

NIH Public Access

Author Manuscript

Am J Med Genet B Neuropsychiatr Genet. Author manuscript; available in PMC 2011 April 5.

Published in final edited form as:

Am J Med Genet B Neuropsychiatr Genet. 2010 April 5; 153B(3): 802–811. doi:10.1002/ajmg.b.31050.

Linkage analyses of cannabis dependence, craving, and withdrawal in the San Francisco Family Study

Cindy L. Ehlers¹, Ian R. Gizer², Cassandra Vieten³, and Kirk C. Wilhelmsen²

¹ Department of Molecular and Integrative Neurosciences, The Scripps Research Institute, La Jolla, California

²Department of Genetics and Neurology and the Bowles Center for Alcohol Studies, University of North Carolina, Chapel Hill, North Carolina

³ California Pacific Medical Center Research Institute, San Francisco, California

Abstract

Cannabis is the most widely used illicit drug in the United States. There is ample evidence that cannabis use has a heritable component, yet the genes underlying cannabis use disorders are yet to be completely identified. This study's aims were to map susceptibility loci for cannabis use and dependence and two narrower cannabis-related phenotypes of "craving" and "withdrawal" using a family study design. Participants were 2524 adults participating in the University of California San Francisco (UCSF) Family Alcoholism Study. DSM-IV diagnoses of cannabis dependence, as well as indices of cannabis craving and withdrawal, were obtained using a modified version of the Semi-Structured Assessment for the Genetics of Alcoholism (SSAGA). Genotypes were determined for a panel of 791 microsatellite polymorphisms. Multipoint variance component LOD scores were obtained using SOLAR. Genome-wide significance for linkage (LOD > 3.0) was not found for the DSM-IV cannabis dependence diagnosis, however, linkage analyses of cannabis "craving" and the cannabis withdrawal symptom of "nervous, tense, restless or irritable" revealed five sites with LOD scores over 3.0 on chromosomes 1, 3, 6, 7, 9. These results identify new regions of the genome associated with cannabis use phenotypes as well as corroborate the importance of several chromosome regions highlighted in previous linkage analyses for other substance dependence phenotypes.

Keywords

dependence; genome scan; heritability; marijuana

Introduction

National epidemiological samples have demonstrated that cannabis is the most commonly used illicit drug [Anthony et al., 1994; Stinson et al., 2005, 2006], and that its use has substantially increased over the last decade [Compton et al., 2004]. Chronic use of cannabis is associated with both physical and mental health problems. Persistent use poses health problems similar to those of tobacco [Taylor et al., 2000; Mittleman et al., 2001; Fisher et al., 2005; Hashibe et al., 2005; Tashkin, 2005]. Cannabis use has also been implicated in a syndrome characterized by apathy, loss of goal-directed behavior, and cognitive impairment

Correspondence to: Dr. Cindy L. Ehlers, The Scripps Research Institute, 10550 North Torrey Pines Road, SP30-1501 La Jolla, CA 92037, cindye@scripps.edu.

termed the "amotivational syndrome" [Sharma, 1975; Pope et al., 2001; Solowij et al., 2002; Schuckit, 2006]. Cannabis use is associated with psychotic illness and depression [Degenhardt et al., 2003; Hall et al., 2004], as well as impaired educational and work performance [Kandel and Chen, 2000; Lynskey and Hall, 2000; Swift et al., 2001; Schuckit, 2006]. Use of cannabis, particularly by adolescents and young adults, may also facilitate progression to other illicit drug use (the "gateway" drug hypothesis) [Fergusson and Horwood, 2000; Lynskey et al., 2003].

Twin and family studies have consistently found that cannabis use and use disorders appear to in part have a genetic basis [for review see Agrawal and Lynskey, 2006]. Studies that have evaluated the role of genetic and environmental risk factors on cannabis abuse or cannabis dependence in twin samples have found estimates of heritability that range from 0.45 to 0.78 [see Kendler and Prescott, 1998; Tsuang et al., 1998; van den Bree et al., 1998; Maes et al., 1999; Kendler et al., 2000, McGue et al., 2000; Miles et al., 2001; Rhee et al., 2003]. Taken together, these studies suggest that there are substantial heritable influences on cannabis use disorders and that studies aimed at identifying genes that contribute to cannabis involvement may be warranted.

Disorders of cannabis use may be influenced by a number of genes that are difficult to detect because they may each have a small effect on the broad clinical phenotype. However, the genes influencing cannabis use disorders might be detected if they have a major effect on more narrowly defined phenotypes that more closely index the biological processes associated with addiction. Several biologically based theories have been suggested to explain the compulsion to use addictive drugs that might be profitably used to develop phenotypes useful for the study of cannabis use disorders. A general theory of addiction posits that the neurobiological mechanisms underlying the homeostatic regulation of appetitive drives and instincts becomes dysregulated during the process of drug exposure [Koob, 2000]. Some measures of the strength of this process include an increase or strong desire to take the drug often called 'drug craving' [see Anton, 1999], and ultimately the development of tolerance to and withdrawal from use of the drug [Schuckit, 1995]. Some studies have shown that requiring the presence of craving and withdrawal for the diagnosis of alcohol dependence leads to a better diagnostic distinction between abuse and dependence [De Bruijn et al., 2004]. In the case of cannabis dependence, recent data from large national epidemiologic samples have confirmed that cannabis withdrawal is prevalent, correlated with more symptoms of dependence, and associated with significant distress and impairment [Agrawal et al., 2008c; Hasin et al., 2008].

A few studies have conducted linkage analyses using cannabis dependence, cannabis dependence symptoms, and cannabis dependence subtypes as phenotypes [see Hopfer et al., 2007; Agrawal et al., 2008a,b; Ehlers et al., 2009], and some significant loci have been identified. Yet no studies to date have conducted linkage analyses using cannabis withdrawal and/or craving phenotypes. The aim of the present set of analyses was to identify loci associated with cannabis dependence, withdrawal and craving in the UCSF Family Alcoholism Study. The UCSF Family Alcoholism Study is a project that was designed to identify genetic loci that influence susceptibility to alcohol dependence and related phenotypes. It used a small family design, focusing primarily on sibling pairs and parent-child trios. The study enrolled 2454 individuals from 970 families from December 1995 through January 2003. Test-retest and inter-rater reliability for clinical data have been shown to be very good [Vieten et al., 2004]. Design, methods, recruitment strategies and sample demographics of the UCSF Family Alcoholism Study have been presented previously [see Seaton et al., 2004; Vieten et al., 2004], along with intrafamilial correlations for primary diagnostic phenotypes.

Methods

Participants

The UCSF family study recruited probands with lifetime DSM-IV alcohol dependence and the relatives of those probands, nation-wide. In addition, a community sample was ascertained in order to estimate population means and standard deviations for heritability and linkage analyses. Participants were recruited through the use of semi-targeted direct mail, a web site, press releases and advertisements and from alumni of treatment centers across the nation. Probands were invited to participate if they met screening criteria for alcohol dependence at some point in their lifetime and had at least one sibling or both parents available to participate in the study. With the permission of the proband, relatives were invited by mail to participate.

Probands reporting serious drug dependence (defined as use of stimulants, cocaine, or opiates daily for more than three months or weekly for more than six months), and those who reported any history of intravenous substance use were excluded. Probands were excluded if, upon screening, they reported a current or past diagnosis of schizophrenia, bipolar disorder, or other psychiatric illness involving psychotic symptoms (those with depressive and anxiety disorders were accepted); a life-threatening illness; or an inability to speak and read English.

Two thousand one hundred and fifty-four individuals were enrolled in the UCSF Family Alcoholism Study. The sample had a mean age of 48.8 ± 13.2 years, a mean educational level of 14.4 ± 2.9 years, and a mean annual income of $57,356 \pm 54,656$ (median, 545,000). Racial distribution was 92% Caucasian, 3% each African American and Hispanic, and 1% each Native American and other. No attempt was made to exclude or over sample minorities. Probands were 58% female. Relatives of probands were 38% alcohol dependent.

An unselected general population sample of 147 individuals was also recruited to assess phenotype base rates. Letters were sent to residents of the same geographical areas as the family samples, requesting participation in a study on "health behaviors and characteristics" to avoid a sample biased toward participation in a study on alcoholism. No inclusion/ exclusion criteria were applied aside from the ability to respond to the telephone interview and complete the questionnaires. The details of recruitment of all participants have been previously published [see Seaton et al., 2004; Veiten et al., 2004].

Clinical measures and diagnoses

A remote data collection procedure was developed allowing for blood samples and other questionnaires to be returned by mail, and structured diagnostic interviews to be conducted by telephone, making nationwide data collection possible. Potential participants first had the study explained and gave written informed consent. A modified version of the Semi-Structured Assessment for the Genetics of Alcoholism (SSAGA) [Bucholz et al., 1994] was used to collect demographic, medical, psychiatric, alcohol, nicotine, cannabis, and other drug use history. The SSAGA is a fully structured, poly-diagnostic psychiatric interview that has undergone both reliability and validity testing. Modifications of the SSAGA for the present study included changes in diagnostic criteria and items to approximate DSM-IV criteria. Only the sections of the SSAGA relevant to substance abuse, demographics and medical history were utilized. We have previously demonstrated that a number of cannabis usage phenotypes are significantly heritable including: having ever used cannabis, used more than 21 times in a single year, DSM-IV cannabis dependence, individual criteria of DSM-IV cannabis dependence [Ehlers et al., unpublished work]. Two phenotypes with the highest heritability were cannabis "craving" and withdrawal. Therefore, the phenotypes chosen for the present linkage analyses were: (1) a DSM-IV cannabis dependence diagnosis; (2) cannabis "craving" defined as endorsing: "In situations where you couldn't use marijuana, did you ever have such a strong desire for it that you couldn't think of anything else"; and (3) a measure of cannabis withdrawal defined as: "Did stopping or cutting down after a period of regular use of marijuana ever cause you to feel nervous, tense, restless or irritable". The number of participants with each phenotype, with and without co-morbid alcohol dependence, is presented in table 1. Twenty percent of those enrolled did not complete all study requirements. Details of the interview process and inter-rater reliability of the SSAGA in this study have been reported previously [Vieten et al., 2004].

DNA collection and genotyping

The DNA extraction procedure and genotyping protocol have been previously described [Wilhelmsen et al., 2003]. Briefly, DNA was isolated from whole blood using a commercial kit (Gentra, Minneapolis, MN), and genotypes for a panel of microsatellite polymorphisms were generated using fluorescently labeled polymerase chain reaction (PCR) primers (HD5, version 2.0; Applied Biosystems, Foster City, CA). The HD5 panel set consisted of 811 markers with an average marker-to-marker distance of 4.6 cM (maximum, 14 cM) and an average heterozygosity of greater than 77%. A small subset of markers was omitted from the panel because of null alleles, irregular allele spacing or other problems with reproducibility. None of the omitted markers were adjacent to other omitted markers. The sizes of the PCR products for the markers were determined from electropherograms produced with an ABI 3700 (Applied Biosystems, Foster City, CA), a 96-channel capillary electrophoresis device.

The sizes of marker amplimers were determined (blinded to pedigree structure and subject characteristics) from the electropherogram using the Genotyper software package (ABI). All electropherograms were visually inspected and exported from Genotyper in base pair sizes relative to the standard measured to one hundredth of a base pair. Fragment sizes were binned to alleles by using an automated algorithm developed by one of the authors (K.C.W.), which assumes that the distribution of allele sizes will have a sine-squared distribution with a fitted periodicity near two base pairs. The program determines the best periodicity and phase for the modeled distribution relative to the observed distribution. Fragments that are distributed between minimums of the modeled distribution are assumed to be the same allele. Allele frequencies observed in the founders were used for all analysis. The sex-averaged marker map order obtained from the manufacturer was used and verified with the family data from the current sample.

The genotypes for all of the autosomal markers were analyzed for each family by using Pedigree Relationship Statistical Test (PREST) [see McPeek and Sun, 2000] to detect sample and pedigree structure errors. DNA was re-isolated from a stored frozen blood specimen, and the genotyping was repeated for any individual for whom PREST detected a probable error. If regenotyping failed to resolve the error, the problematic genotype was subsequently treated as missing. The program Pedcheck was used to detect non-Mendelian inheritance [O'Connell et al., 1998]. Markers with a high frequency of Mendelian segregation errors were excluded from analysis, and for isolated Mendelian errors, the genotypes for the entire family were excluded for the specific marker that yielded the error. To further reduce errors, the probability that each genotype was correct was assessed by using the error-checking algorithm implemented in Merlin [Abecasis et al., 2002], in which genotypes that had a probability of less than 0.025 of being correct were removed from further consideration.

Data analyses

Both genotype and phenotype data were available for 1647 individuals, and phenotype but not genotype data was available for an additional 720 individuals. Of these 720 individual

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87 were relatives that could not b e contacted or did not provide DNA samples and 462 were probands that did not provide a sufficient number of genetically informative relatives for inclusion and the remaining 171 subjects were the relatives of these probands. Seven hundred and thirteen families were considered genetically informative for the heritability analyses. Families that contained sibling, half-sibling, avuncular or cousin pairs were included as being potentially genetically informative. These families ranged in size from 3 to 20 subjects (average 4.63 ± 2.13). The data includes: 563 parent-child, 1085 sibling, 40 half-sibling, 17 grandparent-grandchild, 238 avuncular, and 32 cousin relative pairs. An additional 177 families contained only a single individual with phenotype data. These individuals were included within some analyses to the extent that they contribute information about trait means and variance and the impact of covariates.

Analyses were previously conducted to estimate the heritability of the three phenotypes of interest: DSM-IV cannabis dependence, cannabis craving, and irritable/anxious during cannabis withdrawal using SOLAR [http://solar.sfbrgenetics.org/] [Almasy and Blangero, 1998] [Ehlers et al., unpublished work]. Participant's age at the time of evaluation and sex were evaluated as potential covariates and retained if they accounted for at least 5% of the total variance. The total additive genetic heritability (h²) and its standard error were estimated, and the probability that h² was greater than zero was determined using a Student's t-test for each scale, and all three phenotypes were found to be heritable and as such suitable for linkage analyses.

The variance components method implemented in SOLAR [http://solar.sfbrgenetics.org/] was then used to calculate multipoint LOD scores across the genome at 1 cM intervals for the three cannabis phenotypes. All traits were analyzed under a continuous trait model rather than using the latent threshold model as the former approach provided a more conservative estimate of linkage in the present study. A LOD threshold of 3.0 was used to identify loci that provided evidence for linkage, and a LOD threshold of 2.0 was used to identify loci yielding suggestive evidence of linkage. For each reported linkage peak, simulation analyses were then conducted using SOLAR to derive nominal p-values. Specifically, a genetic locus was simulated under the null hypothesis of no linkage across 20,000 trials, and the distribution of LOD scores across those trials was used to derive a nominal p-value for the observed LOD score.

Because the current sample was selected for alcohol dependence, it should be noted that prevalence rates for the cannabis dependence diagnoses and respective symptoms in the unselected control sample were estimated and included in the tested models to correct for ascertainment bias when calculating h^2 and linkage. In addition, the high marker density of the HD5 microsatellite panel allows for the extraction of a large proportion of IBD information among relative pairs as participants' genotypes can be almost completely phased, thus reducing the reliance on population allele frequencies to estimate allele-sharing probabilities. As a result, and given the very low number of ethnic minority participants in the UCSF sample, we chose to include all participants in the analyses without correcting for ethnic background.

Results

Demographics

The demographic characteristics of this population have been presented previously [see Vieten et al., 2004]. In brief, the UCSF family sample had a mean age of 48.4 ± 13.4 years, a mean educational level of 14.4 ± 2.9 years, and an annual income of $$57,356 \pm $54,656$. Racial distribution was: 92% Caucasian, 3% each African American and Hispanic, and 1% each Native American and other. Fifty-four percent were married. Probands were 58%

female, with 97% of probands meeting lifetime criteria for alcohol dependence, whereas, 38% of the relatives of probands were found to be alcohol dependent. The random sample had similar demographic characteristics to the Family Study sample with 15% having a lifetime history of DSM-IV alcohol dependence. The prevalence rates for DSM-IV lifetime cannabis dependence were .16 for probands and .07 for the random sample. Prevalence rates for cannabis withdrawal and cannabis craving were .09 in the probands and .03 in the random sample.

Linkage analyses

Analyses of multipoint variance component LOD scores for the dichotomous DSM-IV cannabis dependence phenotype did not yield any peaks that achieved genome-wide significance. However, two sites were identified with LOD scores over 2.0, a site on chromosome 1 between 142 and 159 cM with a peak at 154 cM (LOD = 2.1), and a site on chromosome 2 between 194 and 216 cM with a peak at 211 cM (LOD = 2.6). These findings are presented in Figure 1 and Table 2.

Linkage analyses of cannabis "craving" e.g. endorsing: "In situations where you couldn't use marijuana, did you ever have such a strong desire for it that you couldn't think of anything else" and the cannabis withdrawal symptom of: "Did stopping or cutting down after a period of regular use of marijuana ever cause you to feel nervous, tense, restless or irritable" revealed 5 sites with LOD scores over 3.0. The symptom of cannabis craving yielded the strongest evidence for linkage (LOD = 5.7) on chromosome 7 between 70 and 83 cM with a peak at 79 cM. At that same site there was suggestive evidence of linkage within that same support interval with a peak at 71 cM for the withdrawal phenotype (LOD = 2.2) as illustrated in Figure 2.

The symptom of cannabis craving also yielded strong evidence for linkage on chromosome 3 between 164 and 176 cM with a peak at 171 cM (LOD = 4.4). There was additional evidence for linkage for cannabis craving on chromosome 1 between 36 and 48 cM with a peak at 43 cM (LOD = 3.6), and on chromosome 6 between 195 an 202 cM with a peak at 202 cM (LOD = 3.2). There was suggestive evidence on chromosome 9 between 25 an 44 cM with a peak at 35 cM (LOD = 2.6), and on chromosome 15 between 75 and 84 cM with a peak at 81 cM (LOD = 2.3).

The symptom "nervous, tense, restless or irritable" during cannabis withdrawal also demonstrated evidence for linkage on chromosome 9 between 167 and 177 cM with a peak at 174 cM (LOD = 3.6). Additionally, suggestive evidence for linkage for this withdrawal phenotype was found on chromosome 3 between 89 and 102 cM with a peak at 96 cM (LOD = 2.5) and on chromosome 7 between 65 and 90 cM with a peak at 71cM (LOD = 2.2) (see Table 2 for complete results). Nominal p-values generated by simulation analyses are presented for all three cannabis phenotypes in table 3.

Secondary analyses were conducted in order to determine whether these genome locations were specific only to the "nervous, tense, restless or irritable" symptom of withdrawal or whether the "sleeplessness" and a "number of withdrawal symptoms count" phenotype could also produce similar LOD scores in those locations. Evidence for linkage on chromosome 9 was found for the "sleeplessness" phenotype at 158 cM (LOD = 4.2) and the symptom count phenotype (LOD = 3.2). Additionally, suggestive evidence for linkage for the sleeplessness (LOD = 2.6) and symptom count (LOD = 3.0) phenotype was found on chromosome 3 with a peak at 95 cM. Thus it appears that similar regions in the genome may harbor genes for several withdrawal symptoms.

In order to determine whether the sites identified for the cannabis phenotypes were mainly due to comorbid alcohol dependence a separate linkage analysis was conducted using alcohol dependence as a covariate and few differences were found. The results of those analyses are presented in table 4. The linkage results for cannabis dependence, craving and withdrawal were also compared to the results obtained from previous linkage scans for alcohol and nicotine dependence that were also conducted in the UCSF family study population. None of the linkage regions reported in a previous linkage scan for alcohol dependence [Gizer et al., 2009] yielded neither significant nor suggestive linkage in a subsequent scan for nicotine dependence [Gizer et al., unpublished work] in the UCSF family study population. However, in the present set of analyses, suggestive evidence for linkage to cannabis dependence was found at a site on chromosome 2 previously identified in a scan for nicotine dependence [Gizer et al., unpublished work], and another site identified for cannabis craving on chromosome 9 in the present study was also identified in a previous scan for alcohol dependence [Gizer et al., 2009] in the UCSF family study population. These results provide evidence suggesting that two of the regions identified in the present set of analyses may also confer risk towards nicotine and alcohol dependence in this population.

Discussion

Several studies suggest that there is a moderate genetic influence on cannabis dependence, and recently there have been additional reports that have identified regions in the genome that may be linked to cannabis dependence [see Kendler and Prescott, 1998; Tsuang et al., 1998; Maes et al., 1999; Miles et al., 2001; Lynskey et al., 2002; Kendler et al., 2003; Rhee et al., 2003; Wilhelmsen and Ehlers, 2005; Agrawal and Lynskey, 2006; Hopfer et al., 2007; Agrawal et al., 2008a,b]. Only two sites, one on chromosome 1 and one on chromosome 2, were identified in the present study for DSM-IV cannabis dependence. The site on chromosome 1 at 154 cM (LOD = 2.1) does not appear to be within 30 cM of other sites identified using substance dependence phenotypes. However, this peak was between two sites; one at around 197 cM that was identified by Dick et al. [2002] in the Collaborative Study for the Genetics of Alcoholism (COGA) dataset for a quantitative alcohol-related phenotype and one at around 102 cM for a cannabis problems factor score in an autosomal linkage scan for cannabis use disorders in the nicotine addiction genetics project [Agrawal et al, 2008a].

The site identified on chromosome 2 near 211 cM (LOD = 2.6) for DSM-IV cannabis dependence in the UCSF Family Alcoholism Study appears to be in the same general region (within 30 cM) as a site identified in the COGA dataset for comorbid alcohol dependence and major depression [Nurnberger et al., 2001], and for subjective response to alcohol [Schuckit et al., 2001] using microsatellite markers and for alcohol dependence using SNP genotyping [Agrawal et al., 2008d]. It also appears to be near a site for alcohol dependence identified in multiplex families in Pittsburgh [Hill et al., 2004], and for heavy cocaine use [Gelernter et al., 2005] and opioid dependence [Gelernter et al., 2006] in African Americans. Additionally, this site was identified by a previous genome scan for nicotine dependence in the UCSF family study [Gizer et al., unpublished work]. Thus, this site appears to harbor genes that confer risk for a number of different substance-related phenotypes.

It has been suggested that the effort to identify genetic factors and the mechanisms whereby they influence addiction may be aided by the use of phenotypes that may be more closely related to the biological processes underlying risk for use disorders [Gottesman and Gould, 2003]. One phenotype that most DSM-IV substance dependence syndromes have in common is a withdrawal syndrome defined as a combination of physiologic and psychological processes that occurs after cessation of its use [Edwards, 1986; Weisbeck et

al., 1996; Hasin et al., 2006, 2008]. Although withdrawal is not required to meet DSM-IV criteria for drug/alcohol dependence it is associated with a more severe clinical course and a poorer prognosis [Schuckit et al., 1999; Budney et al., 1999, 2003, 2004, ²⁰⁰⁸; Budney and Hughes, 2006; Hasin et al., 2000, 2008; Ehlers et al., 2004a]; and as such, has been shown to be more heritable than the clinical diagnosis itself in some populations [Ehlers et al., 2004b]. Another phenotype that is common to many drugs of abuse is craving. Human and animal studies have demonstrated that craving is an important element in the addictive process and that control of craving may improve efforts at abstinence [see Wise, 1988; Robinson and Berridge, 1993; Anton, 1999; Sinha and O'Malley, 1999; Field et al., 2004; Heishman and Singleton, 2006; Haughey et al., 2008]. Evaluation of the heritability of cannabis withdrawal (h² =0.28, p<0.001) and craving (h² =0.36, p<0.0001) demonstrated that these two phenotypes were more heritable than the DSM-IV diagnosis of cannabis dependence (h² =0.20, p<0.0001) in the UCSF family study population [Ehlers et al, unpublished work]. Additionally, five sites in the genome that provided evidence for linkage to these phenotypes were identified.

Two of the chromosome sites that were identified in the UCSF family study population for cannabis craving and cannabis withdrawal were on chromosomes 3q and 9q. These locations have been identified previously in genome scans for cannabis and other drug dependence phenotypes [see Stallings et al., 2003, 2005; Hopfer et al., 2007]. Hopfer and colleagues [2007] conducted a genome wide scan for loci influencing adolescent cannabis dependence symptoms in 324 sibling pairs from 192 families. In that study, probands (52.1% of whom were EuroAmerican, 36.5% of whom were Hispanic, and 7.8% of whom were African-American) were identified from consecutive admissions to substance abuse treatment facilities. The authors found evidence for suggestive linkage on chromosomes 3q21 (LOD = 2.61) and 9q34 (LOD = 2.57). Using the same population of adolescents, Stallings et al. [2003, 2005], conducted linkage scans for the average number of dependence symptoms, conduct disorder symptoms and a composite index of antisocial substance dependence, and also found evidence for linkage to the chromosome 9q34 region (LOD = 3.15) and 3q24-3q25 (LOD = 3.27) for the composite index. The site on chromosome 3q was within 30 cM of a site identified for antisocial personality disorder/conduct disorder in an American Indian community [Ehlers et al., 2008]. The site identified on chromosome 3g for cannabis craving in the UCSF family study (LOD = 4.5) was also within 30 cM of the site identified on that chromosome by Stallings et al. [2005]. Additionally, a site on chromosome 9q (LOD=3.5) for cannabis withdrawal in the UCSF family study was also within 30 cM of the site identified on that chromosome by Stallings et al. [2005]. Additionally, a site on chromosome 9q (LOD=3.5) for cannabis withdrawal in the UCSF family study was also within 30 cM of the site identified on that chromosome by Stallings et al., [2005]. Taken together these findings suggest that these sites on chromosome 3q and 9q may harbor genes that confer risk for severe phenotypes associated with drug dependence including antisocial symptoms.

The region of the genome that provided the best evidence for linkage in the UCSF family study was on chromosome 7q (LOD = 5.6) for the cannabis craving phenotype. This region has consistently emerged with significant evidence of linkage in the COGA project. In the initial COGA sample this region on chromosome 7q provided the strongest evidence of linkage to alcohol dependence with a multipoint LOD score of 3.49 [Reich et al., 1998]. An additional linkage analysis with expanded microsatellite markers in the full COGA sample revealed a LOD score of 2.9 [Wang et al., 2004], and a set of SNPs genotyped in the region that were not in linkage disequilibrium with adjacent markers generated a LOD score at the peak of 4.1 [Dunn et al., 2005]. Other population samples have also found evidence for linkage on 7q. A report of linkage to this region on 7q has been made by the Nicotine Addiction Genetics project for alcohol consumption phenotypes [Agrawal et al., 2008a], and

for alcohol dependence in a sample of multiplex families ascertained at Pittsburgh [Hill et al., 2004]. A systematic SNP screen to fine map this region on chromosome 7 using the COGA dataset identified an association with *ACN9* a gene involved in gluconeogenesis and the assimilation of ethanol or acetate into carbohydrate [Dick et al., 2008].

There were several areas of the genome that were reported previously as being linked to cannabis use that were not identified in the current linkage scan for cannabis dependence phenotypes in the UCSF Family study. For instance, Agrawal and colleagues [2008d] conducted a linkage scan in a sample from the COGA study for six DSM-IV cannabis dependence criteria considered as a continuous variable (0-6) and found a maximum LOD score of 1.9 at 95 cM on chromosome 14. A site on chromosome 14 at 122 cM was also found for a severe cannabis subtype generated by a cluster analysis in an American Indian community [Ehlers et al., 2009]. This region on chromosome 14 was also identified previously in a genome-wide linkage analysis for ASPD/CD in that American Indian community [Ehlers et al., 2008]. Two areas of the genome were also identified in that American Indian community study with LOD scores that provide evidence for linkage for a severe cannabis use/antisocial subtype on chromosome 16 (@139 cM, LOD score = 4.4), and on chromosome 19 (@74 cM, LOD score = 6.4) [Ehlers et al., 2009]. There have also been some findings in genome scans for cannabis dependence and other drug use related phenotypes on chromosome 19. A region has been identified on chromosome 19 at 17 cM for early-onset cannabis use, and for frequency of use on chromosome 18, in a study of 2314 Australian families [Agrawal et al., 2008b]. A site on chromosome 4 for a cannabis problems factor score has been identified on chromosome 4 in the region of the gamma aminobutyric acid type A gene cluser with a LOD score of 2.2 in the Nicotine Addiction Genetics Project [Agrawal et al., 2008a].

In summary, data from the UCSF Family study did not reveal evidence for genome-wide significance for linkage (LOD > 3.0) to the DSM-IV cannabis dependence diagnosis, however, linkage analyses of cannabis "craving" and the cannabis withdrawal symptom of "nervous, tense, restless or irritable" revealed five sites with LOD scores over 3.0 on chromosomes 1, 3, 6, 7, 9. These results have identified new regions of the genome associated with cannabis use phenotypes as well as corroborated the importance of several chromosome regions highlighted in previous linkage analyses for other substance dependence phenotypes. The results of this study should, however, be interpreted in the context of several limitations. First, multiple phenotypes were tested in the present study, thus increasing the possibility that a false positive result was interpreted as substantive. To protect against this possibility, empirically-derived p-values were reported for each linkage signal allowing for the objective evaluation of the linkage evidence. Second, the findings may not generalize to other large studies that were not ascertained for alcoholic probands. Third, only retrospective and cross-sectional data on cannabis use and use disorders were assessed. Fourth, comparisons to other large samples may be limited by differences in a number of genetic and environmentally determined variables. The exclusion criteria for this study eliminated participants reporting serious drug dependence on stimulants, cocaine, or opiates, current diagnosis of schizophrenia, bipolar disorder, or other psychiatric illness involving psychotic symptoms. This exclusion was included to create a more homogeneous sample for genetic analyses. However, this limitation reduces external validity and clinical applicability since the co-occurrence of mental health disorders and substance abuse is common. Additionally, this sample was comprised of almost entirely Caucasians that also limits applicability to other ethnic groups.

Supported in part by funds from the State of California for medical research on alcohol and substance abuse through the University of California at San Francisco, the Ernest Gallo Clinic and Research Center (KCW), the National Institute of Alcohol Abuse and Alcoholism grants T32 AA007573 (KCW) and AA010201 and Clinical Translational Studies U54 RR0250204. (CLE).

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Figure 1.

Multipoint Linkage Analysis for the Cannabis dependence (upper graph), Cannabis craving (middle graph) and Cannabis withdrawal (lower graph) phenotypes for the entire genome. Results for each chromosome are aligned end to end with the p terminus on the left. Log of the Odds (LOD) score is plotted on the Y-axis. Vertical lines indicate the boundaries between the chromosomes. The numbers above on the X-axis indicate the chromosome number.



Figure 2.

Multipoint Linkage Analysis for Cannabis dependence (dotted line), Cannabis craving (solid gray line) and Cannabis withdrawal (solid black line) phenotype for chromosome 7. The analysis assumes a latent normally distributed variable with a threshold above which an individual is affected. Log of the Odds (LOD) score (Y-axis) is plotted for the chromosome location map (in centimorgans (cM), X-axis). Nearest markers to the peak are presented within the support interval.

Table 1

Number of subjects with and without Alcohol Dependence that were genotyped for the linkage analysis of MJ phenotypes

| | No AlcDep Dx | Yes AlcDep Dx | Total |
|----------------------|--------------|---------------|-------|
| DSM-IV MJ Dependence | 763 | 850 | 1613 |
| MJ Withdrawal | 761 | 850 | 1611 |
| MJ Craving | 754 | 847 | 1601 |

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Table 2

| study |
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| Chr | Phenotype | LOD | Chr loc | Nearest Marker(s) | Supporting reference, phenotype |
|-----|---------------------|-----|-------------------|-------------------|--|
| 1 | Cannabis cravin | 3.6 | 43cM (36-48cM) | D1S199-D1S2864 | |
| | Cannabisdependence | 2.1 | 154cM (142-159cM) | D1S498 | |
| 7 | Cannabis dependence | 2.6 | 211cM (194-216cM) | D2S2361 | Numberger et al., (2001) alcohol dep and depression; Schuckit et al., (2001) subjective response to alcohol; Hill et al., (2004) alcohol dep; Gelernter et al., (2005, 2006) cocaine, opiod dep; Agrawal et al., (2008d), alc dep; Gizer et al.,(unpublished work), nicotine dep |
| 3 | Cannabis withdrawal | 2.5 | 96cM (89-102cM) | D3S1566-D3S3681 | Stallings et al., (2003), substance dep; |
| | Cannabis craving | 4.4 | 171cM (164-176cM) | D3S1279-D3S3668 | Enters et al., (2005), ASTD/CD; Hopfer et al., (2007) cannabis dep |
| 9 | Cannabis craving | 3.2 | 202cM (195-202cM) | D6S281 | |
| 7 | Cannabis withdrawal | 2.2 | 71cM (65-90cM) | D7S506 | Reich et al., (1998) alcohol dep; |
| | Cannabis craving | 5.7 | 79cM (70-83cM) | D7S502-D7S2476 | Agrawat et al., (2004) alcohol dep Hill et al., (2004) alcohol dep |
| 6 | Cannabis craving | 2.6 | 35cM (25-44cM) | D9S157 | Gizer et al., (unpublished work) alcohol dep; |
| | Cannabis withdrawal | 3.6 | 174cM (167-177cM) | D9S1838 | Atamings et au, (2005), substance dep, Hopfer et al., (2007) cannabis dep |
| 15 | Cannabis craving | 2.3 | 81cM (75-84cM) | D15S127 | Dick et al., (2002) alcohol dep subtype; Ehlers et al., (2004b) alcohol withdrawal |
| | | | | | |

| Table 3 |
|---|
| Nominal p-values for cannabis phenotypes generated by simulation analyses |

| Chr | Loc | Phenotype | LOD | Nominal p-value |
|-----|-------|-----------------------------|-----|-----------------|
| 1 | 43cM | Cannabis craving | 3.6 | 0.0077 |
| | 154cM | Cannabis Dependence | 2.1 | 0.004 |
| 2 | 211cM | Cannabis Dependence | 2.6 | 0.0016 |
| 3 | 96cM | Cannabis | 2.5 | 0.0041 |
| | 171cM | Withdrawal Cannabis craving | 4.4 | 0.0031 |
| 6 | 202cM | Cannabis craving | 3.2 | 0.0109 |
| 7 | 71cM | Cannabis | 2.2 | 0.0062 |
| | 79cM | Withdrawal Cannabis craving | 5.7 | 0.0007 |
| 9 | 35cM | Cannabis craving | 3.6 | 0.0077 |
| | 174cM | Cannabis Withdrawal | 3.6 | 0.0007 |
| 15 | 81cM | Cannabis craving | 2.3 | 0.0249 |

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Table 4

Linkage results for MJ phenotypes with and without Alcohol Dependence included as a covariate

| Chr | Phenotype | Chr loc (cM) | LOD | LOD w/AlcDen as covar | Difference |
|-----|------------|--------------|-----|-----------------------|------------|
| 1 | Craving | 43 | 3.6 | 3.5 | -0.1 |
| | Dependence | 154 | 2.1 | 2.6 | +0.5 |
| 7 | Dependence | 211 | 2.6 | 2.4 | -0.2 |
| 3 | Withdrawal | 96 | 2.5 | 2.0 | -0.5 |
| | Craving | 171 | 4.4 | 4.4 | 0 |
| 9 | Craving | 202 | 3.2 | 3.3 | +0.1 |
| 7 | Withdrawal | 71 | 2.2 | 2.6 | +0.4 |
| | Craving | 79 | 5.7 | 6.1 | +0.4 |
| 6 | Craving | 35 | 3.6 | 2.4 | -1.2 |
| | Withdrawal | 174 | 3.6 | 3.7 | +0.1 |
| 15 | Craving | 81 | 2.3 | 2.1 | -0.2 |