

Alveolar eosinophilia in current smokers with chronic obstructive pulmonary disease in the SPIROMICS cohort

To the Editor:

Novel therapies for chronic obstructive pulmonary disease (COPD) are urgently needed. Eosinophilic inflammation is an appealing target, because blood or sputum eosinophils in stable COPD may predict responses to systemic or inhaled corticosteroid therapy.¹ Titrating steroid therapy in the stable state on the basis of sputum eosinophils reduced severe exacerbations² and has been recommended for clinical practice.³ However, the prevalence of eosinophilic inflammation in COPD and its uniformity between systemic and lung compartments remain incompletely defined. Controversy exists on whether sputum analysis (reflecting large airway events) is required, or whether blood eosinophilia can suffice, on the basis of strong correlation between the 2 found by 1 group.⁴ Thus, better understanding of eosinophils in COPD is needed.

The Subpopulations and Intermediate Outcomes in COPD Study (SPIROMICS) ([ClinicalTrials.gov](https://clinicaltrials.gov/ct2/show/study/NCT01969344) NCT01969344T4),

which recently completed enrolling 2981 participants, provides a unique opportunity to address these controversies.⁵ This analysis was performed on a subset (n = 139) of SPIROMICS participants who agreed to a bronchoscopy substudy that used multicolor flow cytometry to identify leukocyte subsets and to define their activation states. All investigations were conducted according to the principles of the Declaration of Helsinki. The protocol was approved by the institutional review boards of the 8 participating clinical centers. Briefly, participants underwent sputum induction, then 2 to 4 weeks later, returned for a bronchoscopy visit at which we collected peripheral blood and bronchoalveolar lavage (BAL). Sputum, blood, and BAL samples were stained on the day of collection at the clinical centers, then fixed and shipped to a central laboratory for flow cytometry analysis. Additional details can be found in this article's Online Repository at www.jacionline.org. We categorized participants as never-smokers (NS), current smokers (CS-NAO) and former smokers (FS-NAO) with no airflow obstruction, and current smokers (CS-COPD) and former smokers (FS-COPD) with COPD (Table I).

Using our described staining protocol and flow cytometric gating,⁶ we identified eosinophils as CD45⁺, CCR3⁺, CD16⁻ cells with high forward scatter and side scatter (see Fig E1 in this article's Online Repository at www.jacionline.org). When grouping subjects by smoking status and COPD disease status, we found no differences between eosinophil percentages in peripheral blood (Fig 1, A) or sputum (Fig 1, B). In contrast, BAL eosinophils were significantly increased as a percentage of all CD45⁺ cells in current smokers with COPD, relative to other groups (Fig 1, C). However, the percentage of BAL eosinophils did not correlate significantly in any sample type with FEV₁% predicted or imaging variables (percent emphysema or Pi10) (not shown). IgE levels were largely within the normal range (geometric mean, 46.7 IU/mL; 95% CI, 35.3-60.2). Log₁₀-transformed IgE levels neither differed between groups nor correlated with eosinophil percentages in any of the 3 sample types, whether among all subjects or only the CS-COPD group (not shown). We found no correlation of BAL eosinophil percentages with plasma levels of the CCR3 ligands CCL5, CCL8, and CCL24 (not shown), and did not measure CCL11 or IL-5.

To better understand the correlation between current smoking and elevated BAL eosinophil percentages in COPD, we performed multivariate modeling. We adjusted for demographic characteristics (age, sex, black race), smoking intensity (per 10 pack-year exposure), obstruction and smoking status (never vs former vs current), chronic bronchitis, histories of asthma or gastroesophageal reflux, and inhaled corticosteroid use. We also controlled for the presence of self-reported eye/nose allergies (defined as both any diagnosis and current diagnosis, with positive response [presence of allergy symptoms] reported by 18.5% of participants) and self-reported seasonal allergies (positive response [presence of allergy symptoms] reported by 13.3% of participants). Results showed a significant association between eosinophil percentage in BAL with current smoking plus COPD, with a 2.5-fold increase in eosinophil percentage in smokers with COPD ($\beta = 2.5$; 95% CI, 1.0-3.9).

To gain insights into eosinophil activation states in various compartments, we examined cell-surface expression of the adhesion molecules CD11b (Clone CBRM1/5, which recognizes an activation-specific epitope), CD34, and CD49d; the activation receptor CD69; and CD125, the IL-5 receptor alpha chain. Within

TABLE I. Subjects' characteristics

Group	NS	FS-NAO	CS-NAO	FS-COPD	CS-COPD	P value
Subjects, n	21	29	30	37	22	
Sex ratio, M/F	8/13	13/16	16/14	24/13	12/10	.32
Age (y)	52 ± 8	63 ± 8	51 ± 6	65 ± 7	60 ± 7	<.0001
Smoking (pack-years)	0 ± 0	39 ± 16	35 ± 12	51 ± 24	53 ± 20	.0007*
FEV ₁ (% predicted)	100 ± 0	102 ± 12	97 ± 13	79 ± 19	78 ± 17	<.0001
FEV ₁ /FVC	0.82 (0.06)	0.76 (0.06)	0.76 (0.05)	0.58 (0.09)	0.58 (0.10)	<.0001
ICS use (yes/no)	1/20	1/28	3/26†	15/22	6/16	.0002

Data are presented as mean ± SD except for sex ratios and ICS use; former smoker defined as having quit for more than 6 mo. One-way ANOVA with Holm-Sidak *post hoc* testing was used to determine significant differences between groups.

ICS, Inhaled corticosteroid; F, female; M, male.

*The NS group omitted, no single group statistically different by *post hoc* testing.

†Data missing from 1 subject.

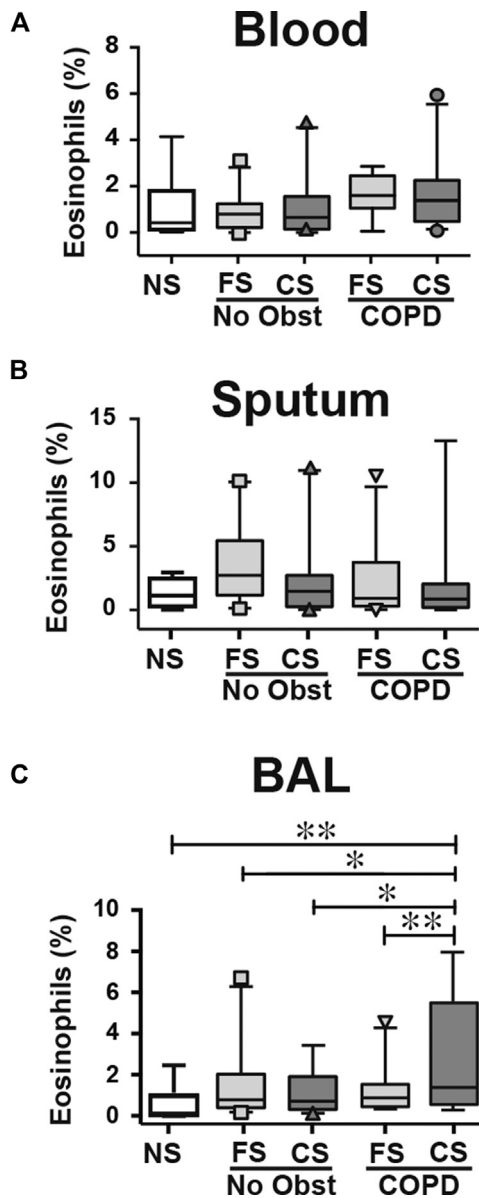


FIG 1. Active smoking significantly increases BAL eosinophil percentages in COPD. Flow cytometric analysis of eosinophils in blood (n = 97) (A), sputum (n = 94) (B), and BAL (n = 91) (C). Box and whiskers plot showing median ± interquartile range and 5th and 95th percentiles, with outliers shown as symbols. *P < .05; **P < .01; ANOVA with Holm-Sidak *post hoc* testing.

a given subject group, every receptor showed significant differences between sample types in percentages of positive eosinophils (see Table E1, column P values, in this article's Online Repository at www.jacionline.org). In contrast, within sample types, only CD125 in BAL differed between groups (significantly lower in CS-COPD) (Table E1, row P values).

Accordingly, we analyzed eosinophil receptor-positivity in the 3 sample types regardless of subject groups (see Fig E2 in this article's Online Repository at www.jacionline.org). Relative to blood, significantly more eosinophils were positive for CD11b, CD34, and CD69 in BAL and for CD34 and CD69 in sputum. CD69 may contribute to intrapulmonary eosinophil retention as shown for lung-resident T memory cells,⁷ although for both cell types, CD69 might be upregulated by the lung environment but not causing retention.

Finally, we examined the magnitude of receptor expression by mean fluorescence intensity (see Table E2 and Fig E3, A, in this article's Online Repository at www.jacionline.org), which, as with the percent of eosinophils expressing a receptor, differed significantly between sample types for every receptor. In terms of subject comparisons, there were no significant differences in the expression of CD34, CD69, or CD125 by BAL eosinophils, but the expression of CD11b was significantly greater in FS-NAO relative to FS-COPD (Fig E3, B). Moreover, the expression of CD49d was significantly higher in BAL eosinophils of CS-COPD than in groups other than CS-NAO, which was also elevated relative to FS-COPD (but not to other groups) (Fig E3, B).

This analysis demonstrates that active smoking increases steady-state localization of eosinophils to the distal lung in COPD, relative to smokers (current or former) without airflow obstruction and to NS. This interaction was not observed in FS-COPD, implying that smoking reversibly impacts eosinophil recruitment or retention (or both). Eosinophilic inflammation was compartmentalized because eosinophils as a percentage of all leukocytes in BAL showed no correlation with results in blood or in sputum (data not shown). These data extend 2 previous studies that used cytopsin differential counts to enumerate BAL eosinophils. Both found, as we did, that induced sputum results neither differed between groups nor correlated with BAL percentages.^{8,9} Our results are also congruent with those of Wen et al,⁹ who found increased BAL eosinophils in current smokers with COPD, relative to ex-smokers with COPD.⁹

Strengths of our study include its size, inclusion of extensively phenotyped subjects at multiple clinical centers, and rigorous

analytic plan, using a single flow cytometry instrument and analysis by a limited number of scientists blinded to clinical data. Limitations of our entire immunophenotyping study include the absence of viability staining at the time of flow acquisition and the duration (usually 2-4 weeks) between sputum and bronchoscopy visits. Smoking status was determined by self-report, albeit in temporal proximity to the bronchoscopy visit, but was not verified by objective measurements. Finally, subjects who agreed to the bronchoscopy substudy were self-selected and may not be representative of the general population.

Carlos H. Martinez, MD, MPH^a
Sara X. Li, MD^c
Andrew J. Hirzel, BS^d
Valerie R. Stolberg, MPH^b
Neil E. Alexis, PhD^e
R. Graham Barr, MD, PhD^f
Eugene R. Bleeker, MD^g
Elizabeth E. Carretta, MPH^h
Stephanie A. Christenson, MDⁱ
Christopher B. Cooper, MD⁸
David J. Couper, PhD^c
Claire M. Doerschuk, MD^c
MeiLan K. Han, MD, MS^a
Nadia N. Hansel, MD, MPH^h
Annette T. Hastie, PhD^c
Eric A. Hoffman, PhD^j
Robert J. Kaner, MD^j
Fernando J. Martinez, MD, MSⁱ
Deborah A. Meyers, PhD^c
Wanda K. O'Neal, PhD^c
Robert Paine III, MD^k
Nirupama Putcha, MD, MHS^h
Stephen I. Rennard, MD^l
Prescott G. Woodruff, MD, MPH^f
Michelle Zeidler, MD⁸
Jeffrey L. Curtis, MD^{a,b}
Christine M. Freeman, PhD^{a,b}
for the SPIROMICS Investigators

From ^athe University of Michigan and ^bVA Ann Arbor Healthcare System, Ann Arbor, Mich; ^cthe University of North Carolina at Chapel Hill, Chapel Hill, NC; ^dColumbia University, New York, NY; ^eWake Forest University, Winston-Salem, NC; ^fUniversity of California at San Francisco, San Francisco, Calif; ^gUniversity of California at Los Angeles, Los Angeles, Calif; ^hJohns Hopkins University, Baltimore, Md; ⁱthe University of Iowa, Iowa City, Iowa; ^jWeill Cornell Medical College, New York, NY; ^kthe University of Utah, Salt Lake City, Utah; and ^lAstraZeneca, Cambridge, United Kingdom. E-mail: cmfreema@umich.edu.

SPIROMICS was supported by contracts from the National Institutes of Health (NIH)/National Heart, Lung, and Blood Institute (NHLBI) (grant nos. HHSN268200900013C, HHSN268200900014C, HHSN268200900015C, HHSN268200900016C, HHSN268200900017C, HHSN268200900018C, HHSN268200900019C, and HHSN268200900020C), which were supplemented by contributions made through the Foundation for the NIH from AstraZeneca, Bellerophon Therapeutics, Boehringer Ingelheim Pharmaceuticals, Inc, Chiesi Farmaceutici SpA, Forest Research Institute, Inc, GSK, Grifols Therapeutics, Inc, and Ikaria, Inc; and Nycomed GmbH, Takeda Pharmaceutical Company, Novartis Pharmaceuticals Corporation, Regeneron Pharmaceuticals, Inc, and Sanofi. Additional support came from the Department of Veterans Affairs through Merit Review Awards I01 BX001389 (C.M.F.) and I01 CX000911 (J.L.C.) and the NIH/NHLBI (grant nos. K2 HL128936 and R01 HL122438-S1 to C.H.M.).

Disclosure of potential conflict of interest: V. R. Stolberg has received a grant from the Department of Defense. R. G. Barr has received grants from the National Institutes of Health (NIH), the Foundation for the NIH, and the Alpha1 Foundation; has received personal fees from UpToDate; and has received travel support from the COPD Foundation. E. R. Bleeker and D. A. Meyers have received a grant from the National Heart, Lung, and Blood Institute. E. E. Carretta has received grants from the NIH. S. A. Christenson has a board membership with AstraZeneca; has received grants from MedImmune and Genentech; and has received payment for lectures from AstraZeneca. C. B. Cooper has received grants from Equinox Health Clubs, Amgen, and Spiration; has received personal fees from Equinox Health Clubs, PulmonX,

Boehringer Ingelheim, GlaxoSmithKline, and Spiration; and works part-time on scientific engagement for GlaxoSmithKline Global Respiratory Franchise. D. J. Couper has received a grant from the NIH. C. M. Doerschuk has received grants from the NIH. M. K. Han has received a grant from the National Heart, Lung, and Blood Institute and has consultant arrangements with GlaxoSmithKline, Boehringer Ingelheim, Novartis, Sunovion, and AstraZeneca. N. N. Hansel has a board membership with AstraZeneca, GlaxoSmithKline, and Boehringer Ingelheim; has consultant arrangements with AstraZeneca, GlaxoSmithKline, and Boehringer Ingelheim; and has received grants from the NIH, the National Institute of Environmental Health Sciences, the National Heart, Lung, and Blood Institute, the Environmental Protection Agency, National Jewish, University of Iowa, Washington University, American Asthma Foundation, Boehringer Ingelheim, COPD Foundation, University of Pittsburgh, University of California San Francisco, University of Nebraska, and Temple University. A. T. Hastie has received grants from the National Heart, Lung, and Blood Institute and the Foundation for the NIH. E. A. Hoffman has received grants and travel support from the NIH and the COPD Gene Foundation. R. J. Kaner has received a grant and travel support from the NIH; has consultant arrangements with Gilead and AstraZeneca; and has received payment for lectures from Roche and Boehringer Ingelheim. F. J. Martinez has received grants from the NIH; has received personal fees from Forest, Janssen, GlaxoSmithKline, Nycomed/Takeda, Amgen, AstraZeneca, Boehringer Ingelheim, Ikaria/Bellerophon, Genentech, Novartis, Pearl, Pfizer, Roche, Sunovion, Theravance, Axon, CME Incite, California Society for Allergy and Immunology, Annenberg, Integritas, InThought, Miller Medical, National Association for Continuing Education, Paradigm, Peer Voice, UpToDate, Haymarket Communications, Western Society of Allergy and Immunology, Informa, Bioscale, Unity Biotechnology, ConCert, Lucid, Methodist Hospital, Prime, WebMD, Bayer, Ikaria, Kadmon, Vercyte, American Thoracic Society, Academic CME, Falco, Axon Communication, Johnson & Johnson, Clarion, Continuing Education, Potomac, Afferent, and Adept; has received nonfinancial support from Boehringer Ingelheim, Centocor, Gilead, and Biogen/Stromedix; and declares other interests with Mereo, Boehringer Ingelheim, and Centocor. W. K. O'Neal has received a grant from the National Heart, Lung, and Blood Institute. R. Paine III has received grants from the National Heart, Lung, and Blood Institute and the Foundation for the NIH. S. I. Rennard has received personal fees from the American Board of Internal Medicine, Able Associates, Advantage Healthcare, Align2Action, Almirall, APT, the American Thoracic Society, AstraZeneca, Baxter, Boehringer Ingelheim, Cheisi, CIPLA, ClearView Healthcare, Cleveland Clinic, CME Incite, Complete Medical Group, COPD Foundation, Cory Paeth, CSA, CSL, CTS Carmel, Daiichi Sankyo, Decision Resources, Dunn Group, Easton Associates, Elevation Pharma, FirstWord, Forest, Frankel Group, Gerson, GlaxoSmithKline, Gilead, Grifols, GroupH, Guidepoint Global, Haymarket, HealthStar, Huron Consulting, Incite, Inthought, IntraMed (Forest), Johnson & Johnson, LEK, McKinsey, Medical Knowledge, MedImmune, Methodist Health System-Dallas, Navigant, NCI Consulting, Novartis, Nuvis, Pearl, Penn Technology, Pfizer, PlanningShop, Prescott, Pro Ed Comm, ProMed, PSL FirstWord, Pulmatrix, Quadrant, Qwessential, Regeneron, Saatchi and Saatchi, Schlesinger Associates, Strategic North, Synapse, Takeda, Theron, and WebMD; has received grants from the National Heart, Lung, and Blood Institute, Nebraska Department of Health and Human Services, Otsuka, Pfizer, GlaxoSmithKline, Boehringer Ingelheim, Nycomed, AstraZeneca, Centocor, and Almirall; is employed by AstraZeneca and also retains professorship and a part-time appointment at the University of Nebraska Medical Center; and has received funding from the tobacco industry in the past but all ties with tobacco industry companies and entities supported by tobacco companies were terminated in 2007. P. G. Woodruff has consultant arrangements with AstraZeneca, Theravance, Regeneron, Sanofi, Genentech, Novartis, Janssen, and Neostem and has received grants from Genentech and MedImmune. M. Zeidler has received grants from the NIH. J. L. Curtis has received grants from the NIH/National Heart, Lung, and Blood Institute, the NIH/National Institute of Allergy and Infectious Disease, the Department of Veterans Affairs, the Department of Defense, and MedImmune. C. M. Freeman has received grants from the NIH, MedImmune, and the Department of Veterans Affairs. The rest of the authors declare that they have no relevant conflicts of interest.

REFERENCES

- Ernst P, Saad N, Suissa S. Inhaled corticosteroids in COPD: the clinical evidence. *Eur Respir J* 2015;45:525-37.
- Siva R, Green RH, Brightling CE, Shelley M, Hargadon B, McKenna S, et al. Eosinophilic airway inflammation and exacerbations of COPD: a randomised controlled trial. *Eur Respir J* 2007;29:906-13.
- McDonald VM, Higgins I, Wood LG, Gibson PG. Multidimensional assessment and tailored interventions for COPD: respiratory utopia or common sense? *Thorax* 2013;68:691-4.
- Bafadhel M, McKenna S, Terry S, Mistry V, Reid C, Haldar P, et al. Acute exacerbations of chronic obstructive pulmonary disease: identification of biologic clusters and their biomarkers. *Am J Respir Crit Care Med* 2011;184:662-71.

5. Couper D, Lavange LM, Han M, Barr RG, Bleecker E, Hoffman EA, et al. Design of the Subpopulations and Intermediate Outcomes in COPD Study (SPIROMICS). *Thorax* 2013;69:491-4.
6. Freeman CM, Crudgington S, Stolberg VR, Brown JP, Sonstein J, Alexis NE, et al. Design of a multi-center immunophenotyping analysis of peripheral blood, sputum and bronchoalveolar lavage fluid in the Subpopulations and Intermediate Outcome Measures in COPD Study (SPIROMICS). *J Transl Med* 2015;13:19.
7. Clark RA. Resident memory T cells in human health and disease. *Sci Transl Med* 2015;7:269rv1.
8. Rutgers SR, Timens W, Kaufmann HF, van der Mark TW, Koeter GH, Postma DS. Comparison of induced sputum with bronchial wash, bronchoalveolar lavage and bronchial biopsies in COPD. *Eur Respir J* 2000;15:109-15.
9. Wen Y, Reid DW, Zhang D, Ward C, Wood-Baker R, Walters EH. Assessment of airway inflammation using sputum, BAL, and endobronchial biopsies in current and ex-smokers with established COPD. *Int J Chron Obstruct Pulmon Dis* 2010;5:327-34.

Available online September 12, 2017.
<http://dx.doi.org/10.1016/j.jaci.2017.07.039>