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COX-1 (*PTGS1*) and COX-2 (*PTGS2*) polymorphisms, NSAID interactions, and risk of colon and rectal cancer in two independent populations

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Abstract

Purpose—Nonsteroidal anti-inflammatory drugs (NSAIDs) target the prostaglandin H synthase enzymes, cyclooxygenase (COX)-1 and -2, and reduce colorectal cancer risk. Genetic variation in the genes encoding these enzymes may be associated with changes in colon and rectal cancer risk and in NSAID efficacy.

Methods—We genotyped candidate polymorphisms and tagSNPs in *PTGS1* (COX-1) and *PTGS2* (COX-2) in a population-based case-control study (Diet, Activity and Lifestyle Study, DALS) of colon cancer (n=1470 cases/1837 controls) and rectal cancer (n=583/775), and independently among cases and controls from the Colon Cancer Family Registry (CCFR; colon n=959/1535, rectal n=505/839).

Results—In *PTGS2*, a functional polymorphism (−765G>C; rs20417) was associated with a 2-fold increased rectal cancer risk (p=0.05) in the DALS study. This association replicated with a significant nearly 5-fold increased risk of rectal cancer in the CCFR study (OR_{CC vs GG}=4.88; 95%CI=1.54–15.45; OR_{GC vs GG}=1.36; 95%CI: 0.95–1.94). Genotype-NSAID interactions were observed in the DALS study for *PTGS1* and rectal cancer risk, and for *PTGS2* and colon cancer risk, but were no longer significant after correcting for multiple comparisons and did not replicate in the CCFR. No significant associations between *PTGS1* polymorphisms and colon or rectal cancer risk were observed.

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Conclusions—These findings suggest that polymorphisms in *PTGS2* may be associated with rectal cancer risk and impact the protective effects of NSAIDs.

Keywords

colorectal cancer; PTGS; COX; genetic association; NSAID; aspirin; polymorphism

INTRODUCTION

Inflammation is thought to play a major role in the development and progression of colorectal cancer. The use of nonsteroidal anti-inflammatory drugs (NSAIDs) such as aspirin reduces the risk of colorectal cancer (1). NSAIDs inhibit the prostaglandin H synthase (PTGS) enzymes, which convert arachidonic acid into prostaglandins. Several prostaglandins, primarily PGE₂, have been implicated in colorectal carcinogenesis (2). Although both PTGS isoforms, cyclooxygenase (COX)-1 and -2, catalyze the same reactions and share approximately 60% amino acid identity, they are encoded by distinct genes and differ substantially in their expression and regulation (3, 4). COX-1 is constitutively expressed and is important for “housekeeping” functions, whereas COX-2 is typically an inducible enzyme expressed in cells responding to inflammatory or proliferative stimuli (3).

Several lines of evidence indicate that COX-2 facilitates colorectal carcinogenesis. COX-2 is overexpressed in up to 90% of colon carcinomas and 40% of a precursor lesion, colorectal adenoma (5–7). Aspirin decreases the risk of colorectal cancers that express high levels of COX-2 but has little effect on the risk of tumors that have little or no COX-2 (8). Further, in Min mice, selective inhibition of COX-2 or deletion of the *PTGS2* gene results in a substantial reduction in polyp development and tumorigenesis, providing evidence for COX-2 involvement in carcinogenesis that is already at the stage of precursor lesions (9). COX-2 has also been shown to activate co-carcinogens through oxidation [14].

There is also accumulating evidence to support the proposal (10) that COX-1, specifically the platelet enzyme, is involved in colorectal tumorigenesis. First, there is the decreased incidence and mortality of colorectal cancer that are associated with low doses of aspirin (11), doses that selectively and persistently inhibit COX-1 in anucleate platelets (12). Oral low-dose aspirin (80–100 mg) produces a transient pulse of the drug in the blood that peaks at only 1–3 μM, with a $t_{1/2}$ of ~20 min (13, 14). Given that aspirin’s IC₅₀ for human COX-2 is ~15 μM (15, 16), low dose aspirin is likely to give little if any prolonged inhibition of COX-2 activity in nucleated cells, which readily replace any acetylated COX-2 protein. A second observation linking COX-1 to colorectal carcinogenesis is that knockout of the *PTGS1* gene markedly decreases the incidence of polyposis in Min mice (17). Thus, COX-1 and -2 appear to have distinct roles in colorectal carcinogenesis, and polymorphisms in the genes encoding these enzymes (*PTGS1* and *PTGS2*) might plausibly affect cancer risk. We have previously shown that polymorphisms related to prostaglandin synthesis affect the risk of colorectal adenoma and may modify the preventive associations with NSAID use (1, 18–22). In the current analysis, we investigated *PTGS1* and *PTGS2* polymorphisms in relation to the risk of colon and rectal cancer and their potential interactions with NSAID use in a large population-based study of colon and rectal cancer risk, and validated those findings in a second, independent, study. The results indicate that a *PTGS2* functional promoter variant is reproducibly associated with a two-to-four fold increased risk of rectal, but not colon, cancer.

MATERIALS AND METHODS

Study Design and Data Collection

The analyses are based on a case/unrelated-control study of colon and rectal cancer and a population-based case/unaffected-sibling-control study, here restricted to non-Hispanic whites (NHW). Methods, described in detail elsewhere (23–27), are described briefly here. Study population characteristics are in Table 1.

Diet, Activity and Lifestyle Study (DALIS) colon and rectal cancer populations (Discovery Study)—NHW colon cancer cases (n=1470) and controls (n=1837) and rectal cancer cases (n=583) and controls (n=775) were recruited from Utah, the Northern California Kaiser Permanente Medical Care Program (KPMCP), and metropolitan Minneapolis-St. Paul, Minnesota (colon cases only). Eligible participants were aged 30–79 years with no previous diagnosis of colorectal cancer, familial adenomatous polyposis, Crohn’s disease, or ulcerative colitis. Colon cancer cases were diagnosed between 1991 and 1994 (23), and rectal cancer cases between 1997 and 2001 (24, 25), respectively. Diet, physical activity, smoking, anthropometry, medical history, NSAID use, family history of cancer, demographics, race/ethnicity, and reproductive history data were obtained by questionnaire (23, 24, 26, 28–32). The referent period for the study was two years prior to diagnosis for cases and two years prior to selection for controls. NSAID use was defined as aspirin/NSAID use at least three times per week for one month or more. The colon and rectal cancer populations were recruited separately at different time periods, but are collectively referred to in this manuscript as the DALIS study as they were parallel study designs.

Colon Cancer Family Registry study (Validation Study)—Participants were recruited to the Colon Cancer Family Registry (CCFR) from six registry centers: University of Hawaii, Honolulu, Hawaii, USA; Fred Hutchinson Cancer Research Center, Seattle, Washington, USA; Mayo Clinic, University of Southern California Consortium (Dartmouth Medical School, University of Southern California, University of Colorado, University of Arizona, Cleveland Clinic Foundation, University of North Carolina and University of Minnesota); Cancer Care Ontario, Toronto, Ontario, Canada and the University of Melbourne, Victoria, Australia. Both population-based and clinic-based ascertainment strategies were used (27), with some centers recruiting all incident cases from population-based cancer registries (i.e. population-based recruiting), whereas others oversampled cases with a family history of colorectal cancer or cases who were diagnosed at a young age (i.e. family-based recruiting), as described in detail previously (27). The current study includes only population-based participants. All cases were interviewed within five years of diagnosis; 73% of cases were interviewed within two years of diagnosis. Standardized questionnaires were used to collect epidemiologic data from study participants on demographic characteristics, race/ethnicity, medical history, NSAID use, family history of cancer, smoking history, selected diet, physical activity, height and weight, and, in women only, reproductive history and hormone use. “Regular NSAID use” was defined as use of aspirin or ibuprofen at least twice per week for one month or more (27, 33, 34). The CCFR study used a case/unaffected-sibling-control design restricted to NHWs. Analyses of population-based families included 1,464 cases and 2,374 unaffected siblings after exclusion criteria were applied (see below). Cases included probands and affected relatives diagnosed with primary invasive colorectal cancer from 1998–2002. Controls were siblings of cases without a colorectal cancer diagnosis at the time of ascertainment. There were 1534 sibships in our study. Because some sibships have multiple cases and/or controls, the number of sibships can exceed the total number of cases.

Sibships lacking either a case or an unaffected sibling, and cases for whom time-to-interview was more than five years were excluded. Also excluded were individuals whose samples were not available for genotyping, who did not have epidemiologic data, duplicate samples, or who had missing genotypes. Informed consent was obtained from all participants. This study was approved by the Institutional Review Board at each CCFR site.

Genotyping

For the DALS study, a linkage-disequilibrium (LD)-based tagSNP-selection algorithm (35) was used to identify tagSNPs ($r^2=0.90$, $MAF>4\%$) representing common genetic variation in *PTGS1* and *PTGS2* in the CEPH population (Utah residents with ancestry from northern and western Europe) (36). We genotyped 19 polymorphisms in *PTGS1*, including 13 tagSNPs, five candidate SNPs: R8W (rs1236913), P17L (rs3842787), R149L (rs10306140), L237M (rs5789), and R108Q (rs5787); and one deletion polymorphism (L15–L16del) identified previously through sequencing (37). *PTGS2* polymorphisms included 15 tagSNPs and two candidate SNPs from the promoter region: $-765G>C$ (rs20417) and $-163 C>G$ (rs5270). The targeted polymorphisms are shown in Supplementary Table 1. Genotype quality control and exclusion criteria were as described (21). To ensure adequate gene coverage, multiple SNPs were genotyped from LD bins containing a large number of SNPs. After genotyping was completed, redundant SNPs were removed from the analysis based on LD value ($r^2>0.9$) among NHW controls (Supplementary Figure 1). The CCFR study was genotyped for 6 *PTGS1* and 8 *PTGS2* SNPs to provide independent validation of findings from the DALS study. SNPs were chosen for the validation study if preliminary analyses in the discovery dataset resulted in an unadjusted p-value <0.10 . In addition, all candidate SNPs were genotyped in the CCFR study unless they were monomorphic in the DALS study (Supplementary Table 1). The p-value cutoff was determined during the preliminary analyses of the DALS dataset, and chosen to minimize the number of false negative SNPs from DALS while reducing the number of SNPs to be tested in the CCFR.

We used the Illumina™ GoldenGate assay to genotype blood-derived DNA in both the DALS and CCFR studies. *PTGS1* SNPs rs5789 (L237M) and rs1236913 (R8W) were confirmed by Taqman allelic discrimination assay in the DALS colon cancer study. The $-765G>C$ polymorphism in *PTGS2* (rs20417) was genotyped in the CCFR study using a Taqman allelic discrimination assay (18). *PTGS1* rs3842787 (P17L) and the L15–L16del were genotyped by Sanger sequencing in all studies. Two *PTGS1* tagSNPs that failed QC and three candidate SNPs with a $MAF <0.2\%$: R149L (rs10306140), R108Q (rs5787), $-163 C>G$ (rs5270) were excluded from subsequent analysis (Supplementary Table 1).

Statistical analysis

Single SNP Main Effects—Odds ratios (ORs) and 95% confidence intervals (CIs) were calculated in the DALS study using unconditional logistic regression. Because of the case-unaffected sibling control design for the CCFR, conditional logistic regression was used with each sibling set treated as a matched set. All models were restricted to NHWs, as they represented $>90\%$ of all study populations and the tagSNP-selection algorithm used was based on the LD structure of the CEPH population, which has ancestry from northern and western Europe. All models were adjusted for continuous age and sex. DALS models were also adjusted for study site. “Main effects” analyses examined the association between each individual SNP and colon or rectal cancer risk. For each of these analyses, likelihood ratio tests (LRTs) were from a 2 degree of freedom (df) test where genotypes were modeled using indicator variables for the heterozygous and the homozygous variant genotypes (co-dominant models) and from a 1 df test where homozygous variant and heterozygous genotypes were grouped for analysis (dominant models), which was the case only for SNPs where fewer than ten cases or controls had the homozygous variant genotype. If fewer than

five cases or controls had the heterozygous variant genotype, the statistical model was not run. Significance was assessed using LRTs. All tests of statistical significance used a two-sided p-value and $\alpha=0.05$.

Interaction Analyses—Interactions were evaluated by taking the product of indicator variables for NSAID use (current vs. never/former) and for genotypes. For SNP-NSAID interactions a 2 df test was used to evaluate the multiplicative interaction term for co-dominant SNPs and binary NSAID use (current vs. never/former) and a 1 df test was used to evaluate the multiplicative interaction term for dominant SNPs and binary NSAID use. Because use of NSAIDs may be associated with other known risk factors for CRC, NSAID interactions were adjusted for additional variables within each study. DALIS interactions were additionally adjusted for the following continuous variables: BMI, smoking (cigarettes/day), physical activity (hours/week), dietary calcium (mg/day), calories (kilocalories/day), and dietary fiber (g/day). CCFR interactions were additionally adjusted for BMI (continuous), smoking in pack-years (continuous), and physical activity (categorized from average MET hours into inactive, less active, active, and very active). Aspirin use was also investigated independently. To avoid small cell counts, the dominant model was used if there were less than ten homozygous variant cases or controls in either NSAID category. If fewer than five cases or controls had the heterozygous variant genotype in either NSAID category, the statistical model was not run. Significance was assessed using likelihood ratio tests (LRT). Table 3 includes only SNPs genotyped in both studies with a p-interaction less than 0.05 in at least one study prior to any multiple testing corrections. The p-values presented in Tables 2–4 are prior to correction for multiple testing. All analyses were performed using SAS 9.3 or R version 2.13.2.

Multiple Testing Corrections—The DALIS study was treated as the discovery data set and multiple comparison corrected p-values were attained for all polymorphisms using minP permutation tests with 10,000 replications (38). Candidate functional polymorphisms have pre-specified hypotheses to impact cancer risk; therefore multiple testing corrections are not necessarily applicable to those polymorphisms. The CCFR study served as an independent validation data set and was not subject to multiple comparison correction in the primary analyses. A secondary, post hoc multiple testing correction was performed in the CCFR colon cancer study for genotype-NSAID interactions (Table 3).

RESULTS

Genetic associations

Characteristics (age, sex, site, NSAID use) of the DALIS and CCFR study populations are presented in Table 1. In our analysis, the DALIS study served as the discovery dataset, and the CCFR study was an independent validation dataset. Table 2 includes only SNPs that were genotyped in both studies. After correcting for multiple comparisons, there were no statistically significant (minP 0.05) associations between SNPs in *PTGS1* and risk of colon or rectal cancer in the DALIS discovery dataset (Table 2). The rare L15–L16 deletion did show a trend towards increased risk in both the DALIS and CCFR colon and rectal studies, consistent with previous observations for adenoma (19), but this did not reach statistical significance. Post hoc analyses combining colon and rectal cancers within each study also did not reach significance for this polymorphism (data not shown).

In *PTGS2*, we observed a nearly two-fold increase in risk of rectal cancer in the DALIS study for individuals with the rs20417 CC genotype (–765 G>C, $OR_{CC \text{ vs } GG} = 1.95$; 95% CI: 0.89–4.26; LRT $p=0.05$). Although this association was not significant after correcting for multiple testing (minP>0.05), it did replicate in the independent CCFR study population,

with a statistically significant (LRT $p=0.01$) increased risk of rectal cancer for individuals with the GC or CC genotype (Table 2). Individuals with the CC genotype had an almost five-fold increase in rectal cancer risk ($OR_{CC \text{ vs } GG} = 4.88$; 95% CI 1.54–15.45). A comparison of rectal cancer risk for the homozygous variant CC genotype to the GG common genotype resulted in p -value of 0.09 in the DAL5 study, and a p -value of 0.01 in the CCFR study (data not shown). In both study populations, the increased risk was limited to rectal cancer. A polytomous regression model found a significant difference between colon and rectal cancer risk (global $p<0.0001$, data not shown) for this SNP in the DAL5 study. There were no other statistically significant associations between *PTGS2* SNPs and risk of colon or rectal cancer in the DAL5 study.

NSAID Interactions

We observed nominally significant (LRT $p = 0.05$) genotype-NSAID interactions for SNPs in *PTGS1* in the DAL5 discovery study (Table 3). First, the benefit of regular NSAID use for reducing rectal cancer risk was limited to those with the PP (CC) genotype for P17L (rs3842787; LRT $p=0.05$). This is consistent with our previous finding that the benefit of regular NSAID use for reducing adenoma risk was limited to those with the PP genotype (19). Additionally, we observed that NSAID use was of greater benefit for reducing rectal cancer risk among those carrying the variant allele of either rs10306135 (4331 A>T, LRT $p=0.01$) or rs6478565 (15268 A>G, LRT $p=0.03$). There is modest linkage disequilibrium between these two SNPs, which may contribute to the similar findings ($r^2=0.56$, Supplementary Figure 1). These associations were no longer statistically significant after correcting for multiple testing ($\text{min}P>0.05$), and did not replicate in the CCFR independent validation study. No significant genotype-NSAID interactions were observed in *PTGS1* in relation to colon cancer risk. Aspirin use alone also showed no significant interactions with *PTGS1* genotypes for colon or rectal cancer in either study (data not shown).

For *PTGS2*, one significant genotype-NSAID interaction was seen in the DAL5 colon cancer discovery dataset (rs20424; LRT $p=0.01$), but it was no longer significant after correcting for multiple testing ($\text{min}P>0.05$), and did not replicate in the CCFR validation study. No other significant genotype-NSAID interactions were observed in *PTGS2* in the DAL5 study.

Examination of aspirin use alone showed nominally significant associations with three SNPs in *PTGS2* in the DAL5 study (Supplementary Table 3). However, two of the three interactions seen between *PTGS2* genotype and aspirin use did not replicate in the CCFR validation study. A third, rs2745557, had a significant interaction with aspirin use for rectal cancer in the DAL5 study (int $p=0.03$) and for colon cancer in the CCFR study (int $p=0.001$) but there was no association for rectal cancer in DAL5 or colon cancer in the CCFR. In both studies, the variant allele carriers not currently taking aspirin were at increased risk compared to wildtype non-users, but appeared to benefit more from aspirin than wildtype individuals. No observed genotype-NSAID interactions reached statistical significance in both the CCFR and DAL5 populations. Several genotype-NSAID interactions were observed in the CCFR validation study but not in the DAL5 discovery study (Supplementary Table 4) Associations observed in the CCFR validation study but not in the DAL5 discovery study may be due to chance, inadequately controlling for interactions in the DAL5 discovery study in the original analysis, or due to the sib-pair design of the CCFR study (39). To partly address this, we performed an exploratory post hoc stratification of DAL5 by family history, but were unable to replicate associations seen in the CCFR (data not shown). In addition, statistically significant findings were typically limited to either colon or rectal cancer, but not seen in both.

DISCUSSION

We comprehensively assessed the importance of genetic variability in the two primary prostaglandin synthesis genes, *PTGS1* and *PTGS2*, using two independent study populations of colon and rectal cancer risk. We describe a significant association between the rs20417 variant C allele in *PTGS2* and increased risk of rectal cancer in DALIS, a large population-based study, which replicated in a second, independent, large population-based study of rectal cancer from the CCFR. Genotype-NSAID interactions were observed in the DALIS study for *PTGS1* and rectal cancer risk, and for *PTGS2* and colon cancer risk; however, these interactions were no longer statistically significant after correcting for multiple comparisons and did not replicate in the CCFR validation study. Interactions between aspirin use alone and *PTGS2* genotypes were also inconsistent between the DALIS and CCFR studies.

The first report of an association between *PTGS2* rs20417 and risk of colorectal cancer was in a Japanese study (40), but other studies in Caucasian populations did not confirm the finding (41, 42). Recent meta-analyses have indicated that the variant C allele may be a risk factor for colorectal cancer in Asian but not in Caucasian populations (43–47). Importantly, these earlier analyses did not examine colon and rectal cancer separately, and thus it is unknown whether previous studies would have seen an association with rectal cancer risk in Caucasians. In addition, large genome-wide association studies generally have not genotyped rs20417 directly and also have not been stratified by colon and rectal cancer. As our studies were restricted to NHWs, our results suggest that the rs20417 C allele may be a risk factor for this group, but only for rectal cancer and not colon cancer. We have previously reported a possible reduced risk of colorectal adenoma associated with this allele in Caucasians, although sample sizes were too small to distinguish between adenomas in colon and rectal sites (18).

We observed an interaction between this rs20417 SNP, NSAID use, and rectal cancer risk in the CCFR study, where the variant C allele carriers had a greater protective benefit from NSAID use. Observing statistically significant NSAID interactions in the CCFR and not in the DALIS may be due to chance, or may be due to differences in the study designs. As a case/sibling-control study in which shared genetics and environment are matched between siblings, the CCFR study is potentially more efficient for studying gene-environment interactions (39), which could be one reason why this interaction was observed in the CCFR but not in the DALIS study. Alternatively, there could be confounding factors in the DALIS study that were not adequately controlled for in our analysis. We did not observe an association between rs20417 and colon cancer risk in either study. A polytomous regression model indicated that the difference in risk between colon and rectal cancer for rs20417 in the DALIS study was significant, with a global $p < 0.0001$ for both main association and NSAID interactions models (data not shown). In general, we saw little reproducibility in statistically significant genetic associations between colon vs. rectal cancer risk.

These findings add further data to evidence that colon and rectal cancer have different etiologies. In addition, a significant interaction between *PTGS2* rs20417 and NSAID use suggests that, in contrast to colorectal adenoma (18), rs20417 interacts with NSAIDs for rectal cancer risk. The interaction was only seen in the CCFR, so this also could be a chance finding. A study of colorectal cancer from Rotterdam reported that NSAID users carrying the rs20417 C allele lived longer than non-users with the wildtype G allele (48). The Rotterdam study did not see an association between colorectal cancer risk and rs20417 genotype, but they did not analyze colon and rectal cancer separately.

COX-2, encoded by the *PTGS2* gene, catalyzes a key step in the conversion of arachidonic acid to bioactive prostaglandins. The *PTGS2* candidate polymorphism, rs20417 (–765 G>C), is known to affect gene expression and prostaglandin production (49, 50). The functional impact of rs20417 has been studied by several groups; their studies suggest a proinflammatory effect of the CC genotype via increased prostanoids. A more than ten-fold increase in PGE₂ and PGD₂ production was observed in monocytes from asthma patients homozygous for rs20417 CC compared to monocytes from GG homozygotes, with monocytes from heterozygotes displaying an intermediate phenotype of elevated PGE₂ and PGD₂ (50, 51). This is consistent with the observation of increased urinary PGE₂ metabolites and biomarkers of monocyte/macrophage activation in stable coronary artery disease patients with the CC genotype (52). Further, AML patients have been found to have increased *PTGS2* mRNA levels in bone marrow and increased COX-2 protein levels in serum (53). There has been a report of a 30% reduction in gene expression associated with the CC variant in an initial study with a reporter-gene system (49). However, subsequent findings were mixed (52), suggesting that *in vitro* models of promoter activity do not fully capture the complex regulation of *PTGS2* transcription.

We previously reported an increased adenoma risk for carriers of the *PTGS1* L15–L16 deletion (19). The current analyses found similar trends for both colon and rectal cancer. However, the trend is not statistically significant, possibly because this deletion is rare and there is lack of power for validating the association. We also present a replication of our previously reported genotype-NSAID interaction for the *PTGS1* P17L polymorphism (rs3842787). Consistent with our findings in colorectal adenoma (19), we found in the DALS study that the NSAID-associated risk reduction for rectal cancer was limited to the wildtype genotype for P17L. The functional impact of *PTGS1* P17L (rs3842787) may be direct, due to the amino acid change in the signal peptide, or indirect via the near-complete linkage disequilibrium in Caucasians between rs3842787 and seven 5' polymorphisms (54).

In general, statistically significant genotype-NSAID interactions did not replicate between the DALS and CCFR studies. The one significant interaction seen in both studies, between aspirin use and *PTGS2* rs2745557, was inconsistent in that it was seen in rectal cancer in the DALS study and in colon cancer in the CCFR study (Supplementary Table 3). This may be a chance finding or may be due to these studies' somewhat different study designs and definitions of NSAID use, different adjustment variables, limited sample size and a weaker NSAID effect in the CCFR (Table 1). Both DALS and CCFR are large population-based case-control studies of colon and rectal cancer risk. The DALS study uses population-based controls and the CCFR uses unaffected siblings as controls. This is both a strength and a limitation of using the CCFR as a replication dataset for DALS. On one hand, the CCFR sib-pair design helps avoid false positives that may result from population stratification and increases the power to detect gene-NSAID interactions. The CCFR sib-pair design, under which the shared genetics and environment are matched between siblings, can have greater power to detect gene-environment interactions than a case-control study design (39). On the other hand, the family-based study design may have reduced the power of the main effect analyses. We felt that the potential benefit of the CCFR in replicating NSAID interactions outweighed the limitations in the main-effect analysis. The main effects are adjusted for age and sex in both studies. DALS is further adjusted for center, which is not necessary in the CCFR due to the sib-pair design. For the NSAID analysis, both studies were also adjusted for the known CRC risk factors of BMI, physical activity, and smoking. We were not able to adjust the CCFR for dietary risk factors as such data are not available within the CCFR. However, the additional adjustments to the DALS study did not substantially alter the results. Larger-scale investigations are needed to address NSAID, and particularly aspirin, pharmacogenetics with more certainty. Given the new results from randomized-controlled

trials of aspirin, which demonstrated strong cancer preventive effects (1, 11, 55), this issue deserves further attention.

CONCLUSIONS

One polymorphism in the *PTGS2* gene ($-765G>C$; rs20417) was associated with a statistically significantly increased risk of rectal cancer in two large, independent, population-based studies in the US. Our results suggest that the rs20417 C allele may be a risk factor for non-Hispanic whites, but only for rectal cancer, not colon cancer. No significant associations were observed between the targeted *PTGS1* polymorphisms and colon or rectal cancer risk. A number of genotype-NSAID interactions were noted; however, no genotype-NSAID interactions reached statistical significance in both the discovery and validation studies or for both colon and rectal cancer. An interaction between rs2745557 in *PTGS2* and aspirin use was suggestive – showing similar gene-aspirin interaction patterns and reaching significance in the DAL5 rectal cancer study and the CCFR colon cancer study, but not vice versa. These findings suggest that further validation is needed.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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REFERENCES

1. Ulrich CM, Bigler J, Potter JD. Non-steroidal anti-inflammatory drugs for cancer prevention: promise, perils and pharmacogenetics. *Nat Rev Cancer*. 2006; 6:130–140. [PubMed: 16491072]
2. Wang D, Dubois RN. Eicosanoids and cancer. *Nat Rev Cancer*. 2010; 10:181–193. [PubMed: 20168319]
3. Ricciotti E, FitzGerald GA. Prostaglandins and inflammation. *Arterioscler Thromb Vasc Biol*. 2011; 31:986–1000. [PubMed: 21508345]
4. Smith WL, DeWitt DL, Garavito RM. Cyclooxygenases: structural, cellular, and molecular biology. *Annual review of biochemistry*. 2000; 69:145–182.
5. Eberhart CE, Coffey RJ, Radhika A, Giardiello FM, Ferrenbach S, DuBois RN. Up-regulation of cyclooxygenase 2 gene expression in human colorectal adenomas and adenocarcinomas. *Gastroenterology*. 1994; 107:1183–1188. [PubMed: 7926468]
6. Kutchera W, Jones DA, Matsunami N, et al. Prostaglandin H synthase 2 is expressed abnormally in human colon cancer: evidence for a transcriptional effect. *Proc Natl Acad Sci U S A*. 1996; 93:4816–4820. [PubMed: 8643486]

7. Hull MA, Fenwick SW, Chapple KS, Scott N, Toogood GJ, Lodge JP. Cyclooxygenase-2 expression in colorectal cancer liver metastases. *Clin Exp Metastasis*. 2000; 18:21–27. [PubMed: 11206834]
8. Chan AT, Ogino S, Fuchs CS. Aspirin and the risk of colorectal cancer in relation to the expression of COX-2. *N Engl J Med*. 2007; 356:2131–2142. [PubMed: 17522398]
9. Oshima M, Dinchuk JE, Kargman SL, et al. Suppression of intestinal polyposis in Apc delta716 knockout mice by inhibition of cyclooxygenase 2 (COX-2). *Cell*. 1996; 87:803–809. [PubMed: 8945508]
10. Patrono C, Patrignani P, Garcia Rodriguez LA. Cyclooxygenase-selective inhibition of prostanoid formation: transducing biochemical selectivity into clinical read-outs. *J Clin Invest*. 2001; 108:7–13. [PubMed: 11435450]
11. Rothwell PM, Wilson M, Elwin CE, et al. Long-term effect of aspirin on colorectal cancer incidence and mortality: 20-year follow-up of five randomised trials. *Lancet*. 2010; 376:1741–1750. [PubMed: 20970847]
12. Patrignani P, Filabozzi P, Patrono C. Selective cumulative inhibition of platelet thromboxane production by low-dose aspirin in healthy subjects. *J Clin Invest*. 1982; 69:1366–1372. [PubMed: 7045161]
13. Pedersen AK, FitzGerald GA. Preparation and analysis of deuterium-labeled aspirin: application to pharmacokinetic studies. *J Pharm Sci*. 1985; 74:188–192. [PubMed: 3989690]
14. Tsikas D, Tewes KS, Gutzki FM, Schwedhelm E, Greipel J, Frolich JC. Gas chromatographic-tandem mass spectrometric determination of acetylsalicylic acid in human plasma after oral administration of low-dose aspirin and guaiacum. *Journal of chromatography. B, Biomedical sciences and applications*. 1998; 709:79–88.
15. Cromlish WA, Kennedy BP. Selective inhibition of cyclooxygenase-1 and-2 using intact insect cell assays. *Biochem Pharmacol*. 1996; 52:1777–1785. [PubMed: 8986141]
16. Cryer B, Feldman M. Cyclooxygenase-1 and cyclooxygenase-2 selectivity of widely used nonsteroidal anti-inflammatory drugs. *Am J Med*. 1998; 104:413–421. [PubMed: 9626023]
17. Chulada PC, Thompson MB, Mahler JF, et al. Genetic disruption of PtgS-1, as well as PtgS-2, reduces intestinal tumorigenesis in Min mice. *Cancer Res*. 2000; 60:4705–4708. [PubMed: 10987272]
18. Ulrich CM, Whitton J, Yu JH, et al. PTGS2 (COX-2) –765G > C promoter variant reduces risk of colorectal adenoma among nonusers of nonsteroidal anti-inflammatory drugs. *Cancer Epidemiol Biomarkers Prev*. 2005; 14:616–619. [PubMed: 15767339]
19. Ulrich CM, Bigler J, Sparks R, et al. Polymorphisms in PTGS1 (=COX-1) and risk of colorectal polyps. *Cancer Epidemiol Biomarkers Prev*. 2004; 13:889–893. [PubMed: 15159324]
20. Liu W, Poole EM, Ulrich CM, Kulmacz RJ. Polymorphic human prostaglandin H synthase-2 proteins and their interactions with cyclooxygenase substrates and inhibitors. *Pharmacogenomics J*. 2011; 11:337–347. [PubMed: 20548327]
21. Poole EM, Hsu L, Xiao L, et al. Genetic variation in prostaglandin E2 synthesis and signaling, prostaglandin dehydrogenase, and the risk of colorectal adenoma. *Cancer Epidemiol Biomarkers Prev*. 2010; 19:547–557. [PubMed: 20086108]
22. Poole EM, Bigler J, Whitton J, et al. Genetic variability in prostaglandin synthesis, fish intake and risk of colorectal polyps. *Carcinogenesis*. 2007; 28:1259–1263. [PubMed: 17277229]
23. Slattery ML, Potter J, Caan B, et al. Energy balance and colon cancer--beyond physical activity. *Cancer Research*. 1997; 57:75–80. [PubMed: 8988044]
24. Slattery ML, Caan BJ, Benson J, Murtaugh M. Energy balance and rectal cancer: an evaluation of energy intake, energy expenditure, and body mass index. *Nutr Cancer*. 2003; 46:166–171. [PubMed: 14690792]
25. Slattery ML, Edwards SL, Ma KN, Friedman GD, Potter JD. Physical activity and colon cancer: a public health perspective. *Ann Epidemiol*. 1997; 7:137–145. [PubMed: 9099401]
26. Kampman E, Potter JD, Slattery ML, Caan BJ, Edwards S. Hormone replacement therapy, reproductive history, and colon cancer: a multicenter, case-control study in the United States. *Cancer Causes Control*. 1997; 8:146–158. [PubMed: 9134238]

27. Newcomb PA, Baron J, Cotterchio M, et al. Colon Cancer Family Registry: an international resource for studies of the genetic epidemiology of colon cancer. *Cancer Epidemiol Biomarkers Prev.* 2007; 16:2331–2343. [PubMed: 17982118]
28. Murtaugh MA, Ma KN, Benson J, Curtin K, Caan B, Slattery ML. Antioxidants, carotenoids, and risk of rectal cancer. *Am J Epidemiol.* 2004; 159:32–41. [PubMed: 14693657]
29. Slattery ML, Edwards S, Curtin K, et al. Physical activity and colorectal cancer. *Am J Epidemiol.* 2003; 158:214–224. [PubMed: 12882943]
30. Slattery ML, Potter JD, Duncan DM, Berry TD. Dietary fats and colon cancer: assessment of risk associated with specific fatty acids. *Int J Cancer.* 1997; 73:670–677. [PubMed: 9398044]
31. Slattery ML, Levin TR, Ma K, Goldgar D, Holubkov R, Edwards S. Family history and colorectal cancer: predictors of risk. *Cancer Causes Control.* 2003; 14:879–887. [PubMed: 14682445]
32. Friedman GD, Coates AO, Potter JD, Slattery ML. Drugs and colon cancer. *Pharmacoepidemiol Drug Saf.* 1998; 7:99–106. [PubMed: 15073733]
33. Seufert BL, Poole EM, Whitton J, et al. IkappaBKbeta and NFkappaB1, NSAID Use and Risk of Colorectal Cancer in the Colon Cancer Family Registry. *Carcinogenesis.* 2012
34. Coghil AE, Newcomb PA, Campbell PT, et al. Prediagnostic non-steroidal anti-inflammatory drug use and survival after diagnosis of colorectal cancer. *Gut.* 2010
35. Carlson CS, Eberle MA, Rieder MJ, Yi Q, Kruglyak L, Nickerson DA. Selecting a maximally informative set of single-nucleotide polymorphisms for association analyses using linkage disequilibrium. *Am J Hum Genet.* 2004; 74:106–120. [PubMed: 14681826]
36. Thorisson GA, Smith AV, Krishnan L, Stein LD. The International HapMap Project Web site. *Genome Res.* 2005; 15:1592–1593. [PubMed: 16251469]
37. Ulrich CM, Bigler J, Sibert J, et al. Cyclooxygenase 1 (COX1) polymorphisms in African-American and Caucasian populations. *Hum Mutat.* 2002; 20:409–410. [PubMed: 12402351]
38. Dudoit S, van der Laan MJ, Pollard KS. Multiple testing. Part I. Single-step procedures for control of general type I error rates. *Stat Appl Genet Mol Biol.* 2004; 3 Article 13.
39. Witte JS, Gauderman WJ, Thomas DC. Asymptotic bias and efficiency in case-control studies of candidate genes and gene-environment interactions: basic family designs. *Am J Epidemiol.* 1999; 149:693–705. [PubMed: 10206618]
40. Hamajima N, Takezaki T, Matsuo K, et al. Genotype Frequencies of Cyclooxygenase 2 (COX2) Rare Polymorphisms for Japanese with and without Colorectal Cancer. *Asian Pacific journal of cancer prevention : APJCP.* 2001; 2:57–62. [PubMed: 12718655]
41. Iglesias D, Nejda N, Azcoita MM, Schwartz S Jr, Gonzalez-Aguilera JJ, Fernandez-Peralta AM. Effect of COX2 -765G>C and c.3618A>G polymorphisms on the risk and survival of sporadic colorectal cancer. *Cancer Causes Control.* 2009; 20:1421–1429. [PubMed: 19468846]
42. Thompson CL, Plummer SJ, Merkulova A, et al. No association between cyclooxygenase-2 and uridine diphosphate glucuronosyltransferase 1A6 genetic polymorphisms and colon cancer risk. *World J Gastroenterol.* 2009; 15:2240–2244. [PubMed: 19437564]
43. Cao H, Xu Z, Long H, Li XQ, Li SL. The-765C allele of the cyclooxygenase-2 gene as a potential risk factor of colorectal cancer: a meta-analysis. *Tohoku J Exp Med.* 2010; 222:15–21. [PubMed: 20808059]
44. Dong J, Dai J, Zhang M, Hu Z, Shen H. Potentially functional COX-2-1195G>A polymorphism increases the risk of digestive system cancers: a meta-analysis. *J Gastroenterol Hepatol.* 2010; 25:1042–1050. [PubMed: 20594217]
45. Zhu W, Wei BB, Shan X, Liu P. 765G>C and 8473T>C polymorphisms of COX-2 and cancer risk: a meta-analysis based on 33 case-control studies. *Mol Biol Rep.* 2010; 37:277–288. [PubMed: 19669667]
46. Pereira C, Medeiros RM, Dinis-Ribeiro MJ. Cyclooxygenase polymorphisms in gastric and colorectal carcinogenesis: are conclusive results available? *European journal of gastroenterology & hepatology.* 2009; 21:76–91. [PubMed: 19060633]
47. Theodoratou E, Montazeri Z, Hawken S, et al. Systematic meta-analyses and field synopsis of genetic association studies in colorectal cancer. *J Natl Cancer Inst.* 2012; 104:1433–1457. [PubMed: 23019048]

48. Siemes C, Visser LE, Coebergh JW, Hofman A, Uitterlinden AG, Stricker BH. Protective effect of NSAIDs on cancer and influence of COX-2 C(-765G) genotype. *Curr Cancer Drug Targets*. 2008; 8:753–764. [PubMed: 19075598]
49. Papafili A, Hill MR, Brull DJ, et al. Common promoter variant in cyclooxygenase-2 represses gene expression: evidence of role in acute-phase inflammatory response.[comment]. *Arteriosclerosis, Thrombosis & Vascular Biology*. 2002; 22:1631–1636.
50. Sanak M, Szczeklik W, Szczeklik A. Association of COX-2 gene haplotypes with prostaglandins production in bronchial asthma. *J Allergy Clin Immunol*. 2005; 116:221–223. [PubMed: 15990799]
51. Szczeklik W, Sanak M, Szczeklik A. Functional effects and gender association of COX-2 gene polymorphism G-765C in bronchial asthma. *J Allergy Clin Immunol*. 2004; 114:248–253. [PubMed: 15316498]
52. Sanak M, Plutecka H, Szczeklik W, Piwowarska W, Rostoff P, Szczeklik A. Functional promoter polymorphism of cyclooxygenase-2 modulates the inflammatory response in stable coronary heart disease. *Polskie Archiwum Medycyny Wewnętrznej*. 2010; 120:82–88. [PubMed: 20332714]
53. Zheng J, Chen S, Jiang L, You Y, Wu D, Zhou Y. Functional Genetic Variations of Cyclooxygenase-2 and Susceptibility to Acute Myeloid Leukemia in a Chinese Population. *Eur J Haematol*. 2011
54. Lee CR, Bottone FG Jr, Krahn JM, et al. Identification and functional characterization of polymorphisms in human cyclooxygenase-1 (PTGS1). *Pharmacogenet Genomics*. 2007; 17:145–160. [PubMed: 17301694]
55. Cross JT, Poole EM, Ulrich CM. A review of gene-drug interactions for nonsteroidal anti-inflammatory drug use in preventing colorectal neoplasia. *Pharmacogenomics J*. 2008; 8:237–247. [PubMed: 18195728]

^b Current NSAID use is defined as current, regular use three times per week for at least one month in the DALIS study and as current, regular use of at least two pills per week for at least one month for the CCFR study.

^c NA – this was a matching factor.

Table 2

Association between selected *PTGS1* and *PTGS2* polymorphisms and risk of colon and rectal cancer.^a

	Colon Cancer						Rectal Cancer									
	Controls			Cases			Controls			Cases						
	N	%		N	%		N	%		N	%					
			OR	95% CI	p ^d				OR	95% CI	p ^d					
<i>PTGS1</i> Candidate polymorphisms																
rs1236913 (R8W C>T)																
DALS ^b	1608	87.6	1273	86.6	ref	-	-	0.35	668	87.3	491	85.7	ref	-	-	0.34
	227	12.4	197	13.4	1.10	0.90	1.35		97	12.7	82	14.3	1.17	0.85	1.60	
CCFR ^c	1306	87.4	808	87.1	ref	-	-	0.91	720	87.3	414	86.6	ref	-	-	0.28
	188	12.6	120	12.9	1.02	0.72	1.44		105	12.7	64	13.4	1.32	0.79	2.18	
L15-L16 deletion																
DALS ^b	1770	98.9	1411	98.7	ref	-	-	0.63	752	98.3	559	97.9	ref	-	-	0.55
	19	1.1	18	1.3	1.17	0.61	2.25		13	1.7	12	2.1	1.28	0.58	2.83	
CCFR ^c	1478	98.9	918	98.9	ref	-	-	0.21	816	98.9	472	98.7	ref	-	-	0.22
	16	1.1	10	1.1	2.19	0.61	7.83		9	1.1	6	1.3	2.89	0.51	16.48	
rs3842787 (P17L C>T)																
DALS ^b	1546	86.6	1231	86.2	ref	-	-	0.94	668	87.2	490	85.4	ref	-	-	0.35
	228	12.8	187	13.1	1.04	0.84	1.28		98	12.8	84	14.7	1.16	0.85	1.59	
CCFR ^c	1308	87.6	792	85.3	ref	-	-	0.09	724	87.8	413	86.4	ref	-	-	0.89
	176	12.4	136	14.6	1.41	0.95	2.09		101	12.2	65	13.6	1.04	0.62	1.73	
rs5789 (L237M C>A)																
DALS ^b	1738	94.7	1394	94.8	ref	-	-	0.82	732	94.6	551	94.5	ref	-	-	0.97
	97	5.3	76	5.2	0.96	0.71	1.32		42	5.5	32	5.5	1.01	0.63	1.62	

	Colon Cancer										Rectal Cancer											
	Controls					Cases					Controls					Cases						
	N	%	N	%	OR	95% CI	p ^d	N	%	N	%	OR	95% CI	p ^d	N	%	N	%	OR	95% CI	p ^d	
CCFR ^c	C/C	1410	93.9	883	94.4	ref	-	0.17	776	94.4	456	93.4	ref	-	0.35							
	C/A+A/A	92	6.1	52	5.6	0.72	0.44	1.15	46	5.6	32	6.6	1.42	0.67	3.00							
PTGS1 tag SNPs																						
rs10306135 (4331 A>T)																						
DALS ^b	A/A	1295	73.5	1009	71.7	ref	-	0.55	577	74.5	432	74.4	ref	-	0.91							
	A/T	428	24.3	367	26.1	1.09	0.93	1.29	197	25.4	149	25.6	1.01	0.79	1.30							
	T/T	40	2.3	32	2.3	1.02	0.64	1.64	*	*	*	*	*	*								
CCFR ^c	A/A	1113	74.1	700	74.4	ref	-	0.42	595	72.3	355	72.4	ref	-	0.27							
	A/T	352	23.4	223	23.7	0.95	0.73	1.24	208	25.3	121	24.7	1.03	0.72	1.48							
	T/T	38	2.5	18	1.9	0.61	0.29	1.29	20	2.4	14	2.9	2.04	0.84	4.94							
rs6478565 (15268 A>G)																						
DALS ^b	A/A	1197	67.9	972	68.8	ref	-	0.77	536	69.2	397	68.2	ref	-	0.18							
	A/G	507	28.8	398	28.2	0.96	0.82	1.12	209	27.0	172	29.6	1.11	0.88	1.42							
	G/G	58	3.3	42	3.0	0.89	0.59	1.34	30	3.9	13	2.2	0.60	0.31	1.17							
CCFR ^c	A/A	1073	69.9	664	69.2	ref	-	0.77	580	69.1	343	67.9	ref	-	0.30							
	A/G	418	27.2	272	28.4	0.97	0.76	1.23	233	27.8	143	28.3	1.07	0.76	1.52							
	G/G	44	2.9	23	2.4	0.79	0.42	1.48	26	3.1	19	3.8	1.82	0.85	3.93							
PTGS2 candidate SNP																						
rs20417 (-765 G>C)																						
DALS ^b	G/G	1232	69.6	979	68.9	ref	-	0.95	553	71.4	433	74.3	ref	-	0.05							
	G/C	495	28.0	404	28.5	1.02	0.87	1.20	211	27.2	134	23.0	0.81	0.63	1.04							
	C/C	44	2.5	37	2.6	1.04	0.67	1.63	11	1.4	16	2.7	1.95	0.89	4.26							
CCFR ^c	G/G	1059	71.0	648	69.8	ref	-	0.67	576	70.4	314	65.4	ref	-	0.01							

	Colon Cancer										Rectal Cancer											
	Controls					Cases					Controls					Cases						
	N	%	N	%	OR	95% CI	p ^d	N	%	N	%	OR	95% CI	p ^d	N	%	N	%	OR	95% CI	p ^d	
<i>PTGS2</i> tag SNPs																						
rs4648250 (-1740 A>G)																						
DALS ^b	A/A	1715	97.9	1376	98.6	ref	-	0.09	748	98.6	575	99.7	-	-	ND							
	A/G	37	2.1	19	1.4	0.63	0.36	1.10	11	1.4	2	0.3	-	-								
CCFR ^c	A/A	1343	99.2	841	98.6	ref	-	0.13	726	99.5	427	99.1	-	-	ND							
	A/G	11	0.8	12	1.4	2.32	0.75	7.15	4	0.5	4	0.9	-	-								
rs689466 (-1195 A>G)																						
DALS ^b	A/A	1198	67.5	910	64.0	ref	-	0.15	509	65.7	376	64.7	ref	-	0.87							
	A/G	509	28.7	455	32.0	1.16	1.00	1.36	237	30.6	185	31.8	1.06	0.83	1.33							
	G/G	67	3.8	57	4.0	1.10	0.76	1.58	29	3.7	20	3.4	0.94	0.52	1.68							
CCFR ^c	A/A	958	63.2	619	65.9	ref	-	0.38	558	67.5	338	68.0	ref	-	0.42							
	A/G	496	32.7	287	30.6	0.83	0.64	1.08	249	30.1	138	27.8	0.91	0.63	1.31							
	G/G	63	4.2	33	3.5	0.84	0.49	1.44	20	2.4	21	4.2	1.46	0.66	3.25							
rs20424 (-196 C>G)																						
DALS ^b	C/C	* 1726	97.2	1379	97.2	ref	-	0.91	748	96.6	556	95.4	ref	-	0.22							
	C/G+G/G	49	2.8	40	2.8	1.02	0.67	1.57	26	3.3	27	4.6	1.41	0.81	2.45							
	G/G	1	0.1	0	0.0				1	0.1	0	0.0										
CCFR ^c	C/C	1327	97.1	831	96.5	ref	-	0.03	720	97.0	424	97.0	ref	-	0.24							
	C/G+G/G	40	3.0	30	3.5	2.44	1.05	5.67	22	3.0	13	3.0	0.56	0.21	1.48							
rs2745557 (201 C>A)																						
DALS ^b	G/G	1233	69.6	981	69.5	ref	-	0.98	534	68.9	392	67.5	ref	-	0.45							

	Colon Cancer										Rectal Cancer										
	Controls					Cases					Controls					Cases					
	N	%	N	%	OR	95% CI	p ^d	N	%	N	%	OR	95% CI	p ^d	N	%	N	%	OR	95% CI	p ^d
rs4648261 (418 G>A)																					
G/A	492	27.8	394	27.9	1.01	0.86	1.18	214	27.6	174	29.9	1.10	0.87	1.40							
A/A	47	2.7	36	2.6	0.96	0.62	1.50	27	3.5	15	2.6	0.75	0.39	1.42							
CCFR ^c	1048	68.4	649	67.7	ref	-	-	556	66.3	322	63.9	ref	-	-	0.41	0.19					
G/A	431	28.1	281	29.3	1.10	0.86	1.42	258	30.8	162	32.1	1.28	0.90	1.84							
A/A	54	3.5	29	3.0	0.78	0.42	1.44	25	3.0	20	4.0	1.88	0.86	4.11							
rs4648268 (2284 G>A)																					
DALS ^b	1664	94.0	1332	93.9	ref	-	-	0.90	732	94.6	546	94.1	ref	-	-	0.70					
G/A+A/A	107	6.1	86	6.1	1.02	0.76	1.37	42	5.4	34	5.9	1.10	0.69	1.75							
CCFR ^c	1432	93.3	904	94.3	ref	-	-	0.04	769	91.7	478	94.7	ref	-	-	0.07					
G/A+A/A	103	6.7	55	5.7	0.58	0.35	0.98	70	8.3	27	5.3	0.52	0.25	1.08							
rs4648268 (2284 G>A)																					
DALS ^b	1443	81.2	1148	80.7	ref	-	-	0.14	626	80.8	462	79.2	ref	-	-	0.47					
G/A	322	18.1	253	17.8	0.99	0.82	1.19	149	19.2	121	20.7	1.10	0.84	1.44							
A/A	13	0.7	21	1.5	1.98	0.99	3.98	*	*	*	*	*	*	*							
CCFR ^c	1051	79.0	690	81.2	ref	-	-	0.01	581	81.0	344	80.4	ref	-	-	0.41					
G/A	269	20.2	149	17.5	0.61	0.43	0.86	136	19.0	84	19.7	1.22	0.76	1.95							
A/A	11	0.8	11	1.3	1.39	0.50	3.87	*	*	*	*	*	*	*							
rs5275 (6364 T>C)																					
DALS ^b	725	40.8	599	42.2	ref	-	-	0.20	342	44.2	252	43.2	ref	-	-	0.91					
T/C	805	45.4	655	46.1	0.99	0.86	1.16	344	44.4	265	45.5	1.05	0.84	1.32							
C/C	245	13.8	166	11.7	0.82	0.66	1.03	88	11.4	66	11.3	1.02	0.71	1.47							
CCFR ^c	588	43.4	370	43.2	ref	-	-	0.71	299	41.0	182	41.9	ref	-	-	0.97					
T/C	601	44.3	378	44.1	0.90	0.69	1.18	339	46.5	204	47.0	1.01	0.70	1.46							
C/C	167	12.3	109	12.7	0.98	0.64	1.51	91	12.5	48	11.1	1.08	0.59	1.97							

* Dominant model.

^a Only SNPs genotyped in both DAL5 and CCFR are shown. Additional SNPs genotyped in DAL5 are in Supplementary Table 2. The dominant model was used when <10 cases or controls had the homozygous variant genotype, modeling was not run when <5 cases or controls were heterozygotes.

^b Adjusted for age, sex, and center.

^c Adjusted for age and sex.

^d Global p-value from a likelihood ratio test prior to correction for multiple comparisons. No SNPs remained significant in the DAL5 studies after correcting for multiple comparisons using minP permutation tests (minP 0.05).

Table 3
Interactions between selected *PTGS1* and *PTGS2* SNPs, NSAID use, and risk of colon and rectal cancer.^a

NSAID Use: ^d	Colon Cancer										Rectal Cancer										
	Never/Former					Current					Never/Former					Current					
	Controls	Cases	OR	95% CI	p ^e	Controls	Cases	OR	95% CI	p ^e	Controls	Cases	OR	95% CI	p ^e	Controls	Cases	OR	95% CI	p ^e	
<i>PTGS1</i>																					
rs3842787 (P17L S0 C>T)																					
DALS ^b	C/C	914	824	ref	-	623	396	0.65	0.55	0.77	0.40	350	310	ref	-	314	178	0.61	0.48	0.78	0.05
	C/T+T/T	128	132	1.16	0.89	1.51	63	0.63	0.45	0.87	0.87	57	44	0.88	0.57	1.34	41	37	1.03	0.64	1.66
CCFR ^c	C/C	969	605	ref	-	327	182	0.83	0.64	1.07	0.26	570	335	ref	-	149	78	0.86	0.57	1.28	0.46
	C/T+T/T	147	104	1.34	0.85	2.11	32	1.66	0.83	3.32	3.32	76	52	1.20	0.66	2.19	23	12	0.71	0.28	1.78
rs10306135 (4331 A>T)																					
DALS ^b	A/A	723	662	ref	-	565	337	0.62	0.52	0.73	0.65	318	256	ref	-	255	174	0.80	0.62	1.04	0.01
	A/T	266	247	1.03	0.84	1.27	160	0.75	0.57	0.98	0.98	93	101	1.33	0.96	1.85	104	47	0.55	0.38	0.82
	T/T	25	22	1.00	0.55	1.82	15	0.69	0.30	1.56	*	*	*	*	*	*	*	*	*	*	*
CCFR ^c	A/A	822	542	ref	-	279	157	0.82	0.62	1.07	0.69	467	290	ref	-	123	64	0.68	0.43	1.06	0.21
	A/T+T/T	302	182	0.85	0.62	1.18	86	0.78	0.49	1.23	1.23	182	112	0.86	0.58	1.29	44	23	0.97	0.50	1.88
rs6478565 (15268 A>G)																					
DALS ^b	A/A	663	634	ref	-	526	330	0.61	0.51	0.73	0.88	294	236	ref	-	238	160	0.80	0.61	1.04	0.03
	A/G	309	272	0.92	0.75	1.12	197	0.61	0.47	0.79	0.79	118	123	1.31	0.96	1.79	121	60	0.60	0.42	0.87
	G/G	36	28	0.88	0.53	1.48	22	0.58	0.29	1.17	1.17	*	*	*	*	*	*	*	*	*	*
CCFR ^c	A/A	801	514	ref	-	260	149	0.86	0.65	1.14	0.89	456	281	ref	-	120	61	0.75	0.48	1.16	0.61
	A/G+G/G	347	222	0.85	0.63	1.14	113	0.71	0.46	1.08	1.08	203	131	1.00	0.68	1.48	53	31	0.91	0.48	1.75
<i>PTGS2</i>																					
rs20424 (-62 C>G)																					
DALS ^b	C/C	986	918	ref	-	731	448	0.61	0.53	0.72	0.01	397	342	ref	-	347	211	0.68	0.54	0.85	0.97
	C/G+G/G	34	22	0.65	0.37	1.14	15	1.24	0.61	2.49	2.49	15	17	1.35	0.66	2.76	11	10	0.94	0.39	2.26

NSAID Use: ^d	Colon Cancer						Rectal Cancer												
	Never/Former			Current			Never/Former			Current									
	Controls	Cases	OR	95% CI	Controls	Cases	OR	95% CI	p ^e	Controls	Cases	OR	95% CI	p ^e					
CCFR ^c	992	637	ref	-	323	191	0.90	0.69	1.17	0.31	560	338	-	-	154	85	-	-	ND
	25	22	2.75	1.07	7.04	8	1.22	0.30	4.90		17	11	-	-	5	2	-	-	

* Dominant model.

^a Only SNPs with an interaction p-value <0.05 in the DALSS study are shown. The dominant model was used when <10 cases or controls had the homozygous variant genotype, modeling was not run if <5 cases or controls were heterozygotes.

^b Adjusted for age, sex, center, BMI, smoking, physical activity, calcium, calories, and dietary fiber.

^c Adjusted for age, sex, BMI, smoking and physical activity.

^d Current NSAID use is defined as current, regular use three times per week for at least one month in the DALSS study and as current, regular use of at least two pills per week for at least one month for the CCFR study.

^e Interaction p-value from a likelihood ratio test prior to correction for multiple comparisons. No SNPs remained significant in the DALSS study after correcting for multiple comparisons using minP permutation tests (minP 0.05).