolstad AI. Gene expression profiling of minor salivary glands clearly distinguishes primary Sjogren's syndrome patients from healthy control subjects. Arthritis and rheumatism. 2005 May; 52(5):1534–44. [PubMed: 15880807]
- 44. Groom J, Kalled SL, Cutler AH, Olson C, Woodcock SA, Schneider P, et al. Association of BAFF/ BLyS overexpression and altered B cell differentiation with Sjogren's syndrome. The Journal of clinical investigation. 2002 Jan; 109(1):59–68. [PubMed: 11781351]
- 45. Mackay F, Browning JL. BAFF: a fundamental survival factor for B cells. Nature reviews Immunology. 2002 Jul; 2(7):465–75.
- De Paiva CS, Chotikavanich S, Pangelinan SB, Pitcher JD 3rd, Fang B, Zheng X, et al. IL-17 disrupts corneal barrier following desiccating stress. Mucosal immunology. 2009 May; 2(3):243– 53. [PubMed: 19242409]
- 47. Perez P, Goicovich E, Alliende C, Aguilera S, Leyton C, Molina C, et al. Differential expression of matrix metalloproteinases in labial salivary glands of patients with primary Sjogren's syndrome. Arthritis and rheumatism. 2000 Dec; 43(12):2807–17. [PubMed: 11145040]
- 48. Goicovich E, Molina C, Perez P, Aguilera S, Fernandez J, Olea N, et al. Enhanced degradation of proteins of the basal lamina and stroma by matrix metalloproteinases from the salivary glands of Sjogren's syndrome patients: correlation with reduced structural integrity of acini and ducts. Arthritis and rheumatism. 2003 Sep; 48(9):2573–84. [PubMed: 13130477]
- 49. Lee SY, Han SJ, Nam SM, Yoon SC, Ahn JM, Kim TI, et al. Analysis of Tear Cytokines and Clinical Correlations in Sjogren Syndrome Dry Eye Patients and Non-Sjogren Syndrome Dry Eye Patients. American journal of ophthalmology. 2013 Jun 7.
- Nguyen CQ, Yin H, Lee BH, Carcamo WC, Chiorini JA, Peck AB. Pathogenic effect of interleukin-17A in induction of Sjogren's syndrome-like disease using adenovirus-mediated gene transfer. Arthritis research & therapy. 2010; 12(6):R220. [PubMed: 21182786]
- Holtta V, Klemetti P, Sipponen T, Westerholm-Ormio M, Kociubinski G, Salo H, et al. IL-23/ IL-17 immunity as a hallmark of Crohn's disease. Inflammatory bowel diseases. 2008 Sep; 14(9): 1175–84. [PubMed: 18512248]
- Williams RO, Paleolog E, Feldmann M. Cytokine inhibitors in rheumatoid arthritis and other autoimmune diseases. Current opinion in pharmacology. 2007 Aug; 7(4):412–7. [PubMed: 17627887]
- 53. Bernard F, Romano A, Granel B. Regulatory T cells and systemic autoimmune diseases: systemic lupus erythematosus, rheumatoid arthritis, primary Sjogren's syndrome. La Revue de medecine interne / fondee par la Societe nationale francaise de medecine interne. 2010 Feb; 31(2):116–27. Lymphocytes T regulateurs et maladies auto-immunes systemiques : lupus erythemateux

systemique, polyarthrite rhumatoide et syndrome de Gougerot-Sjogren primaire. [PubMed: 19962219]

- 54. Chauhan SK, El Annan J, Ecoiffier T, Goyal S, Zhang Q, Saban DR, et al. Autoimmunity in dry eye is due to resistance of Th17 to Treg suppression. Journal of immunology. 2009 Feb 1; 182(3): 1247–52.
- Katsifis GE, Rekka S, Moutsopoulos NM, Pillemer S, Wahl SM. Systemic and local interleukin-17 and linked cytokines associated with Sjogren's syndrome immunopathogenesis. The American journal of pathology. 2009 Sep; 175(3):1167–77. [PubMed: 19700754]
- Marsh P, Pflugfelder SC. Topical nonpreserved methylprednisolone therapy for keratoconjunctivitis sicca in Sjogren syndrome. Ophthalmology. 1999 Apr; 106(4):811–6. [PubMed: 10201607]
- 57. Avunduk AM, Avunduk MC, Varnell ED, Kaufman HE. The comparison of efficacies of topical corticosteroids and nonsteroidal anti-inflammatory drops on dry eye patients: a clinical and immunocytochemical study. American journal of ophthalmology. 2003 Oct; 136(4):593–602. [PubMed: 14516798]
- Donnenfeld E, Pflugfelder SC. Topical ophthalmic cyclosporine: pharmacology and clinical uses. Survey of ophthalmology. 2009 May-Jun;54(3):321–38. [PubMed: 19422961]
- Tsubota K, Fujita H, Tadano K, Takeuchi T, Murakami T, Saito I, et al. Improvement of lacrimal function by topical application of CyA in murine models of Sjogren's syndrome. Investigative ophthalmology & visual science. 2001 Jan; 42(1):101–10. [PubMed: 11133854]
- 60. Sall K, Stevenson OD, Mundorf TK, Reis BL. CsA Phase 3 Study Group. Two multicenter, randomized studies of the efficacy and safety of cyclosporine ophthalmic emulsion in moderate to severe dry eye disease. Ophthalmology. 2000 Apr; 107(4):631–9. [PubMed: 10768324]
- Samarkos M, Moutsopoulos HM. Recent advances in the management of ocular complications of Sjogren's syndrome. Current allergy and asthma reports. 2005 Jul; 5(4):327–32. Epub 2005/06/22. eng. [PubMed: 15967078]
- 62. Gao J, Sana R, Calder V, Calonge M, Lee W, Wheeler LA, et al. Mitochondrial permeability transition pore in inflammatory apoptosis of human conjunctival epithelial cells and T cells: effect of cyclosporin a. Investigative ophthalmology & visual science. 2013; 54(7):4717–33. [PubMed: 23778874]
- Fleck M, Zhang HG, Kern ER, Hsu HC, Muller-Ladner U, Mountz JD. Treatment of chronic sialadenitis in a murine model of Sjogren's syndrome by local fasL gene transfer. Arthritis and rheumatism. 2001 Apr; 44(4):964–73. [PubMed: 11315936]
- 64. Nakamura S, Shibuya M, Saito Y, Nakashima H, Saito F, Higuchi A, et al. Protective effect of Dbeta-hydroxybutyrate on corneal epithelia in dry eye conditions through suppression of apoptosis. Investigative ophthalmology & visual science. 2003 Nov; 44(11):4682–8. [PubMed: 14578386]
- 65. Ono M, Takamura E, Shinozaki K, Tsumura T, Hamano T, Yagi Y, et al. Therapeutic effect of cevimeline on dry eye in patients with Sjogren's syndrome: a randomized, double-blind clinical study. American journal of ophthalmology. 2004 Jul; 138(1):6–17. [PubMed: 15234277]
- 66. Fife RS, Chase WF, Dore RK, Wiesenhutter CW, Lockhart PB, Tindall E, et al. Cevimeline for the treatment of xerostomia in patients with Sjogren syndrome: a randomized trial. Archives of internal medicine. 2002 Jun 10; 162(11):1293–300. [PubMed: 12038948]
- 67. Petrone D, Condemi JJ, Fife R, Gluck O, Cohen S, Dalgin P. A double-blind, randomized, placebo-controlled study of cevimeline in Sjogren's syndrome patients with xerostomia and keratoconjunctivitis sicca. Arthritis and rheumatism. 2002 Mar; 46(3):748–54. Epub 2002/03/29. eng. [PubMed: 11920411]
- Abdulahad WH, Kroese FG, Vissink A, Bootsma H. Immune regulation and B-cell depletion therapy in patients with primary Sjogren's syndrome. Journal of autoimmunity. 2012 Aug; 39(1– 2):103–11. [PubMed: 22341852]
- 69. Rocha EM, Di Pasquale G, Riveros PP, Quinn K, Handelman B, Chiorini JA. Transduction, tropism, and biodistribution of AAV vectors in the lacrimal gland. Investigative ophthalmology & visual science. 2011; 52(13):9567–72. \* This study establishes that adenoviral-associated vectors can be delivered to murine lacrimal glands, supporting gene therapy as a potential treatment method for KCS. [PubMed: 22110082]

- 70. Rocha EM, Cotrim AP, Zheng C, Riveros PP, Baum BJ, Chiorini JA. Recovery of radiationinduced dry eye and corneal damage by pretreatment with adenoviral vector-mediated transfer of erythropoietin to the salivary glands in mice. Human gene therapy. 2013 Apr; 24(4):417–23. [PubMed: 23402345]
- 71. Contreras-Ruiz L, Zorzi GK, Hileeto D, Lopez-Garcia A, Calonge M, Seijo B, et al. A nanomedicine to treat ocular surface inflammation: performance on an experimental dry eye murine model. Gene therapy. 2013 May; 20(5):467–77. [PubMed: 22809996]