HIV/AIDS

# Less Bone Loss With Maraviroc- Versus Tenofovir-Containing Antiretroviral Therapy in the AIDS Clinical Trials Group A5303 Study

# Babafemi O. Taiwo,<sup>1</sup> Ellen S. Chan,<sup>2</sup> Carl J. Fichtenbaum,<sup>3</sup> Heather Ribaudo,<sup>2</sup> Athe Tsibris,<sup>4</sup> Karin L. Klingman,<sup>5</sup> Joseph J. Eron,<sup>6</sup> Baiba Berzins,<sup>1</sup> Kevin Robertson,<sup>7</sup> Alan Landay,<sup>8</sup> Igho Ofotokun,<sup>9</sup> and Todd Brown<sup>10</sup>; for the AIDS Clinical Trials Group A5303 Study Team

<sup>1</sup>Division of Infectious Diseases, Northwestern University, Chicago, Illinois; <sup>2</sup>Statistical and Data Analysis Center, Harvard School of Public Health, Boston, Massachusetts; <sup>3</sup>Division of Infectious Diseases, University of Cincinnati, Ohio; <sup>4</sup>Division of Infectious Diseases, Brigham and Women's Hospital, Harvard Medical School, Boston, Massachusetts; <sup>5</sup>HIV Research Branch, Division of AIDS, National Institute of Allergy and Infectious Diseases, Bethesda, Maryland; Departments of <sup>6</sup>Infectious Diseases, and <sup>7</sup>Neurology, University of North Carolina at Chapel Hill; <sup>8</sup>Department of Immunology/Microbiology, Rush University Medical Center, Chicago, Illinois; <sup>9</sup>Division of Infectious Diseases, Department of Medicine, Emory University, Atlanta, Georgia; and <sup>10</sup>Division of Endocrinology, Diabetes, and Metabolism, Johns Hopkins University, Baltimore, Maryland

**Background.** There is a need to prevent or minimize bone loss associated with antiretroviral treatment (ART) initiation. We compared maraviroc (MVC)- to tenofovir disoproxil fumarate (TDF)–containing ART.

*Methods.* This was a double-blind, placebo-controlled trial. ART-naive subjects with human immunodeficiency virus type 1 RNA load (viral load [VL]) >1000 copies/mL and R5 tropism were randomized to MVC 150 mg or TDF 300 mg once daily (1:1), stratified by VL <100 000 or  $\geq$ 100 000 copies/mL and age <30 or  $\geq$ 30 years. All subjects received darunavir 800 mg, ritonavir 100 mg, and emtricitabine 200 mg daily. Dual-energy X-ray absorptiometry scanning was done at baseline and week 48. The primary endpoint was percentage change in total hip bone mineral density (BMD) from baseline to week 48 in the as-treated population.

**Results.** We enrolled 262 subjects. A total of 259 subjects (130 MVC, 129 TDF) contributed to the analyses (91% male; median age, 33 years; 45% white, 30% black, 22% Hispanic). Baseline median VL was 4.5  $\log_{10}$  copies/mL and CD4 count was 390 cells/µL. The decline in hip BMD (n = 115 for MVC, n = 109 for TDF) at week 48 was less with MVC (median [Q1, Q3] of -1.51% [-2.93%, -0.11%] vs -2.40% [-4.30%, -1.32%] for TDF (*P* < .001). Lumbar spine BMD decline was also less with MVC (median -0.88% vs -2.35%; *P* < .001). Similar proportions of subjects in both arms achieved VL ≤50 copies/mL in as-treated and ITT analyses.

*Conclusions.* MVC was associated with less bone loss at the hip and lumbar spine compared with TDF. MVC may be an option to attenuate ART-associated bone loss.

Clinical Trials Registration. NCT01400412.

Keywords. maraviroc; tenofovir; bone; darunavir.

Bone mineral density (BMD) declines by 2%–6% in the 2 years following antiretroviral therapy (ART) initiation, and the magnitude depends in part on the ART

Clinical Infectious Diseases® 2015;61(7):1179–88

regimen [1–5]. Use of tenofovir disoproxil fumarate (TDF) has been linked to a 1%–2% greater bone loss than other nucleos(t)ide reverse transcriptase inhibitors (NRTIs) [3, 5–8]. The bone loss after ART initiation is greatest within the first 24–48 weeks [5, 9, 10], suggesting that it may be linked to systemic immunologic and inflammatory changes that are also pronounced in the initial weeks to months after ART initiation [11, 12].

Maraviroc (MVC) has virologic activity against R5tropic human immunodeficiency virus type 1 (HIV-1) through inhibition of CC chemokine receptor 5 (CCR5), the main coreceptor for HIV-1 [13]. Studies

Received 10 April 2015; accepted 22 May 2015; electronically published 9 June 2015.

Correspondence: Babafemi O. Taiwo, MBBS, Division of Infectious Diseases, Northwestern University, 645 N Michigan Ave, Ste 900, Chicago, IL 60611 (b-taiwo@northwestern.edu).

<sup>©</sup> The Author 2015. Published by Oxford University Press on behalf of the Infectious Diseases Society of America. All rights reserved. For Permissions, please e-mail: journals.permissions@oup.com. DOI: 10.1093/cid/civ455

of potential immunomodulatory and antiinflammatory effects of MVC have produced conflicting results [14–18]. CCR5 is expressed on both osteoblasts and osteoclasts, and one of CCR5's primary ligands is macrophage inflammatory protein 1 alpha (MIP-1 $\alpha$ ), which has been shown to increase osteoclastogenesis and inhibit osteoblast function in preclinical models [19–21]. CCR5 may play an important role in the signaling between osteoblasts and osteoclasts [19]. Thus, it is plausible that MVC would have a beneficial effect on bone mineralization in the context of ART initiation.

The AIDS Clinical Trials Group (ACTG) study A5303 was conducted to investigate the effects of MVC on bone loss after ART initiation in treatment-naive HIV-1–infected patients. Our a priori hypothesis was that initiating a MVCcontaining ART regimen would be associated with less bone loss compared to a regimen containing TDF. We also compared the antiretroviral efficacy of the regimens.

# **METHODS**

# **Study Design**

A5303 was a phase 2, prospective, double-blind, placebocontrolled, multicenter, 48-week clinical trial conducted at 33 ACTG and 4 Adolescent Trials Network research sites in the United States. Eligible subjects were ART-naive patients (aged ≥18 years) with plasma HIV-1 RNA concentration (viral load [VL]) >1000 copies/mL and R5 tropism by Trofile phenotypic assay (Monogram Biosciences, South San Francisco, California). The inclusion and exclusion criteria are listed at Clinical-Trials.gov (NCT01400412). The institutional review board of each study site approved the protocol. Each participant provided written informed consent.

# **Study Procedures**

Eligible subjects were randomized (1:1) to MVC 150 mg plus TDF placebo or TDF 300 mg plus MVC placebo, stratified by screening VL <100 000 or  $\geq$ 100 000 copies/mL and age <30 or  $\geq$ 30 years. Each subject also received darunavir 800 mg, ritonavir 100 mg, and emtricitabine 200 mg. Although the recommended dose of MVC is 150 mg twice daily when combined with ritonavir-boosted darunavir (DRV/r) [22], we selected a dose of 150 mg once daily based on pharmacokinetic and clinical studies supporting this lower dosing [23–27]. Subjects were instructed to take the study drugs with food once daily.

Within 4 weeks prior to randomization, each subject underwent a baseline dual-energy X-ray absorptiometry (DXA) scan of the left hip and lumbar spine (L1–L4) at the study site, using either a Lunar (GE Healthcare, Fairfield, Connecticut) or Hologic (Hologic Incorporated, Bedford, Massachusetts) DXA scanner. A second DXA scan was performed at week 48 (±4 weeks) using the same DXA scanning system. For subjects who had a permanent change in the MVC or TDF component of their randomized regimen or who prematurely discontinued the study, DXA scanning was performed at the time of discontinuation. All DXA scans were read centrally at the Body Composition Analysis Center at Tufts University. The European Spine Phantom was used for cross-calibration of DXA machines and quality assurance at each site. The *z* scores—the number of standard deviations a subject's BMD falls from the mean BMD—were calculated from the site-specific BMD measures using normative data matched for age, sex, and race. The *z* scores were chosen over *t* scores given the relatively young age of the study population [28].

Routine study visits after randomization occurred at week 4 ( $\pm$ 7 days), and weeks 16, 24, 36, and 48 (all  $\pm$ 14 days). Viral load (Abbott Real*Time* assay HIV-1, lower detection limit of 40 copies/mL), CD4 cell count, hematology, liver function tests, and blood chemistry were measured at each visit. Adherence to study medications was assessed by self-report at all study visits postentry except week 36.

# **Study Endpoints**

The primary endpoint was the percentage of change in total hip BMD from baseline to week 48. The main secondary endpoint was the percentage of change in lumbar spine BMD from baseline to week 48. Other secondary endpoints included time to virologic failure, proportion of subjects with VL <50 copies/mL, changes in CD4 cell count from baseline, emergent resistance during failure, and incidence plus severity of adverse events. Virologic failure was defined as 2 consecutive VL results >1000 copies/mL at or after week 16 and before week 24, or >200 copies/mL at or after week 24. A confirmatory VL measurement was obtained within 30 days of receiving an initial virologic failure result. Subjects who discontinued the study with an unconfirmed virologic failure result were considered to have virologic failure at the visit week of the initial result. Time to virologic failure was defined as the time from study entry to the visit week of the initial failure; subjects without evidence of virologic failure had their time to virologic failure censored at the study week of their last VL measurement. Emergent resistance was assessed using plasma samples obtained at the virologic failure confirmation visit by genotyping the HIV-1 reverse transcriptase and protease genes.

# **Statistical Analyses**

The target sample size of 127 subjects per arm (total of 254) provided 90% power to detect a difference of 1.5% or larger in total hip BMD change from baseline to week 48 between the 2 arms, assuming that 20% of subjects would be nonevaluable due to scan failure or loss to follow-up. This sample size also provided 87% power to claim noninferiority of the MVC arm for the virologic efficacy aim, assuming a cumulative probability of virologic failure of 15% in both arms by week 48, a maximum allowable difference of 15%, and 10% loss to follow-up. The primary analysis was as-treated and included only subjects who remained on their randomized treatment without any interruption of >10 weeks. Intent-to-treat (ITT) analyses that included outcomes regardless of status on randomized treatment were also performed using 3 different approaches to handle missing BMD data. The first approach assumed that missing data occurred completely at random, and thus only included subjects with total hip BMD measurements available at both baseline and week 48 (complete case). The other approaches used to handle missing data assumed informative missing data. Specifically, missing week 48 measurements were imputed with (1) the last available DXA scan measurement while on randomized regimen after at least 12 weeks of study treatment (last observation carried forward), and (2) an arbitrary value less than any percentage week 48 change from baseline, that is, largest decrease from baseline (worst rank). Stratified Wilcoxon rank-sum tests were used to test for differences between the 2 treatment groups, stratified by age (<30 vs  $\geq$ 30 years). Wilcoxon signed-rank tests were used to test for within-treatment-group changes greater than zero; 95% confidence intervals (CIs) for



Figure 1. Consolidated Standards of Reporting Trials (CONSORT) diagram. Abbreviations: BMD, bone mineral density; DRV/r, darunavir/ritonavir; DXA, dual-energy X-ray absorptiometry; FTC, emtricitabine; IVDU, intravenous drug user; MVC, maraviroc; TDF, tenofovir disoproxil fumarate.

median changes within treatment group were estimated using distribution-free method via percentiles. Linear regression models were used to evaluate interactions between treatment arm and age, baseline VL, and race/ethnicity (post hoc).

Product-limit estimates were used to estimate the cumulative probability of virologic failure over time and its corresponding 95% CI for each treatment group. The difference in these estimated probabilities at week 48 was estimated with a 95% CI stratified by VL at screening; stratum specific variances on the estimated 48-week failure probability were used to define the stratum weights and compared (upper bound) against the noninferiority boundary of 15 percentage points. The proportion of subjects in each arm with VL  $\leq$ 50 copies/mL at weeks 24 and 48 was calculated using the as-treated approach described above as well as 2 ITT analyses (missing VL ignored; missing VL equals failure [>50 copies/mL]).

Analyses of CD4 count used the same as-treated population as the BMD as-treated analysis. Safety analyses included all subjects who started study treatment. All statistical tests were 2-sided and interpreted at the 5% nominal level of significance without adjustment for multiple comparisons. Analyses were conducted using SAS statistical software version 9.4.

# RESULTS

Figure 1 shows the disposition of the 262 subjects enrolled in the trial. Three subjects (all from the TDF arm) never initiated study treatment and were excluded from all analyses. The analyzed population (N = 259 [130 MVC, 129 TDF]) was 91% male, with a median age of 33 years; 45% non-Hispanic white (white), 30% non-Hispanic black (black), and 22% Hispanic. At baseline, median VL was 4.5 log<sub>10</sub> copies/mL, and CD4 count was 390 cells/ $\mu$ L. Baseline demographics and disease characteristics were generally similar between the study arms, but there was a chance imbalance with more black subjects and more

# Table 1. Baseline Characteristics and Bone Mineral Density

Characteristic	MVC (n = 130)	TDF (n = 129)	Total (N = 259)				
Age, y							
Median (Q1, Q3)	33 (26, 42)	33 (26, 42)	33 (26, 42)				
<30 y	49 (38)	48 (37)	97 (37)				
≥30 y	81 (62)	81 (63)	162 (63)				
Sex							
Male	115 (88)	120 (93)	235 (91)				
Female	15 (12)	9 (7)	24 (9)				
Race/ethnicity							
White non-Hispanic	57 (44)	59 (46)	116 (45)				
Black non-Hispanic	45 (35)	33 (26)	78 (30)				
Hispanic	24 (18)	34 (26)	58 (22)				
Other	4 (4)	3 (3)	7 (2)				
HIV-1 RNA, log <sub>10</sub> copies/mL, median (Q1, Q3)	4.59 (3.91, 5.07)	4.47 (4.02, 4.91)	4.50 (3.97, 5.00)				
HIV-1 RNA, copies/mL							
<100 000	92 (71)	102 (80)	194 (75)				
≥100 000	38 (29)	26 (21)	64 (25)				
CD4 count, cells/µL, median (Q1, Q3)	389 (295, 496)	392 (290, 518)	390 (294, 517)				
Hepatitis C antibody positive	10 (8)	12 (9)	22 (8)				
Creatinine clearance, mL/min							
Median (Q1, Q3)	124 (105, 152)	126 (106, 139)	124 (106, 145)				
>90 mL/min	90%	89%	90%				
Body mass index, kg/m <sup>2</sup> , median (Q1, Q3)	25 (22, 29)	26 (23, 29)	25 (23, 29)				
BMD, median (Q1, Q3)							
Total hip BMD, g/cm <sup>2</sup>	1.05 (0.96, 1.18)	1.03 (0.95, 1.15)	1.04 (0.95, 1.17)				
z score	-0.2 (-0.9, 0.6)	-0.1 (-0.8, 0.6)	-0.1 (-0.8, 0.6)				
Lumbar spine BMD, g/cm <sup>2</sup>	1.16 (1.03, 1.28)	1.11 (1.00, 1.20)	1.14 (1.02, 1.25)				
z score	-0.2 (-1.0, 0.6)	-0.3 (-1.3, 0.6)	-0.3 (-1.1, 0.6)				
Femoral neck BMD, g/cm <sup>2</sup>	1.01 (0.88, 1.13)	0.94 (0.85, 1.07)	0.98 (0.86, 1.12)				
z score	0.0 (-0.8, 0.8)	-0.2 (-0.9, 0.6)	-0.1 (-0.8, 0.7)				

Data are presented as No. (%) unless otherwise specified.

Abbreviations: BMD, bone mineral density; HIV-1, human immunodeficiency virus type 1; MVC, maraviroc; TDF, tenofovir disoproxil fumarate.

subjects with baseline VL >100 000 copies/mL in the MVC arm (Table 1). Subjects in both study arms had similar baseline BMD.

# **BMD Changes**

The primary as-treated analysis of the percentage of change from baseline to week 48 in hip BMD included 224 subjects (115 subjects in the MVC group and 109 in the TDF group). As shown in Figure 2A, there was a decline in hip BMD in both arms, which was smaller in the MVC group (P < .001): the median (Q1, Q3) percentage of change in BMD was -1.51% (-2.93%, -0.11%) for the MVC group compared with -2.40%(-4.30%, -1.32%) for the TDF group. Lumbar spine BMD also declined less in the MVC group than in the TDF group (P = .001); median (Q1, Q3) percentage of change was -0.88% (-2.93%, 1.30%) for the MVC group and -2.35% (-4.25%, -0.45%) for the TDF group. ITT analyses yielded similar conclusions (all P < .001; data not shown).

Given the chance racial and VL imbalance at randomization, a post hoc linear regression analysis adjusting for race (black vs other) was performed. Following initial model diagnostics, 2 outlying data points were excluded. Analyses were adjusted for age and baseline VL as well as baseline BMD at the relevant body site. Although adjustment for age stratum, baseline VL, and race/ethnicity did not alter the primary finding of a smaller decline in hip BMD in the MVC group compared with TDF  $(P \leq .001)$ , a differential effect of MVC by race/ethnicity was apparent (interaction between study treatment and race/ethnicity; P = .034, Figure 3). This interaction appears to have been driven by a smaller decline in BMD with MVC among black participants (Figure 2). The estimated difference between MVC vs TDF in percentage of bone loss in hip over 48 weeks among nonblack participants was 0.71% (95% CI, -0.13% to 1.55%; P = .096), compared with 2.34% (95% CI, 1.10%-3.58%; P = .0003) in black participants. The observed difference in BMD loss at the spine between the MVC and TDF groups was larger in black participants compared with nonblack participants; however, this difference was not statistically significant (P = .31). The estimated difference between MVC vs TDF in percentage of spine BMD loss over 48 weeks in nonblack participants was 1.15% (95% CI, .13%-2.18%; P = .028) compared with 2.09% (95% CI, .58%-3.61%; P = .007) in black participants. No evidence of treatment interactions with age or baseline VL was apparent at either the hip or spine (P > .59).

# Efficacy and Safety

There were 14 virologic failures (8 in the MVC group and 6 in the TDF group) by week 48, 10 of which were confirmed (8 in the MVC group and 2 in the TDF group) and 4 (all in the TDF arm) who were lost to follow-up after initial failure. The median difference between the arms (MVC minus TDF) in the



**Figure 2.** Percentage of change in bone mineral density from baseline to week 48 among all participants (*A*), nonblack participants (*B*), and non-Hispanic black participants (*C*). The line inside the box indicates the median value. The lower and upper edges of the box indicate the first and third quartiles (the 25th and 75th percentiles). The lower and upper whiskers are the first and third quintiles  $\pm 1.5$  times interquartile range. Stratified Wilcoxon rank-sum tests were used to test for differences between the 2 treatment groups, stratified by age (<30 vs  $\geq$ 30 years). Abbreviations: MVC, maraviroc; TDF, tenofovir disoproxil fumarate.



**Figure 3.** Adjusted treatment effects on the percentage of change in hip (*A*) and spine (*B*) bone mineral density (BMD) from baseline to week 48 with 95% confidence intervals (CIs). Following initial model diagnostics, 2 extreme outlying and influential data points were excluded: an extreme decrease (-32.1%) in the maraviroc (MVC) group and extreme increase (+26.6%) in the tenofovir disoproxil fumarate (TDF) group. Estimates from simple linear regression analyses are (1) unadjusted, (2) stratified by age (<30 y,  $\geq 30$  y) and baseline human immunodeficiency virus type 1 (HIV-1) RNA ( $<100\ 000$ ,  $\geq 100\ 000$  copies/mL), (3) adjusted for baseline BMD at the specific site, (4) adjusted for race (nonblack vs non-Hispanic black), and (5) adjusted by race. Models 3–5 are also stratified by age and baseline HIV-1 RNA level; models 4 and 5 also adjust for baseline BMD.

cumulative probability of virologic failure while on randomized treatment (as-treated) was 2% (95% CI, -4% to 5%), which was well within the predefined noninferiority margin (Table 2). In as-treated analysis, VL  $\leq$ 50 copies/mL was achieved in 85% and 93% of subjects in the MVC and TDF arms, respectively, at week 24 (P = .061), whereas 94% had VL  $\leq$ 50 copies/mL in both arms at week 48 (P = .893). ITT analyses yielded similar results (Table 2). Four subjects (3 in the MVC group and 1 in the TDF group) underwent successful genotyping, of whom 1 subject (on MVC) had the M184V NRTI mutation and the polymorphic mixture V118I/V. There were no protease inhibitor (PI) resistanceassociated mutations.

Significant within-group increases in CD4 count occurred from baseline to week 48 in both groups (P < .001). The median (Q1, Q3) increase in the MVC group was 234 (131, 327) cells/ µL, which was greater than the increase of 188 (94, 304) cells/µL in the TDF group (P = .036). Both regimens were well tolerated. Grade 3 adverse events occurred in 10% of subjects in the MVC arm and 14% of those on TDF, whereas 2% and 3%, respectively, experienced grade 4 adverse events (Supplementary Table 1). There were no deaths.

# DISCUSSION

We determined BMD changes in HIV-1-infected patients initiating MVC vs TDF, each combined with DRV/r and emtricitabine. Similar to other studies [1-5], BMD declined over the first 48 weeks in both treatment groups. However, the magnitude of the decline was less in the MVC arm, with a median of -1.5% at the hip and -0.9% at the lumbar spine, compared with -2.4% and -2.4%, respectively, in the TDF arm. These results are consistent with randomized trials of TDF- vs abacavircontaining regimens that found approximately 1%-2% greater BMD decline with TDF use [3, 5, 7, 8]. The results also recapitulate the findings of a recent clinical trial of TDF vs tenofovir alafenamide (TAF), an oral tenofovir prodrug that achieves high intracellular levels of the active metabolite, tenofovir diphosphate, while maintaining 90% lower plasma levels of tenofovir than TDF. At week 48, TAF combined with elvitegravircobicistat resulted in -0.7% BMD loss at the hip and -1.3% decline at the lumbar spine compared with -3.0% and -2.9%, respectively, with TDF [29].

Post hoc analysis motivated by a chance racial imbalance at randomization revealed that the difference in BMD loss with

Table 2. Cumulative Probability of Virologic Failure and Proportion With Human Immunodeficiency Virus Type 1 RNA <50 Copies/mL

	Study Week	(As-Treated) Cumulative Probability of Virologic Failure, % (95% CI) <sup>a</sup>	Proportion With HIV-1 RNA ≤50 copies/mL, %			
Treatment Group			On-Treatment	ITT, Missing Date Ignored	ITT, Missing Equals Failure	
MVC						
	16	0 (0–0)				
	24	4 (2–9)	85	85	83	
	36	5 (2–10)				
	48	5 (2–10)	94	93	86	
TDF						
	16	1 (0–5)				
	24	2 (1–7)	93	93	87	
	36	2 (1–7)				
	48	3 (1–9)	94	92	81	
			95% CI for between-arm difference (MVC-TDF) at 24 wk			
			(–16%, 0%)	(-16%, 0%)	(–13%, 5%)	
Between-arm difference (MVC-TDF) by 48 wk <sup>b</sup>		95% CI for between-arm difference (MVC-TDF) at 48 wk				
		2 (-4, 5)	(-6%, 7%)	(-6%, 8%)	(-4%, 15%)	

Abbreviations: CI, confidence interval; HIV-1, human immunodeficiency virus type 1; ITT, intent-to-treat; MVC, maraviroc; TDF, tenofovir disoproxil fumarate.

<sup>a</sup> Product limit estimate of the cumulative probability of virologic failure while on randomized treatment. Subjects without prior virologic failure are censored at the earliest of discontinuation of randomized treatment or end of study.

<sup>b</sup> Stratified by screening HIV-1 RNA stratum.

MVC compared to TDF was greater in black participants compared with other racial/ethnic groups, reaching statistical significance at the hip but not the lumbar spine. This observation appears to have been driven by less BMD loss with MVC among black subjects.

Nonuse of TDF is the putative explanation for the lower BMD decline in the MVC arm of our study. The mechanisms of TDF's negative impact on BMD remain uncertain, but may include direct effects of tenofovir on bone cells, or indirect effects on calcium-phosphate homeostasis, resulting in inadequate bone mineralization [30, 31]. It is less clear whether MVC had a protective effect, although the interaction between BMD changes in the MVC arm and race is intriguing. Race was not evaluated in a small (N = 27) 48-week study that showed a 2.1% increase in proximal femur BMD after switching from virally suppressive triple ART to MVC 300 mg plus DRV/r [32]. The divergence of MVC's effect in racial/ethnic groups is unlikely to be explained by any of the theoretical pathways for a beneficial bone effect of MVC. One proposed pathway is that MVC may modulate factors that contribute to skeletal deterioration such as proinflammatory and immunomodulatory cytokines [33-35]. The evidence on whether MVC attenuates systemic immune activation and inflammation independent of its virologic effects, however, remains mixed [14-16]. Another hypothetical pathway for a beneficial skeletal effect of MVC involves the proosteoclastogenic MIP-1 $\alpha$ , a CCR5 ligand [19–21]. Because bone is continuously remodeled by bone-forming osteoblasts and bone-resorbing osteoclasts, inhibition of MIP-1 $\alpha$  may tip the balance toward bone formation. Further examination of the effects of MVC on biomarkers of inflammation and immune activation and the associations with BMD changes and race/ethnicity is planned.

MVC was combined with emtricitabine and DRV/r in our study. Emtricitabine has not been independently implicated in bone loss, although its bone effects have not been previously investigated to our knowledge. In contrast, use of some ritonavir-boosted PIs has been associated with more bone loss in multiple studies [3, 8, 36, 37]. In the randomized trial of 3 initial regimens containing TDF and emtricitabine (A5260s), for example, BMD loss at 96 weeks was comparable between DRV/r and atazanavir/ritonavir recipients, whereas both of these groups experienced a greater decline than the raltegravir group [37]. Whereas the greater bone loss with boosted PIs has been attributed to higher tenofovir levels when TDF and boosted PIs are coadministered [38], the relationship between boosted PIs and bone health is more complex and has not been fully elucidated. In A5224s, independent effects of atazanavir/ritonavir vs efavirenz were seen in the lumbar spine but not the hip, and the effect of TDF on bone did not differ between the atazanavir/ritonavir and efavirenz arms [3]. In vitro studies have shown evidence of PI effects on osteoblast and osteoclast function, as well as inhibition of 1- $\alpha$  hydroxylase activity, which converts 25-hydroxyvitamin D to bioactive 1,25-dihydroxyvitamin D [39-41]. Studies of MVC-containing regimens that do not include boosted PIs and tenofovir may further illuminate the interactions between MVC and bone loss.

An unapproved MVC dose of 150 mg daily was used in this study. This dosing was based on several studies [23-27], and is affirmed by our finding of similar virologic efficacy between the experimental MVC regimen and the TDF-containing standard of care. Of note, the MODERN (Maraviroc Once-Daily with Darunavir Enhanced by Ritonavir in a New Regimen) study was terminated prematurely due to inferiority of MVC 150 mg plus DRV/r 800/100 mg once daily vs the same TDFcontaining comparator used in our study [42]. Addition of emtricitabine to MVC 150 mg plus DRV/r 800/100 mg in our study likely contributed to the regimen's impressive efficacy. It is unknown whether conventional MVC dosing would amplify any beneficial effect of MVC or uncover untoward effects. Another limitation of our study is the dearth of information on the clinical significance of observed differences in BMD decline between MVC vs TDF. However, cumulative TDF exposure has been independently associated with osteoporotic fracture [36]. We did not evaluate the impact of smoking and alcohol use as these data were not collected prospectively. Finally, the participants were relatively young (median age, 33 years) and only 9% were female, limiting the study's generalizability.

Taken together, our findings demonstrate BMD differences that may be expected with MVC- vs TDF-based initial ART.

# **Supplementary Data**

Supplementary materials are available at *Clinical Infectious Diseases* online (http://cid.oxfordjournals.org). Supplementary materials consist of data provided by the author that are published to benefit the reader. The posted materials are not copyedited. The contents of all supplementary data are the sole responsibility of the authors. Questions or messages regarding errors should be addressed to the author.

#### Notes

Acknowledgments. We thank all the patients who volunteered for the study. We acknowledge members of the ACTG A5303 team: Denise Barr (clinical trials specialist), Ana I. Martinez (Division of AIDS pharmacist), Robert Kalayjian and Cara Wilson (investigators), Edward Acosta (pharmacologist), David Rusin (data manager), Amy Gonzales (laboratory data manager), Orlando Roman (community representative), Kathy Melbourne and James Rooney (Gilead), Alex Reinhart (ViiV), Bryan Baugh (Janssen), Roula Qaqish (AbbVie), Heera R. Jayvant (Pfizer), and Laura Napolitano, Charles Walworth, and Christos Petropoulos (LabCorp). We gratefully acknowledge the research sites and personnel who participated in this study: Ohio State University (John Davis, Mark Hite); University of California, San Diego Antiviral Research Center (Edward Seefried, Constance Benson); Northwestern University (Nina Lambert, Karen Coleman); University of Alabama (Messer, Tamara James); University of Cincinnati (Amy Dill, Jenifer Baer); University of Colorado Hospital (Christine Griesmer, Cathi Basler); University of Miami (Hector Bolivar, Margaret A. Fischl); Houston AIDS Research Team (Roberto C. Arduino, Aristoteles E. Villamil); Rush University (Beverly Sha, Tondria Green); Vanderbilt University (Brenda Jackson, Fred Nicotera); Washington University in St Louis (GeYoul Kim, Mark Rodrieguez); University of California, San Francisco (Annie Luetkemeyer, Jay

Dwyer); The Miriam Hospital (Pamela Poethke, Aadia Rana); University of Carolina, Chapel Hill (Miriam Chicurel-Bayard, Megan Telfer); University of North Carolina at Greensboro research site (Cornelius van Dam, Timothy Lane); Ponce de Leon Center, Emory University (Ighovwerha Ofotokun, Melody Palmore); Case Western University (Patricia Walton, Felicia Williams); Puerto Rico Clinical Trials Unit (Jorge L. Santana, Olga I. Mendez); University of Pennsylvania (Pablo Tebas, Aleshia Thomas); Massachusetts General Hospital (Teri Flynn, ANP-BC, Amy Sbrolla); University of California, Los Angeles CARE Center (Raphael Landovitz, Vanessa Cajahuaringa); University of Washington AIDS CRS (Shelia Dunaway, Sheryl Storey); Johns Hopkins University (Ilene Wiggins, Andrea Weiss); University of Colorado (Daniel Reirden, Hannah Bernath); Columbia Physicians and Surgeons (Michael Yin, Jolene Noel-Connor); Cooper University Hospital (Rose Kim, Yolanda Smith); Brigham and Women's Hospital (Paul Sax, Cheryl Keenan); Georgetown University (Princy Kumar, Joseph Timpone); University of Southern California (Michael P. Dubé, Bartolo Santos); and University of Rochester (Mary Adams, Christine Hurley).

**Disclaimer.** The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health (NIH).

Financial support. This work was supported by the National Institute of Allergy and Infectious Diseases (award number U01AI068636); the National Institute of Mental Health; and the National Institute of Dental and Craniofacial Research. This work was also supported by grants from the NIH (grant numbers AI068634 and AI068636 to the ACTG Statistical Data Analysis Center [to E. S. C. and H. R.] and to the research sites that participated in the study [grant numbers UMAI069494, UMAI069432, UM1AI 069471, UM1AI069452, UM1 AI069501, 2UM1AI069432, UL1 TR001082, 1U01AI069477-01, P30AI073961, 5UM1 AI068636, 2UM1AI069503, UM1 AI069471, 2UM1AI069439-08, and UL1 TR000445 from the National Center for Advancing Translational Sciences/NIH, AI69439, UM1 AI069496, 5UM1AI069412, UM1 AI069423, 1UL1TR001111, P30 AI50410, 2UMIA1069423-08, 2UM1AI069418-08, 2P30 AI 50409-10, UL1TR000454, AI069501, 5UM1AI069415-10, 2UM1AI069412-08, AI069424, UL1 RR025780, 2UM1-AI069470-08, UM1AI069472, 2UMAI069432, AI 69501, UM1AI069471, UM1A 068636-09, 5 P30 AI-045008-15, U01AI069447, NO1-HD-3-3345, UMI AI069511, UM1 AI069465, UL1TR001079, UL1 RR024160, and UL1 TR000042]). ViiV, Gilead, and AbbVie provided study drugs. ViiV provided funding for DXA scanning.

**Potential conflicts of interest.** B. O. T. has served as a consultant to ViiV, Pfizer, Janssen, GlaxoSmithKline (GSK), and Gilead, and has received research support to Northwestern University from ViiV and Pfizer. C. J. F.'s institution receives research funding from Gilead, Pfizer, and Cubist. J. J. E has received grants from and/or served as a consultant for Bristol-Myers Squibb (BMS), Merck, ViiV, GSK, Gilead, and AbbVie. A. L. has served on the scientific advisory board for Merck and has provided scientific consultation to GSK, BMS, and Tobira. I. O. receives research funding to Emory University from BMS. T. B. has served as a consultant for BMS, GSK, Merck, AbbVie, Gilead, ViiV, Theratechnologies, and EMD-Serono and has received research funding from Merck and GSK. All other authors report no potential conflicts.

All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed.

#### References

- Duvivier C, Kolta S, Assoumou L, et al. Greater decrease in bone mineral density with protease inhibitor regimens compared with nonnucleoside reverse transcriptase inhibitor regimens in HIV-1 infected naive patients. AIDS 2009; 27:817–24.
- van Vonderen MG, Lips P, van Agtmael MA, et al. First line zidovudine/ lamivudine/ lopinavir/ritonavir leads to greater bone loss compared to nevirapine/lopinavir/ritonavir. AIDS 2009; 23:1367–76.

- McComsey GA, Kitch D, Daar E, et al. Bone mineral density and fractures in antiretroviral-naive persons randomized to receive abacavirlamivudine or tenofovir disoproxil fumarate-emtricitabine along with efavirenz or atazanavir-ritonavir: AIDS Clinical Trials Group A5224s, a substudy of ACTG A5202. J Infect Dis 2011; 203:1791–801.
- Brown TT, McComsey GA, King MS, et al. Loss of bone mineral density after antiretroviral therapy initiation, independent of antiretroviral regimen. J Acquir Immune Defic Syndr 2009; 51:554–61.
- Stellbrink HJ, Orkin C, Arribas JR, et al. Comparison of changes in bone density and turnover with abacavir-lamivudine versus tenofovir-emtricitabine in HIV-infected adults: 48-week results from the ASSERT study. Clin Infect Dis 2010; 51:963–72.
- Gallant JE, Staszewski S, Pozniak AL, et al; 903 Study Group. Efficacy and safety of tenofovir DF vs. stavudine in combination therapy in antiretroviral-treatment naive patients: a 3-year randomized trial. JAMA 2004; 292:191–201.
- Moyle GJ, Stellbrink HJ, Compston J, et al. 96-week results of abacavir/ lamivudine versus tenofovir/emtricitabine, plus efavirenz, in antiretroviralnaive, HIV-1-infected adults: ASSERT study. Antivir Ther 2013; 18:905–13.
- Huang JS, Hughes MD, Riddler SA, et al. Bone mineral density effects of randomized regimen and nucleoside reverse transcriptase inhibitor selection from ACTG A5142. J Infect Dis 2011; 203:1791–801.
- 9. Hansen AB, Obel N, Nielsen H, Pedersen C, Gerstoft J. Bone mineral density changes in protease inhibitor-sparing vs. nucleoside reverse transcriptase inhibitor-sparing highly active antiretroviral therapy: data from a randomized trial. HIV Med **2011**; 12:157–65.
- Grant PM, Kitch D, McComsey GA. Low baseline CD4+ count is associated with greater bone mineral density loss after antiretroviral therapy initiation. Clin Infect Dis 2013; 57:1483–8.
- Gandhi RT, Spritzler J, Chan E, et al. Effect of baseline- and treatmentrelated factors on immunologic recovery after initiation of antiretroviral therapy in HIV-1 positive subjects: results from ACTG 384. J Acquir Immune Defic Syndr 2006; 42:426–34.
- Grant PM, Komarrow L, Andersen J, et al. Risk analyses for immune reconstitution inflammatory syndrome in a randomized study of early vs. deferred ART during an opportunistic infection. PLoS One 2010; 5: e11416.
- Dorr P, Westby M, Dobbs S, et al. Maraviroc (UK-427,857), a potent, orally bioavailable, and selective small-molecule inhibitor of chemokine receptor CCR5 with broad-spectrum anti-human immunodeficiency virus type 1 activity. Antimicrob Agents Chemother 2005; 49:4721–32.
- Funderburg N, Kalinowska M, Eason J, et al. Effects of maraviroc and efavirenz on markers of immune activation and inflammation and associations with CD4+ cell rises in HIV-infected patients. PLoS One 2010; 5:e13188.
- Wilkin TJ, Lalama CM, McKinnon J, et al. A pilot trial of adding maraviroc to suppressive antiretroviral therapy for suboptimal CD4+T-cell recovery despite sustained virologic suppression: ACTG A5256. J Infect Dis 2012; 206:534–42.
- Arberas H, Guardo AC, Bargalló ME, et al. In vitro effects of the CCR5 inhibitor maraviroc on human T cell function. J Antimicrob Chemother 2013; 68:577–86.
- Pozo-Balado MM, Martínez-Bonet M, Rosado I, et al. Maraviroc reduces the regulatory T-cell frequency in antiretroviral-naive HIV-infected subjects. J Infect Dis 2014; 210:890–8.
- Hunt PW, Shulman NS, Hayes TL, et al. The immunologic effects of maraviroc intensification in treated HIV-infected individuals with incomplete CD4+ T-cell recovery: a randomized trial. PLoS One 2013; 8:e80157.
- Yano S, Mentaverri R, Kanuparthi D, et al. Functional expression of beta-chemokine receptors in osteoblasts: role of regulated upon activation, normal T cell expressed and secreted (RANTES) in osteoblasts and regulation of its secretion by osteoblasts and osteoclasts. Endocrinology 2005; 146:2324–35.
- Han JH, Choi SJ, Kurihara N, et al. Macrophage inflammatory proteinlalpha is an osteoclastogenic factor in myeloma that is independent of receptor activator of nuclear factor kappaB ligand. Blood 2001; 97:3349–53.

- 21. Oba Y, Lee JW, Ehrlich LA, et al. MIP-1∝ utilizes both CCR1 and CCR5 to induce osteoclast formation and increase adhesion of myeloma cells to marrow stromal cells. Exper Hematol **2005**; 33:272–8.
- 22. US Department of Health and Human Services. Drug interactions between CCR5 antagonists and other drugs. In: Guidelines for the use of antiretroviral agents in HIV-1 infected adults and adolescents. Available at: http://aidsinfo.nih.gov/guidelines/html/1/adult-and-adolescent-arvguidelines/288/ccr5-antagonist-drug-interactions. Accessed 27 January 2015.
- 23. Okoli C, Siccardi M, Thomas-William S, et al. Once daily maraviroc 300 mg or 150 mg in combination with DRV/r 800/100 mg. J Antimicrob Chemother **2012**; 67:671–4.
- Abel S, Back DJ, Vourvahis M. Maraviroc: pharmacokinetics and drug interactions. Antivir Ther 2009; 14:607–18.
- 25. Kakuda TN, Abel S, Davis J, et al. Pharmacokinetic interactions of maraviroc with darunavir-ritonavir, etravirine and etravirine-darunavir ritonavir in healthy volunteers: results of two drug interaction trials. Antimicrob Agents Chemother 2011; 55:2290–6.
- McFadyen L, Jacqmin P, Wade J, et al. Maraviroc exposure-efficacy (50 copies/mL) analysis in HIV-1-infected treatment-naive subjects—ITT population (MERIT study) [abstract TUPE0053]. In: Program and abstracts of the XVII International AIDS Conference, Mexico City, 3–8 August 2008.
- Gulick RM, Lalezari J, Goodrich J, et al. Maraviroc for previously treated patients with R5 HIV-1 infection. N Engl J Med 2008; 359:1429–41.
- National Osteoporosis Foundation. Clinician's guide to treatment and prevention of osteoporosis. Available at: http://nof.org/files/nof/public/ content/file/344/upload/159.pdf. Accessed 28 January 2015.
- 29. Sax PE, Saag MS, Yin MT, et al. Renal and bone safety of tenofovir alafenamide vs tenofovir disoproxil fumarate [abstract 143LB]. In: Program and abstracts of the 22nd Conference on Retroviruses and Opportunistic Infections, Seattle, WA, 23–26 February 2015.
- Grigsby IF, Pham L, Mansky LM, Gopalakrishnan R, Mansky KC. Tenofovir-associated bone density loss. Ther Clin Risk Manag 2010; 6:41–7.
- 31. Havens PL, Kiser JJ, Stephensen CB, et al. Association of higher plasma vitamin d binding protein and lower free calcitriol levels with tenofovir disoproxil fumarate use and plasma and intracellular tenofovir pharmacokinetics: cause of a functional vitamin D deficiency? Antimicrob Agents Chemother 2013; 57:5619–28.
- Bianco C, Rossetti B, Gagliardini R, et al. Bone mineral density improvement after 48 weeks of switch to maraviroc+darunavir/ritonavir 300/800/100 mg QD, preliminary results of GUSTA study. J Int AIDS Soc 2014; 17(4 suppl 3):19816.
- Kwan TS, Padrines M, Theoleyre S, Heymann D, Fortun Y. IL-6, RANKL, TNF alpha/IL-1: interrelations in bone resorption pathophysiology. Cytokine Growth Factor Rev 2004; 15:49–60.
- 34. Siggelkow H, Eidner T, Lehmann G, et al. Cytokines, osteoprotegerin, and RANKL in vitro and histomorphometric indices of bone turnover in patients with different bone diseases. J Bone Miner Res 2003; 18:529–38.
- 35. Ding C, Parameswaran V, Udayan R, Burgess J, Jones G. Circulating levels of inflammatory markers predict change in bone mineral density and resorption in older adults: a longitudinal study. J Clin Endocrinol Metab **2008**; 93:1952–8.
- Bedimo RJ, Maalouf NM, Zhang S, Dreschler H, Tebas P. Osteoporotic fracture risk associated with cumulative exposure to tenofovir and other antiretroviral agents. AIDS 2012; 26:825–31.
- 37. Brown TT, Moser C, Currier JS, et al. Bone density changes after antiretroviral initiation with protease inhibitors or raltegravir [abstract 779LB]. In: Program and Abstract of the 21st Conference of Retroviruses and Opportunistic Infections, Boston, MA, 3–7 March 2014.
- Hoetelmans RM, Mariën K, De Pauw M, et al. Pharmacokinetic interaction between TMC 114/ritonavir and tenofovir disoproxil fumarate in healthy volunteers. Br J Clin Pharmacol 2007; 64:655–61.
- Jain RG, Lenhard JM. Select HIV protease inhibitors alter bone and fat metabolism ex vivo. J Biol Chem 2002; 277:19247–50.

- Wang MW, Wei S, Faccio R, et al. The HIV protease inhibitor ritonavir blocks osteoclastogenesis and function by impairing RANKL-induced signaling. J Clin Invest 2004; 114:206–13.
- Cozzolino M, Vidal M, Arcidiacono MV, Tebas P, Yarasheski KE, Dusso AS. HIV-protease inhibitors impair vitamin D bioactivation to 1,25dihydroxyvitamin D. AIDS 2003; 17:513–20.
- 42. Stellbrink HJ, Pulik P, Szlavik J, et al. Maraviroc (MVC) dosed once daily with darunavir/ritonavir (DRV/r) in a 2 drug-regimen compared to emtricitabine/tenofovir (TDF/FTC) with DRV/r; 48-week results from MODERN (Study A4001095) [abstract TUAB0101]. In: Program and abstracts of the 20th International AIDS Conference, Melbourne, Australia, 20–25 July 2014.