

Systematic Review and Meta-Analysis To Estimate Antibacterial Treatment Effect in Acute Bacterial Skin and Skin Structure Infection

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A systematic literature review and meta-analysis were conducted to estimate the antibacterial treatment effect for linezolid and ceftaroline to inform on the design of acute bacterial skin and skin structure infection (ABSSSI) noninferiority trials. The primary endpoints included an early clinical treatment response (ECTR) defined as cessation of lesion spread at 48 to 72 h postrandomization and the test-of-cure (TOC) response defined as total resolution of the infection at 7 to 14 days posttreatment. The systematic review identified no placebo-controlled trials in ABSSSI, 4 placebo-controlled trials in uncomplicated skin and soft tissue infection as a proxy for placebo in ABSSSI, 12 linezolid trials in ABSSSI, 3 ceftaroline trials in ABSSSI, and 2 trials for non-antibacterial treatment. The ECTR rates at 48 to 72 h and corresponding 95% confidence intervals (CI) were 78.7% (95% CI, 61.1 to 96.3%) for linezolid, 74.0% (95% CI, 69.7 to 78.3%) for ceftaroline, and 59.0% (95% CI, 52.8 to 65.3%) for nonantibacterial treatment effect could not be estimated, given no available placebo or proxy for placebo data for this endpoint. Clinical, methodological, and statistical heterogeneity influenced the selection of trials for the meta-analysis of the TOC treatment effect estimation. The pooled estimates of the TOC treatment response were 31.0% (95% CI, 62.2 to 55.9%) for the proxy for placebo, 88.1% (95% CI, 81.0 to 95.1%) for linezolid, and 86.1% (95% CI, 83.7 to 88.6%) for ceftaroline. The TOC clinical treatment effect estimation was 25.1% for linezolid and 27.8% for ceftaroline. The antibacterial treatment effect estimation at TOC will inform on the design and analysis of future noninferiority ABSSSI clinical trials.

ver the past decade, robust clinical, scientific, and regulatory debate has emerged for initiatives to improve the design, execution, and analysis of antibacterial clinical trials (1-4). New trials for acute bacterial skin and skin structure infections (ABSSSI), previously referred to as complicated skin and skin structure infections (cSSTI), remain important, given the rise in incidence of methicillin-resistant Staphylococcus aureus infections and reports of treatment failure (5-8). Per guidance from the U.S. Food and Drug Administration (FDA), patient eligibility for enrollment in ABSSSI trials should be restricted to those with erysipelas, cellulitis, major cutaneous abscesses, and wound infections having a minimal lesion surface area involvement of 75 cm^2 (9). For trial endpoints, the traditional test-of-cure (TOC) endpoint, with the treatment success defined as total resolution of the infection at 7 to 14 days posttreatment, remains aligned with the European regulatory guidance, yet the treatment success for the primary efficacy endpoint aligned with the FDA guidance is defined as cessation of lesion spread after 48 to 72 h of treatment (9-11). Revisions to the enrollment and endpoint criteria in recent regulatory guidance for ABSSSI trials necessitate reevaluation of the antibacterial treatment effect estimation calculated from across-trials comparisons of existing data for noninferiority trial design (4, 12–16).

To inform on future noninferiority ABSSSI trial design, we conducted a systematic review and meta-analysis of antibacterial treatment effect estimation. Linezolid and ceftaroline were *a priori* selected as drugs representative of potential active comparators for hospitalized adults with ABSSSI in a global phase 3 clinical development program. The trial data extracted for the systematic review were aligned with regulatory guidance for the enrollment and endpoint criteria in ABSSSI trials. A predefined meta-analysis plan defined the efficacy variables, primary endpoints of interest, and computational methods for antibacterial treatment effect estimation in ABSSSI historical trials for noninferiority margin justification.

MATERIALS AND METHODS

Study design. The systematic review and meta-analysis were designed and executed in compliance with the Preferred Reporting Items for Systematic Reviews and Meta-Analysis (PRISMA) (17-19). Two independent reviewers conducted computer-based literature searches and a systematic review using the MEDLINE search engine (PubMed, U.S. National Library of Medicine, National Institutes of Health; http://www.ncbi.nlm .nih.gov). Ideally, treatment effect estimation for a noninferiority antibacterial trial should be calculated from placebo-controlled trials to allow for within-trial comparison of the antibacterial treatment response compared to the placebo treatment response (11, 14). Given a prior report that placebo-controlled trials were nonexistent for complicated skin infections (20), we partitioned the literature searches to identify placebo-controlled trials of any antibacterial treatment for ABSSSI (Appendix, search A1) as well as randomized clinical trials of active comparators in ABSSSI for linezolid (search B1) and ceftaroline (search B2). Due to nomenclature changes over time, studies that assessed ABSSSI, complicated skin and soft tissue infections (cSSTI), or complicated skin and skin structure infections (cSSSI) were included. While it was previously reported that there were no historical data for placebo-controlled trials of any antibacterial treatment in complicated skin infections, our search strategy, by design, would have captured any placebo-controlled ABSSSI trials, inclusive of

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trials for linezolid and ceftaroline. The placebo search was extended to include clinical trials reporting a placebo treatment response in uncomplicated skin and soft tissue infections (uSSTI) as a proxy for placebo in ABSSSI (search A2). This strategy, which comprised use of a proxy for placebo effect estimation, has been recommended in the FDA guidance for noninferiority trials and has been used for placebo treatment effect estimation for trials in complicated urinary tract infections (13, 21, 22). Given that historical studies published in 1937 reported relatively high response rates in the treatment of erysipelas with UV light, an additional search (search C) was conducted to identify clinical trials of nonantibacterial treatment in uSSTI and in ABSSSI (4, 23, 24). This search was *a priori* conducted to identify potentially informative data and not as a proxy for placebo treatment response estimation.

Identification of trials and full-screen review for eligibility. For each search, the publications were restricted to dates before 28 February 2013. Search terms included abbreviations and plural and full-phrase versions of the selected search terms (listed in Appendix 1) combined with Boolean operators (AND, OR) used in succession to narrow or widen the respective searches. Two reviewers independently assessed the identified publications for trial eligibility, data quality, and efficacy data. Eligible studies included randomized clinical trials for the respective searches of placebocontrolled trials (search A1), active comparator trials of linezolid (search B1) and ceftaroline (search B2) in ABSSSI, and trials of nonantibacterial treatment in uSSTI and ABSSSI (search C). Case series, observational studies, review papers, duplicate studies, and studies with incomplete data were not eligible for full-text review. The publications identified in searches A1, B1, B2, and C were excluded for one or more of the following criteria: pediatric exclusivity, lack of reported measure of endpoints of interest, >15% of subjects with either diabetic foot infections, animal or human bites, necrotizing fasciitis, decubitus ulcer infection, myonecrosis or ecthyma gangrenosum, and pooled treatment response rates for multiple infection types. Publications of uSSTI studies in search A2 were eligible for full review if the report was a placebo-controlled trial in impetigo, furunculosis, carbunculosis, folliculitis, ecthyma, erysipelas, or secondarily infected traumatic lesions (SITL). Publications of uSSTI studies were excluded for one of the following criteria: placebo add-on treatment, pediatric exclusivity, prevention or colonization trials, or inclusion of subjects with minor cutaneous abscesses treated via incision and drainage as surgical treatment. The latter group was excluded as any potential estimate of placebo effect could not be assessed independently of the surgical treatment effect (25-27).

Primary endpoints. For the meta-analysis, the two primary endpoints of interest were in the intention-to-treat (ITT) population and defined as the early clinical treatment response at 48 to 72 h after randomization and the TOC clinical treatment response. Outcome definitions of clinical success, cure, or treatment failure were evaluated from each of the published studies on the basis of criteria available and assessed for methodological and clinical heterogeneity. The early clinical treatment response was defined as the proportion of subjects who experienced cessation of spread or the reduction in the total surface area of the lesion at 48 to 72 h after randomization or after the first dose of the study drug (9). The TOC clinical treatment response varied in definition based on the location and year of the trial execution but was typically defined as the proportion of subjects who experienced total resolution of all signs and symptoms of the infection at 1 to 2 weeks after completion of the therapy (9). Studies that incorporated improvement in the definition of success were included in the meta-analysis, yet noted for this composite definition of success. Treatment was divided into three broad categories: placebo (with extension to a proxy for placebo), antibacterial, and nonantibacterial treatment for the ITT population unless otherwise noted.

Data extraction. Data were systematically extracted for general study characteristics, which included author, publication date, study drug and duration of therapy, active comparator, study population description, study years, countries or regions, study centers, and outcomes. Baseline demographic data were abstracted to assess age, gender, geographic region, proportion of baseline infection types, and lesion size when available. Methodological details were abstracted to assess randomization procedures, sequence generation, blinding in subjects and in investigators, eligibility criteria, inclusion and exclusion criteria, subject withdrawals, allocation concealment, primary outcome variable, secondary outcome variable, study definitions, timing of outcome assessment (including early clinical treatment response at 48 to 72 h after randomization and TOC clinical treatment response), incomplete reporting of outcomes, and other potential sources of bias.

Assessments of trial heterogeneity and publication bias. Publications reporting trials meeting the inclusion criteria were assessed for clinical, methodological, and statistical heterogeneity. Clinical heterogeneity was defined by patient selection, interventions, and outcomes, methodological heterogeneity was defined by study design and execution, and statistical heterogeneity was defined as a variation in the results beyond sampling variability (28-31). Given the qualitative decisions associated with clinical and methodological heterogeneity, each trial meeting the eligibility criteria was systematically assessed by each reviewer for inclusion and exclusion criteria, types and distributions of skin infections, timing of the outcome assessments, treatment duration, inclusion of lesion improvement as an indicator of success, and the proportion of subjects with methicillin-resistant Staphylococcus aureus (MRSA) pathogens. Each of two independent reviewers employed the Cochrane Risk of Bias Tool for assessment of publication bias, and any potential discrepancies were adjudicated via consensus with a third assessor (32).

Study selection for meta-analysis. Extensive clinical, methodological, and statistical heterogeneity influenced the selection of studies included in the meta-analysis (31). The proxy for placebo trials selected for inclusion in the meta-analysis was restricted to studies that did not exclusively enroll subjects with SITL, given the differential regulatory approval of antibacterial agents for this infection type. For the linezolid and ceftaroline trials, selection for inclusion in the meta-analysis was based on trial parameters in the historical data that approximated recent ABSSSI guidance to minimize the clinical and methodological heterogeneity of studies pooled in the meta-analysis (9, 31). These selection criteria were trials with total surface lesion area of >75 cm², a non-MRSA-specific population, and abscesses in <50% of the population. The trials reporting UV light treatment response were included in the meta-analysis as pooled nonantibacterial treatment (20, 23, 24).

Data analysis. Pooled treatment response rates and 95% confidence intervals (CI) were calculated for each treatment group using the DerSimonian-Laird methodology, a noniterative, random effects model to account for interstudy variability that utilized the metaphor package in R software (33). This method is commonly employed in meta-analyses of clinical trials, as it accounts for the heterogeneity of studies through a statistical parameter that represents the interstudy variation of the trials in the model (33). The proportion of total variation in the study estimates that was due to interstudy statistical heterogeneity was assessed using I^2 values and associated P values for trials included in the meta-analysis. Given the small number of the proxy for placebo and active comparator trials, meta-regression was not feasible. The indirect, across-trial comparisons to estimate the antibacterial treatment effect were calculated as the difference between the lower bound of the 95% CI of the response rate estimate for each antibacterial treatment and the upper bound of the 95% CI of the proxy for the placebo response rate estimate (14, 21). This methodology is aligned with the FDA guidance on the design of noninferiority trials and is acknowledged for yield of intrinsically conservative noninferiority margin estimates (14). Publication bias and heterogeneity were assessed using funnel plots (34).

RESULTS

Systematic review. No placebo-controlled trials of ABSSSI were identified (Appendix, search A1). There were 800 placebo records for uSSTI (search A2), 477 linezolid records (search B1), 73 ceftaroline records (search B2), and 2,390 records for trials defined as

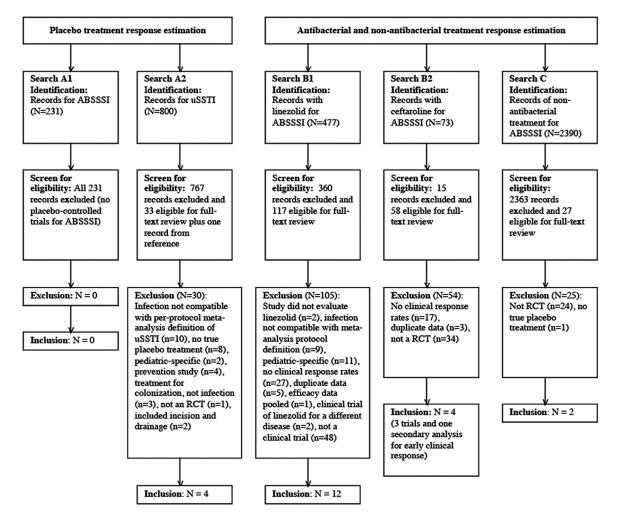


FIG 1 Flow chart depicting the systematic literature search for trials of treatment effect estimation in acute bacterial skin and skin structure infection (ABSSSI): publication identification, screening, eligibility, and trial inclusion. RCT, randomized clinical trial.

nonantibacterial treatments for skin infection (search C). The final selection of publications included 4 placebo-controlled uSSTI trials (35–38), 12 linezolid trials (39–50), 3 ceftaroline trials (51– 53), 1 retrospective ceftaroline secondary analysis (54), and 2 nonantibacterial treatment trials of UV light for erysipelas (23, 24) (Fig. 1).

(i) Proxy trials of uSSTI for placebo treatment of ABSSI (searches A1 and A2). The four trials as proxy for placebo included 234 subjects with a range of uncomplicated infections reported as impetigo, folliculitis, SITL, and pyoderma (Table 1). Publication dates were from 1974 to 2013, three of the trials enrolled a subset of subjects <18 years of age, and one publication required certified translation from French to English (35–38). Each trial reported a TOC clinical treatment response; none reported an early clinical treatment response.

(ii) Antibacterial trials of ABSSSI (searches B1 and B2). Among the 12 linezolid trials published between the years 2000 and 2013, three were phase 2 trials, three restricted enrollment to subjects with MRSA infections (44, 46, 47), three were specific to China and the Asia-Pacific region (43, 44, 48), and one (45) reported only the treatment response rate for the clinically evaluable population (Table 1). The ceftaroline publications included one phase 2 trial, two registrational phase 3 trials, and a secondary subset analysis of the two phase 3 CANVAS (ceftaroline versus vancomycin in skin and skin structure infections) trials reporting an early clinical treatment response (51–54).

(iii) Nonantibacterial treatment trials (search C). The openlabel controlled trials of subjects with erysipelas compared UV light to either prontosil or sulfanilamide (23, 24). The clinical data available did not specify lesion size, yet provided descriptive data for severely ill subjects with erysipelas and endpoints aligned with an early clinical treatment response at 48 to 72 h.

Meta-analysis. Data on the primary treatment response rate, lesion size, and treatment duration from the 21 trials and the one secondary analysis deemed eligible for the meta-analysis are summarized in Table 2.

(i) Treatment response at the TOC endpoint. The four proxy for placebo uSSTI trials reported TOC clinical treatment response rates ranging from 8% to 66% (Table 2) (35–38). Clinical and methodological heterogeneity was evident for lesion size, infection type, and timing of the TOC assessment (Table 3). For the meta-analysis, the trial that restricted enrollment to subjects with SITL was excluded (35), resulting in a pooled proxy for placebo TOC clinical treatment response rate of 31.0% (95% CI, 6.2% to

Treatment and study	Trial and design ^b	Population age and infection type ^c	Comparator
Placebo in uncomplicated SSTI, $n = 4$			
Tomayko et al., 2013 (35)	Phase 3, RCT, DB	Age ≥ 9 mo with SITL	Topical retapamulin
Koning et al., 2008 (36)	RCT, DB	Age ≥ 9 mo with impetigo	Topical retapamulin
Colin and Avon, 1988 (37)	RCT, DB	Subjects with SSTI	Topical mupirocin
Zaynoun et al., 1974 (38)	RCT, DB	Age ≥ 4 mo with pyoderma ^d	Topical gentamicin
Linezolid, $n = 12$			
Prokocimer et al., 2013 (39)	Phase 3, RCT, DB	Adults with ABSSSI	Tedizolid
Noel et al., 2012 (40)	Phase 2, RCT, EB phase 2, RCT, EB	Age ≥ 18 yr with cSSSI	Omadacycline
Covington et al., 2011 (41)	Phase 2, RCT, EB	Age ≥ 18 yr with ABSSSI	JNJ-QR
Craft et al., 2011 (42)	Phase 2, RCT, EB	Age ≥ 18 yr with ABSSSI	CEM-102
Lin et al., 2008 (43)	Phase 3, RCT, DB	Age ≥18 yr in China with cSSTI or pneumonia	Vancomycin
Kohno et al., 2007 (44)	RCT, OL	Adults in Japan with cSSTI, pneumonia, or sepsis due to MRSA	Vancomycin
Jauregui et al., 2005 (45)	Phase 3, RCT, DB	Adults with cSSSI	Dalbavancin
Sharpe et al., 2005 (46)	RCT, OL	Age ≥ 18 yr with MRSA cSSTI	Vancomycin
Weigelt et al., 2005 (47)	RCT, OL	Adults with MRSA cSSTI	Vancomycin
Chen et al., 2004 (48)	One arm, OL	Adults with cSSTI in the Asia-Pacific region	No comparator
Stevens et al., 2002 (49)	RCT, OL	Age ≥13 yr with MRSA cSSTI, pneumonia, UTI, or bacteremia	Vancomycin
Stevens et al., 2000 (50)	RCT, DB	Age ≥ 18 yr with cSSTI	Oxacillin-dicloxacillin
Ceftaroline, $n = 4$			
Friedland et al., 2012 (54)	Secondary analysis phase 3 RCT, DB	Adults with ABSSSI	Vancomycin
Corey et al., 2010 (51)	Phase 3, RCT, DB	Age ≥ 18 yr with cSSSI	Vancomycin
Wilcox et al., 2010 (52)	Phase 3, RCT, DB	Age ≥ 18 yr with cSSSI	Vancomycin
Talbot et al., 2007 (53)	Phase 2, RCT, EB	Age ≥ 18 yr with cSSSI	Vancomycin
UV light, $n = 2$			
Snodgrass and Anderson, 1937 (23)	Clinical, OL	Subjects with erysipelas	Prontosil
Snodgrass and Anderson, 1937 (24)	Clinical, OL	Subjects with erysipelas	Sulfanilamide

TABLE 1 Study design, population, and active comparator of 22 published trials identified in a systematic review for antibacterial treatment effect estimation for acute bacterial skin and skin structure infections^{*a*}

^a Intention-to-treat population assigned to linezolid, ceftaroline, placebo, or UV light for complicated skin and soft tissue infections (cSSTI), complicated skin and skin structure infections (cSSSI), or acute bacterial skin and skin structure infections (ABSSSI).

^b RCT, randomized control trial; DB, double blinded; EB, evaluator blinded; OL, open label.

^c Geographic region mentioned where relevant. SITL, secondarily infected traumatic lesion; MRSA, methicillin-resistant *Staphylococcus aureus*; UTI, urinary tract infection. ^d Infections were impetigo, insect bites, and dermatitis.

55.9%); the variance across trials ($I^2 = 90.6\%$, P < 0.0001) was high (Fig. 2a) (36-38). Eleven of the 12 linezolid trials reported TOC clinical treatment response rates ranging from 50% to 95% (Table 2) (39-44, 46-50). These trials, published over a 13-year interval, had evidence of clinical and methodological heterogeneity for lesion size, type of infection, geographic region, MRSA etiology, per-protocol treatment duration, definition of clinical success, and timing of TOC assessment (Tables 2 and 3). For the meta-analysis, the selection of the three linezolid trials comprised subjects with total surface area lesion size of \geq 75 cm², non-MRSA-specific populations, and <50% of subjects with abscesses (39, 41, 42). The pooled linezolid TOC clinical treatment response rate was 88.1% (95% CI, 81.0% to 95.1%) (Fig. 2b), with evidence of variation across the trials ($I^2 = 80.4$, P =0.006). The three ceftaroline trials had consistent distributions of infection types, durations of treatment, definitions of clinical success, and timing of TOC assessment (Tables 2 and 3). The pooled ceftaroline TOC clinical treatment response rate

was 86.1% (95% CI, 83.7% to 88.6%) (Fig. 2c), with no evidence of clinical, methodological, and statistical heterogeneity ($I^2 = 0, P = 0.74$) (51–53). The TOC endpoint was not reported in the trials of UV light (23, 24).

(ii) Treatment response at the early clinical endpoint. No early clinical treatment response was reported in the proxy for placebo uSSTI trials. The early clinical treatment response for linezolid in the ITT population was reported in three trials, with a pooled estimate of 78.7% (95% CI, 61.1% to 96.3%); a high variance across trials ($I^2 = 96.7\%$, P < 0.0001) was evident (39, 41, 42). This response rate approximated the early clinical treatment response rate of 79.4% (95% CI, 75.1% to 83.7%) reported in the most robust of the three linezolid trials with these data, as well as the early clinical treatment response of ceftaroline at 74.0% (95% CI, 69.7% to 78.3%) reported in one trial (39, 54). The pooled UV light treatment response in erysipelas at the early endpoint was 59.0% (95% CI, 52.8% to 65.3%), with low variance between the two trials ($I^2 = 0$, P = 0.482), as well as low methodological and

		Mean (SD) Rx ^a duration	Intention-to-treat response (no. events/total [%])	
Treatment and study	Skin infection lesion size	(days)	Early	TOC
Placebo in uncomplicated SSTI, $n = 4$				
Tomayko et al., 2013 (35)	Total area $\leq 100 \text{ cm}^2$	NA^b	NA	75/113 (66.4)
Koning et al., 2008 (36)	Total area ≤100 cm ²	NA	NA	28/71 (39.4)
Colin and Avon, 1988 (37)	Subgroups: 1–2, 2–4, 4–6, >6 cm ²	NA	NA	2/25 (8.0)
Zaynoun et al., 1974 (38)	NA	NA	NA	12/25 (48.0)
Linezolid, $n = 12$				
Prokocimer et al., 2013 (39)	\geq 75 cm ²	NA	266/335 (79.4)	288/335 (86.0)
Noel et al., 2012 (40)	$13.5 \text{ cm} (14.2)^c$	9.6 (4.4)	NA	82/108 (75.9)
Covington et al., 2011 (41)	\geq 75 cm ²	NA	45/78 (57.7)	64/78 (82.1)
Craft et al., 2011 (42)	$\geq 100 \text{ cm}^2$	11.5 (NA)	63/65 (97)	73/77 (94.8)
Lin et al., 2008 (43)	NA	12.2 (5.4)	NA	30/33 (90.9)
Kohno et al., 2007 (44)	NA	10.9 (5.0)	NA	$9/17 (52.9)^d$
Jauregui et al., 2005 (45)	NA	NA	NA	NA ^e
Sharpe et al., 2005 (46)	NA	NA	NA	$15/30 (50.0)^d$
Weigelt et al., 2005 (47)	NA	11.8 (4.9)	NA	439/583 (75.3)
Chen et al., 2004 (48)	NA	NA	NA	72/77 (93.5)
Stevens et al., 2002 (49)	51.2% with lesions ≥ 28 cm ²	12.6 (7.1)	NA	$37/53 \ (69.8)^d$
Stevens et al., 2000 (50)	NA	13.4 (5.4)	NA	279/400 (69.8)
Ceftaroline, $n = 4$				
Friedland et al., 2012 (54)	\geq 75 cm ²	NA	296/400 (74.0)	NA
Corey et al., 2010 (51)	83.5% had ≥1 dimension >5 cm	Median, 7	NA	304/351 (86.6)
Wilcox et al., 2010 (52)	94.3% had ≥ 1 dimension >5 cm	Median, 6.5	NA	291/342 (85.1)
Talbot et al., 2007 (53)	NA	Median, 7.8 (range, 0.4–19.5)	NA	59/67 (88.1)
UV light, $n = 2$				
Snodgrass and Anderson, 1937 (23)	NA	2.6 exposures ^f	64/104 (61.5)	NA
Snodgrass and Anderson, 1937 (24)	NA	1.4 exposures ^f	77/135 (57.0)	NA

TABLE 2 Trial parameters and treatment response estimates for placebo treatment in uncomplicated skin and soft tissue infection and antibacterial and nonantibacterial treatment for acute bacterial skin and skin structure infection

^a Values are mean (SD) unless otherwise specified. Rx, treatment.

^b NA, not available.

^c Mean maximum linear dimension (SD).

^d Microbiological ITT (MITT) reported for Sharpe et al., 2005 (46), and Stevens et al., 2002 (49), but microbiological evaluable (ME) was the only outcome available in Kohno et al., 2007 (44).

^e Jauregui et al., 2005 (45), report results for the clinically evaluable (CE) and microbiological evaluable (ME) population at TOC.

f Exposures per case.

clinical heterogeneity in the trial design or execution (Tables 2 and 3) (23, 24).

Antibacterial treatment effect estimation for ABSSSI. Given the biological plausibility of UV light treatment against bacteria, the response rates to UV light treatment in erysipelas were considered inappropriate as proxy for placebo responses (23, 24, 55). Given no available proxy for placebo early clinical treatment response data, an early clinical treatment effect estimate could not be calculated for linezolid or ceftaroline. In contrast, the available proxy for placebo uSSTI treatment response rate (31.0%; 95% CI, 6.2% and 55.9%) permitted calculation of the TOC clinical treatment effect estimate for linezolid and ceftaroline. This acrosstrials comparison, defined as the difference between the upper bound of the 95% CI for the proxy for placebo treatment response and the respective lower bound of the 95% CI for the treatment response in linezolid (81.0%) and in ceftaroline (83.7%), resulted in TOC clinical treatment effect estimation of 25.1% for linezolid and 27.8% for ceftaroline (Table 4).

Risk of bias assessment. From the assessment of publication quality and risk of bias, the four placebo-controlled trials of uSSTI all had low or uncertain bias scores, 7 of the 16 active comparator trials had one or more high bias scores, and the 2 nonantibacterial trials of UV light had high bias in three of the six categories (Table 5). Funnel plots were assessed (data not shown) to determine potential publication bias; however, the assessment was inconclusive, given the small number of trials and the heterogeneity in treatment response rates.

DISCUSSION

The findings from this systematic review and meta-analysis of the antibacterial treatment effect estimation in ABSSSI inform on future trial designs for this group of infections. The early clinical

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	Assessment	Assessment			Infection t	Infection type distribution $(\%)^a$	n (%) ^a					
Treatment and study	Early	TOC	Duration	Improvement ^b	Impetigo	Folliculitis	SITL	Other	Abscess	Cellulitis/erysipelas	Wound/other	MRSA
Proxy for placebo Tomavko et al., 2013 (35)	NAc	$12-14^{d}$	J	No	0	D	100	D				
Koning et al., 2008 (36)	NA	14^d	J	Yes	100	0	0	0				
Colin and Avon, 1988 (37)	NA	7^d	U	No	28	36	0	36				
Zaynoun et al., 1974 (38)	NA	$7 - 10^{d}$	7	No	48	0^e	0	52				
Linezolid												
Prokocimer et al., 2013 (39)	$48-72^{d}$	$7 - 14^{f}$	10	No					29.3	41.5	29.3	43.1
Noel et al., 2012 (40)	NA	$10-17^{g}$	NA	No					67	9	24	50.8
Covington et al., 2011 (41)	$48-72^{h,i}$	$2 - 14^{g}$	7-14	No					35.9	34.6	29.5	NA
Craft et al., 2011 (42)	At 72 ^h	$7 - 14^{d}$	10 - 14	No					0	64	36	76
Lin et al., 2008 (43)	NA	7–28 ^g	7-21	Yes					NA	NA	NA	NA
Kohno et al., 2007 (44)	NA	7-218	7-21	No					NA	NA	NA	100
Jauregui et al., 2005 (45)	NA	14 ± 2	14	Yes					30	30	40	50.5
Sharpe et al., 2005 (46)	NA	10^{-7}	7-21	No					NA	NA	NA	100
Weigelt et al., 2005 (47)	NA	Þf	4-21	No					26.7	47.6	25.7	41.3
Chen et al., 2004 (48)	NA	$14-21^{g}$	7-28	Yes					NA	NA	NA	3.2
Stevens et al., 2002 (49)	NA	$15-21^{g}$	7-14	No					33.7	13.0	53.3	100
Stevens et al., 2000 (50)	NA	15–21 ^g	10-21	Yes					14.6	44.8/10.3	30.3	NA
Ceftaroline												
Friedland et al., 2012 (54)	$48-72^{d}$	NA	5-14	NA					28.7	54.2	17.1	42.3
Corey et al., 2010 (51)	NA	8–15 ^g	5-14	Yes					28.2	34.5	37.4	34.3
Wilcox et al., 2010 (52)	NA	8–15 ^g	5-14	Yes					40.6	30.1	29.3	32
Talbot et al., 2007 (53)	NA	$8 - 14^{g}$	7-14	Yes					44.8	34.3	20.9	NA
UV light												
Snodgrass and Anderson, 1937a (23)	72'	NA	NA	NA					0	100	0	0
Snodgrass and Anderson, 1937b (24)	72'	NA	NA	NA					0	100	0	0
^a SITL, secondarily infected traumatic lesions defined as abrasions, lacerations, and wounds; MRSA, methicillin-resistant Staphylococcus aureus. ^b Improvement: successful treatment response at test of cure (TOC).	defined as abr e at test of cure	asions, lacera e (TOC).	tions, and wou	nds; MRSA, methicilli	n-resistant Sta	phylococcus aureu	s.					
^d A G and a C and a G and a												
[•] After the first dose of study drug. [•] Folliculitis cases enrolled yet not possible to discern the case number assigned to the placebo arm.	discern the cas	e number ass	signed to the pl:	acebo arm.								
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TABLE 3 Evidence of methodological and clinical heterogeneity in 22 treatment trials representing proxy for placebo, antibacterial, and nonantibacterial UV light treatment for acute bacterial skin and skin structure infection

 f At the end of the treatment visit. g After last dose of study drug.

^h Postrandomization.
 ⁱ Also includes the assessment from 36 to 84 h.
 ^j Hours after initiation of therapy.

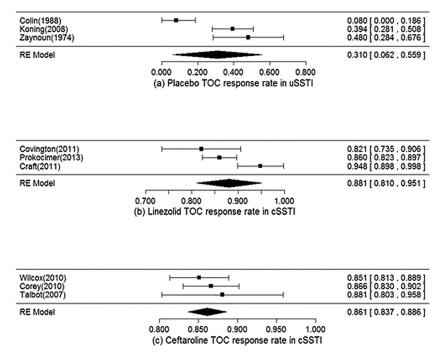


FIG 2 Forest plots of the primary meta-analysis for clinical response at test of cure for proxy-for-placebo treatment (a), linezolid treatment (b), and ceftaroline treatment (c). RE, random effects.

treatment responses for linezolid (78.7%), ceftaroline (74.0%), and UV light (59.0%), provide reference data for the recently recommended primary endpoint in ABSSSI trials (9). The TOC treatment effect estimations for linezolid (25.1%) and ceftaroline (27.8%) provide historical data for noninferiority margin justification in future noninferiority trial designs and for superiority trial designs that may include adaptive models and futility stopping criteria (56, 57). Overall, our findings provide historical evidence of the sensitivity to drug effect (HESDE) at the TOC endpoint for ABSSSI and are consistent with prior estimations of the treatment effect (13).

These TOC treatment effect estimations are conservative, yet are based on the calculations recommended in the FDA guidance (9). These estimates are conservative for two reasons. First, the definition of the antibacterial treatment effect as the difference between the upper bound of the 95% CI for the proxy for placebo treatment response and the respective lower bound of the 95% CI for the antibacterial treatment response is intrinsically conservative. An alternative calculation based on the difference in the point estimates of the placebo and treatment group response rates would yield treatment effect estimations of 57.1% for linezolid and 55.1% for ceftaroline (Table 4). Second, these estimates are further conservative, given that the placebo treatment response for uSSTI, assumed to be higher than the placebo treatment response in ABSSSI, was used as a proxy for the placebo response in ABSSSI. As in most antibacterial trials involving serious infection indications, where there is existing effective therapy as the standard of care, placebo treatment is medically and ethically inappropriate and historically not reported. Hence, randomized, parallelgroup noninferiority trials are employed to demonstrate drug efficacy and meet the regulatory guidance for approval of a new test or antibacterial agent in serious infection indications (7-9, 12). To inform on noninferiority margins, we followed the FDA guidance for use of a proxy for placebo in the absence of a true placebo response for ABSSSI. While the placebo response rate in uSSTI should not be interpreted as the true placebo response rate

TABLE 4 Summary of meta-analysis results for the clinical treatment response at the test-of-cure endpoint for placebo in uncomplicated skin and soft tissue infection, linezolid, and ceftaroline, and estimated antibacterial treatment effect estimation for acute bacterial skin and skin structure infection

Treatment group (no. of trials) ^{<i>a</i>}	Point estimate response rate (% [95% CI ^b])	Treatment effect estimate (%) ^c	Alternate treatment effect estimate $(\%)^d$
Placebo in uSSTI (3)	31.0 (6.2–55.9)		
Linezolid in ABSSSI (3)	88.1 (81.0-95.1)	25.1	57.1
Ceftaroline in ABSSSI (3)	86.1 (83.7–88.6)	27.8	55.1

^a uSSTI, uncomplicated skin and soft tissue infection; ABSSSI, acute bacterial skin and skin structure infection.

^b CI, confidence interval.

^c Calculated as the difference between the lower bound of the 95% CI of the response rate estimate for each antibacterial treatment and the upper bound of the 95% CI of the proxy for placebo response rate estimate (13, 20).

^d Calculated as the difference in the point estimate of the response rate between each antibacterial treatment and placebo.

	Selection bias		Performance and	Attrition bias:	Reporting bias:	
Treatment and study	Random sequence generation	Allocation concealment	detection bias: blinding ^b	incomplete outcome data	selective reporting	Other biases
Placebo in uncomplicated SSTI, $n = 4$						
Tomayko et al., 2013 (35)	L	L	L	L	U	U
Koning et al., 2008 (36)	U	U	L	L	U	U
Colin and Avon, 1988 (37)	U	U	L	L	U	U
Zaynoun et al., 1974 (38)	U	L	L	L	U	U
Linezolid, $n = 12$						
Prokocimer et al., 2013 (39)	L	L	L	L	L	U
Noel et al., 2012 (40)	L	L	L	L	U	U
Covington et al., 2011 (41)	U	U	L	L	U	U
Craft et al., 2011 (42)	U	L	L	U	U	U
Lin et al., 2008 (43)	U	U	L	U	U	U
Kohno et al., 2007 (44)	U	Н	Н	L	L	U
Jauregui et al., 2005 (45)	L	L	L	L	Н	U
Sharpe et al., 2005 (46)	U	Н	Н	L	L	U
Weigelt et al., 2005 (47)	U	Н	Н	U	U	U
Chen et al., 2004 (48)	Н	Н	Н	L	L	U
Stevens et al., 2002 (49)	U	Н	Н	L	U	U
Stevens et al., 2000 (50)	U	L	L	L	U	U
Ceftaroline, $n = 4$						
Friedland et al., 2012 (54)	L	L	L	L	L	U
Corey et al., 2010 (51)	L	L	L	L	U	U
Wilcox et al., 2010 (52)	L	L	L	L	U	U
Talbot et al., 2007 (53)	U	L	H ^c	L	U	U
UV light, $n = 2$						
Snodgrass and Anderson, 1937 (23)	Н	Н	Н	L	U	U
Snodgrass and Anderson, 1937 (24)	Н	Н	Н	L	U	U

TABLE 5 Assessment of publication bias in 22 publications of placebo treatment for uncomplicated skin and soft tissue infection and antibacterial and nonantibacterial treatment for acute bacterial skin and skin structure infection^a

^a Cochrane Risk of Bias Tool and PRISMA guidelines. H, high bias; U, uncertain bias or unable to determine; L, low bias.

^b Blinding of participants, personnel, and outcome assessment.

^c Evaluator blinded only.

in ABSSSI, we assumed that the use of this proxy for the placebo treatment response, resulting in an overestimation of the ABSSSI placebo treatment response, is a conservative approach in estimating antibacterial efficacy relative to that of placebo. Based on the uncertainty of the true placebo effect and the use of the FDA guidance, each ABSSSI treatment effect estimate is conservative, given that the overestimation of the ABSSSI placebo treatment response results in a conservative underestimation of the ABSSSI treatment effect (14, 22, 58).

The second major finding from our study is the evidence and assessment of heterogeneity in these historical clinical trials as relevant and informative to future ABSSSI clinical trials. For the meta-analysis, all data from the ceftaroline trials were included, given minimal evidence for clinical, methodological, and statistical heterogeneity. For the linezolid trials, data from the maximum number of trials (n = 12) offered the potential benefit of increased robustness for the treatment effect estimate, but this was offset by significant evidence of clinical, methodological, and statistical heterogeneity (31). The meta-analysis trial selection process for the linezolid TOC clinical treatment response was prioritized to identify the historical trials aligned with the recent FDA regulatory guidance for future ABSSSI trials. This process yielded a selection of just three trials and represented the best reproducibility for the

linezolid treatment response to be observed in future noninferiority trials, along with the potential to minimize the deviations in the study operating characteristics, inclusive of type I error and power.

As with all systematic reviews and meta-analyses, we acknowledge the potential bias inherent in our study design and reported findings. First, the publications identified in the systematic review were contingent upon the search terms used, and the inclusion of open-label studies may have introduced bias to the reported outcomes of effectiveness (59). Second, other active comparator treatment effect estimations are plausible for ABSSSI and can be assessed in future meta-analyses. Tedizolid, as an example, was in late-stage development for ABSSSI during the conduct of our systematic review and meta-analysis. Active comparator data for the linezolid treatment response, from the tedizolid phase 3 trial, was included in our meta-analysis (Fig. 2b), and a recent safety summary of tedizolid suggests supportive evidence of preclinical and clinical data for tedizolid treatment in ABSSSI (39, 60). Given the recent approval for use of tedizolid in ABSSSI, the tedizolid treatment effect estimation can be systematically examined for future trials that utilize tedizolid as the active comparator agent. Third, debate continues for the primary endpoint measures in ABSSSI clinical trials (9-11, 13, 15, 16, 20, 60-63). In alignment with the

regulatory guidance, we have categorized the early clinical treatment response at 48 to 72 h, when available, as well as the TOC clinical treatment response, when available. Both the timing of the TOC endpoint and the characterization of the clinical treatment response at this endpoint were limited by nonstandardized definitions across studies (Table 3). Fourth, the early outcome measure for ABSSSI recommended by the Foundation of the National Institutes of Health, of at least 20% reduction in total surface area at day 2 or day 3 versus baseline was not definitely reported in the trials that met our final inclusion criteria, and, therefore, cessation of spread at 48 to 72 h was substituted for this endpoint (20, 62, 63). The comparative early endpoint analysis for cessation of spread versus at least 20% reduction in total surface areas was, therefore, beyond the scope of our systematic review. Fifth, only three linezolid studies and one ceftaroline study reported the 48- to 72-h early clinical treatment response, and, hence, the small sample size affects the robustness of this measure of treatment response. Sixth, it is plausible that successful early clinical treatment responses in ABSSSI subjects (48 to 72 h) might potentially become failures at later efficacy time point measurements (7 to 14 days posttreatment). Future clinical trials and nonrandomized studies are needed to assess subjects at both early and later endpoints, in order to further understand the potential discordances between early successes and late failures and subsequent clinical implications (39, 45). Seventh, publication bias may exist, but assessment using funnel plot analysis was inconclusive for such bias. Last, we acknowledge potential secular changes in the standard of medical care and hygiene practices, which may have influenced treatment effect estimates from these historical studies.

In summary, the results of this systematic review and metaanalysis for the antibacterial treatment effect estimation for ABSSSI inform the design, execution, and analysis of future ABSSSI trials. The across-trials comparison enables noninferiority margin justification in the absence of placebo-controlled trials for ABSSSI. We have characterized the vast amount of clinical, methodological, and statistical heterogeneity for ABSSSI as reported in trials of proxy for placebo treatment as well as trials of linezolid and ceftaroline.

APPENDIX

Search terms for the systematic review for antibacterial treatment effect estimation for acute bacterial skin and skin structure infection. Publications were restricted to dates before 28 February 2013. Searches included abbreviations, plural, and full-phrase versions of the terms.

Search A1. For placebo-controlled trials for ABSSSI, the search terms were "placebo" combined with "SSTI," "SSSI," "ABSSSI," "cellulitis," "abscess," "wound infection," "burn infection," and "erysipelas."

Search A2. For uSSTI publications, the search terms were "placebo" combined with "SSTI," "SSSI," "ABSSSI," "impetigo," "abscess," "cellulitis," "furunculosis," "carbunculosis," "folliculitis," "traumatic lesion," "erysipelas," and "ecthyma."

Searches B1 and B2. The searches for linezolid and ceftaroline were conducted separately, with each drug name, respectively, combined with "SSTI," "SSSI," "ABSSSI," "cellulitis," "abscess," "wound infection," "burn infection," and "erysipelas."

Search C. For trials reporting nonantibacterial treatment response, the terms included all previously listed skin infection terms, along with a review of the available reports of nonantibacterial treatment in skin infections.

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The research study was conducted by all authors within our defined research, epidemiology (J.E.C., F.S.M.-G., L.M.M.), and statistical (G.L.) roles and scope of work for Quantitative Sciences and our antibacterial development program at GlaxoSmithKline (GSK). J.E.C. is a GSK-UNC research assistant, F.S.M.-G, is a GSK consultant, L.M.M. is a GSK employee and eligible for stock ownership, and G.L. was an employee at GSK, with stock options during the study execution; he is currently an employee at Pfizer.

REFERENCES

- Infectious Diseases Society of America. 2010. The 10 × '20 initiative: pursuing a global commitment to develop 10 new antibacterial drugs by 2020. Clin Infect Dis 50:1081–1083. http://dx.doi.org/10.1086/652237.
- 2. Boucher HW, Talbot GH, Bradley JS, Edwards JE, Gilbert D, Rice LB, Scheld M, Spellberg B, Bartlett J. 2009. Bad bugs, no drugs: no ESKAPE! An update from the Infectious Diseases Society of America. Clin Infect Dis 48:1–12. http://dx.doi.org/10.1086/595011.
- 3. European Centre for Disease Prevention and Control and the European Medicines Agency. 2009. Joint technical report: the bacterial challenge: time to react. Updated September 2009. ECDC/EMEA, London, United Kingdom. http://www.emea.europa.eu/pdfs/human/anti microbial_resistance/EMEA-576176-2009.pdf.
- Itani KM, Shorr AF. 2014. FDA guidance for ABSSSI trials: implications for conducting and interpreting clinical trials. Clin Infect Dis 58(Suppl 1):S4–S9. http://dx.doi.org/10.1093/cid/cit612.
- Moran GJ, Abrahamian FM, LoVecchio F, Talan DA. 2013. Acute bacterial skin infections: developments since the 2005 Infectious Diseases Society of America (IDSA) Guidelines. J Emerg Med 44:e397–e412. http: //dx.doi.org/10.1016/j.jemermed.2012.11.050.
- Dryden MS. 2010. Complicated skin and soft tissue infection. J Antimicrob Chemother 65(Suppl 3):iii35–iii44. http://dx.doi.org/10.1093/jac /dkq302.
- Johnson JK, Khoie T, Shurland S, Kriesel K, Stine OC, Roghmann MC, Jernigan JA, Harriman K, Harrison LH, Farley MM. 2007. Skin and soft tissue infections caused by methicillin-resistant *Staphylococcus aureus* USA300 clone. Emerg Infect Dis 13:1195–1200. http://dx.doi.org/10.3201 /eid1308.061575.
- Fridkin SK, Hageman JC, Morrison M, Sanza LT, Como-Sabetti K, Jernigan JA, Harriman K, Harrison LH, Farley MM, Active Bacterial Core Surveillance Program of the Emerging Infections Program Network. 2005. Methicillin-resistant *Staphylococcus aureus* disease in three communities. N Engl J Med 352:1436–1444. http://dx.doi.org/10.1056 /NEJMoa043252.
- U.S. Food and Drug Administration Division of Anti-Infective Products in the Center for Drug Evaluation and Research. 2013. Guidance for Industry. Acute bacterial skin and skin structure infections: developing drugs for treatment. Silver Spring, MD. http://www.fda.gov/downloads /Drugs/GuidanceComplianceRegulatoryInformation/Guidances /UCM071185.pdf.
- European Medicines Agency/Committee for Human Medicinal Products. 2013. Addendum to the guideline on the evaluation of medicinal products indicated for treatment of bacterial infections (EMA/CHMP/ 351889/2013). EMA/CHMP, London, United Kingdom. http://www.ema .europa.eu/docs/en_GB/document_library/Scientific_guideline/2013/11 /WC500153953.pdf.
- European Medicines Agency/Committee for Human Medicinal Products. 2012. Addendum to the note for guidance on evaluation of medicinal products indicated for treatment of bacterial infections to address indication-specific clinical data (CPMP/EWP/558/95 REV 2).

EMA/CHMP, London, United Kingdom. http://www.ema.europa.eu /docs/en_GB/document_library/Scientific_guideline/2009/09/WC50 0003417.pdf.

- Bassetti M, Baguneid M, Bouza E, Dryden M, Nathwani D, Wilcos M. 2014. European perspective and update on the management of complicated skin and soft tissue infections due to methicillin-resistant *Staphylococcus aureus* after more than 10 years of experience with linezolid. Clin Microbiol Infect 20(Suppl 4):3–18. http://dx.doi.org/10.1111/1469-0691 .12463.
- 13. Spellberg B, Talbot GH, Boucher HW, Bradley JS, Gilbert D, Scheld WM, Edwards J, Bartlett JG, Antimicrobial Availability Task Force of the Infectious Diseases Society of America. 2009. Antimicrobial agents for complicated skin and skin structure infections: justification of nonin-feriority margins in the absence of placebo-controlled trials. Clin Infect Dis 49:383–391. http://dx.doi.org/10.1086/600296.
- U.S. Food and Drug Administration. 2010. Guidance for industry noninferiority clinical trials. U.S. Food and Drug Administration, Silver Spring, MD. http://www.fda.gov/downloads/Drugs/Guidances/UCM202 140.pdf.
- Spellberg B. 2010. Skin and soft-tissue infections: modern evolution of an ancient problem. Clin Infect Dis 51:904–906. http://dx.doi.org/10.1086 /656432.
- Spellberg B. 2011. Acute bacterial skin and skin structure infection trials: the bad is the enemy of the good. Clin Infect Dis 53:1308–1309. http://dx .doi.org/10.1093/cid/cir741.
- Moher D, Simera I, Schulz KF, Hoey J, Altman DG. 2008. Helping editors, peer reviewers and authors improve the clarity, completeness, and transparency of reporting health research. BMC Med 6:13. http://dx.doi .org/10.1186/1741-7015-6-13.
- Moher D, Liberati A, Tetzlaff J, Altman DG. 2009. Preferred reporting items for systematic reviews and meta-analyses: The PRISMA statement. Ann Intern Med 151:264–269. http://dx.doi.org/10.7326/0003-4819-151 -4-200908180-00135.
- Liberati A, Altman DG, Tetzlaff J, Moher D. 2009. The PRISMA statement for reporting systematic reviews and meta-analyses of studies that evaluate health care interventions: explanations and elaboration. Ann Intern Med 151:W65–W94. http://dx.doi.org/10.7326/0003-4819-151-4 -200908180-00136.
- Corey GR, Stryjewski ME. 2011. New rules for clinical trials of patients with acute bacterial skin and skin-structure infections: do not let the perfect be the enemy of the good. Clin Infect Dis 52(Suppl 7):S469–S476. http://dx.doi.org/10.1093/cid/cir162.
- U.S. Food and Drug Administration. 2012. Guidance for industry: complicated urinary tract infections: developing drugs for treatment. U.S. Food and Drug Administration, Silver Spring, MD. http://www.fda.gov /downloads/Drugs/Guidances/ucm070981.pdf.
- Singh KP, Li G, Mitrani-Gold FS, Kurtinecz M, Wetherington J, Tomayko J, Mundy LM. 2013. A systematic review and meta-analysis of antimicrobial treatment effect estimation in complicated urinary tract infection. Antimicrob Agents Chemother 57:5284–5290. http://dx.doi.org /10.1128/AAC.01257-13.
- Snodgrass WR, Anderson T. 1937. Prontosil in erysipelas. Br Med J 2:101–104. http://dx.doi.org/10.1136/bmj.2.3993.101.
- Snodgrass WR, Anderson T. 1937. Sulphanilamide in the treatment of erysipelas. Br Med J 2:1156–1159. http://dx.doi.org/10.1136/bmj.2.4014 .1156.
- 25. Duong M, Markwell S, Peter J, Barenkamp S. 2010. Randomized, controlled trial of antibiotics in the management of community-acquired skin abscesses in the pediatric patient. Ann Emerg Med 55:401–407. http: //dx.doi.org/10.1016/j.annemergmed.2009.03.014.
- Llera JL, Levy RC. 1985. Treatment of cutaneous abscess: a double-blind clinical study. Ann Emerg Med 14:15–19. http://dx.doi.org/10.1016 /S0196-0644(85)80727-7.
- 27. Rajendran PM, Young D, Maurer T, Chambers H, Perdreau-Remington F, Ro P, Harris H. 2007. Randomized, double-blind placebo-controlled trial of cephalexin for treatment of uncomplicated skin abscesses in a population at risk for community-acquired methicillin-resistant *Staphylococcus aureus* infection. Antimicrob Agents Chemother 51:4044–4048. http://dx.doi.org/10.1128/AAC.00377-07.
- Petitti DB. 2001. Approaches to heterogeneity in meta-analysis. Stat Med 20:3625–3633. http://dx.doi.org/10.1002/sim.1091.
- Patsopoulos NA, Evangelou E, Ioannidis JPA. 2008. Sensitivity of between-study heterogeneity in meta-analysis: proposed metrics and empir-

ical evaluation. Int J Epidemiol 37:1148–1157. http://dx.doi.org/10.1093 /ije/dyn065.

- Higgins JPT. 2008. Commentary: heterogeneity in meta-analysis should be expected and appropriately quantified. Int J Epidemiol 37:1158–1160. http://dx.doi.org/10.1093/ije/dyn204.
- Berlin JA, Crowe BJ, Whalen E, Xia HA, Koro CE, Kuebler J. 2013. Meta-analysis of clinical trial safety data in a drug development program: answers to frequently asked questions. Clin Trials 10:20–31. http://dx.doi .org/10.1177/1740774512465495.
- 32. Higgins JPT, Green S (ed). March 2011. Cochrane handbook for systematic reviews of interventions, version 5.1.0. The Cochrane Collaboration. www.cochrane-handbook.org.
- DerSimonian R, Laird N. 1986. Meta-analysis in clinical trials. Control Clin Trials 7:177–188. http://dx.doi.org/10.1016/0197-2456(86)90046-2.
- DerSimonian R, Kacker R. 2007. Random-effects model for metaanalysis of clinical trials: an update. Contemp Clin Trials 28:105–114. http: //dx.doi.org/10.1016/j.cct.2006.04.004.
- 35. Tomayko JF, Li G, Breton JJ, Scangarella-Oman N, Dalessandro M, Martin M. 2013. The safety and efficacy of topical retapamulin ointment versus placebo ointment in the treatment of secondarily infected traumatic lesions: a randomized, double-blind superiority study. Adv Skin Wound Care 26:113–121. http://dx.doi.org/10.1097/01.ASW.0000427922 .12498.c4.
- Koning S, van der Wouden JC, Chosidow O, Twynholm M, Singh KP, Scangarella N, Oranje AP. 2008. Efficacy and safety of retapamulin ointment as treatment of impetigo: randomized double-blind multicentre placebo-controlled trial. Br J Dermatol 158:1077–1082. http://dx.doi.org/10 .1111/j.1365-2133.2008.08485.x.
- Colin M, Avon P. 1988. A double-blind comparative evaluation of a new topical antibacterial agent, mupirocine, and placebo in the treatment of skin and soft tissue infections. Pharmatherapeutica 5:198–203.
- Zaynoun ST, Matta MT, Uwayda MM, Kurban AK. 1974. Topical antibiotics in pyodermas. Br J Dermatol 90:331–334. http://dx.doi.org/10 .1111/j.1365-2133.1974.tb06411.x.
- Prokocimer P, De Anda C, Fang E, Mehra P, Das A. 2013. Tedizolid phosphate vs linezolid for treatment of acute bacterial skin and skin structure infections: the ESTABLISH-1 randomized trial. JAMA 309:559–569. http://dx.doi.org/10.1001/jama.2013.241.
- Noel GJ, Draper MP, Hait H, Tanaka SK, Arbeit RD. 2012. A randomized, evaluator-blind, phase 2 study comparing the safety and efficacy of omadacycline to those of linezolid for treatment of complicated skin and skin structure infections. Antimicrob Agents Chemother 56:5650–5654. http://dx.doi.org/10.1128/AAC.00948-12.
- 41. Covington P, Davenport JM, Andrae D, O'Riordan W, Liverman L, McIntyre G, Almenoff J. 2011. Randomized, double-blind, phase II, multicenter study evaluating the safety/tolerability and efficacy of JNJ-Q2, a novel fluoroquinolone, compared with linezolid for treatment of acute bacterial skin and skin structure infection. Antimicrob Agents Chemother 55:5790–5797. http://dx.doi.org/10.1128/AAC.05044-11.
- 42. Craft JC, Moriarty SR, Clark K, Scott D, Degenhardt TP, Still JG, Corey GR, Das A, Fernandes P. 2011. A randomized, double-blind phase 2 study comparing the efficacy and safety of an oral fusidic acid loading-dose regimen to oral linezolid for the treatment of acute bacterial skin and skin structure infections. Clin Infect Dis 52(Suppl 7):S520–S526. http://dx.doi.org/10.1093/cid/cir167.
- 43. Lin D, Zhang Y, Wu J, Wang F, Zheng JC, Sheng RY, Zhou X, Shen HH, Iizerman MM, Croos-Dabrera RV, Sheng W. 2008. Linezolid for the treatment of infections caused by Gram-positive pathogens in China. Int J Antimicrob Agents 32:241–249. http://dx.doi.org/10.1016 /j.ijantimicag.2008.04.004.
- 44. Kohno S, Yamaguchi K, Aikawa N, Sumiyama Y, Odagiri S, Aoki N, Niki Y, Watanabe S, Furue M, Ito T, Croos-Dabrera R, Tack KJ. 2007. Linezolid versus vancomycin for the treatment of infections caused by methicillin-resistant *Staphylococcus aureus* in Japan. J Antimicrob Chemother 60:1361–1369. http://dx.doi.org/10.1093/jac/dkm369.
- 45. Jauregui LE, Babazadeh S, Seltzer E, Goldberg L, Krievins D, Frederick M, Krause D, Satilovs I, Endzinas Z, Breaux J, O'Riordan W. 2005. Randomized, double-blind comparison of once-weekly dalbavancin versus twice-daily linezolid therapy for the treatment of complicated skin and skin structure infections. Clin Infect Dis 41:1407–1415. http://dx.doi.org /10.1086/497271.
- 46. Sharpe JN, Shively EH, Polk HC. 2005. Clinical and economic outcomes of oral linezolid versus intravenous vancomycin in the treatment of

MRSA-complicated, lower-extremity skin and soft-tissue infections caused by methicillin-resistant *Staphylococcus aureus*. Am J Surg **189:**425–428. http://dx.doi.org/10.1016/j.amjsurg.2005.01.011.

- 47. Weigelt J, Itani K, Stevens D, Lau W, Dryden M, Knirsch C, Linezolid Study Group CSSTI. 2005. Linezolid versus vancomycin in treatment of complicated skin and soft tissue infections. Antimicrob Agents Chemother 49:2260-2266. http://dx.doi.org/10.1128/AAC.49 .6.2260-2266.2005.
- Chen Y, Lee S, Kim W. 2004. Efficacy and tolerability of linezolid in treating severe skin and soft tissue infections caused by Gram-positive pathogens. J Formos Med Assoc 103:349–354.
- Stevens DL, Herr D, Lampiris H, Hunt JL, Batts DH, Hafkin B. 2002. Linezolid versus vancomycin for the treatment of methicillin-resistant *Staphylococcus aureus* infections. Clin Infect Dis 34:1481–1490. http://dx .doi.org/10.1086/340353.
- Stevens DL, Smith LG, Bruss JB, McConnell-Martin MA, Duvall SE, Todd WM, Hafkin B. 2000. Randomized comparison of linezolid (PNU-100766) versus oxacillin-dicloxacillin for treatment of complicated skin and soft tissue infections. Antimicrob Agents Chemother 44:3408–3413. http://dx.doi.org/10.1128/AAC.44.12.3408-3413.2000.
- 51. Corey GR, Wilcox MH, Talbot GH, Thye D, Friedland D, Baculik T, CANVAS 1 investigators. 2010. CANVAS 1: the first phase III, randomized, double-blind study evaluating ceftaroline fosamil for the treatment of patients with complicated skin and skin structure infections. J Antimicrob Chemother 65(Suppl 4):iv41–51.
- 52. Wilcox MH, Corey GR, Talbot GH, Thye D, Friedland D, Baculik T, CANVAS 2 investigators. 2010. CANVAS 2: the second phase III, randomized, double-blind study evaluating ceftaroline fosamil for the treatment of patients with complicated skin and skin structure infections. J Antimicrob Chemother 65(Suppl 4):iv53–65.
- Talbot GH, Thye D, Das A, Ge Y. 2007. Phase 2 study of ceftaroline versus standard therapy in treatment of complicated skin and skin structure infections. Antimicrob Agents Chemother 51:3612–3616. http://dx .doi.org/10.1128/AAC.00590-07.
- 54. Friedland HD, O'Neal T, Biek D, Eckburg PB, Rank DR, Llorens L, Smith A, Witherell GW, Laudano JB, Thye D. 2012. CANVAS 1 and 2: analysis of clinical response at day 3 in two phase 3 trials of ceftaroline fosamil versus vancomycin plus aztreonam in treatment of acute bacterial skin and skin structure infections. Antimicrob Agents Chemother 56: 2231–2236. http://dx.doi.org/10.1128/AAC.05738-11.
- 55. Guerrero-Beltrán JA, Barbosa-Cánovas GV. 2004. Review: advantages

and limitations on processing foods by UV light. Food Sci Technol Int 10:137–147. http://dx.doi.org/10.1177/1082013204044359.

- 56. U.S. Food and Drug Administration Center for Drug Evaluation and Research. 2013. Guidance for industry. Antibacterial therapies for patients with unmet medical need for the treatment of serious bacterial diseases. U.S. Food and Drug Administration, Silver Spring, MD. http://www .fda.gov/downloads/Drugs/GuidanceComplianceRegulatoryInformation /Guidances/UCM359184.pdf.
- U.S. Food and Drug Administration Center for Drug Evaluation and Research. 2014. Guidance for industry. Adaptive design clinical trials for drugs and biologics. U.S. Food and Drug Administration, Silver Spring, MD. http://www.fda.gov/downloads/Drugs/Guidances/ucm201790.pdf.
- 58. U.S. Food and Drug Administration Division of Anti-Infective Products in the Center for Drug Evaluation and Research. 2010. Draft guidance for industry. Acute bacterial skin and skin structure infections: developing drugs for treatment. U.S. Food and Drug Administration, Silver Spring, MD. http://www.fda.gov/downloads /Drugs/GuidanceComplianceRegulatoryInformation/Guidances /UCM071185.pdf.
- Jadad AR, Moore RA, Carroll D, Jenkinson C, Reynolds DJ, Gavaghan DJ, McQuay HJ. 1996. Assessing the quality of reports of randomized clinical trials: is blinding necessary? Control Clin Trials 17:1–12. http://dx .doi.org/10.1016/0197-2456(95)00134-4.
- Das D, Tulkens PM, Mehra P, Fang E, Prokocimer P. 2014. Tedizolid phosphate for the management of acute bacterial skin and skin structure infections: safety summary. Clin Infect Dis 58(Suppl 1):S51–S57. http://dx .doi.org/10.1093/cid/cit618.
- Shlaes DM, Sahm D, Opiela C, Spellberg B. 2013. The FDA reboot of antibiotic development. Antimicrob Agents Chemother 57:4605–4607. http://dx.doi.org/10.1128/AAC.01277-13.
- Toerner JG, Burke L, Komo S, Papadopoulos E. 2012. A collaborative model for endpoint development for acute bacterial skin and skin structure infections and community-acquired bacterial pneumonia. Clin Infect Dis 55:1122–1123. http://dx.doi.org/10.1093/cid/cis567.
- 63. Talbot GH, Powers JH, Fleming TF, Siuciak JA, Bradley J, Boucher H, Project Team CABP-ABSSSI. 2012. Progress on developing endpoints for registrational clinical trials of community-acquired bacterial pneumonia and acute bacterial skin and skin structure infections: update from the biomarkers consortium of the Foundation for the National Institutes of Health. Clin Infect Dis 55:1114–1121. http://dx.doi .org/10.1093/cid/cis566.