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Prevalence of human papillomavirus among oesophageal squamous cell carcinoma cases: systematic review and meta-analysis

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Background: Oncogenic human papillomavirus (HPV) has been hypothesised as a risk factor for oesophageal squamous cell carcinoma (OSCC), but aetiological research has been limited by the varying methodology used for establishing HPV prevalence. The aims of this systematic review and meta-analysis were to estimate the prevalence of HPV DNA detected in OSCC tumours and the influence of study characteristics.

Methods: Study-level estimates of overall and type-specific HPV prevalence were meta-analysed to obtain random-effects summary estimates.

Results: This analysis included 124 studies with a total of 13 832 OSCC cases. The average HPV prevalence (95% confidence interval) among OSCC cases was 0.277 (0.234, 0.320) by polymerase chain reaction; 0.243 (0.159, 0.326) by *in situ* hybridisation; 0.304 (0.185, 0.423) by immunohistochemistry; 0.322 (0.154, 0.490) by L1 serology; and 0.176 (0.061, 0.292) by Southern/slot/dot blot. The highest HPV prevalence was found in Africa and Asia, notably among Chinese studies from provinces with high OSCC incidence rates.

Conclusions: Future research should focus on quantifying HPV in OSCC cases using strict quality control measures, as well as determining the association between HPV and OSCC incidence by conducting large, population-based case-control studies. Such studies will provide a richer understanding of the role of HPV in OSCC aetiology.

Oesophageal cancer is the eighth most common cancer worldwide, with an estimated 481 000 annual incident cases, and is the sixth most common cause of cancer mortality, with an estimated 406 000 annual deaths (Ferlay *et al*, 2010). Incidence rates of oesophageal cancer vary greatly by geographic region (Corley and Buffler, 2001; Ferlay *et al*, 2010), with markedly high rates in the 'Asian Oesophageal Cancer Belt', which extends from northern Iran, east to China and north into Russia (Glenn, 2001; Kamangar *et al*, 2007; Wei *et al*, 2011). Reasons for the geographic variability are not well understood (Hongo *et al*, 2009).

Squamous cell carcinoma is the most common form of oesophageal cancer (Corley and Buffler, 2001). Epidemiological studies conducted in the Western countries, Asia, and Africa have concluded that tobacco and alcohol consumption are the main known risk factors for OSCC (IARC, 1986; IARC, 1988; Munoz and Castellsague, 1994), accounting for approximately 90% of OSCC in men and 29% in women (Castellsague *et al*, 1999). Yet, it has been suggested that alcohol and tobacco consumption are not the major risk factors for OSCC in one of the highest risk areas of northern China (Linxian County) (Yu *et al*, 1993). Additional

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established risk factors include poverty and nutritional deficiencies (Hongo *et al*, 2009; Mao *et al*, 2011).

Infection with oncogenic HPV as a contributor to OSCC was hypothesised over three decades ago (Syrjanen, 1982). However, the International Agency for Cancer Research (IARC) has concluded that there is inadequate evidence in humans for HPV carcinogenicity in association with OSCC (IARC, 2012).

The aims of this systematic review and meta-analysis were to estimate the prevalence of HPV detected in OSCC cases and the influence of type of HPV detection methods and other study characteristics on this measure.

MATERIALS AND METHODS

Search strategy and inclusion criteria. A systematic literature search was conducted using MEDLINE (via PubMed) and Excerpta Medica database (EMBASE) through 12 July 2013, with no specified start date or language restrictions. This systematic review was conducted in accordance with PRISMA guidelines (Moher *et al*, 2009).

Broad search terms included: HPV, oesophagus, and cancer (e.g., neoplasm, carcinoma, tumour) (specific search terms available upon request). Eligible studies included peer-reviewed publications with HPV prevalence data from a minimum of 20 adult human cases of primary, invasive, and oesophageal squamous cell carcinoma (OSCC). Review articles and editorials were included if they contained original data. Abstracts were excluded. Primary and secondary references were searched to identify additional studies. A medical reference librarian verified the search strategy, and independently repeated the search. If multiple publications presented data from the same population, only the publication with the largest sample size was included. Articles written in languages other than English were translated and reviewed. Because articles were excluded based on multiple criteria, we assigned a primary exclusion criterion based on the following rank-order: study not conducted in humans (i.e., *in vitro* or animal study), not oesophageal cancer, histology other than squamous cell carcinoma, not primary tumour, not invasive cancer (i.e., *in situ* only), study population included only children (i.e., <18 years old), letter to the editor or review (without original data), case report, abstract, histology unidentified, <20 cases, and no quantification of HPV.

Data extraction. For studies meeting our inclusion criteria, the following data were extracted: first author, publication year, study location, dates of sample collection, sample size, specimen type, HPV detection method, and overall and type-specific prevalence of HPV. Continent, country, and region were recorded. For studies with samples from multiple countries, data were extracted separately by country when possible.

Studies were categorised into five geographical areas: Africa, Asia, Europe and Australia, North America, or South and Central America. In addition, we analysed separately the studies performed in Iran and the high-OSCC-incidence province-level divisions of China (i.e., provinces with higher than the overall crude incidence of oesophageal cancer for China of 22.1/100 000) (Chen *et al*, 2013). These included Fujian, Gansu, Hebei, Henan, Jiangsu, Shandong, Shanxi, and Sichuan provinces, as well as Xinjiang Uyghar Autonomous Region.

Specimen type was classified as paraffin-embedded fixed biopsies, fresh or frozen biopsies, or blood samples. HPV detection method was classified as polymerase chain reaction (PCR), *in situ* hybridisation (ISH), immunohistochemistry, serology, and Southern blot (including dot and slot blot). For studies that used multiple HPV detection methods, overall and type-specific prevalence of HPV were extracted independently. For studies that utilised PCR, information on specific HPV primers used was also

extracted. For studies that reported use of multiple PCR primers, only the three most sensitive primers were recorded. The following rank order of sensitivity (from most to least sensitive), based on a meta-analysis of invasive cervical cancer, was assumed: SPF10, GP5+/6+, L1C1/2, MY09/11, PU1M/2R, GP5/6, L1, E6, E7, HPV-16 specific, and HPV-18 specific (Clifford *et al*, 2003). HPV-type-specific estimates were only recorded if the authors reported which HPV types were tested. For studies that utilised serology, data on HPV prevalence were extracted separately by serological marker (i.e., L1 vs E6/E7).

Data extraction was performed for each article by two of five independent reviewers (AMB, ABW, CC, JLP, XS) using a standardized data extraction form, and discrepancies were resolved by consensus. Study authors were contacted to obtain specific needed information. This occurred when the reviewers were unable to determine 1) HPV prevalence by detection method or 2) HPV prevalence by HPV subtype.

Statistical analysis. For each study, overall HPV prevalence estimates were calculated as the percentage of OSCC cases who tested positive for any HPV type. In some studies, the total number of available OSCC cases differed from the number of OSCC cases actually tested for HPV (e.g., cases were not tested if they were beta-globin-negative). Additionally, if one HPV detection method was utilised as a screening method for another, we did not include the results of the secondary screening method in our analyses. Information on all HPV-based studies is presented in Supplementary Table 1. The number of OSCC cases tested was used as the denominator. For studies reporting type-specific HPV prevalence, estimates were calculated as percentages of OSCC cases (same denominator as for overall prevalence) that tested positive for a single type of HPV. Infections for multiple types of HPV were separated into types; thus, type-specific prevalence represents types for tumours with either single or multiple infections. Data on overall and type-specific HPV prevalence are presented for each study in the appendices by HPV detection method. We calculated the variance of each prevalence estimate as pq/n , where p is the prevalence, q is $1-p$, and n is the number of OSCC cases (Barendregt *et al*, 2013).

For studies reporting HPV prevalence equal to zero we used an empirical continuity correction method described by Sweeting *et al* (2004) to smooth the zero values. Specifically, we estimated the pooled prevalence, by detection method, for studies with non-zero prevalence estimates. These estimates divided by 100 were then used as the number of HPV-positive OSCC cases, and 1 minus the value added to the HPV-positive OSCC cases was added to the number of HPV-negative OSCC cases. A similar approach was used to smooth HPV type-specific estimates in studies that explicitly reported zero OSCC cases that were positive for a particular HPV type (e.g., 0 OSCC cases who were HPV-16 positive). If studies did not explicitly report type-specific HPV prevalence, this information was considered missing and not smoothed.

Two studies (Ma *et al*, 2000; Sitas *et al*, 2012) provided only type-specific HPV prevalence estimates, and overall HPV prevalence estimates could not be determined (i.e., unable to reconcile single vs multiple type infections). In such instances, the overall HPV prevalence was considered to be missing and was not smoothed but these studies were included in the type-specific analyses.

For the analyses, we used three models: (1) crude model, (2) univariable model, and (3) multivariable model. The crude model was run by HPV detection type for overall prevalence of HPV (Supplementary Table 2). The univariable model was used to explore the heterogeneity of overall random-effects estimates (Table 2). Both overall (Table 2) and prevalence estimates stratified by HPV detection type (Supplementary Table 2) were considered

from models that included HPV prevalence and disjoint indicator variables for a single study characteristic that included: geographic location, publication date, DNA source, and primer set (for PCR only). For example, running an overall HPV prevalence model for HPV detection type would include HPV prevalence and a disjoint indicator variable for the HPV detection types (i.e., PCR, ISH, immunohistochemistry, serology, and Southern blot) but no other covariates. Median values of HPV prevalence were also calculated (Supplementary Table 2). The multivariable model of HPV prevalence was adjusted for all study characteristics (i.e., HPV detection method, continent, publication date, and study size) (Table 2). For all models, study-level estimates of overall and type-specific HPV prevalence were meta-analysed using the *metareg* command in STATA to obtain random-effects summary estimates (DerSimonian and Laird, 1986; Thompson and Sharp, 1999).

In the crude models, we also estimated the among-population variance (τ^2). Tau-squared is the variance of the presumptively normal distribution of true values among populations in which each population has its own true value (i.e., its own true HPV prevalence). We utilised τ^2 to calculate a 95% population effects interval (PEI) within which 95% of the populations' true values are estimated to lie (i.e., mean prevalence $\pm 1.96\tau$) (Mosteller and Colditz, 1996; Ludema *et al*, 2011). To calculate the PEI, we used a logit transformation (Barendregt *et al*, 2013).

Two sensitivity analyses were performed to assess the robustness of the results. First, the analysis was performed excluding the PCR studies with beta-globin-negative cases (i.e., the studies where the total number of available OSCC cases differed from the number of OSCC cases actually tested for HPV) to assess the overall influence of these data on the estimates. Second, summary estimates of overall HPV prevalence were calculated for the serology studies, stratified by L1 and E6/E7 antigen status.

Meta-analyses were performed using STATA version 12 for Windows (StataCorp LP, College Station, TX, USA). Ancillary analyses were performed using SAS version 9.2 (SAS Institute, Inc., Cary, NC, USA).

RESULTS

Systematic review and study characteristics. Our systematic search of PubMed and EMBASE resulted in 724 unique papers. Of these, 124 (17.1%) studies were eligible and included in this systematic review and meta-analysis. Of the remaining 600 publications, 585 (97.5%) were excluded based on various exclusion criteria, 14 (2.3%) were duplicate study populations, and 1 (0.2%) was ineligible after contacting the author. The primary reasons for exclusion included the following: the outcome studied not being oesophageal cancer ($n=185$ publications) and the paper being a review article or letter to the editor without original data ($n=137$ publications) (Figure 1).

Individual characteristics of the included 124 studies are summarised in Supplementary Table 1. Study sample sizes ranged from 20 to 1561 OSCC cases (median=60). Summing across studies, a total of 14 047 OSCC cases were identified. However, as 215 case samples did not have sufficient DNA for analysis, a total of 13 832 OSCC cases were included in the analysis. As reported in Table 1, the majority of studies were conducted in Asian countries ($n=77$, 62.1%), with the remaining studies spanning five other continents as follows: 20 (16.1%) studies in Europe, 9 (7.3%) studies in Africa, 8 (6.5%) studies in North America, 4 (3.2%) studies in South America, 1 (0.8%) study in Australia, and 5 (4.0%) studies included populations from more than one continent. The majority (70.9%) of the Chinese studies were from the high-OSCC-incidence provinces of China. For multi-country studies, information was extracted separately by country for five multi-continent studies (Togawa *et al*, 1994; Suzuk *et al*, 1996; Wang *et al*, 2010; Castillo *et al*, 2011; Sitas *et al*, 2012) and for five single-continent studies (Mori *et al*, 1989; Turner *et al*, 1997; Kawaguchi *et al*, 2000; Castillo *et al*, 2006; Goto *et al*, 2011).

The majority of studies ($n=90$, 72.6%) used only one laboratory detection method to detect HPV, whereas 29 (23.4%) studies used two detection methods and 5 (4.0%) studies

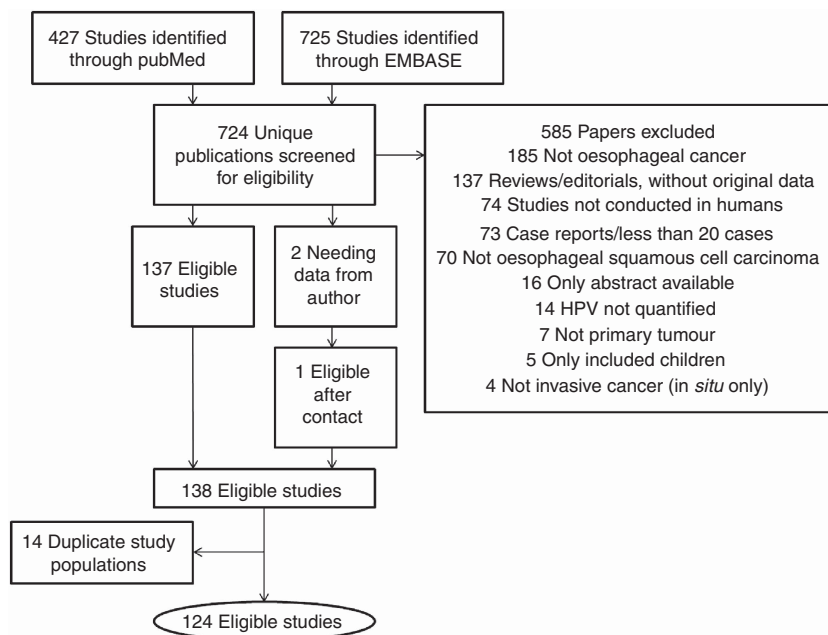


Figure 1. Flow diagram of inclusion and exclusion of identified studies for the meta-analysis of overall prevalence of human papillomavirus (HPV) among oesophageal squamous cell carcinoma (OSCC) cases. Although many articles were excluded based on multiple criteria, we assigned primary exclusion criteria according to the following rank order: study not conducted in humans (i.e., *in vitro* or animal study), not oesophageal cancer, not squamous cell carcinoma (histology known), not primary tumour, not invasive (i.e., *in situ* only), study population children only (i.e., <18 years old), letter to the editor or review (without original data), case report, abstract, histology unidentified, <20 cases, and no quantification of HPV.

used three detection methods. The total number of estimates was 174 summing the across-study populations and detection methods. The majority of the estimates ($n = 109$, 62.6%) were based on PCR methods (Table 2).

Meta-analysis of HPV prevalence in OSCC cases. The pooled prevalence estimates for studies with non-zero prevalence estimates, stratified by detection method, were 0.325 by PCR; 0.312 by ISH; 0.359 by immunohistochemistry; 0.321 by L1 serology; and 0.201

Table 1. Selected characteristics of 124 studies included in the meta-analysis of overall prevalence of human papillomavirus (HPV) among oesophageal squamous cell carcinoma (OSCC) cases

Continent	Studies, N	OSCC Cases, N ^a	Countries represented
Africa	9	1235	Egypt, Kenya, South Africa
Asia	77	9861	China ^b , India, Iran, Japan, Pakistan, South Korea, Taiwan, Turkey
Australia	1	433	Australia
Europe	20	1275	Belgium, Czech Republic, Finland, France, Germany, Greece, Hungary, Italy, Netherlands, Norway, Poland, Romania, Russia, Slovenia, Sweden, United Kingdom
North America	8	280	Canada, Mexico, United States
South America	4	748	Brazil, Chile, Colombia
Multi-continent	5	—	
Total	124	13 832	

^aFor the five multi-continent studies, data extraction was performed separately by country (Togawa *et al*, 1994; Suzuk *et al*, 1996; Wang *et al*, 2010; Castillo *et al*, 2011; Sitas *et al*, 2012). Therefore, the OSCC cases from these studies were reported by continent.

^bOf 55 studies performed in China, 39 (70.9%) were performed in high OSCC incidence provinces of China.

Table 2. Multiple meta-regression of overall prevalence^a of human papillomavirus (HPV) among oesophageal squamous cell carcinoma (OSCC) cases

Characteristic ^b	Studies N	Estimate N	OSCC cases N	Univariable meta-regression model ^c	Multivariable meta-regression model ^d
					Prevalence (95% CI)
Intercept ^e					0.403 (0.337, 0.470)
HPV detection method				Prevalence (95% CI)	Prevalence Difference (95% CI)
PCR	95	109	9257	0.277 (0.234, 0.320)	REF
<i>In situ</i> hybridisation	25	28	2631	0.243 (0.159, 0.326)	-0.034 (-0.126, 0.059)
Immunohistochemistry	7	7	773	0.304 (0.185, 0.423)	0.067 (-0.191, 0.058)
Serology	14	14	862	0.322 (0.154, 0.490)	0.112 (-0.054, 0.277)
Southern blot	11	16	556	0.176 (0.061, 0.292)	-0.067 (-0.191, 0.058)
Continent					
Asia	84	116	10 985	0.314 (0.274, 0.354)	REF
Europe/Australia	22	27	1410	0.156 (0.073, 0.238)	-0.180 (-0.277, -0.084)
North America	8	11	350	0.166 (0.036, 0.296)	-0.166 (-0.304, -0.027)
South America	5	8	664	0.194 (0.042, 0.346)	-0.169 (-0.327, -0.011)
Africa	10	12	670	0.198 (0.075, 0.322)	-0.113 (-0.244, 0.018)
Publication date					
2000 to present	83	105	10 621	0.310 (0.267, 0.352)	REF
1990–1999	36	65	3322	0.204 (0.149, 0.259)	-0.093 (-0.168, -0.017)
Prior to 1990	3	4	136	0.103 (-0.114, 0.321)	-0.282 (-0.536, -0.029)
Study size					
≤60 OSCC cases	65	98	3410	0.272 (0.226, 0.318)	REF
>60 OSCC cases	63	76	10 669	0.261 (0.211, 0.311)	-0.092 (-0.162, -0.023)

^aTwo studies that did not report overall HPV were excluded (Ma *et al*, 2000; Sitas *et al*, 2012).

^bCategories are exclusive for estimates but not for studies because some studies presented results for multiple detection methods.

^cUnivariable meta-regression of a single study characteristic, where the HPV prevalence estimates are not controlled for other study characteristics. For example, the prevalence of PCR is adjusted for other HPV detection types (i.e., *in situ* hybridisation, immunohistochemistry, serology and Southern blot) but no other study characteristics (i.e., continent, publication date, or study size).

^dMultivariable meta-regression, where the HPV prevalence estimates are controlled for all other study characteristics in the table.

^eIntercept: Asian PCR studies conducted 2000 to present with ≤60 OSCC cases.

by Southern and other blotting techniques. We utilised the unrounded values of these prevalence estimates to smooth values for studies with zero HPV-positive cases.

The overall random-effects summary estimates from the crude model for HPV were similar across detection methods. The average prevalence of HPV (95% confidence interval (CI)) was 0.278 (95% CI: 0.233, 0.322) by PCR; 0.242 (95% CI: 0.157, 0.328) by ISH; 0.305 (95% CI: 0.160, 0.450) by immunohistochemistry; 0.321 (95% CI: 0.156, 0.486) by L1 serology; and 0.166 (95% CI: 0.097, 0.234) by Southern blot (Supplementary Table 2). Utilising the logit transformation, the average prevalence of HPV (95% population effects interval (PEI)) was 0.292 (95% PEI: 0.039, 0.809) by PCR; 0.280 (95% PEI: 0.045, 0.763) by ISH; 0.329 (95% PEI: 0.050, 0.821) by immunohistochemistry; 0.302 (95% PEI: 0.075, 0.698) by L1 serology; and 0.213 (95% PEI: 0.084, 0.442) by Southern blot (data not shown in tables). In general, the median per cent positive estimates were similar to the summary medians. The median prevalence of overall HPV positivity was 0.211 by PCR; 0.221 by ISH; 0.232 by immunohistochemistry; 0.275 by L1 serology; and 0.180 by Southern blot (Supplementary Table 2).

In the univariable model, the mean HPV prevalence was 0.277 (95% CI: 0.234, 0.320) for PCR, 0.314 (95% CI: 0.274, 0.354) for Asia, 0.310 (95% CI: 0.267, 0.352) for publications after 1999, and 0.272 (95% CI: 0.226, 0.318) for studies with ≤ 60 cases. In the multivariable meta-regression analysis, the mean HPV prevalence for the intercept (Asian PCR studies conducted after 1999 with ≤ 60 cases) was 0.403 (95% CI: 0.337, 0.470). By detection method, estimates for L1 serology were the most different from PCR, with a prevalence difference of 0.112 higher than the intercept. Among continents, Europe/Australia was the most different from Asia, with a prevalence difference of -0.180 . Studies conducted prior to 1990 were the most different from studies conducted from 2000 to present, with a prevalence difference of -0.282 . Finally, studies with >60 cases had a prevalence difference of -0.092 compared with studies with ≤ 60 cases (Table 2).

The range for HPV prevalence was 0–84.1% for PCR; 0–72.3% for ISH; 0–73.3% for immunohistochemistry; 3.2–58.5% for serology; and 0–45.0% for Southern/slot/dot blot (Supplementary Table 1). The prevalence of high-risk types of HPV, namely HPV-16 and -18, was fairly similar across PCR, ISH, and Southern blot, but differed slightly for serology and immunohistochemistry (Supplementary Table 2). HPV-16 prevalence was 0.185 (95% CI: 0.142, 0.228) for PCR; 0.255 (95% CI: 0.114, 0.397) for ISH; 0.359 (95% CI: -0.101 , 0.819) for immunohistochemistry; 0.173 (95% CI: 0.087, 0.259) for L1 serology; and 0.187 (95% CI: 0.116, 0.259) for Southern blot. Median prevalence estimates for HPV-16 were 0.132 for PCR; 0.190 for ISH; 0.430 for immunohistochemistry; 0.084 for L1 serology; and 0.198 for Southern blot.

Results stratified by detection method for HPV prevalence according to continent, publication year, DNA source, and primer set (for PCR only) are provided in Supplementary Table 2. As with the meta-regression, striking differences were noted for study location across HPV detection methods. Trends across publication year, tissue source, and primer set were noted but were not as consistent across detection methods (Supplementary Table 2).

We also examined within-study comparison for studies that had both PCR and ISH ($n = 11$), PCR and Southern blot ($n = 9$), PCR and immunohistochemistry ($n = 3$), ISH and immunohistochemistry ($n = 4$), ISH and Southern blot ($n = 3$), and HPV 16 and 18 ($n = 62$). The results from these analyses were similar to our overall results. For example, the within-study meta-analysis for ISH compared with PCR resulted in a prevalence difference of -0.038 (95% CI: -0.024 , 0.164), based on a mean prevalence of 0.202 (95% CI: 0.059, 0.345) for PCR (data not shown in tables).

In our sensitivity analyses, results were similar when excluding the PCR studies with beta-globin-negative cases. When the PCR

analysis is restricted to only studies with all beta-globin-positive cases, the overall HPV prevalence is 0.288 (95% CI: 0.238, 0.338). However, results were dissimilar for serology studies when stratified by L1 or E6/E7 antigen status. For example, the overall HPV prevalence for E6/E7 antigen status studies was 0.100 (95% CI: -0.777 , 0.978). These results were based on only two studies that utilised E6/E7 antigen status (Dai *et al*, 2007; Guo *et al*, 2012). Therefore, only L1 serology results are presented in the tables.

DISCUSSION

This systematic review and meta-analysis of the literature on HPV prevalence combined data on 13 832 OSCC cases from 124 studies. In our study, we utilised multivariable meta-regression methods to analyse HPV prevalence in OSCC tumours. We summarised the influence of several important factors on HPV prevalence in OSCC cases, including HPV detection method, geographic location, publication date, and study size.

The prevalence of overall HPV varied widely across detection methods, ranging from 17.6% for Southern blot to 32.2% for L1 serology. Our assessment of heterogeneity suggested meaningful variation in HPV prevalence across HPV detection methods. The estimates of overall HPV prevalence from our meta-analysis are consistent with a recent meta-analysis by Syrjanen, which reported an overall summary HPV prevalence of 29.0% (95% CI: 25.1–33.1%), after combining data on histological biopsies, dot blot hybridisation, filter ISH, hybrid capture 2 assay, ISH, PCR, and Southern blot hybridisation (Syrjanen, 2013). In contrast, we have only presented stratum-specific estimates by detection method in order to elucidate the heterogeneity in HPV prevalence estimates. Each detection method has strengths and limitations with regard to measurement of possible infection at different time points, technical requirements, and test characteristics. For instance, PCR, Southern blot, and ISH have high specificity, ISH has low sensitivity, and PCR has a high rate of false positives (Kimple *et al*, 2012).

Further, it is uninformative to estimate an overall (i.e., worldwide) HPV prevalence, as it is a function of the number of HPV types that are tested and a host of other methodologic determinants as well as highly variable characteristics of person, place, and time. Testing for additional HPV types could partially account for the increasing prevalence across time. However, this seems unlikely as HPV-16 is the predominant type detected and is nearly always tested for as a high-risk HPV type.

The observed differences in HPV prevalence by geographic location warrant further attention. Most notably, our study uniquely reports estimates for overall HPV prevalence in Chinese regions characterised by the high incidence of OSCC. Our estimates of HPV prevalence in these Chinese regions, ranging from 26.4% (95% CI: 10.8%, 42.0%) for Southern blot to 41.6% (95% CI: 33.1%, 50.2%) for PCR, are substantially higher than any continent-specific estimates, including other regions of Asia. Interestingly, all of our detection method-specific estimates for high-risk provinces of China are lower than the estimate of 41.8% (95% CI: 34.3%, 49.7%) reported by Syrjanen (2013) for Chinese studies, which includes both high- and low-risk Chinese regions. This is possible due to the inclusion of additional detection types not included in our analysis (e.g., histological biopsies, hybrid capture 2 assay). In a continent-specific analysis stratified by the HPV detection method, we found that HPV prevalence was highest in Asia and Africa and similar across all other continents. These results are consistent with (1) the estimates reported by Syrjanen that show high prevalence in China and South Africa (Syrjanen, 2013) and (2) a systematic review by Liyanage *et al* (2013) that reported higher levels of HPV detection in regions with high OSCC risk.

Although the geographic differences in oral HPV prevalence are notable (Kreimer *et al*, 2010), HPV cannot be expected to fully account for the geographical variation seen in OSCC incidence (Ferlay *et al*, 2013). Even when an infectious agent participates in the aetiology of every case of a disease (e.g., *tubercle bacillus* and tuberculosis, *Plasmodium* protozoa and malaria, or HIV and AIDS), aetiologic co-factors help determine the disease's geographic variation.

Further, these findings need to be cautiously interpreted, as they do not account for the background prevalence of HPV. Future studies need to account for the background prevalence of HPV, which varies widely by region. A recent meta-analysis examined the prevalence of oral HPV, ranging from approximately 2.6% in Canada to 18.9% in Brazil (Kreimer *et al*, 2010). Additionally, this review cannot account for other established risk factors, including tobacco, alcohol, poverty, and nutritional deficiencies (IARC, 1986; IARC, 1988; Munoz and Castellsague, 1994; Hongo *et al*, 2009; Mao *et al*, 2011).

HPV-16 was the predominant type of HPV assayed and identified across detection methods, with the exception of L1 serology. In stratified analyses, HPV-16 prevalence ranged from 17.3% for L1 serology to 35.9% for immunohistochemistry. HPV-18 was the predominate type of HPV identified for L1 serology, with a prevalence of 26.0%. HPV-18 prevalence ranged from 0% for immunohistochemistry to 26.0% for L1 serology. Our results for PCR were consistent with a recent meta-analysis by Yong *et al* (2013), which reported a HPV-16 prevalence of 11.7% (95% CI: 7.7%, 16.2%) and a HPV-18 prevalence of 1.8% (95% CI: 0.9%, 3.0%) in OSCC tumours detected with PCR alone. Unlike previous studies, our analysis also summarises the differences in HPV type-specific prevalence across multiple detection methods.

Characterising the prevalence of HPV in OSCC cases is an important preliminary step to assessing the association between HPV and OSCC. The prevalence of HPV among OSCC cases in this meta-analysis is lower than the prevalence of HPV previously observed for other cancers with more established epidemiologic evidence for HPV involvement, such as cervical, anal, oropharyngeal, or tonsillar (IARC, 2012). Nonetheless, the HPV prevalence in OSCC cases is high enough, and correlated strongly enough with OSCC incidence ecologically, suggesting that an aetiological connection might be present between the virus and the cancer. By way of analogy, we note that the prevalence of cigarette smoking is lower in bladder cancer cases than in cases of lung cancer. A causal connection between smoking and bladder cancer was established nonetheless. The requisite research took longer, largely because smoking's association with bladder cancer, though causal, was weaker than its association with lung cancer.

An association between HPV and OSCC is biologically plausible on the basis of (1) the close proximity and histologic similarities between oesophageal and oral squamous epithelium – suggesting that the oesophagus could be exposed to HPV in a similar fashion as the oral cavity and oropharynx (Gillison and Shah, 2003); (2) the detection of HPV DNA in both benign oesophageal squamous cell papillomas and malignant OSCC tumour samples (Syrjanen, 2002); (3) *in vivo* studies in cattle, showing that bovine papillomavirus leads to the development of papillomas, then to squamous cell carcinoma (Borzacchiello and Roperto, 2008); and (4) *in vitro* experiments of human oesophageal epithelial cells where HPV-18 was shown to be an initiating factor in cancer transformation (Shen *et al*, 2000). However, counterarguments have been suggested, including the lack of association between OSCC and immunosuppression or previous HPV-associated disease (Gillison and Shah, 2003) and wide variability in case-series report of HPV positivity in OSCC cases (Syrjanen, 2002) – ranging worldwide from over 70% positivity (Liu *et al*, 2000; Cao *et al*, 2005; Zhu *et al*, 2005; Yang *et al*, 2008; Ding *et al*, 2009; Qu *et al*, 2012) to 0% (Hale *et al*, 1989; Loke *et al*, 1990; Benamouzig *et al*, 1995; Smits *et al*,

1995; Saegusa, 1997; Morgan *et al*, 1997; Kamath *et al*, 2000; Talamini *et al*, 2000; Awerkiew *et al*, 2003; White *et al*, 2005; Koh *et al*, 2008; Bellizzi *et al*, 2009; Song *et al*, 2009; Patel *et al*, 2011; Noori *et al*, 2012; Haeri *et al*, 2013). Even within countries, the prevalence of HPV varies widely. For example, in one Chinese study, HPV positivity in OSCC tumours has been reported to range from 2.0% in Linxian province to 22.2% in Shantou province (Si *et al*, 2003). This variation is unlikely due to differences in the study methodology. The large variation in HPV DNA prevalence estimates in OSCC tumours across geographical sites has not been fully explained, but it could possibly be due to small sample sizes, differing HPV detection methods and PCR primers, cut points utilised for HPV positivity, variability between laboratories, and contamination of samples (Hubbard, 2003; Koshiol *et al*, 2010).

Contamination of samples is of paramount concern, as studies have reported that dehydrated HPV maintains 100% infectivity for as long as a day (Roden *et al*, 1997). Additional studies have found HPV DNA on fomites and various medical surfaces (Ferenczy *et al*, 1989; Strauss *et al*, 2002). This is problematic because HPV DNA is resistant to cleaning detergents (Strauss *et al*, 2003), which leads to concerns that fomites could be responsible for contaminating samples (Roden *et al*, 1997). Although we attempted to evaluate quality control measures, this was difficult as many studies do not report on these issues, and we were concerned about biasing our results by including only studies that report on quality control measures. Future studies should strive to carefully avoid contamination and to report quality control measures.

Limitations of the current meta-analysis should be considered. Bias could arise from the study estimates because the accuracy of these estimates depends on the detection method utilised and the types of HPV assessed. That is, some studies utilised broad primers or multiple probes to detect multiple types of HPV, whereas other studies only detected a specific HPV type (e.g., HPV-16). Additionally, there is potential publication bias. This potential is seen in the multivariable meta-regression, which suggests that smaller studies may have higher reported prevalence. Finally, we used a continent-level variable to categorise studies, in addition to high-risk China and Iran. This classification may have masked important variability between studies on the same continent.

The studies included in this meta-analysis are heterogeneous. We directly examined heterogeneity by describing the prevalence of HPV in OSCC cases by HPV type, continent, publication date, DNA source, and primer set. Of course, we were not fully able to explain the heterogeneity. Prevalence estimates remained heterogeneous even in stratified results (e.g., PCR studies conducted in Asia). This is to be expected, as a full explanation of heterogeneity would require knowledge of every determinant of HPV prevalence in OSCC cases, as well as every determinant of HPV measurement validity and reliability, and a sufficient number of studies to explore possible combinations of all such variables. Nonetheless, we identified several variables that account for substantial portions of the heterogeneity.

Strengths of the current meta-analysis include large overall sample size, use of multivariable meta-regression to assess study characteristics, examination of type-specific HPV prevalence, inclusion of studies published in all languages, and exclusion of case reports or studies with small sample size. The use of multivariable meta-regression allowed us to determine study characteristics that influence heterogeneity of HPV prevalence estimates. By examining type-specific HPV prevalence, we were able to determine what is the predominate type of HPV detected in OSCC cases. However, we are still dependent on the HPV types that are included in the primer sets, which are largely optimised for the detection of HPV genital types. By including studies of all languages, we avoided selection bias due to publication language. Finally, by excluding case reports and studies with <20 cases, we

attempted to exclude studies that were a convenience sample or not representative of all OSCC cases.

Although a high proportion of OSCC cases with prevalent HPV does not determine aetiologic significance, the data presented in this paper strongly indicate that such a hypothesis warrants further investigation. Similar to that which has been done for head and neck cancer (Agrawal *et al*, 2011), advances in aetiologic understanding of the role of HPV in OSCC tumour development will come from exome sequencing, which will ideally be conducted in a large number of tumours from differing geographic locals and differing tumour sites.

In summary, our results suggest that HPV prevalence in OSCC tumours is highest in Africa and Asia, especially in the regions of China with a high incidence of OSCC, and that high-risk HPV-16 is the most commonly identified HPV type in OSCC tumours. Our meta-analysis demonstrates that HPV detection method, continent, publication date, and study size contribute to the variability among individual study findings. Utilising multivariable meta-regression, this systematic review highlights study characteristics that need to be considered when examining the association between HPV infection and OSCC. Although this review is a preliminary step in assessing the relationship between HPV and OSCC, it may also be used to better tailor monitoring and public health interventions (e.g., HPV vaccinations) if that relationship becomes established as causal. Future research should focus on quantifying HPV in OSCC cases using strict quality control measures to avoid contamination, as well as determining the association between HPV and OSCC incidence by conducting large, population-based case-control studies. Such studies will provide a richer understanding of the role of HPV in OSCC aetiology.

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