# AMERICAN THORACIC SOCIETY DOCUMENTS

## An Official American Thoracic Society Workshop Report: Translational Research in Rare Respiratory Diseases

Arnold S. Kristof, Basil J. Petrof, Qutayba Hamid, Martin Kolb, Jennifer S. Landry, Alex MacKenzie, Francis X. McCormack, Inga J. Murawski, Joel Moss, Frank Rauch, Ivan O. Rosas, Adam J. Shapiro, Benjamin M. Smith, David Y. Thomas, Bruce C. Trapnell, Lisa R. Young, and Maimoona A. Zariwala; on behalf of the ATS Assembly on Respiratory Cell and Molecular Biology

THIS OFFICIAL WORKSHOP REPORT OF THE AMERICAN THORACIC SOCIETY WAS APPROVED MAY 2017

### Abstract

Rare respiratory diseases (RRDs) are a heterogeneous group of disorders that collectively represent a significant health care burden. In recent years, strong advocacy and policy initiatives have led to advances in the implementation of research and clinical care for rare diseases. The development of specialized centers and research networks has facilitated support for affected individuals as well as emerging programs in basic, translational, and clinical research. In selected RRDs, subsequent gains in knowledge have informed the development of targeted therapies and effective diagnostic tests, but many gaps persist. There was therefore a desire to identify the elements contributing to an effective translational research program in RRDs. To this end, a workshop was convened in October 2015 with a focus on the implementation of effective transnational research networks and collaborations aimed at developing novel diagnostic and therapeutic tools. Key elements included an emphasis on molecular pathogenesis, the continuing engagement of patient advocacy groups and policy makers, the effective use of preclinical models in the translational research pipeline, and the detailed phenotyping of patient cohorts. During the course of the workshop, current logistical and knowledge gaps were identified, and new solutions or opportunities were highlighted.

The October 2015 Rare Respiratory Diseases Workshop was supported in part by the Meakins-Christie Laboratories and the Translational Research in Respiratory Diseases Program at the Research Institute of the McGill University Health Centre.

Correspondence and requests for reprints should be addressed to Arnold S. Kristof, M.D., Meakins-Christie Laboratories and Program for Translational Research in Respiratory Diseases, Research Institute of the McGill University Health Centre, 1001 Décarie Boulevard, Montreal, PQ, H4A 3J1, Canada. E-mail: arnold.kristof@mcgill.ca

This article has an online supplement, which is accessible from this issue's table of contents at www.atsjournals.org

Ann Am Thorac Soc Vol 14, No 8, pp 1239–1247, Aug 2017 Copyright © 2017 by the American Thoracic Society DOI: 10.1513/AnnalsATS.201705-406WS Internet address: www.atsjournals.org

Contents	Section 3: Preclinical Models and	Using biomarkers to identify
Methods	Mechanisms of Disease	disease in presymptomatic
Section 1: The Organization of	Cell-based assays and	patients
Translational Research in	chemical screening	Phenotyping by unbiased
RRDs	Murine transgenic models	biomarker discovery
Section 2: Advances in Clinical	for diseases of known	Phenotyping to discover rare
Infrastructure for Translational	genetic etiology	variants of common diseases
Research in RRDs	Murine models for RRDs	Genotype phenotype
Registries and natural	with no unique molecular	correlation
history studies	etiology	Section 5: New Opportunities:
Longitudinal cohorts that	Section 4: Phenotyping and	Respiratory Manifestations of
encompass the pediatric-	Biomarker Development	Rare Neuromusculoskeletal
to-adult spectrum	Phenotyping by respiratory	Diseases
Clinical trials	function	Summary

Rare or orphan diseases are defined as those with a prevalence of less than 1 in 2,000, or less than 200,000 cases in the United States. When combined, they affect approximately 5 to 8% of the population, but these disorders have been underrepresented in the spectrum of health care services delivery, as well as in the distribution of resources for clinical and translational research. Rare respiratory diseases (RRDs) make up an important subset (1, 2), and respiratory manifestations have a significant impact on patients' quality of life, function, and mortality. Although prevalence estimates vary widely, RRDs are estimated to affect up to 3 million persons in each of Europe and the United States (3). Recent reviews and statements (3-5) have identified gaps in the implementation of effective research and care delivery for patients with RRDs. The discovery and evaluation of biological tools for clinical use (i.e., translational research) were identified as priorities. To accelerate advances in the field, there was a perceived need to formalize the approach to effective translational research infrastructures and methodologies in RRDs, using selected RRDs as illustrative examples. The purpose of this document is to summarize discussion from a workshop convened in October 2015 at which the optimization of translational research in RRDs was addressed, in addition to current gaps and emerging opportunities.

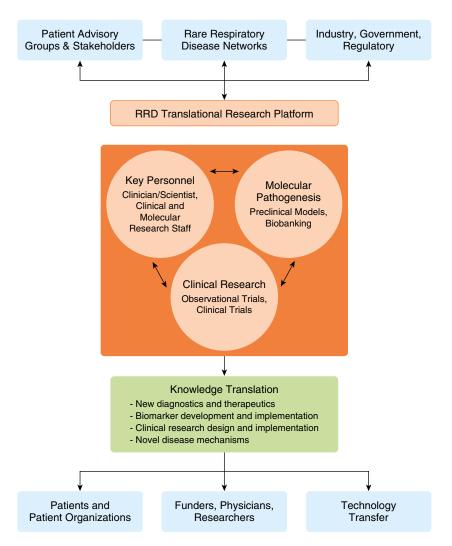
## Methods

An RRD workshop was held in Montreal, Quebec, Canada, on October 22-23, 2015. The goal was to build upon existing knowledge based on RRD research and provide consensus on its optimal organization and required infrastructure. The planning group and workshop were chaired by A.S.K., B.J.P., and Q.H. Workshop participants were chosen for their leadership roles in existing RRD research networks and programs, and they included representatives of patient advisory groups and local networks, patients with RRDs, clinician scientists, respiratory therapists, nurses, researchers, and trainees. Speakers were responsible for reviewing emerging infrastructures, methodologies, and areas of need in facilitating translational research in RRDs. Each

workshop speaker contributed to the writing of this article. The workshop consisted of individual sections that covered five research-driven themes. All planning committee members and presenters completed a Declaration of Potential Conflict of Interest Form prior to the workshop.

### Section 1: The Organization of Translational Research in RRDs

The goal of translational research is to exploit knowledge gained from experimental or clinical models to develop new diagnostics or therapeutics for patients



**Figure 1.** Optimizing translational research platforms for rare respiratory diseases (RRDs). Effective translational research on RRDs requires an integrated infrastructure that interfaces with critical stakeholders. Patient advisory groups and referral centers form a critical entry point and represent end users in the research pipeline (*blue*). The development of patient cohorts and recruitment into observational studies or clinical trials is facilitated by dedicated centers that participate in organized networks (*orange*). Key personnel (i.e., clinician scientists, clinical research coordinator/nurse, core experts in molecular pathophysiology) manage complex, multisystem chronic diseases, sample and data collection and analysis, as well as communicate with stakeholders and network partners. Understanding of disease pathogenesis is crucial, and the research exploits preclinical models, access to biobanks and registries, detailed clinical and phenotypic analysis in observational studies, and incorporation of diagnostic and prognostic biomarkers. Obstacles in RRD trial design include small sample sizes, chronicity, and limited effects over time. Emerging techniques include adaptive designs, N-of-1 and crossover trials, the incorporation of diagnostic or prognostic biomarkers, and clinical surrogate markers (e.g., the use of baseline decline in lung function prior to the intervention).

(6) (Figure 1). Traditional therapeutic development pathways are focused on screening-based technologies to identify new compounds and biomarkers and to target large populations for phase III clinical trials and postmarketing analysis. Although these populations are small for RRDs, the genetic or molecular defect is often known and can be exploited to define accessible therapeutic targets. This also permits the parallel development of biomarkers that serve as strong surrogate outcome variables in clinical trials. For instance, studies in model organisms to elucidate the role of the mechanistic target of rapamycin (mTOR) in the control of cell growth (7) complemented the interrogation of patient-derived biopsy samples for aberrant mTOR signaling and its role in the pathogenesis of lymphangioleiomyomatosis (LAM) (8). Within 10 years, a phase III clinical trial demonstrated the efficacy of the mTOR inhibitor rapamycin (sirolimus) in improving lung function and quality of life for patients with LAM (9, 10), and it helped establish clinical guidelines for the medical community (11, 12). In pulmonary alveolar proteinosis, studies on granulocytemacrophage colony-stimulating factor signaling in isolated macrophages informed the characterization of granulocytemacrophage colony-stimulating factor deficiency in patients and led to the development of diagnostic bioassays and replacement therapies (13). Similarly, studies on the genetic and molecular bases of cystic fibrosis (CF) have identified molecular targets for cystic fibrosis transmembrane conductance regulator (CFTR) corrector or potentiator therapies (14, 15).

RRD advocates have implemented innovative policy initiatives and research infrastructures to address the shortfalls in available clinical tools. One key step was the incorporation of all rare diseases under umbrella organizations that represent common clinical and research needs. In the United States, the National Institutes of Health Office of Rare Diseases was established in 1993, in part because of efforts by the National Organization for Rare Disorders advocacy group. The Rare Diseases Clinical Research Network (251 institutions worldwide and >200 rare diseases) was established to create disease-specific consortia; cost-sharing arrangements for research infrastructures; uniform data acquisition protocols; and an umbrella organization for disease registries,

cohort studies, and randomized controlled trials. Additional missions include the direct engagement of patients and their advocates, as well as the training of new investigators. As a platform, the network facilitates clinical and translational research by establishing standard operating procedures for data collection, coordinating patient recruitment for large-scale studies, and providing a data management and coordinating center that supports study design and data analysis.

The Rare Lung Diseases Consortium is a subset of the Rare Diseases Clinical Research Network that collaborates with patient advocacy groups and supports the inclusion of those with no existing advocacy vehicle. The National Center for Advancing Translational Sciences was established to support the preclinical and clinical development of drugs and diagnostics for rare diseases. Similar advocacy organizations operate globally, including those represented at the workshop (see Table E1 in the online supplement). Moreover, a formalized, consortium-based approach to translational research in rare diseases of pediatric onset has been successfully implemented (16). Therefore, rare disease research networks, in collaboration with patient advocacy groups and policy makers, serve as critical drivers and coordinators of clinical and translational research.

## Section 2: Advances in Clinical Infrastructure for Translational Research in RRDs

Successful clinical and translational research platforms for RRDs benefit from networkbased recruitment of patients into prospective and longitudinal cohort studies that ideally encompass the pediatric-to-adult spectrum. The collection and analysis of granular (i.e., detailed) datasets allows for precise phenotyping, which supports research on basic disease mechanisms and the design of clinical trials by developing robust surrogate physiological or molecular outcomes (i.e., biomarkers). The elements of the translational research backbone discussed are described in the subsections below.

## Registries and natural history studies

Registries are data collection systems suited for tracking outcomes in chronic diseases that can be accurately defined and rely on voluntary provision of secondary patient data from multiple centers to central

repositories. Estimates of disease prevalence, practice patterns, and health care use can be inferred. However, the accuracy, homogeneity, and timeliness of clinical data cannot be guaranteed, and coordination for banking of biological samples is challenging. Natural history studies can be designed to collect a highly granular clinical and biological dataset over time. The quality and consistency of the data are optimized by centralization of protocols, banking repositories, and consistent measurement and analytic techniques. Genetic and molecular determinants can be tracked over time and correlated with clinical outcomes. Given the relative homogeneity of the patient populations, the collection of multiple data parameters over time can reduce bias and permit inference from retrospective analyses.

For instance, a National Institutes of Health-funded registry and natural history study were initiated in parallel. Registry data described the clinical spectrum of LAM (17). The NHLBI LAM natural history study was designed to frequently assess the evolution of disease and included the collection of biological samples that could be linked to clinical data. As a result, mTOR, along with multiple other putative therapeutic targets, was validated as an important disease modifier (18). Isolation of circulating tumor cells revealed that the abnormal cells in LAM exhibit metastatic properties (19). A number of biomarkers were identified in the blood (e.g., vascular endothelial growth factor-D [VEGF-D]) (20) and correlated with surrogate physiological or radiological outcomes. Detailed clinical and molecular phenotyping identified subsets of patients with differences in prognosis (e.g., rate of decline in  $FEV_1$ ) or response to treatment (e.g., VEGF-D), which can inform the interpretation and design of translational or clinical studies.

## Longitudinal cohorts that encompass the pediatric-to-adult spectrum

Because most rare diseases begin in childhood, the identification of early determinants of disease progression and developmental trends in longitudinal cohorts using adaptable study designs is desirable. Potential challenges include their long duration and expense, subject attrition, and requirement for large sample sizes. Attrition can be especially problematic during the transition from pediatric to adult care. However, these are often minimized in RRD research by the availability of

molecular testing and biomarkers, as well as the involvement of patient advisory groups in research networks. Illustrative successes and gaps were discussed for infant respiratory distress syndrome and bronchopulmonary dysplasia (BPD), which occur most commonly in preterm infants and with effects spanning the pediatric-toadult continuum. Gaps include lack of genetic and molecular markers of disease progression and suboptimal characterization of lung function over time. Potential solutions include the establishment of pediatric-to-adult transition and RRD clinics that minimize the loss to follow-up that occurs as patients exit pediatric care models and facilitate multidisciplinary translational research studies. For instance, the maximum potential  $FEV_1$ , as defined by the peak  $FEV_1$ before decline begins (approximately age 24 yr), may differ among RRDs and could serve as an important surrogate endpoint for future decline or for assessing molecular and genetic markers of disease progression. In a cohort of 21-year-old patients with premature birth and BPD, the median maximum FEV1 was 80% of predicted and was associated with greater airflow obstruction and bronchial hyperreactivity than in those subjects without BPD (21). The detailed characterization of transition cohorts would inform the development of surrogate markers of disease and the evaluation of novel therapeutic modalities.

Primary ciliary dyskinesia (PCD) is another RRD that begins in childhood and leads to chronic upper and lower respiratory infections caused by loss of ciliary function and defective mucociliary clearance (22). Little is known regarding the progression of lung function abnormalities into adulthood, and studies have thus far been limited by suboptimal pediatric-to-adult transition follow-up. The Genetic Disorders of Mucociliary Clearance Consortium and the PCD Foundation are currently focused on prospective evaluation of disease heterogeneity and genotype-phenotype correlation throughout the pediatric-toadult spectrum (see section 4). Successes include establishing nasal nitric oxide as an effective screening test for PCD, evaluating inhaled hyperosmolar agents as a potential therapy, and identifying novel mutations in ciliary structure/function genes (see section 4) (23). The PCD Foundation has recently sponsored clinical guidelines for diagnosis

and management (23), which promote awareness in the community and referral to tertiary clinical and translational research centers.

### **Clinical trials**

As controlled experiments, randomized clinical trials can be leveraged to discover and evaluate novel disease mechanisms and biomarkers. The Multicenter International Lymphangioleiomyomatosis Efficacy of Sirolimus (MILES) trial investigators evaluated the efficacy of sirolimus for the prevention of lung function decline in patients with LAM (10). VEGF-D levels were elevated in patients with LAM but not in those with other cystic lung diseases (24), indicating its potential use as a diagnostic biomarker. In the MILES trial, the authors demonstrated a correlation between VEGF-D levels and decline in lung function or response to treatment (9). Researchers in future trials may use elevated VEGF-D levels as an enrollment criterion to both target the most appropriate patient population and optimize statistical power. Key points discussed are summarized in Figure 1.

## Section 3: Preclinical Models and Mechanisms of Disease

The use of preclinical cell-based and animal models represents a crucial component in the translational research pipeline. Although they may not fully replicate human disease, preclinical models can be employed to (1) test mechanistic hypotheses generated from clinical observations; (2) discover new mechanisms of disease initiation, susceptibility, or progression; (3) conduct screens for novel therapies; and/or (4) identify or validate biomarkers and test the effectiveness of therapeutic approaches. In addition, these models can incorporate environmental factors (e.g., nutrition, stress) that interact with genetic determinants. Given the emphasis on molecular pathogenesis in RRDs, these models are well suited to complement RRD translational research platforms. Selected examples were discussed, and these are summarized below.

## Cell-based assays and chemical screening

Translational investigators in CF, a proteintrafficking disease, have conducted cell-based

drug screens using CFTR activity or subcellular localization as endpoints to identify potential "potentiator" or "corrector" drugs, respectively (14). Medium- to high-throughput ratiometric and electrophysiological cell-based assays (OPatch; Sophion Bioscience, Woburn, MA) were used to screen for CFTR correctors before validation of potential leads in standard epithelial ion transport assays. A primary airway cell biobank was established (Trans Canadian Network Initiative and McGill University Cystic Fibrosis Translational Research Centre) with seven distinct CF mutations represented, permitting the validation and testing of lead compounds and derivatives in patient-derived cultures. This platform was used for the preclinical development of latonduine, a proteostasis modulator and bone fide CF $\Delta$ 508 corrector (25).

More broadly, systems biology and personalized medicine approaches provide a strong rationale for lead development and subsequent clinical trials. Genome-wide transcriptional profiling of CF cell lines with known CFTR mutations can complement chemical screens that both identify mutation-specific expression profiles and screen for drugs that may modulate these responses.

Stem cell and tissue engineering technologies can also be used to establish preclinical models and represent the developmental and genetic backgrounds of the aberrant disease-causing cell. Induced pluripotent stem cells can be engineered from patients before undergoing lineagedependent differentiation to propagate cell cultures. In one study, induced pluripotent stem cell technology was used to re-create macrophages originally derived from a patient with hereditary pulmonary alveolar proteinosis (26). Reduced surfactant clearance in hereditary pulmonary alveolar proteinosis macrophages could be corrected by lentiviral expression of wild-type CSF2RA, thereby complementing the endogenous mutated form. This model can be used for drug screening, mechanistic studies, or transplant therapy (27).

# Murine transgenic models for diseases of known genetic etiology

Murine models can be used to explore mechanisms of disease susceptibility arising from known human genetic mutations. Mutations in the *HPS1* gene cause Hermansky-Pudlak syndrome, in which affected individuals develop severe pulmonary fibrosis. Although *Hps*-transgenic models do not spontaneously develop disease, they are more susceptible to injury-induced (e.g., bleomycin) fibrosis via a mechanism that requires epithelial cell apoptosis, as well as the subsequent profibrotic activation of alveolar macrophages (28–31). Genetic mutations seen in human subjects can be replicated in mouse models to explore mechanisms of disease and reveal additional cell-based or molecular therapeutic targets.

# Murine models for RRDs with no unique molecular etiology

The absence of a defined disease-causing genetic mutation and the inability to fully reproduce pathological features of the human disease in mice present significant challenges. In idiopathic pulmonary fibrosis (IPF), the initiating event is not known, and murine models of fibrosis may use a variety of artificial induction agents to reproduce the pathophysiology. The routine use of inbred mice reduces experimental variability but does not address the genetic heterogeneity in human cohorts. Often, the timing or context of model initiation (e.g., age or sex of mice), interventions, or measurement of endpoints is not applicable to the clinical setting. Chronicity of the phenotype may hinder the use of fully representative experimental models because of animal care and use ethical guidelines, experimental complexity, or resource limitations. In IPF, researchers in translational studies have nonetheless attempted to focus on mechanisms of fibrosis and resulting loss of pulmonary function. These include aging-related susceptibility factors, aberrant epithelial cell function, and factors leading to excessive wound healing and lung remodeling (32). Murine models have been refined significantly by the use of repetitive induction (e.g., bleomycin [33]) and adenoviral gene transfer (e.g., adenoviral vector expressing human transforming growth factor- $\beta$  [34]), as well as *in vivo* lineage tracing and stem cell technologies that permit the assessment of individual cell types over time (35). Contemporary murine studies incorporate chronicity, factors that initiate or promote fibrosis, interventional trial design, and clinically relevant endpoints that render the models translatable and amenable to

the development of diagnostic biomarkers and therapeutics. As a result, translational animal experiments can complement successful large-scale human phase III studies (36, 37). Key points discussed are summarized in Table 1.

# Section 4: Phenotyping and Biomarker Development

RRDs often exhibit significant phenotypic heterogeneity, which can be exploited to better understand mechanisms of disease or to design clinical studies that incorporate the analysis of outcomes in population subgroups. Modern high-throughput technologies permit large-scale DNA sequencing, which can link clinical phenotypes to a defined molecular etiology. Workshop participants discussed the paradigms described below to illustrate the utility of population phenotyping in RRDs and how this informs the development of well-defined surrogate endpoints for the effective design of clinical trials.

## Phenotyping by respiratory function

The collection of granular physiological data over time (section 2) is crucial for understanding patient heterogeneity in RRDs. Patients with LAM exhibit different rates of decline in  $FEV_1$  and diffusing capacity of the lung for carbon monoxide

## Table 1. Preclinical models

## Key Messages

- Preclinical cell-based and animal models represent a critical component of the translational pipeline in RRDs.
- New technologies, improved study design, and the incorporation of chronicity permit the establishment of preclinical models that enhance the translational research effort even if they do not precisely replicate the human disease.
- Induced pluripotent stem cells and *in* vivo cell lineage tracking are promising technologies to characterize pathogenesis from early stages of disease.

over time, which correlates directly with disease-related morbidity (38). These surrogate endpoints can be exploited to identify and characterize other biomarkers of disease severity, as well as to differentiate between responders and nonresponders to therapy. The surrogate endpoint approach can greatly reduce the sample size in clinical trials. Importantly, clinical trials in which changes in lung function in the placebo or treatment arms exhibit variability may yield negative results even if many of the patients respond to the intervention. In one study of patients with Hermansky-Pudlak syndrome, a salutary effect of pirfenidone on lung function did not achieve statistical significance; patient selection by prior rate of decline in lung function might have resulted in positive results, and pirfenidone could have been approved for this indication (39). Finally, novel clinical trial designs (e.g., N-of-1, crossover, adaptive) can complement the use of surrogate physiological endpoints in optimizing clinical and translational protocols (40, 41).

# Using biomarkers to identify disease in presymptomatic patients

Another major challenge is to identify patients at risk for developing clinically significant disease so that prognostic and therapeutic tools can be employed early on. For instance, screening computed

**Current Challenges** 

#### The development of appropriate animal models to study the effect of genetic variation and/or environmental factors on disease initiation or progression

- The establishment of single-center or networked translational research platforms that incorporate patient care, cohort development, and preclinical investigations in the same pipeline to support multidirectional interactions in development of diagnostics and therapeutics
- The formulation of standard protocols and endpoints for rapidly assessing the regulatory requirements for entry into human clinical trials

Definition of abbreviation: RRD = rare respiratory disease.

tomographic (CT) scans or serum VEGF-D levels in patients with tuberous sclerosis (i.e., at risk for LAM) might permit the identification and prognostication of patients who subsequently develop LAM. In a recent study with patients deemed at risk for familial pneumonitis syndromes on the basis of screening relatives of affected patients, investigators identified panels of clinical and molecular markers that predict the onset of CT scan abnormalities (42). These types of phenotyping studies can greatly facilitate the classification of patients in RRD research protocols.

## Phenotyping by unbiased biomarker discovery

RRD networks and patient-based advocacy groups have contributed to the successful collection of biological samples with linked clinical data from natural history studies, registries, and clinical trials. Samples can be interrogated for genetic, transcriptomic, or molecular biomarkers in an unbiased fashion. In IPF, clinical outcomes were associated with molecular "endotypes" that reflect epithelial dysfunction and senescence, innate immune responses, and aberrant lung remodeling (43). Genetic, transcriptomic, and proteomic patterns can be used as quantitative surrogate endpoints for stratification in clinical trials and as clinical tools for prognostication (i.e., personalized medicine). This was illustrated in a study of *N*-acetylcysteine for IPF: A single-nucleotide polymorphism (rs3750920) in the TOLLIP gene predicted improved clinical outcomes in patients with IPF receiving N-acetylcysteine, whereas clinical outcomes were worse in those with the alternate allele (44, 45). In separate studies, transcriptomic patterns in peripheral blood mononuclear cells and telomere length have been associated with disease progression and/or acute exacerbations (46, 47). The enrichment of cohorts with potential responders to treatment might be achieved using biomarker panels that discriminate between IPF and other forms of interstitial pneumonitis (48) or those who are predicted to exhibit biological responses to therapy.

## Phenotyping to discover rare variants of common diseases

The detailed phenotyping of common diseases can reveal rare variants that

potentially shed important insights into disease pathogenesis. For instance, the interrogation of DNA from families with pulmonary fibrosis led to the identification of candidate genes for therapeutics and biomarker development (49). Detailed CT image analysis of patients within chronic obstructive pulmonary disease cohorts (i.e., the Multi-Ethnic Study of Atherosclerosis Lung Study) revealed that specific airway segmental branching variants were more common in patients with chronic obstructive pulmonary disease (B. M. Smith, oral communication, October 2015). These anatomical variants were in turn associated with single-nucleotide polymorphisms in a number of candidate genes, which encode protein mediators of lung development. Current studies addressing mechanisms of disease in respective mutant mice, as well as the natural history of obstructive airway disease in patients with these mutations, might provide important insights into factors that determine susceptibility to the initiation or progression of common airway diseases.

### Genotype-phenotype correlation

Genotype-phenotype correlation is a promising approach to understanding mechanisms of disease and to classifying patients within cohorts that are genetically and phenotypically heterogeneous. A translational approach was formalized by the Genetic Disorders of Mucociliary Clearance Consortium and the PCD Foundation. Significant heterogeneity in the clinical features of PCD can arise from age of presentation, the presence and nature of anatomical lateralization defects, the severity of airway obstruction, upper airway infections, infertility, and gastrointestinal dysfunction (50, 51). Current diagnostic modalities are focused on ciliary structure and function, but they are often cumbersome and subjective; fail to capture the full spectrum of disease; and, especially in young children, do not effectively distinguish PCD from other syndromes with similar clinical presentations (e.g., CF). The complexity of ciliary composition amplifies the number of potential causative genes. Until 2008, identification of the first eight genes associated with PCD relied largely upon candidate gene-based approaches using model organisms and gene mapping by linkage analysis. Profiling of the two initially discovered genes

(i.e., DNAI1, DNAH5) identified mutations in nine exons that were used in the first diagnostic test for PCD. Since 2009, the pace of discovery has been accelerated by next-generation sequencing technologies. Currently, 35 genes are known to be associated with PCD, accounting for approximately two-thirds of all PCD cases. Emerging insights into associations between genotype and phenotype are reviewed elsewhere (52). For example, there is a strong association between genotype and changes in ciliary structure and/or function, and nasal nitric oxide levels are usually low. DNAH11 mutations, however, were observed in patients with normal ciliary ultrastructure (53). Patients harboring RSPH1 mutations exhibited a milder pulmonary phenotype, and symptoms became prevalent with age (54). The identification of all disease-causing mutations in PCD is now feasible using current sequencing technologies, and correlations with phenotype will permit the optimization of counseling, prognostication, and design of future clinical and translational studies. Now commercially available, next-generation sequencing panels will also allow identification of cases that require whole-exome sequencing for novel gene discoveries. Key points discussed are summarized in Table 2.

## Section 5: New Opportunities: Respiratory Manifestations of Rare Neuromusculoskeletal Diseases

Translational RRD research centers can leverage growing international patient cohorts, the power of patient advocacy, and expertise in preclinical and clinical research that addresses lung pathophysiology and respiratory clinical outcomes. Workshop organizers proposed that RRD translational research platforms can investigate rare neuromusculoskeletal diseases in which respiratory manifestations account for significant morbidity and mortality. As is the case for other multisystem disorders that significantly affect the lung (e.g., CF, tuberous sclerosis complex), RRD translational research programs could develop clinical cohorts and preclinical models, as discussed above, that inform the development of diagnostics and therapeutics for improving the

#### Table 2. Phenotyping and biomarker development

#### **Key Messages**

- Simultaneous innovations in physiological, imaging-based, molecular, and genetic biomarker discovery have greatly improved patient care and facilitated the design of clinical trials using surrogate endpoints.
- A common infrastructure for pediatric and adult RRDs could support bidirectional preclinical and clinical investigations, as well as mechanisms to screen for early markers of disease in otherwise asymptomatic patients.
- Detailed phenotypic analysis of common disease cohorts might reveal otherwise unknown rare diseases that provide translatable genetic and molecular insights.

#### **Current Challenges**

- Health disparities in the overall population related to age, race, sex, socioeconomic class, or geographical location complicate effective patient care and the recruitment of fully representative cohorts. Health disparities research might shed light on mechanisms to account for phenotypic and genetic variation in these populations.
- Despite the establishment of clinical networks, subcontracting and institutional review board agreements represent significant hurdles in conducting interinstitutional research. The formulation of widely adopted standard operating protocols, in addition to the use of centralized, multicenter processes, represents a promising approach.

Definition of abbreviation: RRD = rare respiratory disease.

quality of life and survival of these patients.

Several examples of rare genetically inherited and metabolic myopathies that can present primarily with respiratory manifestations in adults were discussed (e.g., myotonic dystrophy, mitochondrial myopathy, McArdle disease) (55). Pompe disease, a glycogenosis caused by autosomal recessive mutations in the acid  $\alpha$ -glucosidase (GAA) gene (56), was highlighted as a rare neuromuscular disorder that disproportionately affects the diaphragm. Studies indicate a correlation between GAA levels and the age of onset, as well as progression of respiratory muscle dysfunction. Exercise limitation, sleepdisordered breathing, and aspiration pneumonia are also associated with increased morbidity and mortality. Recombinant human GAA therapy slows the loss of pulmonary function (57), but diagnosis and treatment are often delayed. In addition, detailed, long-term, respirationfocused physiological and biological markers of disease progression and response to established or novel therapies are lacking. These can potentially be developed in the context of multinational collaborations,

special needs RRD clinics, and translational research platforms.

Rare disorders of bone formation and aberrant matrix production can also result in chronic respiratory failure and sleep-disordered breathing on the basis of chest wall and/or parenchymal restriction. Osteogenesis imperfecta (OI) is a genetically inherited disease of bone fragility that is usually caused by mutations in the collagen type I genes (COL1A1, COL1A2), with typical skeletal and extraskeletal manifestations (58). Translational research efforts in OI have been championed by the Osteogenesis Imperfecta Foundation and the Shriners Hospitals for Children Network. Consistent genotype-phenotype correlation and sensitivity of genetic testing have defined a rich cohort in which to study natural history and potential therapies. Little is known regarding changes in lung physiology and gas exchange over time in patients with OI, but respiratory dysfunction appears to be a frequent cause of death (59, 60). A crude analysis of 92 adults suggested a correlation between scoliosis and reduced lung function when corrected for arm span (61). Based on spirometry, there

was evidence of obstructive airway abnormalities in a significant proportion of patients, even in the absence of scoliosis, suggesting direct involvement of the lung. The detailed characterization of pulmonary function and gas exchange over time in existing preclinical OI models and genetically well-defined clinical cohorts may provide important insights into the evolution of pulmonary disease in patients with OI or other diseases of chest wall restriction.

## Summary

Translational research in RRDs has led to an explosion in novel diagnostic and therapeutic approaches for previously neglected patients. Optimal RRD translational research programs require a coordinated and multidisciplinary approach that relies on an integrated infrastructure. Effective translational research programs in RRDs are managed by dedicated clinicians/scientists and coordinators who facilitate communications and collaboration with transnational research networks, patient advocacy groups, and policy makers. Preclinical models of RRDs and patient cohort phenotyping are critical in determining the link between molecular pathophysiology and disease classification, respectively. The development of screening assays and surrogate physiological or biological markers facilitates drug development and the design of clinical trials, as well as screening/diagnosis and disease monitoring. Current gaps include the detailed characterization of subjects transitioning from pediatric to adult care; the coordination of special needs multidisciplinary clinics; regulatory hurdles impeding interinstitutional collaboration and drug or diagnostics development; and novel clinical trial designs that overcome chronicity, heterogeneity, and effects of cointerventions. Finally, it was proposed that translational research platforms in RRDs, which include multisystem disorders that impact significantly upon the lung, could incorporate subjects with rare neuromusculoskeletal diseases and potentially lead to the conduct of research that ultimately has a positive impact on their quality of life and survival.

This official Workshop Report was prepared by an *ad hoc* committee of the Assembly on Respiratory Cell and Molecular Biology.

Members of the subcommittee are as follows: ARNOLD S. KRISTOF, M.D. (Chair) QUTAYBA HAMID, M.D., PH.D. (Co-Chair) BASIL J. PETROF, M.D. (Co-Chair) MARTIN KOLB, M.D., PH.D. JENNIFER S. LANDRY, M.D. ALEX MACKENZIE, M.D., Ph.D. FRANCIS X. MCCORMACK, M.D. JOEL MOSS, M.D., PH.D. INGA J. MURAWSKI, PH.D. FRANK RAUCH, M.D. IVAN O. ROSAS, M.D. ADAM J. SHAPIRO, M.D. BENJAMIN M. SMITH. M.D. DAVID Y. THOMAS, PH.D. BRUCE C. TRAPNELL, M.D. LISA R. YOUNG, M.D. MAIMOONA A. ZARIWALA, PH.D.

Author disclosures: Q.H. served on an advisory committee for Teva Pharmaceuticals; received research support from Novartis and GlaxoSmithKline. B.J.P. served as a speaker and received research support from Genzyme. M.K. served as a speaker, consultant, on an

advisory committee, on a data and safety monitoring board, and received research support from Boehringer Ingelheim, F. Hoffman-La Roche/Roche/Intermune, and GlaxoSmithKline; served as a consultant, on an advisory committee, and received research support from Gilead and Prometic; served as a consultant and received research support from Janssen; served on an advisory committee AstraZeneca and Genoa; received research support from Actelion and Sanofi. F.X.M. served as a consultant for LAM Therapeutics; served on an advisory committee and a data and safety monitoring board for Takeda; received research support from Novartis and Pfizer: served on a clinical trial. Multicenter International LAM Efficacy of Sirolimus (Miles); holds an intellectual property on patent 8278060 for VEGF-D. F.R. served on an advisory committee for Alexion and Ultragenyx; received research support from Novartis. I.O.R. served on an advisory committee for Patients Like Me and Kadmon Pharmaceuticals; served as a speaker for Genentech and Pri-Med; holds an intellectual property on a patent for biomarkers of IPF from Celgene; received research support from Stomedix/Biogen, Intermune, Gilead, and Bristol-Meyers Squibb. A.J.S. received research support from Parion

Pharmaceuticals and Vertex Pharmaceuticals. D.Y.T. served as company director and employee for Traffic Therapeutics Inc. and Triple R International Inc.; received other payments from Amorchem Holdings Inc.; received research support from GlaxoSmithKline. B.C.T. received travel support from Takeda; received research support from aTyr Pharma. L.R.Y. has intellectual property in royalties from UpToDate and a patent for VEGF-D. M.A.Z. received research support from Thermo-Fisher Scientific and Life Technologies Corp. A.S.K., J.S.L., A.M., J.M., I.J.M., and B.M.S. reported no relationships with relevant commercial interests.

Acknowledgment: J.M. was supported by the Intramural Research Program, NIH, NHLBI. The authors thank all the patients, Alpha-1 Canada (Jim Mundy), the Canadian Organization for Rare Disorders (Wayne Critchley), and the Regroupement québécois des maladies orphelines (Gail Ouellette) for their participation during the workshop. The authors also thank all workshop participants and consultants for helpful comments on the report.

#### References

- 1 McCormack FX. Rare lung diseases. In: Schraufnagel DE, editor. Breathing in America: diseases, progress, and hope. New York: American Thoracic Society; 2010. pp. 185–195. [Accessed 2016 Aug 9]. Available from: https://www.thoracic.org/patients/patient-resources/breathing-inamerica/resources/chapter-18-rare-lung-diseases.pdf
- 2 Swiss group for Interstitial and Orphan Lung Diseases (SIOLD). List of rare lung diseases. Available from: http://www.siold.ch/en/ siold\_home/siold-patients-famille/siold-liste-maladies-rares.htm, accessed August 9, 2016.
- 3 Cottin V, Cordier JF, Richeldi L. Orphan lung diseases: a clinical guide to rare lung disease. London: Springer-Verlag; 2015.
- 4 Spagnolo P, du Bois RM, Cottin V. Rare lung disease and orphan drug development. *Lancet Respir Med* 2013;1:479–487.
- 5 European Respiratory Society/European Lung Foundation. European lung white book. Sheffield, UK: European Respiratory Society; 2003.
- 6 Mankoff SP, Brander C, Ferrone S, Marincola FM. Lost in translation: obstacles to translational medicine. *J Transl Med* 2004;2:14.
- 7 Loewith R, Hall MN. Target of rapamycin (TOR) in nutrient signaling and growth control. *Genetics* 2011;189:1177–1201.
- 8 Goncharova EA, Goncharov DA, Eszterhas A, Hunter DS, Glassberg MK, Yeung RS, Walker CL, Noonan D, Kwiatkowski DJ, Chou MM, et al. Tuberin regulates p70 S6 kinase activation and ribosomal protein S6 phosphorylation: a role for the *TSC2* tumor suppressor gene in pulmonary lymphangioleiomyomatosis (LAM). *J Biol Chem* 2002;277:30958–30967.
- 9 Young L, Lee HS, Inoue Y, Moss J, Singer LG, Strange C, Nakata K, Barker AF, Chapman JT, Brantly ML, et al.; MILES Trial Group. Serum VEGF-D a concentration as a biomarker of lymphangioleiomyomatosis severity and treatment response: a prospective analysis of the Multicenter International Lymphangioleiomyomatosis Efficacy of Sirolimus (MILES) trial. *Lancet Respir Med* 2013;1:445–452.
- 10 McCormack FX, Inoue Y, Moss J, Singer LG, Strange C, Nakata K, Barker AF, Chapman JT, Brantly ML, Stocks JM, et al.; National Institutes of Health Rare Lung Diseases Consortium; MILES Trial Group. Efficacy and safety of sirolimus in lymphangioleiomyomatosis. N Engl J Med 2011;364:1595–1606.
- McCormack FX, Gupta N, Finlay GR, Young LR, Taveira-DaSilva AM, Glasgow CG, Steagall WK, Johnson SR, Sahn SA, Ryu JH, et al.;

ATS/JRS Committee on Lymphangioleiomyomatosis. Official American Thoracic Society/Japanese Respiratory Society Clinical Practice Guidelines: lymphangioleiomyomatosis diagnosis and management. *Am J Respir Crit Care Med* 2016;194:748–761.

- 12 Johnson SR, Cordier JF, Lazor R, Cottin V, Costabel U, Harari S, Reynaud-Gaubert M, Boehler A, Brauner M, Popper H, et al.; Review Panel of the ERS LAM Task Force. European Respiratory Society guidelines for the diagnosis and management of lymphangioleiomyomatosis. Eur Respir J 2010;35:14–26.
- 13 Trapnell BC, Whitsett JA, Nakata K. Pulmonary alveolar proteinosis. *N Engl J Med* 2003;349:2527–2539.
- 14 Rowe SM, Verkman AS. Cystic fibrosis transmembrane regulator correctors and potentiators. *Cold Spring Harb Perspect Med* 2013;3:a009761.
- 15 Hanrahan JW, Sampson HM, Thomas DY. Novel pharmacological strategies to treat cystic fibrosis. *Trends Pharmacol Sci* 2013;34:119–125.
- 16 Beaulieu CL, Majewski J, Schwartzentruber J, Samuels ME, Fernandez BA, Bernier FP, Brudno M, Knoppers B, Marcadier J, Dyment D, et al.; FORGE Canada Consortium. FORGE Canada Consortium: outcomes of a 2-year national rare-disease gene-discovery project. Am J Hum Genet 2014;94:809–817.
- 17 Ryu JH, Moss J, Beck GJ, Lee JC, Brown KK, Chapman JT, Finlay GA, Olson EJ, Ruoss SJ, Maurer JR, et al.; NHLBI LAM Registry Group. The NHLBI lymphangioleiomyomatosis registry: characteristics of 230 patients at enrollment. Am J Respir Crit Care Med 2006;173:105–111.
- 18 Yu J, Henske EP. mTOR activation, lymphangiogenesis, and estrogenmediated cell survival: the "perfect storm" of pro-metastatic factors in LAM pathogenesis. *Lymphat Res Biol* 2010;8:43–49.
- 19 Crooks DM, Pacheco-Rodriguez G, DeCastro RM, McCoy JP Jr, Wang JA, Kumaki F, Darling T, Moss J. Molecular and genetic analysis of disseminated neoplastic cells in lymphangioleiomyomatosis. *Proc Natl Acad Sci USA* 2004;101:17462–17467.
- 20 Seyama K, Kumasaka T, Souma S, Sato T, Kurihara M, Mitani K, Tominaga S, Fukuchi Y. Vascular endothelial growth factor-D is increased in serum of patients with lymphangioleiomyomatosis. *Lymphat Res Biol* 2006;4:143–152.
- 21 Landry JS, Tremblay GM, Li PZ, Wong C, Benedetti A, Taivassalo T. Lung function and bronchial hyperresponsiveness in adults born prematurely: a cohort study. Ann Am Thorac Soc 2016;13:17–24.
- 22 Fitzgerald DA, Shapiro AJ. Primary Ciliary Dyskinesia. *Paediatr Respir Rev* 2016;18:1–2.

- 23 Shapiro AJ, Zariwala MA, Ferkol T, Davis SD, Sagel SD, Dell SD, Rosenfeld M, Olivier KN, Milla C, Daniel SJ, et al.; Genetic Disorders of Mucociliary Clearance Consortium. Diagnosis, monitoring, and treatment of primary ciliary dyskinesia: PCD Foundation consensus recommendations based on state of the art review. *Pediatr Pulmonol* 2016;51:115–132.
- 24 Young LR, Vandyke R, Gulleman PM, Inoue Y, Brown KK, Schmidt LS, Linehan WM, Hajjar F, Kinder BW, Trapnell BC, et al. Serum vascular endothelial growth factor-D prospectively distinguishes lymphangioleiomyomatosis from other diseases. *Chest* 2010;138: 674–681.
- 25 Carlile GW, Keyzers RA, Teske KA, Robert R, Williams DE, Linington RG, Gray CA, Centko RM, Yan L, Anjos SM, *et al.* Correction of F508del-CFTR trafficking by the sponge alkaloid latonduine is modulated by interaction with PARP. *Chem Biol* 2012;19:1288–1299.
- 26 Lachmann N, Happle C, Ackermann M, Lüttge D, Wetzke M, Merkert S, Hetzel M, Kensah G, Jara-Avaca M, Mucci A, et al. Gene correction of human induced pluripotent stem cells repairs the cellular phenotype in pulmonary alveolar proteinosis. Am J Respir Crit Care Med 2014;189:167–182.
- 27 Suzuki T, Arumugam P, Sakagami T, Lachmann N, Chalk C, Sallese A, Abe S, Trapnell C, Carey B, Moritz T, et al. Pulmonary macrophage transplantation therapy. *Nature* 2014;514:450–454.
- 28 Young LR, Gulleman PM, Bridges JP, Weaver TE, Deutsch GH, Blackwell TS, McCormack FX. The alveolar epithelium determines susceptibility to lung fibrosis in Hermansky-Pudlak syndrome. Am J Respir Crit Care Med 2012;186:1014–1024.
- 29 Young LR, Pasula R, Gulleman PM, Deutsch GH, McCormack FX. Susceptibility of Hermansky-Pudlak mice to bleomycin-induced type II cell apoptosis and fibrosis. *Am J Respir Cell Mol Biol* 2007;37:67–74.
- 30 Zhou Y, Huang X, Hecker L, Kurundkar D, Kurundkar A, Liu H, Jin TH, Desai L, Bernard K, Thannickal VJ. Inhibition of mechanosensitive signaling in myofibroblasts ameliorates experimental pulmonary fibrosis. J Clin Invest 2013;123:1096–1108.
- 31 Mahavadi P, Korfei M, Henneke I, Liebisch G, Schmitz G, Gochuico BR, Markart P, Bellusci S, Seeger W, Ruppert C, et al. Epithelial stress and apoptosis underlie Hermansky-Pudlak syndrome-associated interstitial pneumonia. Am J Respir Crit Care Med 2010;182:207–219.
- 32 King TE Jr, Pardo A, Selman M. Idiopathic pulmonary fibrosis. *Lancet* 2011;378:1949–1961.
- 33 Moeller A, Ask K, Warburton D, Gauldie J, Kolb M. The bleomycin animal model: a useful tool to investigate treatment options for idiopathic pulmonary fibrosis? Int J Biochem Cell Biol 2008;40:362–382.
- 34 Sime PJ, Xing Z, Graham FL, Csaky KG, Gauldie J. Adenovectormediated gene transfer of active transforming growth factor-β1 induces prolonged severe fibrosis in rat lung. *J Clin Invest* 1997;100: 768–776.
- 35 Degryse AL, Lawson WE. Progress toward improving animal models for idiopathic pulmonary fibrosis. Am J Med Sci 2011;341:444–449.
- 36 Richeldi L, du Bois RM, Raghu G, Azuma A, Brown KK, Costabel U, Cottin V, Flaherty KR, Hansell DM, Inoue Y, et al.; INPULSIS Trial Investigators. Efficacy and safety of nintedanib in idiopathic pulmonary fibrosis. N Engl J Med 2014;370:2071–2082.
- 37 King TE Jr, Bradford WZ, Castro-Bernardini S, Fagan EA, Glaspole I, Glassberg MK, Gorina E, Hopkins PM, Kardatzke D, Lancaster L, et al.; ASCEND Study Group. A phase 3 trial of pirfenidone in patients with idiopathic pulmonary fibrosis. N Engl J Med 2014;370:2083–2092.
- 38 Taveira-DaSilva AM, Pacheco-Rodriguez G, Moss J. The natural history of lymphangioleiomyomatosis: markers of severity, rate of progression and prognosis. *Lymphat Res Biol* 2010;8:9–19.
- 39 O'Brien K, Troendle J, Gochuico BR, Markello TC, Salas J, Cardona H, Yao J, Bernardini I, Hess R, Gahl WA. Pirfenidone for the treatment of Hermansky-Pudlak syndrome pulmonary fibrosis. *Mol Genet Metab* 2011;103:128–134.
- 40 Augustine EF, Adams HR, Mink JW. Clinical trials in rare disease: challenges and opportunities. J Child Neurol 2013;28:1142–1150.
- 41 Chow SC, Chang M. Adaptive design methods in clinical trials a review. *Orphanet J Rare Dis* 2008;3:11.
- 42 Kropski JA, Pritchett JM, Zoz DF, Crossno PF, Markin C, Garnett ET, Degryse AL, Mitchell DB, Polosukhin VV, Rickman OB, *et al.* Extensive phenotyping of individuals at risk for familial interstitial

pneumonia reveals clues to the pathogenesis of interstitial lung disease. *Am J Respir Crit Care Med* 2015;191:417–426.

- 43 Brownell R, Kaminski N, Woodruff PG, Bradford WZ, Richeldi L, Martinez FJ, Collard HR. Precision medicine: the new frontier in idiopathic pulmonary fibrosis. Am J Respir Crit Care Med 2016;193:1213–1218.
- 44 Oldham JM, Ma SF, Martinez FJ, Anstrom KJ, Raghu G, Schwartz DA, Valenzi E, Witt L, Lee C, Vij R, *et al.*; IPFnet Investigators. *TOLLIP*, *MUC5B*, and the response to *N*-acetylcysteine among individuals with idiopathic pulmonary fibrosis. *Am J Respir Crit Care Med* 2015; 192:1475–1482.
- 45 Chu SG, El-Chemaly S, Rosas IO. Genetics and idiopathic interstitial pneumonias. Semin Respir Crit Care Med 2016;37:321–330.
- 46 Stuart BD, Lee JS, Kozlitina J, Noth I, Devine MS, Glazer CS, Torres F, Kaza V, Girod CE, Jones KD, et al. Effect of telomere length on survival in patients with idiopathic pulmonary fibrosis: an observational cohort study with independent validation. Lancet Respir Med 2014;2:557–565.
- 47 Herazo-Maya JD, Noth I, Duncan SR, Kim S, Ma SF, Tseng GC, Feingold E, Juan-Guardela BM, Richards TJ, Lussier Y, *et al.* Peripheral blood mononuclear cell gene expression profiles predict poor outcome in idiopathic pulmonary fibrosis. *Sci Transl Med* 2013;5:205ra136.
- 48 White ES, Xia M, Murray S, Dyal R, Flaherty CM, Flaherty KR, Moore BB, Cheng L, Doyle TJ, Villalba J, *et al.* Plasma surfactant protein-D, matrix metalloproteinase-7, and osteopontin index distinguishes idiopathic pulmonary fibrosis from other idiopathic interstitial pneumonias. *Am J Respir Crit Care Med* 2016;194:1242–1251.
- 49 Lawson WE, Loyd JE, Degryse AL. Genetics in pulmonary fibrosis familial cases provide clues to the pathogenesis of idiopathic pulmonary fibrosis. Am J Med Sci 2011;341:439–443.
- 50 Lobo J, Zariwala MA, Noone PG. Primary ciliary dyskinesia. Semin Respir Crit Care Med 2015;36:169–179.
- 51 Zariwala MA, Knowles MR, Leigh MW. Primary ciliary dyskinesia. 2007 Jan 24 [updated 2015 Sep 3]. In: Pagon RA, Adam MP, Ardinger HH, Wallace SE, Amemiya A, Bean LJH, Bird TD, Ledbetter N, Mefford HC, Smith RJH, et al., editors. GeneReviews® [Internet]. Seattle: University of Washington, Seattle; 1993–2017. Available from: https://www.ncbi. nlm.nih.gov/books/NBK1122/, accessed August 9, 2016.
- 52 Knowles MR, Daniels LA, Davis SD, Zariwala MA, Leigh MW. Primary ciliary dyskinesia: recent advances in diagnostics, genetics, and characterization of clinical disease. *Am J Respir Crit Care Med* 2013;188:913–922.
- 53 Knowles MR, Leigh MW, Carson JL, Davis SD, Dell SD, Ferkol TW, Olivier KN, Sagel SD, Rosenfeld M, Burns KA, et al.; Genetic Disorders of Mucociliary Clearance Consortium. Mutations of DNAH11 in patients with primary ciliary dyskinesia with normal ciliary ultrastructure. *Thorax* 2012;67:433–441.
- 54 Knowles MR, Ostrowski LE, Leigh MW, Sears PR, Davis SD, Wolf WE, Hazucha MJ, Carson JL, Olivier KN, Sagel SD, et al. Mutations in RSPH1 cause primary ciliary dyskinesia with a unique clinical and ciliary phenotype. Am J Respir Crit Care Med 2014;189:707–717.
- 55 Santamaria F, Montella S, Mirra V, De Stefano S, Andria G, Parenti G. Respiratory manifestations in patients with inherited metabolic diseases. *Eur Respir Rev* 2013;22:437–453.
- 56 Tarnopolsky M, Katzberg H, Petrof BJ, Sirrs S, Sarnat HB, Myers K, Dupré N, Dodig D, Genge A, Venance SL, *et al.* Pompe disease: diagnosis and management. evidence-based guidelines from a Canadian Expert Panel. *Can J Neurol Sci* 2016;43:472–485.
- 57 van der Ploeg AT, Clemens PR, Corzo D, Escolar DM, Florence J, Groeneveld GJ, Herson S, Kishnani PS, Laforet P, Lake SL, *et al.* A randomized study of alglucosidase alfa in late-onset Pompe's disease. *N Engl J Med* 2010;362:1396–1406.
- 58 Trejo P, Rauch F. Osteogenesis imperfecta in children and adolescents—new developments in diagnosis and treatment. Osteoporos Int 2016;27:3427–3437.
- 59 Folkestad L, Hald JD, Canudas-Romo V, Gram J, Hermann AP, Langdahl B, Abrahamsen B, Brixen K. Mortality and causes of death in patients with osteogenesis imperfecta: a register-based nationwide cohort study. *J Bone Miner Res* 2016;31:2159–2166.
- 60 McAllion SJ, Paterson CR. Causes of death in osteogenesis imperfecta. *J Clin Pathol* 1996;49:627–630.
- 61 Wekre LL, Kjensli A, Aasand K, Falch JA, Eriksen EF. Spinal deformities and lung function in adults with osteogenesis imperfecta. *Clin Respir* J 2014;8:437–443.