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The *ERLIN1-CHUK-CWF19L1* gene cluster influences liver fat deposition and hepatic inflammation in the NHLBI Family Heart Study

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Abstract

Objectives—Nonalcoholic fatty liver disease (NAFLD) ranges from simple steatosis to hepatic inflammation to cirrhosis. We sought to identify common genetic variants contributing to NAFLD, using CT measured fatty liver (FL), and alanine aminotransferase levels (ALT), as a biochemical marker of hepatic inflammation.

Methods—We employed a correlated meta-analysis (CMA) to test whether combining FL and ALT genomewide association (GWA) results, using ~2.5 million imputed SNPs, could enhance ability to detect variants influencing both traits.

Results—Variants of the *ERLIN1-CHUK-CWF19L1* gene cluster were associated with concomitant variation of FL and ALT. Nine variants (rs2862954, rs1408579, rs10883451, rs11597086, rs11591741, rs17729876, rs17668255, rs17668357, rs12784396) displayed genomewide significant associations at loci concomitantly influencing FL and ALT $(2.47 \times 10^{-9} \text{ CMA-p } 4.29 \times 10^{-10})$ as compared with the suggestive significance of marginal tests $(4.11 \times 10^{-5} \text{ GWA-p } 2.34 \times 10^{-6})$. For example, the missense variant in *ERLIN1*-rs2862954 was genomewide significant (CMA-p=4.88 \times 10^{-10}) for the combination of FL and ALT, while the respective univariate associations were suggestive (FL:p= 5.74×10^{-6} , ALT:p= 3.71×10^{-6}). Further we investigated whether the concomitant associations diminished but did not vanish (CMA-p 3.3×10^{-7}). Our findings suggest *ERLIN1-CHUK-CWF19L1* variants are associated with early stage of FL accumulation (measured by CT) to hepatic inflammation (ALT levels), and the association enhances when accounting for the correlations between their scans.

Conclusions—CMA approach enhanced the ability to identify novel variants of the *ERLIN1-CHUK-CWF19L1* influencing both simple steatosis and hepatic steatosis with inflammation,

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None.

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which suggest that this gene cluster may regulate the susceptibility of NAFLD in a wide spectrum of disease.

1. Introduction

Nonalcoholic fatty liver disease (NAFLD) is an emerging metabolic-related disorder characterized by hepatic fat accumulation, predominantly in the form of triglycerides (TG), in the absence of significant ethanol consumption or other known causes of liver disease [1, 2]. NAFLD is a common disease and ranges from simple steatosis, which is associated with metabolic abnormalities, to nonalcoholic steatohepatitis (NASH), which presents as inflammatory damage in the liver and can progress ultimately to cirrhosis and liver cancer [1].

Computed tomography (CT) imaging of the liver is a non-invasive procedure which allows for an accurate and reliable assessment of fat content in the hepatic parenchyma. There is an inverse relationship between liver fat burden and liver attenuation, and it has a sensitivity of 82% and 100% specificity for detecting >30% steatosis [3, 4]. Alanine transaminase (ALT) is a liver-derived enzyme, and elevated serum levels of ALT are associated with liver damage. ALT has been used as a biochemical marker of hepatic inflammation to monitor the course and severity of NAFLD [5, 6], although it is a non-specific marker of liver damage.

The heritabilities of these traits have been estimated as 25% – 30% for CT measured fatty liver (FL [7]), and 33% – 55% for serum ALT levels [8, 9]. To date, a limited number of genes has demonstrated associations with susceptibility to FL, including *PNPLA3, NCAN, PPP1R3B, GCKR, and LYPLAL1,* and explain less than 5% of the proportion of variance in FL [7, 10–13]. Also, association of *CPN1-ERLIN1-CHUK* with ALT levels was reported [8]. There is a significant correlation between these traits of NAFLD. The correlation between FL an ALT was reported as 0.405 [14], suggesting common genetic effect. Simultaneous consideration of FL and ALT could enhance the identification of etiologic factors, especially genes. The goal of this study was to search for genetic variants that account for some of the correlated architecture between CT measured FL and levels of ALT by combining the genomewide association scans for these phenotypes in a correlated meta-analysis (CMA) using 2,705 European American participants of the NHLBI Family Heart Study (FamHS).

2. Material and Methods

2.1. Study Design and Sample

The FamHS (https://dsgweb.wustl.edu/fhscc/) recruited 1,200 families, half randomly sampled, and half selected because of an excess of coronary heart disease or related-risk factor abnormalities as compared with age- and sex-specific population rates [15] from four population-based parent studies: the Framingham Heart Study, the Utah Family Tree Study, and two centers for the Atherosclerosis Risk in Communities study (Minneapolis, and Forsyth County, NC). Study participants belonging to the largest pedigrees were invited to undergo CT imaging for assessment of coronary artery calcification; these scans also were used to quantitate hepatic and abdominal fat burden. A total of 2,767 Caucasian subjects in 510 extended families were examined.

2.2. Phenotypes

Participants underwent a cardiac multidetector CT exam with four detectors using a standardized protocol as described previously [16]. In brief, two cardiac gated images of the thorax included the upper third of the liver. The unenhanced CT images were reviewed on a GE Advantage Windows Workstation (General Electric Healthcare, Waukesha, WI). Liver

attenuation (LA) was measured by placing three round regions of interest (ROI's) in two slices of the superior right lobe of the liver. The six ROI's values (three from each slice) were then averaged as a mean LA. All participants were imaged with standardized phantom that contains material that simulates water (HU=0) as well as increasing densities of calcium (50, 100 and 200 mg of calcium). The phantom measures at water density were used to adjust and standardize LA measurements. Lower LA values indicate higher fat content. We inverted the scale of LA by multiplying each value by negative one to produce a phenotype of fatty liver (FL) for which higher values are associated with greater liver fat. ALT (in units of U/L) was measured by reaction rate assay based on the conversion of NADH to NAD using the Vitros analyzer (Johnson & Johnson Clinical Diagnostics, Inc. Rochester NY 14650).

Subjects were excluded from the analyses if they were: (i) heavy drinkers (men: >21 drinks/ week and women: >14 drink drinks/week) or (ii) taking Amiodarone, or (iii) positive for hepatitis C antibodies for ALT analysis (88 subjects for FL and 124 subjects for ALT excluded). Alcohol intake was assessed during interview. The subjects reported whether he/ she "presently drinks alcoholic beverage at all (alcohol intake: 0 = No, 1 =Yes)" and if so, the number of 1.5 oz cocktails, 12 oz glasses (or can) of beer, and 4 oz glasses of wine they consumed per day in 1 week. Total number of alcohol intake in grams per day was calculated from the reported intakes of beer, wine, and spirits. Medications were assessed at the clinic visit in which subjects brought all medications they were taking. Amiodarone has the effect of increasing liver density, and thus we excluded subjects taking this medication. Finally, information of hepatitis C antibodies was obtained from questionnaire.

2.3. Genotypes

A total of 2,705 subjects for the analysis were typed on three Illumina platforms: (i) 974 subjects with 547,353 SNP markers on the Illumina HumMap 550K chip, (ii) 249 subjects with 576,888 SNP markers on the Human 610-Quadv1 Illumina chip, and (iii) 1,482 subjects with 1,111,639 SNP markers on the Human 1M-Duov3 Illumina chip [18]. Quality control was performed before imputation by checking pedigree relationships (using GRR software), and excluding SNPs with low call rates (< 98%). A framework map for imputation was defined by choosing SNPs on all platforms, and restricting to those with minor allele frequency between 1% and 50%, and excluding SNPs with significant (p< 1.0×10^{-6}) departure from Hardy-Weinberg equilibrium. This panel was used to impute ~ 2.5 M SNPs based on phased haplotypes from HapMap (release 22, build 36, CEU, http:// hapmap.ncbi.nlm.nih.gov/downloads/phasing/2007-08_rel22/phased/) using MACH (version 1.0.16, http://www.sph.umich.edu/csg/abecasis/mach/). The final data for the GWA analysis included 2,584 participants for ALT levels and 2,597 participants for CT measured FL.

2.4. Statistical Analysis

2.4.1. Covariate adjustment—Principal components (PC [19]) were estimated using the genotypes of the largest sample of independent subjects (N= 753) and then calculated for family members. The phenotypes were adjusted using two steps: (1) forcing field centers and genotype chip effects into the regression analysis, and then (2) adding age, age^2 , sex, current alcohol intake (Yes/No), phantom, and ten PC using stepwise regression analysis and retaining terms significant at the 5% level. The residuals were standardized to a mean of 0 and variance of 1 (FL and ALT). Finally, an inverse normal rank transformed within sex was performed for FL. In the second adjustment for FL, ALT was forced in the covariate list (in step 1); while FL was forced in the covariate list for ALT (in step 1), similarly to the first adjustment procedure.

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2.4.2. Heritability estimates—The heritabilities were calculated using a variance components model as implemented in the computer program SOLAR (http://txbiomed.org/ departments/genetics/genetics-detail?P=37).

2.4.3. Association analysis-GWA analysis was first carried out for each of ALT and FL assuming additive effects with mixed model linear regression, accounting for dependency among family members as a function of their kinship correlations (R kinship package: http://cran.r-project.org/web/packages/GWAF/). Results were filtered to include SNPs with acceptable imputation quality ($r^{2}hat > 0.3$) and with effect allele frequency between 1-99%. To test whether individual variants influence both FL and ALT, we employed a novel correlated meta-analysis (CMA) approach [20], which combines the SNPwise Z-scores corresponding to the p-values, while accounting for the correlation between scans. In brief, the standard meta-analysis assumes all component scans are independent; however, correlations among scans can arise for a number of reasons, here, mainly because the same set of subjects was used for both phenotypes and subjects' phenotypes are correlated. Province and Borecki [20, 21] developed a method to empirically estimate the covariance among scans to be included in a meta-analysis, and to correct the signal inference in the combined meta p-value. Briefly, all p-values are first converted to corresponding zscores, which are assumed to follow a multivariate normal distribution. Since the vast majority of SNPs in a genome scan is under the null for any given phenotype, the global degree of correlation between the scans can be calculated to assess the degree of nonindependence. A tetrachoric correlation estimator is used on p-values dichotomized as $(P \ 0.5; P>0.5)$ to minimize the effect of the few SNPs that might be under the alternative. The estimated correlation is used to adjust the variance estimate of the combined Z-scores.

2.4.3. Regional plot—It was generated with LocusZoom [22], and Haploview (http:// haploview.software.informer.com/) was used for investigation of linkage disequilibrium (LD) and block structure using the HapMap CEU data.

3. Results

3.1. Characteristics of the Study Sample

Table 1 shows the characteristics of the FamHS participants included in the analysis, in which 56% of subjects were women. The average age was 57 years, and average body mass index (BMI) was 28.9 kg/m². The proportion of subjects with severe obesity (BMI 35.0 (kg/m²) was 12.5% and with diabetes was 13.4%. The mean levels of TG (144.1 \pm 93.9 mg/dL), ALT (21.7 \pm 15.8 U/L [6]) and LA (59.1 \pm 11.2 HU [3]) were within normal range values. The phenotypic correlation between FL and ALT levels was 0.27 (p<0.0001). The heritability estimates were significant (p < 0.0001) for the covariate-adjusted phenotypes, FL (0.27 \pm 0.04) and ALT (0.26 \pm 0.04), suggesting that genetic factors influence the variability levels of these traits and a search for significant genetic variants is warranted.

3.2. Genomewide association analyses

Univariate GWA analysis revealed associated variants near or in *PNPLA3* and *PPP1R3B* at genomewide significance levels ($p < 5.0 \times 10^{-8}$, Supplemental Table 1) in FamHS. The highest association was with a missense locus in *PNPLA3* (rs738409: $\beta = 0.34$ for the G allele, $P = 2.09 \times 10^{-23}$), which was previously reported [7] and also detected for magnetic resonance assessed steatosis [10]. The rs2126259 near-*PPP1R3B* associated with FL ($\beta = -0.26$ for the A allele, $P = 3.28 \times 10^{-8}$) is in complete linkage disequilibrium (LD) with rs4240624 (LD=1, $r^2 = 0.80$, $P = 4.76 \times 10^{-8}$), which was also previously reported associated with FL [7]. No loci were associated with ALT levels at genomewide significance level.

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To identify additional variants influencing both FL and ALT, we carried out a correlated meta-analysis of the two individual scans. Largely, the FL and ALT scans provided independent information (r=0.045 between the scan p-values). This approach yielded evidence for association on 10q24–q25 with variants in *ERLIN1-CHUK-CWF19L1* (endoplasmic reticulum (ER) lipid raft associated 1; conserved helix-loop-helix ubiquitous kinase; CWF19-like 1 cell cycle control; and biogenesis of lysosomal organelles complex-1 subunit 2, respectively) gene cluster. The regional plot depicts correlated meta-analysis associations between variants in *ERLIN1-CHUK-CWF19L1* gene cluster with FL and ALT (Figure 1). Nine SNPs located in two haplotype blocks (HB-1: rs2862954, rs1408579 and rs10883451 in *ERLIN1*; and HB-2: rs11597086 and rs11591741 in *CHUK*, and rs17729876, rs17668255, rs17668357 and rs12784396 in *CWF19L1*, Supplementary Figure 1) were strongly associated with concomitant variation in FL and ALT levels (4.29×10^{-10} CMA-p

 1.84×10^{-9}). For example, the correlated meta-analysis showed evidence of a novel association between the missense-*ERLIN1*-rs2862954 substitution C>T with decreasing levels of both FL ($\beta = -0.13\pm0.03$, C allele frequency=0.48) and ALT ($\beta = -0.14\pm0.03$, Table 2). This finding suggests that the missense-*ERLIN1*-rs2862954 may regulate the susceptibility of NAFLD in a broad spectrum of disease. Thus, the correlated meta-analysis approach enhanced the ability to detect association at genomewide significance level as compared with suggestive individual scans (2.71×10^{-6} GWA-p 2.75×10^{-5} for FL or ALT, Table 2). In addition to these nine SNPs, nineteen SNPs within *CHUK-CWF19L1* or near-*ERLIN1* genes showed suggestive associations with concomitant variation of FL and ALT levels (CMA-p $<1.0 \times 10^{-6}$, Supplementary Table 2, Supplementary Figure 1).

Since diabetic status and severe obesity are metabolic-related disorders with NAFLD and may represent potential confounders with NAFLD, we tested the robustness of these associations while restricting the analysis to subgroups of subjects free of diabetes and/or severe obesity (BMI 35.0 kg/m²). Even though the sample sizes were reduced ~23% after excluding diabetics and severe obese (ALT: 2,597 vs. 2,008 subjects and CT measured FL: 2,584 vs. 1,995 subjects), the association results for CMA and univariate analyses remained similar (Supplementary Table 3A). Also, the results were similar in non-diabetics (Supplementary Table 3B) and in non-extreme obese subjects (Supplementary Table 3C). These findings show evidence that variants within *ERLIN1-CHUK-CWF19L1* gene cluster have concomitant effects on NAFLD traits, even after accounting for some components related to metabolic syndrome (e.g., diabetes and severe obesity).

To test if the correlated associations of *ERLIN1-CHUK-CWF19L1* variants with NAFLD traits were predominantly due to ALT levels, we adjusted LA by ALT (in addition to the other covariates, LA_ALT) and carried out correlated meta-analysis between ALT and FL_ALT. The genomewide significance levels of the correlated meta-analysis were slightly attenuated (CMA-p < 3.3×10^{-7} , Supplementary Table 4A). We also found similar pattern of correlated meta-analysis associations when we used the adjusted ALT by FL (ALT_FL) and FL (CMA-p < 3.7×10^{-8} , Supplementary Table 4B). These findings suggest that variants of *ERLIN1-CHUK-CWF19L1* gene cluster influence both NAFLD traits (FL and ALT), and the significance of association enhances when accounting for the correlation between their scans.

4. Discussion

Variants of the *ERLIN1-CHUK-CWF19L1* gene cluster associated with both FL and ALT levels were identified by correlated meta-analysis. Among the nine SNPs at genomewide significance levels, three of them were in one *ERLIN1* haplotype block (rs2862954, rs1408579 and rs10883451), which includes the missense-*ERLIN1*-rs2862954 associated with lower levels of FL and ALT. The other six SNPs were in a single haplotype block that

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spans *CHUK* (rs11597086 and rs11591741) and *CWF19L1* (rs17729876, rs17668255, rs17668357 and rs12784396). In addition, our findings show that these common variants may influence both FL and ALT levels, even after accounting for some components related to metabolic syndrome, such as extreme obesity and diabetes.

Association of missense-*ERLIN1*-rs2862954 (p= 9.24×10^{-8} [8]) was previously reported with ALT levels in a genomewide meta-analysis study including 12,419 European white subjects (the CoLaus Study from Lausanne Switzerland, the InCHIANTI Study from Tuscany Italy, and the LOLIPOP Study from London UK). ALT levels have been used as a biochemical marker of hepatic inflammation [5, 6]; however it is a non-specific marker of liver damage. Our study shows for the first time that missense-*ERLIN1*-rs2862954 is relevant for the early stage of FL accumulation, even before liver damage occurs. *ERLIN1* encodes a member of the prohibitin family of proteins that define lipid-raft-like domains of the endoplasmic reticulum [23]. Lipid rafts are regions of membranes with a distinct, characteristic structural composition and that appear to act as platforms to colocalize proteins involved in intracellular signaling pathways [24]. *ERLIN1* is expressed in the liver, among other tissues [25].

Variants of CHUK were also previously reported associated with ALT levels (rs11591741: $p = 2.97 \times 10^{-8}$ and rs11597086: $p = 1.83 \times 10^{-8}$ [8]), and NASH fibrosis (rs11591741: p=0.010 [13]) in a multivariate model that accounted for sex, BMI, diabetes, alcohol consumption and PNPLA3 variant (rs738409). The 894 adult patients from the NASH Clinical Research Network and National Institutes of Health Clinical Center studies were predominantly Caucasian (85.7%) and 59.4% had definite histological diagnosis for NASH [13]. CHUK-intronic-rs11591741 is in complete LD with the 5'utr-CWF19L1-rs12784396 (LD=1 and $r^2=1$). These findings suggest a regulatory effect on transcription of *CHUK*-CWF19L1 that may influence the development and severity of NAFLD. The CHUK gene encodes a protein that is an inhibitor of the essential transcription nuclear factor-kappa-beta $(NF-\kappa B)$ complex via ubiquination pathway, and is involved in inflammatory responses triggered by cytokines [6]. It is worth pointing out that NF-B activated by IKK-B (IrcB kinase) is implicated in transcription of proinflammatory mediators (e.g., interleukin-1 (IL-1), interleukin-6 (IL-6) and TNF-a). NF-B dependent inflammatory mediators produced in hepatocytes down-regulate insulin sensitivity and trigger liver injury [26, 27], and is often elevated in patients with NASH [1, 27]. The CWF19L1 gene is conserved in several species; however its function remains unknown.

The SCD gene (10q24.31), located 79 kb downstream from the 5' utr-CWF19L1rs12784396, was previously reported influence NAFLD in mice and in humans. SCD encodes the stearoyl-CoA desaturase 1 (SCD1) and is involved in both the lipogenic as well as the lipid oxidative pathways [28]. SCD1 is a central enzyme in lipid metabolism; it converts saturated fatty acids to monounsaturated fatty acids, which are major substrates for synthesis of triacylglycerols and other lipids [28]. SCD1 knockout mice were resistant to the development of obesity and hepatic steatosis [28], whereas the activity of SCD1 was significantly increased in the fatty livers of ob/ob mice. In addition, hepatic SCD1 activity was increased in nonalcoholic human fatty liver [29], and associated with fatty liver and insulin resistance in obese humans [30]. Although SCD is a plausible candidate gene for NAFLD, we could not detect association of variants within the SCD gene with NAFLD traits (FL and ALT) nor has any other study found such an association, as far as we know. However, as it was previously suggested in mice, the activation of the transcription of SCD1 may occur via the numerous NF-B elements in the SCD1 promoter [28], and/or occur via endoplasmic reticulum stress and inflammatory response mRNAs [31]. If there is a pathway linking activity of SCD1 and CHUK protein-inhibitor NF-kB, and/or activity of SCD1 and

ERLIN-1 protein of lipid-raft-like domain are unknown, and further studies will be necessary to prove or disprove these hypotheses in human.

Presently, it is not possible to accurately predict which subjects with simple steatosis will develop to steatohepatitis and fibrosis, and much less is known about genetic variants influencing the progression of NAFLD. These findings suggest that variants within the *ERLIN1-CHUK-CWF19L1* gene cluster contribute to the increased susceptibility to simple steatosis (current FamHS results), hepatic injury (ALT [8], and the current FamHS results) and NAFLD histological severity (fibrosis [13]). It could be plausible that the evolution of NAFLD, from simple steatosis to hepatic inflammation to fibrosis, is due to the critical accumulation of liver fat, and they do not represent separate effects. Further studies will be necessary to understand the diagnosis and progression of NAFLD. Furthermore, our meta-analysis approach is the first study to report a correlated association between *ERLIN1-CHUK-CWF19L1* variants with the liver fat content and serum plasma ALT levels. Although our study shows evidence that variants of this gene cluster confer increased susceptibility to a broad spectrum of NAFLD, additional investigations will be necessary to understand how genetic effects of *ERLIN1-CHUK-CWF19L1* contribute to individual susceptibility to simple steatosis to the development of hepatic inflammation.

Our study has some limitations but several relevant strengths. Some of the strengths include: ALT levels are known as a sensitive screening test in detecting liver disease better than AST (aspartate aminotransferase) or other biochemical markers [32], and are strongly correlated with biopsy-proven fatty liver disease [6]. ALT levels have also been used to define NAFLD in epidemiological studies. Fatty liver accumulation was directly measured by CT liver attenuation, which is a non-invasive procedure that provides an accurate characterization of fat burden [3, 4]. Our novel correlated meta-analysis approach also provided an objective method to screen variants influencing correlated phenotypes, which resulted in associations that implicate novel variants (e.g., rs12784396 in the utr-5 CWF19L1) for NAFLD. Also, some SNPs that we found associations with concomitant variations of FL and ALT were previously reported associations with ALT levels [8] and with histological fibrosis [13]. The evidence of association between missense-ERLIN1-rs2862954 with NAFLD traits warrant functional followup studies. A limitation of our study is that ALT levels and CT measured FL do not provide the clinical precision of diagnostic forms of liver disease based on histology, which would require biopsies. However, we excluded subjects with high alcohol intake and hepatitis C positive to reduce this possible source of heterogeneity. Moreover, both CT measured FL and ALT levels bring opportunity to study NAFLD.

In brief, our study identified common variants of the *ERLIN1-CHUK-CWF19L1* gene cluster associated with NAFLD in a wide spectrum, simple steatosis, as CT measured FL, and hepatic inflammation measured by plasma levels of ALT. The discovery of these genetic variants may aid in understanding the correlated architecture between liver fat deposition and inflammation process, NAFLD development, and the prospective risk factors including type 2 diabetes and cardiovascular disease.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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Research highlights

We employed a novel correlated meta-analysis to improve the power to detect variants associated with NAFLD.

NAFLD was assessed by using CT measured fatty liver and alanine aminotransferase levels (ALT).

Correlated meta-analysis allowed to identify variants of the *ERLIN1-CHUK-CWF19L1* associated with CT measured fatty liver and ALT levels.

Variants of the *ERLIN1-CHUK-CWF19L1* are associated with the joint effect of steatosis (FL 40 HU) and inflammation (ALT 40 U/L).

Variants of the *ERLIN1-CHUK-CWF19L1* gene cluster may aid in understanding the correlated architecture between liver fat deposition and inflammation process.

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Figure 1.

Regional plot of *ERLIN1-CHUK-CWF19L1* gene cluster based on FamHS imputed genotype data. The CMA – log P value for correlated FL and ALT pair is represented in the ordinate axis, while the chromosome 10 position in Megabase (Mb) using build 35 is represented in the abscissa axis. The bottom panels give the relative positions of genes in the locus. Purple diamond indicates the effect of rs10883451 with the correlated FL and ALT pair. Each circle indicates a SNP with the color of the circle representing the correlation (r²) between that SNP and rs10883451. Blue line indicates estimated recombination hotspots in HapMap.

Table 1

Characteristics of phenotypes in the FamHS

	Number	Mean ± SD	Minimum	Maximum
Sample (% of females)	2,705 (55%)			
Age (years)	2,705	57.1 ± 13.3	30.0	93.0
Alcohol intake (drinks/week)	2,705	3.0 ± 7.1	0	171
CT liver attenuation (HU)	2,597	59.28 ± 11.02	-5.8	88.6
ALT (U/L)	2,584	21.1 ± 13.3	4.0	241.0
Triglycerides (mg/dL)	2,654	144.1 ± 93.9	24.0	1,037.0
Waist circumference (cm)	2,705	98.8 ± 16.3	58.0	187.0
Body mass index (kg/m ²)	2,705	28.9 ± 5.7	14.4	57.6
Severe obesity (N _{Total} ; % of affected)	338 (2,705; 12.5%)			
HOMA-IR	2,613	2.8 ± 2.9	0.3	66.4
Diabetes (N _{Total} ; % of affected)	362 (2,705; 13.4%)			

HOMA-IR: Homeostatic Model Assessment (fasting glucose (mmol/L)*fasting insulin (mU/L)/22.5);

Diabetes: taking hyperglycemic medication and/or fasting glucose 126 ml/dl;

Severe obesity: BMI 35.0 kg/m^2

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Table 2

Summary of correlated meta-analysis for FL and ALT (p-CMA $< 5.0 \times 10^{-8}$) and their univariate genomewide associations

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								FL and ALT		FL			ALT	
SNP	Location	Gene	Type	CAF	CA	Non-CA	П	p-CMA	β	S.D	Ρ	ß	S.D	Р
rs2862954	101902054	ERLINI	missense	0.48	C	Т	z	4.88E-10	-0.13	0.03	5.74E-06	-0.14	0.03	3.71E-06
rs1408579	101902184	ERLINI	intron	0.52	C	Т	Y	4.57E-10	0.13	0.03	5.35E-06	0.14	0.03	3.72E-06
rs10883451	101914408	ERLINI	intron	0.48	C	Т	Y	4.29E-10	-0.14	0.03	4.98E-06	-0.14	0.03	3.73E-06
rs11597086	101943695	CHUK	intron	0.56	A	С	Y	1.46E-09	0.14	0.03	2.34E-06	0.12	0.03	2.76E-05
rs11591741	101966491	CHUK	intron	0.45	C	G	Y	1.84E-09	-0.14	0.03	3.43E-06	-0.12	0.03	2.47E-05
rs17729876	101989736	CWF19L1	intron	0.45	A	G	Y	1.82E-09	-0.14	0.03	3.23E-06	-0.12	0.03	2.59E-05
rs17668255	101990691	CWF19L1	intron	0.55	C	Т	Y	1.82E-09	0.14	0.03	3.14E-06	0.12	0.03	2.65E-05
rs17668357	101993896	CWF19L1	intron	0.44	C	Ð	Y	1.65E-09	-0.14	0.03	2.71E-06	-0.12	0.03	2.75E-05
rs12784396	102017397	CWF19L1	utr-5	0.57	C	Т	z	2.47E-09	0.14	0.03	2.72E-06	0.12	0.03	4.11E-05
Chrom: chrom	osome; Locatic	on: SNP locati	on in Megabi	ase using	; build	35; CA: cod	led alle	ile; CAF: coded a	allele frec	luency;				
β: regression c	oefficient for th	he coded allek	e; SE: Standa	rd error	of the f	3; P: P value	of ass	ociation; IP: Imp	outed SNI	e is rep	resented as "	Y" ("N" :	= typed)	

The genomewide association (GWA) effect sizes of CT liver attenuation was multiplied by -1, which represents fatty liver (FL);

The GWA analysis included 2,584 participants for ALT levels and 2,597 participants for CT measured FL;

The tetrachoric correlation between FL with ALT was 0.045