

resistance training, it does show that quadriceps resistance training during a severe exacerbation of COPD can (1) prevent discharge of patients with muscle function worse than that which they had at admission, and (2) assist such patients in maintaining function during and possibly after the exacerbation phase. Also, even though only the quadriceps muscles were targeted for training, one can speculate that similar training for different muscles would yield comparable results.

This interesting study may have practical consequences for the hospital management of acute COPD exacerbation, beyond usual medical therapy. First, it restores the importance of a nonpharmacological therapy at the earliest onset of disability. Second, it emphasizes the role of physical therapy in the acute hospital setting. However, a study of the same protocol in a more severely ill population (e.g., in patients suffering from respiratory insufficiency, who are more likely to be hospitalized) should be performed to confirm both the feasibility and the effectiveness of this training strategy.

It would have been interesting if the authors had explored recovery of daily activities, which is a direct consequence of better muscle function. Indeed, a rehabilitation course including limb training may positively influence this outcome in severely ill patients (14, 15). Future research in patients with COPD exacerbations should highlight this point, thus conveying an even more powerful message to practitioners.

Conflict of Interest Statement: None of the authors has a financial relationship with a commercial entity that has an interest in the subject of this manuscript.

ENRICO CLINI, M.D.
 PIETRO ROVERSI, M.D.
 ERNESTO CRISAFULLI, M.D., PH.D.
*Department of Oncology, Hematology, and
 Respiratory Diseases
 Ospedale Villa Pineta
 Pavullo, Italy
 and
 University of Modena-Reggio Emilia
 Modena, Italy*

References

1. Donaldson GC, Seemungal TA, Bhowmik A, Wedzicha JA. Relationship between exacerbation frequency and lung function decline in chronic obstructive pulmonary disease. *Thorax* 2002;57:847–852.
2. Seemungal TA, Donaldson GC, Paul EA, Bestall JC, Jeffries DJ, Wedzicha JA. Effect of exacerbation on quality of life in patients

- with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 1998;157:1418–1422.
3. Seemungal TA, Donaldson GC, Bhowmik A, Jeffries DJ, Wedzicha JA. Time course and recovery of exacerbations in patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2000;161:1608–1613.
4. Pitta F, Troosters T, Probst VS, Spruijt MA, Decramer M, Gosselink R. Physical activity and hospitalization for exacerbation of COPD. *Chest* 2006;129:536–544.
5. American Thoracic Society/European Respiratory Society. Statement on pulmonary rehabilitation. *Am J Respir Crit Care Med* 2006;173:1390–1413.
6. Ries AL, Bauldoff GS, Carlin BW, Casaburi R, Emery CF, Mahler DA, Make B, Rochester CL, Zuwallack R, Herrerias C. Pulmonary rehabilitation: joint ACCP/AACVPR evidence-based clinical practice guidelines. *Chest* 2007;131(5, Suppl):4s–42s.
7. Man WDC, Polkey MI, Donaldson N, Gray BJ, Moxham J. Community pulmonary rehabilitation after hospitalisation for acute exacerbations of chronic obstructive pulmonary disease: randomised controlled study. *BMJ* 2004;329:1209–1213.
8. Clini EM, Crisafulli E, Costi S, Rossi G, Lorenzi C, Fabbri LM, Ambrosino N. Effects of early inpatient rehabilitation after acute exacerbation of COPD. *Respir Med* 2009;103:1526–1531.
9. Puhan M, Scharplatz M, Troosters T, Walters EH, Steurer J. Pulmonary rehabilitation following exacerbations of chronic obstructive pulmonary disease. *Cochrane Database Syst Rev* 2009;1:CD005305.
10. Troosters T, Probst VS, Crul T, Pitta F, Gayan-Ramirez G, Decramer M, Gosselink R. Resistance training prevents deterioration in quadriceps muscle function during acute exacerbations of COPD. *Am J Respir Crit Care Med* 2010;181:1072–1077.
11. Rutten EPA, Franssen FME, Engelen M, Wouters EFM, Deutz N, Schols AMW. Greater whole-body myofibrillar protein breakdown in cachectic patients with chronic obstructive pulmonary disease. *Am J Clin Nutr* 2006;83:829–834.
12. Rabinovich RA, Figueras M, Ardite E, Carbó N, Troosters T, Filella X, Barbera JA, Fernandez-Checa JC, Argiles JM, Roca J. Increased TNF α plasma levels during moderate intensity exercise in COPD patients. *Eur Respir J* 2003;21:789–794.
13. Debigare R, Marquis K, Cote CH, Tremblay RR, Michaud A, LeBlanc P, Maltais F. Catabolic/anabolic balance and muscle wasting in patients with COPD. *Chest* 2003;124:83–89.
14. Schweickert WD, Pohlman MC, Pohlman AS, Nigos C, Pawlik AJ, Esbrook CL, Spears L, Miller M, Franczyk M, Deprizio D, et al. Early physical and occupational therapy in mechanically ventilated, critically ill patients: a randomised controlled trial. *Lancet* 2009;373:1874–1882.
15. Burtin C, Clerckx B, Robbeets C, Ferdinande P, Langer D, Troosters T, Hermans G, Decramer M, Gosselink R. Early exercise in critically ill patients enhances short-term functional recovery. *Crit Care Med* 2009;37:2499–2505.

DOI: 10.1164/rccm.201001-0054ED

Bronchiectasis

A Continuum of Ion Transport Dysfunction or Multiple Hits?

Bronchiectasis, a syndrome characterized by chronic cough with purulent sputum and bronchial dilation involving more than one lobe on computed tomography scans, remains in many cases a pathophysiological dilemma. Bronchiectasis is often considered to result from a failure of lung defense that leads to chronic airway infection, airway wall damage, and airway ectasia. In this issue of the *Journal* (pp. 1078–1084), Bienvenu and colleagues investigate the pathogenesis of bronchiectasis in a large cohort of subjects in whom known causes of bronchiectasis were thought to have been excluded (1).

In their study, Bienvenu and colleagues focus on the potential role of abnormal airway ion transport, as indexed by the nasal

potential difference (PD) measurement, in the pathogenesis of the bronchiectatic phenotype, focusing on the role of abnormal cystic fibrosis transmembrane regulator (CFTR) function in governing ion transport rates. Thus, a combination of nasal PD measurements, molecular assessments of CFTR mutation status, sweat Cl⁻ concentrations, and clinical indices, including bacterial colonization and FEV₁, were measured and correlated. The groups were broken down into subjects with bronchiectasis exhibiting no CFTR mutations or sequence variants (DB0, 85 subjects), those with a single CFTR mutation or sequence variant (DB1, 22 subjects), and patients with two CFTR mutations or variants (DB2, 15 patients).

As a group, the subjects with bronchiectasis had evidence of abnormal ion transport. The degree of abnormality correlated with the number of *CFTR* mutations or sequence variants, that is, 0, 1, or 2. Although still subject to debate, there appears to be an emerging consensus that the abnormal ion transport defects in CF reflect abnormalities in both the Na^+ absorptive and Cl^- secretory processes that regulate airway surface hydration (2). It appears that abnormal airway surface hydration in CF leads to collapse of the periciliary layer, adhesion of a concentrated mucus layer to airway surfaces, persistent mucus, bacterial infection, inflammation, and lung damage, including diffuse bronchiectasis. Several key physiological concepts have emerged to explain certain clinical features of CF, including the long periods of relative health during basal states and the profound effects of acute exacerbations on lung function. The first concept is that CF lung disease indeed reflects a vulnerability to exogenous insults that trigger intercurrent exacerbations rather than a persistent failure of lung defense (3). A second concept is that the vulnerability to transient losses of lung defense is a function of (1) the severity of the *CFTR* mutations, and (2) modifier genes and environmental influences, both socioeconomic and viral/inhaled pollutant exposures (4, 5). Analyses of both the severity of *CFTR* mutations and potential modifier gene/environmental influences are instructive in identifying mechanisms relating the postulated continuum of ion transport dysfunction to disease phenotype in the Bienvenu and colleagues bronchiectasis cohort.

The patients described with two *CFTR* mutations or sequence variants (DB2 patients) as a group had intermediate sweat Cl^- values and nasal PD values just outside the range diagnostic for CF, and 11 of 15 had bacterial colonization with organisms typical of CF. Thus, these patients were likely correctly assigned to a "CF" or "CF-related disease" category, and the relatively mild phenotype likely reflected the mild nature of the *CFTR* mutations or sequence variants per se and potentially contributions of modifier genes and/or environmental influences.

The DB1 subjects with bronchiectasis, with a single genetically identified *CFTR* mutation or sequence variant, exhibited nasal ion transport defects intermediate between those of patients with bronchiectasis with two versus no *CFTR* mutations/variants, and importantly, bioelectric properties different from those of phenotypically "normal" *CFTR* obligate heterozygotes. Because the entire *CFTR* gene was not sequenced in this study, some of these DB1 subjects indeed may have had an unrecognized second *CFTR* mutation to account for the differences in nasal PDs from obligate *CFTR* heterozygotes. In parallel, modifier genes, including mutations in the epithelial sodium channel gene (*ENaC*), and environmental influences, for example, respiratory viruses and acid reflux, may play a role in producing the bioelectric and clinical phenotype characteristic of this subgroup.

The most interesting patient population was that of the bronchiectatic patients who exhibited no defined mutations in *CFTR* and nasal PD patterns consistent with abnormal Na^+ transport but normal Cl^- transport (DB0 patients). It is possible that the raised Na^+ transport rates were an acquired phenotype in response to chronic inflammation and infection, although the typical pattern is for reduced Na^+ transport rates in this setting (6). Consequently, it is possible that in these patients, genetic abnormalities in *ENaC*-mediated Na^+ transport may contribute to disease pathogenesis, as has been previously suggested by Fajac and colleagues (7). As the trend in this bronchiectatic subpopulation is for raised nasal PDs and raised amiloride-sensitive PDs (Δ amiloride PDs), the *ENaC* mutations presumably reflect a gain-of-function phenotype. Lessons from another gain-of-function *ENaC* mutation, that is, that associated with Little's syndrome, suggest that this mutation produces a pheno-

type in which the Little's *ENaC* hyperfunction expressed in the kidney is suppressed by CFTR in the respiratory tract, likely explaining the absence of a respiratory phenotype in patients with Little's syndrome (8, 9). Thus, disease-causing *ENaC* mutations in this bronchiectasis subpopulation might be predicted to be ones that escape CFTR regulation. It should be noted that the abnormality in *ENaC* associated with CF lung disease is not the absolute rate of *ENaC*-mediated Na^+ transport, but the failure of *ENaC*-mediated Na^+ absorption to slow as the airway surface becomes dehydrated (volume depleted). This observation suggests that airway surface liquid volume measurements may best assess *ENaC* mutations in the context of causing airway disease (2).

Perhaps equally interesting is the fact that many of the DB0 patients have nasal Na^+ transport and Δ amiloride PDs that overlap the normal range, raising a question concerning why normal subjects in this range did not exhibit disease. This question raises the possibility that "multiple hits" in lung defense are required to produce disease in this subpopulation as well as others. For example, it has been reported that β *ENaC* mice (mice expressing a β *ENaC* transgene), exhibiting a gain of function in airway Na^+ transport and a pulmonary disease phenotype that resembles CF, do not exhibit a spontaneous airway bacterial infection in adulthood (10). However, when β *ENaC* mice are bred with mice that have a second genetic defect in lung defense that also does not produce spontaneous bacterial infection, for example, a genetic deletion of the MyD88 signaling molecule that couples bacterial LPS production to airway luminal neutrophilia, a persistent bacterial infection results in double-mutant mice (11). Thus, it may be instructive in the DB0 patient population to look specifically for modifier genes/second genetic mutations in pathways that are complementary to the mucus clearance pathway in keeping the airway free of bacterial pathogens.

The article by Bienvenu and colleagues also raises three other questions regarding patients with bronchiectasis.

First, are nasal PDs diagnostically useful in this disease group? As the authors argue, although nasal PDs are not part of the currently designated diagnostic workup for CF (12), they appear to be useful in this patient population, including both individual Na^+ versus Cl^- transport parameters and combined indices. Indeed, both Bienvenu and colleagues (1) and Wilschanski and colleagues (13) use diagnostic indices that include the Δ amiloride response as the index of Na^+ transport, and indices of Cl^- transport that include the Δ isoproterenol PD (Δ iso) and Δ Cl^- -free PD/ Δ iso response, respectively. However, it should be noted that the Δ amiloride response measures not only the inhibition of Na^+ transport by amiloride but also the capacity of the epithelium to increase Cl^- secretion in response to the hyperpolarization of the cell interior caused by the amiloride block of *ENaC* (14). Consequently, the Δ amiloride response is somewhat redundant for measuring Cl^- transport with 0 Cl^- /0 Cl^- Δ iso measurements. Indeed, the basal PD is perhaps a better index of Na^+ transport rates alone (14), so perhaps a combination of the basal PD plus the total Cl^- permeability response, as revealed by the sum of the Δ 0 Cl^- and Δ iso responses, may be even more discriminative.

Second, is chronic bacterial infection a typical feature of bronchiectasis? The DB2 patients had a relatively high incidence of bacterial colonization (11 of 15), but the DB1 patients (6 of 22) and the DB0 patients had a relatively low incidence of culture-defined bacterial colonization. This result suggests two quite different possibilities. It is possible that this failure to identify bacterial pathogens reflects the relative insensitivity of classic bacterial hospital cultures to detect the spectrum of bacteria that chronically infect the abnormal lung, as suggested by comparisons of cultures with culture-independent 16S profiling in CF (15). Alternatively, this result could suggest that our current under-

standing of the pathogenesis of bronchiectasis is poor and that processes that directly damage the airway wall may be more important in the bronchiectatic phenotype than chronic infection. Certainly, the results of culture-independent molecular profiling of bacterial species in bronchiectatic patient populations will be important, both for research and for clinical applications.

Finally, are we treating our patients with bronchiectasis optimally? As noted previously, an accurate description of the microbiological communities that may inhabit the bronchiectatic lung will be important to tailor antibiotics. From a broader view, the notion that abnormal ion transport produces a dehydrated airway surface in bronchiectasis, with chronic mucus adhesion and infection, suggests that agents that restore hydration to airway surfaces may be rational for this patient population. Indeed, “hydrating” agents such as inhaled hypertonic saline and other osmolytes appear to be efficacious in subjects with CF (16, 17). Thus, studies of the pathogenesis of bronchiectasis, based on principles of electrolyte transport abnormalities and the spectrum of microbial infections gleaned from CF, may spur a new wave of therapies for bronchiectatic patients that may be more effective in controlling bronchiectatic symptoms and, we hope, eradicating the progression of this disease.

Conflict of Interest Statement: R.C.B. has received consultancy fees from Gilead Sciences (\$1,001–\$5,000), Parion Sciences (\$10,001–\$50,000), Inspire Pharmaceuticals (\$10,001–\$50,000), and Pulmatrix (\$5,001–\$10,000); he has received advisory board fees from Parion Sciences (\$10,001–\$50,000); he holds patents from Inspire Pharmaceuticals and Parion Sciences; he holds stock from Parion Sciences (over \$100,000), and Inspire Pharmaceuticals (over \$100,001).

RICHARD C. BOUCHER, M.D.

*Cystic Fibrosis/Pulmonary Research and Treatment Center
School of Medicine
University of North Carolina at Chapel Hill
Chapel Hill, North Carolina*

References

1. Bienvenu T, Sermet-Gaudelus I, Burgel PR, Hubert D, Crestani B, Bassinet L, Dusser D, Fajac I. CFTR channel dysfunction in non-cystic fibrosis bronchiectasis. *Am J Respir Crit Care Med* 181:1078–1084.
2. Tarran R, Button B, Picher M, Paradiso AM, Ribeiro CM, Lazarowski ER, Zhang L, Collins PL, Pickles RJ, Fredberg JJ, et al. Normal and cystic fibrosis airway surface liquid homeostasis: the effects of phasic shear stress and viral infections. *J Biol Chem* 2005;280:35751–35759.
3. Boucher RC. Airway surface dehydration in cystic fibrosis: pathogenesis and therapy. *Annu Rev Med* 2007;58:157–170.
4. Collaco JM, Vanscoy L, Bremer L, McDougal K, Blackman SM, Bowers A, Naughton K, Jennings J, Ellen J, Cutting GR. Interactions between secondhand smoke and genes that affect cystic fibrosis lung disease. *JAMA* 2008;299:417–424.
5. Collaco JM, Cutting GR. Update on gene modifiers in cystic fibrosis. *Curr Opin Pulm Med* 2008;14:559–566.
6. Gray T, Coakley R, Hirsh A, Thornton D, Kirkham S, Koo JS, Burch L, Boucher R, Nettekheim P. Regulation of MUC5AC mucin secretion and airway surface liquid metabolism by IL-1 β in human bronchial epithelia. *Am J Physiol* 2004;286:L320–L330.
7. Fajac I, Viel M, Sublemontier S, Hubert D, Bienvenu T. Could a defective epithelial sodium channel lead to bronchiectasis. *Respir Res* 2008;9:46.
8. Hopf A, Schreiber R, Mall M, Greger R, Kunzelmann K. Cystic fibrosis transmembrane conductance regulator inhibits epithelial Na⁺ channels carrying Liddle’s syndrome mutations. *J Biol Chem* 1999;274:13894–13899.
9. Stutts MJ, Homolya V, Robinson J, Zhou J, Boucher RC, Knowles MR. Normal Na⁺ absorption and absence of pulmonary disease in the airways of patients with Liddle’s syndrome: potential role of CFTR [abstract]. *Pediatr Pulmonol* 1998;Suppl. 17:217.
10. Mall M, Grubb BR, Harkema JR, O’Neal WK, Boucher RC. Increased airway epithelial Na⁺ absorption produces cystic fibrosis-like lung disease in mice. *Nat Med* 2004;10:487–493.
11. Livraghi A, Klem ER, Hudson EJ, Wilkinson KJ, Wolfgang MC, Boucher RC, Randell SH. Genetic deletion of MYD88-mediated signaling in Δ ENaC over-expressing mice reduces airway neutrophilia but promotes spontaneous, mucus-associated bacterial infection [abstract]. *Pediatr Pulmonol* 2008;Suppl. 31:262.
12. Farrell PM, Rosenstein BJ, White TB, Accurso FJ, Castellani C, Cutting GR, Durie PR, LeGrys VA, Massie J, Parad RB, et al. Guidelines for diagnosis of cystic fibrosis in newborns through older adults: Cystic Fibrosis Foundation consensus report. *J Pediatr* 2008;153:S4–S14.
13. Wilschanski M, Famini H, Strauss-Liviatan N, Rivlin J, Blau H, Bibi H, Bentur L, Yahav Y, Springer H, Kramer MR, et al. Nasal potential difference measurements in patients with atypical cystic fibrosis. *Eur Respir J* 2001;17:1208–1215.
14. Boucher RC. Human airway ion transport. Part 1. *Am J Respir Crit Care Med* 1994;150:271–281.
15. Rogers GB, Carroll MP, Serisier DJ, Hockey PM, Jones G, Kehagia V, Connett GJ, Bruce KD. Use of 16S rRNA gene profiling by terminal restriction fragment length polymorphism analysis to compare bacterial communities in sputum and mouthwash samples from patients with cystic fibrosis. *J Clin Microbiol* 2006;44:2601–2604.
16. Elkins MR, Robinson M, Rose BR, Harbour C, Moriarty CP, Marks GB, Belousova EG, Xuan W, Bye PTP. National Hypertonic Saline in Cystic Fibrosis (NHSCF) Study Group. A controlled trial of long-term inhaled hypertonic saline in patients with cystic fibrosis. *N Engl J Med* 2006;354:229–240.
17. Donaldson SH, Bennett WD, Zeman KL, Knowles MR, Tarran R, Boucher RC. Mucus clearance and lung function in cystic fibrosis with hypertonic saline. *N Engl J Med* 2006;354:241–250.

DOI: 10.1164/rccm.201002-0284ED