<u>Collagen Gene Variants Previously Associated With Anterior Cruciate Ligament Injury</u> <u>Risk Are Also Associated With Joint Laxity</u>

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Abstract:

Background: Genetic association studies demonstrate a relationship between several collagen gene variants and anterior cruciate ligament (ACL) injury, yet the mechanism of these relationships is still unclear. Joint laxity is a heritable trait; increased magnitudes of anterior knee laxity (AKL), genu recurvatum (GR), and general joint laxity (GJL) have been consistently associated with a greater risk of ACL injury. Joint laxity may constitute an important intermediate phenotype for the genetic association with ACL injury that can be measured clinically.

Hypothesis: To determine if genetic variants within the *COL1A1*, *COL5A1*, and *COL12A1* genes, previously associated with ACL injury, were also associated with greater magnitudes of AKL, GR, and GJL.

Study Design: Descriptive laboratory study.

Methods: Blood samples and measures of AKL, GR, and GJL were obtained from 124 (50 male, 74 female) healthy, recreationally active subjects. Genomic DNA was extracted from the blood samples and genotyped for single-nucleotide polymorphisms previously examined relative to ACL injury. Univariate analyses of variance compared the magnitude of each laxity variable across the 3 genotypes for each single-nucleotide polymorphism in both sex-combined and sex-specific models.

Results: Specific genotypes were associated with greater GR in all subjects. Some genotypes were associated with greater magnitudes of GR, AKL, and GJL in females only.

Conclusions: Gene variants previously associated with ACL injury risk were in large part also associated with joint laxity. Sex-specific genetic associations with joint laxity were consistent with those previously reported for ACL injury.

Clinical Relevance: These data provide insight into potential pathways through which genotypic variants in collagen genes have the potential to alter ligament structure and behavior and, thus, ACL injury risk.

Keywords: joint laxity | genetic variations | anterior cruciate ligament injury | collagen genes | sex differences

Article:

Rupture of the anterior cruciate ligament (ACL) is a debilitating injury that is costly both in time loss from sport and in money spent on surgery and rehabilitation.¹⁴ The heightened risk of subsequent ligament injury^{36,41,47} and the development of osteoarthritis within 10 to 15 years after the initial injury are of particular concern.²² Over 70% of ACL injuries are noncontact in nature,⁴ and females more often experience noncontact injuries than similarly trained males.²⁵ While a familial predisposition for ACL injury risk is known,^{10,16,29,31} only recently have specific genetic changes (single-nucleotide polymorphisms [SNPs]) within the genes coding for collagen types I, V, and XII (*COL1A1, COL5A1*, and *COL12A1*) been associated with ACL injury, particularly in females.^{18,29-31} Despite these associations, the mechanisms tying these genetic variants to ACL injury risk have not been elucidated.

The effect of collagen genes and the variations within them may play a critical role in explaining ACL injury risk given the structural importance of collagen within ligaments. Type I collagen is present in the ACL⁹ and is the major fibrillar collagen providing structural stability to ligaments.⁵ Type V is also known to be fibrillar in structure (similar to that of type I)⁵ and to intercalate with type I to affect fibrillogenesis.³ Type XII is a fibril-associated collagen with interrupted triple helices, and, as the name suggests, it is known to interact on the surface of fibrillar collagens and potentially provides a structural role.⁴⁸ Since collagen is the major component of ligaments (75% by dry weight),¹¹ these findings suggest that these collagen gene variants have the potential to alter the structural integrity of the ligament, which may be manifested through changes in joint laxity (a measure that in large part reflects the behavior of ligament structures to an externally applied load). While research has yet to examine joint laxity as a potential intermediate phenotype for these genetic associations with ACL injury, both epidemiology and familial association studies suggest that this potential intermediate role is plausible.

Joint laxity is a highly heritable trait,^{2,6,8,15,40} and increased magnitudes of joint laxity have consistently been implicated as an ACL injury risk factor.^{19,23,26,32,34,43,46} Specifically, studies report a familial predisposition to greater joint laxity or joint laxity pathologies (ie, shoulder dislocation),^{2,6,8,40} and a greater prevalence of general joint laxity (GJL) in female identical twins

compared with fraternal twins¹⁵ supports this heritability. In a prospective injury risk study measuring joint laxity among other variables in 859 military cadets over a 4-year period reported that individuals with greater GJL (> 5 out of 9) and anterior knee laxity (AKL) (1 standard deviation above the mean) had a 2.8- and 2.6-fold greater risk, respectively, of rupturing their ACL.⁴³ When parsed by sex, females who had high AKL remained at a 2.7-fold greater risk, but this risk was not identified in males.⁴³ Another prospective injury risk study measuring knee hyperextension (herein, genu recurvatum [GR]) and AKL reported that greater magnitudes of GR and side-to-side differences in AKL contributed significantly to a multivariate logistic regression model for ACL injury risk.²⁶ Numerous retrospective studies have consistently associated greater magnitudes of GJL,^{19,23} AKL,^{34,46} and GR^{19,32} with ACL injury risk.

Based on these findings, characterizing the relationship between joint laxity and the collagen gene variants previously associated with ACL injury may help elucidate potential mechanisms tying genetic variants to ACL injury risk. Therefore, the purpose of this candidate gene association study was to determine if genetic variants within the *COL1A1*, *COL5A1*, and *COL12A1* genes previously associated with ACL injury1^{8,29-31} were also associated with greater magnitudes of joint laxity (AKL, GR, and GJL). Our expectation was that collagen gene variants that have been previously associated with ACL injury, given their potential to alter ligament structure, would also be associated with greater magnitudes of joint laxity.

Methods

Data for this study were obtained from 124 subjects (50 male: 22.2 ± 2.8 years, 177.9 ± 9.3 cm, 80.9 ± 13.3 kg; 74 female: 21.4 ± 2.6 years, 163.9 ± 6.7 cm, 61.1 ± 8.8 kg), from whom we had previously obtained blood samples and knee laxity data.^{37,39} Participants were recreationally active between 2.5 and 10 hours per week, had a body mass index < 30, and had no history of connective tissue disorders or knee injury. Female participants had normal menstrual cycles and had not used hormone-based medications for the past 6 months. Participants signed a consent form approved by the university institutional review board prior to enrollment.

Because knee laxity can vary across the menstrual cycle in a nonuniform fashion among women,^{28,37} we took repeated measurements across 2 cycles to obtain each individual's minimum (baseline) laxity values.³⁷ AKL was measured in millimeters as the anterior displacement of the tibia relative to the femur when a 133-N posterior-to-anterior directed load was applied to the tibia using the KT-2000 Knee Arthrometer (MEDmetric Corp, San Diego, California).³⁷ GR was measured in degrees with a standard goniometer as the amount of knee hyperextension (positive value) when the subject maximally contracted his or her quadriceps and extended the knee with the distal shank on a 4-in. bolster.³⁷ GJL was measured with the Beighton and Horan Joint Mobility Index,¹ which measures the presence of joint hypermobility at 5 anatomic locations (thumb, wrist, elbow, and knee bilaterally and forward trunk flexion), scoring 1 point for hypermobility at each joint. Measurement reliability values for these laxities were

confirmed prior to testing and found to be 0.97 (0.4 mm) for AKL, 0.97 (0.5°) for GR, and 0.98 (0.3 points) for GJL.³⁷

Blood samples (10 mL) were collected from the subject's antecubital vein during the morning hours and stored at -80°C. DNA was isolated with the DNAeasy Extraction Kit (Qiagen, Valencia, California), and genotyping of the target SNPs was conducted by GeneSeek (Lincoln, Nebraska). Prior to genotyping, a random subset of samples were subjected to standard polymerase chain reaction and electrophoresis to verify the presence of genomic DNA. SNPs were genotyped, including all of those previously investigated for ACL injury,^{18,29-31} whether or not significant associations were reported (Table 1). By including SNPs not previously associated with ACL injury, our goal was to minimize the possibility that there were nonspecific genetic associations with knee laxity and to further strengthen the hypothesis that any genetic associations previously reported for ACL injury.

Polymorphism	Location	Effect	
rs1800012	Promoter	Influences	
	region, Sp1 binding site of	transcription levels of $\alpha 1(I)$	
	COL1A1	chain of type I collagen ²⁴	
rs12722	3' UTR of COL5A1	Regulates mRNA stability ²⁰	
rs13946			
rs240736	Coding region of COL12A1	Missense,	
rs970547		isoleucine \rightarrow threonine27	
		Missense,	
		glycine \rightarrow serine27	
		Within C-terminal NC1	
		binding domain42	

Table 1. Location and effects of candidate single-nucleotide polymorphisms.

Statistical analysis was conducted with PASW Statistics 18 (IBM SPSS Statistics, New York, New York). Race datum was self-reported, and a χ^2 analysis was used to determine effect on population stratification. Univariate, one-way analyses of variance compared mean AKL, GR, and GJL values among the 3 genotypes for each SNP. Significant main effects were followed by post hoc, pairwise comparisons. Given sex-specific genetic associations with ACL injury previously reported for *COL5A1* and *COL12A1* polymorphisms,^{30,31} both sex-combined and sex-specific analyses were performed. Alpha level was set a priori at *P* < 0.05. Genotypic variants at each gene locus were tested to determine if they were in Hardy-Weinberg equilibrium (HWE) based on a χ^2 goodness-of-fit test.³³ A sample in HWE suggests that no genotypes have been disproportionately represented because of a potential selection bias or distortions caused by genetic heterogeneity.

Results

Genotypic distribution and racial makeup of our sample for the candidate SNPs are reported in Table 2. Of the 124 samples analyzed, the success rate varied from 79.8% to 89.5% for each SNP reaction in this study. HWE results and χ^2 analysis of race and genotype are also reported in Table 2. All laxity scores are reported as means and standard deviations.

	Sample Frequency, % (No.)			Racial Makeup, % (No.)					
Polymorphism: Genotype	Population,%	All	Females	Males	Black	Asian	Amer Ind/ Alaska Nat	White	
rs1800012									
TT	N/A	$5(5)^{b}$	$5(3)^{b}$	4 (2)	0 (0)	0 (0)	0 (0)	$6(5)^{b}$	
TG	N/A	21 (22) ^b	16 (10) ^b	26 (12)	25 (4)	9(1)	0 (0)	22 (17) ^b	
GG	N/A	74 (80) ^b	79 (48) ^b	70 (32)	75 (12)	91 (10)	100 (1)	72 (57) ^b	
rs12722									
CC	11b	40 (39) ^b	49 (29) ^b	25 (10)	81 (13) ^c	44 (4)	0 (0)	30 (22) ^b	
СТ	64b	34 (34) ^b	29 (17) ^b	43 (17)	19 (3)	44 (4)	0 (0)	37 (27) ^b	
TT	25b	26 (26) ^b	22 (13) ^b	32 (13)	0 (0) ^c	12 (1)	100 (1)	32 (24) ^b	
rs13946									
CC	4	8 (8)	10 (6)	5 (2)	13 (2)	25 (1)	0 (0)	7 (5)	
СТ	38	33 (35)	30 (18)	39 (18)	19 (3)	20(7)	0 (0)	32 (25)	
TT	58	59 (62)	60 (37)	56 (37)	68 (11)	27 (3)	100(1)	61 (47)	
rs240736									
CC	5	9 (10)	13 (8)	4 (2)	12 (2)	0 (0)	$100(1)^{c}$	8 (7)	
CT	48	38 (42)	39 (25)	37 (17)	50 (8)	27 (3)	0 (0)	39 (31)	
TT	47	53 (58)	48 (31)	59 (27)	38 (6)	72 (8)	0 (0)	53 (44)	
rs970547									
AA	63	53 (57)	49 (31)	58 (26)	50 (9)	36 (4)	0 (0)	53 (44)	
AG	27	43 (46)	44 (28)	40 (18)	44 (8)	55 (6)	100 (1)	43 (31)	
GG	10	5 (5)	6 (4)	2 (4)	7 (1)	9(1)	0 (0)	4 (3)	

Table 2. Population^{*a*,27} and sample genotypic distributions of candidate single-nucleotide polymorphisms.

^aFrequency of HapMap-CEU. ^bSample not in Hardy-Weinberg equilibrium. ^cIndicates a significant over- or underrepresentation within the cell (χ^2 , P \leq 0.05).

The SNP within *COL1A1* (rs1800012) was associated with GR in all subjects (P = 0.05) but not in sex-stratified models (females, P = 0.17; males, P = 0.10). GR was higher in individuals with the TG genotype ($5.0^{\circ} \pm 3.1^{\circ}$) compared to the GG genotype ($2.9^{\circ} \pm 3.6^{\circ}$; P = 0.02). When considering the genotypes that carry the T allele (TT + GT), the association remained significant (TT + GT: $4.8^{\circ} \pm 3.3^{\circ}$ vs GG: $2.9^{\circ} \pm 3.6^{\circ}$; P = 0.02); that is, the presence of the T variant was associated with greater GR in both sexes. The male subset was in HWE, while the female subset was not, and no racial bias was indicated for this SNP.

Of the 2 SNPs analyzed within *COL5A1*, a significant association was observed between rs12722 and GJL in all subjects (P = 0.02), while there were no associations with rs13946. When stratified by sex, significant associations for rs12722 were observed in females for both GR (P < 0.01) and GJL (P=0.05), but no associations were observed in males (P = 0.37-0.48). Females with the CT genotype for rs12722 exhibited greater GR ($4.5^{\circ} \pm 4.2^{\circ}$) than those with either the CC or TT genotypes ($2.2^{\circ} \pm 3.1^{\circ}$ and $1.7^{\circ} \pm 2.8^{\circ}$, respectively; P < 0.03), and those with the CT genotype had a greater GJL score (2.5 ± 1.8) than those with the CC genotype (0.9 ± 1.3 ; P < 0.01). However, GJL score was not different between the CT and TT genotypes (1.5 ± 1.1 ; P = 0.08). Consistent with the inferences of a previous analysis,³¹ the presence of the T allele in females was also associated with greater GJL in CT and TT (2.1 ± 1.6) genotypes versus the CC (0.9 ± 1.3) genotype (P < 0.01). No association was seen for the whole or male groups (P = 0.06 and P = 0.73, respectively).

The male sample population for rs12722 was in HWE, while the female subset was not. The χ^2 analysis of race and genotype indicated that the CC genotype of rs12722 was significantly overrepresented and the TT genotype underrepresented in the black subgroup, indicating a potential bias in our results. To reduce the risk of false positives caused by population stratification, rs12722 was reanalyzed without the black subgroup. The association of rs12722 with GJL was not detected in the nonblack subpopulation, although a trend still existed (*P* = 0.07). When stratified by sex, significant associations remained in females for both GR (*P* = 0.02) and GJL (*P* = 0.02) but, as before, was not seen in males (*P* = 0.59-0.86). Females with the CT genotype for rs12722 exhibited greater GR ($4.6^{\circ} \pm 4.3^{\circ}$) than those with either the CC or TT genotype (respectively, $1.7^{\circ} \pm 2.3^{\circ}$, *P* = 0.01; and $1.7^{\circ} \pm 2.8^{\circ}$, *P* = 0.02) and had a greater GJL score (2.4 ± 1.8) than those with the CC genotype (0.9 ± 1.3 ; *P* < 0.01) but not the TT genotype (1.5 ± 1.1 ; *P* = 0.10). Nonblack females with the T allele of rs12722 had significantly higher GJL scores (CT + TT; 2.0 ± 1.6 vs CC; 0.9 ± 1.3 ; *P* = 0.02), but these differences were not observed in the whole or male group analyses (*P* = 0.15 and *P* = 0.77, respectively).

We noted a trend for the combined sex and female subsets for rs970547 within the coding region of *COL12A1* where all laxity scores tended to be lower for the GG and AG genotypes compared to AA (P = 0.1). As done in a prior analyses of this variant with ACL injury,³⁰ we combined the GG and GA genotypes and compared these to the AA genotype. This resulted in a significant association for AKL when females and males were combined (P = 0.03) and within the female subset (P = 0.03) but not for the male subset (P = 0.64). Specifically, females with the AA genotype had greater AKL (6.7 ± 1.9 mm) than the combined AG and GG genotypes (5.8 ± 2.07 mm). This result suggests that the presence of the G allele is associated with lower magnitudes of AKL.

No genotypic associations were observed for any of the laxity variables for rs240736 within *COL12A1* when males and females were analyzed together (P = 0.10-0.90) or when females were analyzed separately (P = 0.06-0.87). In the male subset, the CC genotype for rs240736 exhibited greater GJL (3.0 ± 1.4) than both the CT and TT genotypes (0.6 ± 0.9 and 1.1 ± 1.4 , respectively; P < 0.04). These results should be interpreted with caution, however, as only 2 males carried the CC genotype. Both *COL12A1* SNPs were in HWE for all subjects and did not show any racial bias.

Table 3 presents a comparative summary of the genetic associations with joint laxity (current study) with prior reports of genetic associations with ACL injury.^{18,29-31}

Gene	Polymorphism	Associations With	Associations With
		Laxity	ACL Injury
COL1A1	rs1800012	TG + TT genotypes	TT genotype was
		were associated with	underrepresented in
		greater GR vs the GG	ACL injured vs
		genotype (all subjects)	controls ^{18,29}
COL5A1	rs12722 rs13946	CC genotype showed	CC genotype was
		decreased GR and	underrepresented in
		GJL than the CT	ACL-injured females
		genotype, and the CT	compared to
		showed greater GR vs	controls ³¹ No
		the TT in females	association with ACL
		only No association	injured ³¹
COL12A1	rs240736 rs970547	CC genotype $(n = 2)$	No association with
		was associated with	ACL injured ³⁰ AA
		greater GJL vs the CT	genotype was
		or TT genotypes in	overrepresented in
		males AA genotype	ACL-injured females
		was associated with	and females with
		greater AKL vs the	family history of ACL
		GA + GG genotypes	injury compared to
		in females only	GA + GG30

Table 3. Comparative findings of associations with target single-nucleotide polymorphisms and joint laxity versus anterior cruciate ligament (ACL) injury.

Discussion

This is the first report to our knowledge investigating genetic polymorphisms associated with ACL injury history^{18,29-31} on specific joint laxity measures that have also been associated with ACL injury history.^{19,23,26,32,34,43,46} These are 2 ACL injury risk factors that to date have been considered independent of each other. The consistency in our findings with joint laxity scores compared to previous studies examining the relative frequency of these genotypes in ACL-

injured populations largely confirms our hypothesis that genotypes associated with a history of ACL injury are also associated with increased joint laxity measures.

Specifically, the CC genotype of rs12722 and the AA genotype of rs970547, which were previously associated with decreased and increased injury risk, respectively,^{30,31} were also associated with lesser and greater magnitudes of joint laxity, respectively. Moreover, the female-specific genetic associations we observed for joint laxity for rs12722 and rs970547 are consistent with female-specific associations with ACL injury.^{30,31} In addition, we observed no association with joint laxity for SNPs that were not associated with ACL injury (rs13964 and rs240736),^{30,31} suggesting that there are specific genetic changes in collagen genes that increase ACL injury risk by altering the amount and structure of collagen proteins.

Relationship Among Collagen, Knee Laxity, and ACL Injury Risk

Given the hierarchical nature of collagen fibers,¹³ the importance of individual molecular bonds on strain capabilities of collagen,⁴⁴ and the interactions between collagen types,¹³ small changes in the amino acid sequence or abundance of these proteins will likely affect the overall structure and function of a ligament. In molecular models of type I collagen, changes in the amino acid sequence and the presence of cross-links between fibers alter the biomechanical properties of the structure.^{44,45} This work directly supports a previous hypothesis that density of cross links effects biomechanical capabilities.¹²

A first look into the genetic variants within the *COL5A1* gene reveals that the T allele of rs12722 confers greater messenger RNA stability than the C allele²⁰ and presumably increases the amount of *COL5A1* protein produced in the cell. This difference, in turn, has been hypothesized to alter fibrillogenesis and overall collagen structure.^{7,20} Type V collagen is known to interact with type I collagen to form a basic subunit upon which further collagen molecules build upon; thus, type V potentially represents an important component of the core of a fiber.^{3,13} The altered abundance of type V protein may change the biochemical and thus biomechanical properties within the ACL that have been measured physiologically. In an ACL-injured group, this CC genotype has been shown to be underrepresented³¹ and is further associated with lower GR and GJL scores in females in this study, both of which implicates the CC genotype as decreasing joint laxity and injury risk.

For rs970547, prior work has shown that the AA genotype was overrepresented in the ACLinjured participants versus controls when compared to the combined AG + GG genotype.³⁰ This finding is consistent with our finding that the AA genotype increased AKL when compared to the AG + GG genotypes. These results are biologically plausible, as the amino acid position that is altered by rs970547 (glycine to serine at position 3058 in the protein sequence; G3058S)²⁷ lies within the NC1 functional binding domain,⁴² which is thought to be essential for collagen matrix organization.¹⁷ This NC1 domain is thought to protrude out of the main structure and provide an interaction site for extracellular matrix molecules and potentially other collagen fibers.¹⁷ If this amino acid change alters the binding domain significantly, the altered collagen structure may be presented clinically through joint laxity and ultimately in ACL rupture.

Our association with elevated GR and the T allele of rs1800012 was inconsistent with previous findings²⁹ but may be explained through the T variant's association with other pathologies, such as lower bone quality and density.²⁴ In bone, the T substitution increases Sp1 binding and mRNA expression, which in turn results in a greater ratio of $\alpha 1(I)$ relative to $\alpha 2(I)$ (the primary structural proteins of type I collagen).²⁴ In turn, this has been associated with lower bone quality and density and reduced fracture strength.²⁴ While the implications of increase in the $\alpha 1(I)$ chain relative to the $\alpha 2(I)$ chain on ligament density and strength is unknown, these properties could have an impact on knee joint laxity. In summary, as molecular differences have been implicated to change collagen fiber biomechanics,^{44,45} overall differential collagen organization may be explained through previously unrelated measures of ligament behavior. Joint laxity provides a starting point for such investigation.

Sex-Specific Associations

The female-specific genetic associations observed between joint laxity and both rs12722 and rs970547 are consistent with the female-specific genetic associations with ACL injury for these SNPs.^{30,31} These associations may reflect a potential hormone-gene interaction,³⁰ given the known effects of sex hormones on collagen metabolism⁴⁹ and the potential for estrogen to interact with mechanical tension to differentially regulate gene expression and, thus, the abundance of collagen.²¹ It is known that GR, GJL, and AKL are greater in females compared to males³⁸ and can vary with hormone concentration changes across the female's menstrual cycle.³⁷ In an effort to better understand these sex-specific findings, further mechanistic work is needed to determine how sex hormones influence the expression of candidate genes associated with collagen regulation and how genotypic differences affect these hormonal responses.

Conclusion

Collectively, our results strengthen a growing body of work that indicates a genetic influence on ACL injury risk. Specifically, these data provide insight into one potential pathway through which genotypic variants in collagen genes have the potential to alter the structure and behavior of ligaments restraining joint motion (thus, joint laxity), thereby influencing injury risk. Moreover, the relative consistency in the associations between these collagen gene variants with both joint laxity and ACL injury risk suggests that screening an individual's joint laxity may have prognostic value in determining an individual's susceptibility to injury risk and initiating tailored intervention strategies to mitigate this risk. However, substantially more work is needed to fully characterize the interaction of these collagen gene variants with ligament structure and function and, ultimately, their effects of knee joint laxity and ACL injury risk.

There are several limitations to this study. Because of the hypothesis-driven nature of this study, the sample size is relatively small. Only 5 SNPs were studied, and given the number of genes

that have the potential to influence the ligament structure, additional relevant genetic variants should be explored. Also, the genotypic frequencies for rs1800012 and rs12722 are disproportionately represented, which is likely explained through our exclusion of previously injured individuals. That is, our healthy sample tended to have lower proportions of genotypic variants (Table 2) that have been associated with ACL injury.^{29,31,35} It will be important to examine these genetic associations in larger populations to validate and refine these findings.

Footnotes

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References

- 1. Beighton P, Solomon L, Soskolne CL. Articular mobility in an African population. Ann Rheum Dis. 1973;32(5):413-418.
- 2. Beighton PH, Horan FT. Dominant inheritance in familial generalised articular hypermobility. J Bone Joint Surg Br. 1970;52(1):145-147.
- 3. Birk DE. Type V collagen: heterotypic type I/V collagen interactions in the regulation of fibril assembly. Micron. 2001;32(3):223-237.
- 4. Boden BP, Dean GS, Feagin JA Jr, Garrett WE Jr. Mechanisms of anterior cruciate ligament injury. Orthopedics. 2000;23(6):573-578.
- 5. Boot-Handford RP, Tuckwell DS. Fibrillar collagen: the key to vertebrate evolution? A tale of molecular incest. Bioessays. 2003;25(2):142-151.
- 6. Bridges AJ, Smith E, Reid J. Joint hypermobility in adults referred to rheumatology clinics. Ann Rheum Dis. 1992;51(6):793-796.
- 7. Collins M, Posthumus M. Type V collagen genotype and exercise-related phenotype relationships: a novel hypothesis. Exerc Sport Sci Rev.2011;39(4):191-198.
- Dowdy PA, O'Driscoll SW. Recurrent anterior shoulder instability. Am J Sports Med. 1994;22(4):489-492.
- 9. Duthon VB, Barea C, Abrassart S, Fasel JH, Fritschy D, Menetrey J. Anatomy of the anterior cruciate ligament. Knee Surg Sports Traumatol Arthrosc. 2006;14(3):204-213.
- 10. Flynn RK, Pedersen CL, Birmingham TB, Kirkley A, Jackowski D, Fowler PJ. The familial predisposition toward tearing the anterior cruciate ligament: a case control study. Am J Sports Med. 2005;33(1):23-28.

- 11. Frank CB. Ligament structure, physiology and function. J Musculoskelet Neuronal Interact. 2004;4(2):199-201.
- Gautieri A, Buehler MJ, Redaelli A. Deformation rate controls elasticity and unfolding pathway of single tropocollagen molecules. J Mech Behav Biomed Mater. 2009;2(2):130-137.
- 13. Gelse K, Poschl E, Aigner T. Collagens: structure, function, and biosynthesis. Adv Drug Deliv Rev. 2003;55(12):1531-1546.
- Griffin LY, Albohm MJ, Arendt EA, et al. Understanding and preventing noncontact anterior cruciate ligament injuries: a review of the Hunt Valley II meeting, January 2005. Am J Sports Med. 2006;34(9):1512-1532.
- 15. Hakim AJ, Cherkas LF, Grahame R, Spector TD, MacGregor AJ. The genetic epidemiology of joint hypermobility: a population study of female twins. Arthritis Rheum. 2004;50(8):2640-2644.
- Harner CD, Paulos LE, Greenwald AE, Rosenberg TD, Cooley VC. Detailed analysis of patients with bilateral anterior cruciate ligament injuries. Am J Sports Med. 1994;22(1):37-43.
- 17. Kania AM, Reichenberger E, Baur ST, et al. Structural variation of type XII collagen at its carboxyl-terminal NC1 domain generated by tissue-specific alternative splicing. J Biol Chem. 1999;274(31):22053-22059.
- Khoschnau S, Melhus H, Jacobson A, et al. Type I collagen alpha1 Sp1 polymorphism and the risk of cruciate ligament ruptures or shoulder dislocations. Am J Sports Med. 2008;36(12):2432-2436.
- 19. Kramer LC, Denegar CR, Buckley WE, Hertel J. Factors associated with anterior cruciate ligament injury: history in female athletes. J Sports Med Phys Fitness. 2007;47:446-454.
- 20. Laguette MJ, Abrahams Y, Prince S, Collins M. Sequence variants within the 3'-UTR of the COL5A1 gene alters mRNA stability: Implications for musculoskeletal soft tissue injuries. Matrix Biol. 2011;30:338-345.
- Lee CY, Liu X, Smith CL, et al. The combined regulation of estrogen and cyclic tension on fibroblast biosynthesis derived from anterior cruciate ligament. Matrix Biol. 2004;23(5):323-329.
- 22. Lohmander LS, Englund PM, Dahl LL, Roos EM. The long-term consequence of anterior cruciate ligament and meniscus injuries: osteoarthritis.Am J Sports Med. 2007;35:1756-1769.

- 23. Loudon JK, Jenkins W, Loudon KL. The relationship between static posture and ACL injury in female athletes. J Orthop Sports Phys Ther.1996;24(2):91-97.
- 24. Mann V, Hobson EE, Li B, et al. A COL1A1 Sp1 binding site polymorphism predisposes to osteoporotic fracture by affecting bone density and quality. J Clin Invest. 2001;107(7):899-907.
- 25. Mihata LC, Beutler AI, Boden BP. Comparing the incidence of anterior cruciate ligament injury in collegiate lacrosse, soccer, and basketball players: implications for anterior cruciate ligament mechanism and prevention. Am J Sports Med. 2006;34(6):899-904.
- 26. Myer GD, Ford KR, Paterno MV, Nick TG, Hewett TE. The effects of generalized joint laxity on risk of anterior cruciate ligament injury in young female athletes. Am J Sports Med. 2008;36(6):1073-1080.
- 27. National Center for Biotechnology Information. dbSNP: short genetic variations. http://www.ncbi.nlm.nih.gov/projects/SNP/.
- Park SK, Stefanyshyn DJ, Ramage B, Hart DA, Ronsky JL. Relationship between knee joint laxity and knee joint mechanics during the menstrual cycle. Br J Sports Med. 2009;43(3):174-179.
- 29. Posthumus M, September AV, Keegan M, et al. Genetic risk factors for anterior cruciate ligament ruptures: COL1A1 gene variant. Br J Sports Med. 2009;43(5):352-356.
- 30. Posthumus M, September AV, O'Cuinneagain D, van der Merwe W, Schwellnus MP, Collins M. The association between the COL12A1 gene and anterior cruciate ligament ruptures. Br J Sports Med. 2010;44(16):1160-1165.
- 31. Posthumus M, September AV, O'Cuinneagain D, van der Merwe W, Schwellnus MP, Collins M. The COL5A1 gene is associated with increased risk of anterior cruciate ligament ruptures in female participants. Am J Sports Med. 2009;37(11):2234-2240.
- 32. Ramesh R, VonArx O, Azzopardi T, Schranz PJ. The risk of anterior cruciate ligament rupture with generalised joint laxity. J Bone Joint Surg Br.2005;87:800-803.
- 33. Rodriguez S, Gaunt TR, Day I. Hardy-Weinberg equilibrium testing of biological ascertainment for Mendelian randomization studies. Am J Epidemiol. 2009;169(4):505-514.
- 34. Scerpella TA, Stayer TJ, Makhuli BZ. Ligamentous laxity and non-contact anterior cruciate ligament tears: A gender based comparison. Orthopaedics. 2005;28(7):656-660.

- 35. September AV, Cook J, Handley CJ, van der Merwe L, Schwellnus MP, Collins M. Variants within the COL5A1 gene are associated with Achilles tendinopathy in two populations. Br J Sports Med. 2009;43(5):357-365.
- 36. Shelbourne KD, Gray T, Haro M. Incidence of subsequent injury to either knee within 5 years after anterior cruciate ligament reconstruction with patellar tendon autograft. Am J Sports Med. 2009;37(2):246-251.
- 37. Shultz SJ, Levine BJ, Nguyen AD, Kim H, Montgomery MM, Perrin DH. A comparison of cyclic variations in anterior knee laxity, genu recurvatum, and general joint laxity across the menstrual cycle. J Orthop Res. 2010;28(11):1411-1417.
- 38. Shultz SJ, Schmitz RJ, Nguyen A, et al. ACL research retreat V: An update on ACL injury risk and prevention, March 25th-27th, 2010, Greensboro, NC. J Athl Train. 2010;45(5):499-508.
- Shultz SJ, Schmitz RJ, Nguyen A, et al. Knee laxity and its cyclic variations influence tibiofemoral joint motion during weight acceptance. Med Sci Sports Exer. 2011;43(2):287-295.
- 40. Silman AJ, Day SJ, Haskard DO. Factors associated with joint mobility in an adolescent population. Ann Rheum Dis. 1987;46(3):209-212.
- 41. Sward P, Kostogiannis I, Roos H. Risk factors for a contralateral anterior cruciate ligament injury. Knee Surg Sports Traumatol Arthrosc.2010;18(3):277-291.
- 42. The UniProt Consortium. Ongoing and future developments at the Universal Protein Resource. Nucleic Acids Res. 2011;39:D214-D219.
- 43. Uhorchak JM, Scoville CR, Williams GN, Arciero RA, St Pierre P, Taylor DC. Risk factors associated with noncontact injury of the anterior cruciate ligament: a prospective four-year evaluation of 859 West Point cadets. Am J Sports Med. 2003;31(6):831-842.
- 44. Uzel SG, Buehler MJ. Molecular structure, mechanical behavior and failure mechanism of the C-terminal cross-link domain in type I collagen. J Mech Behav Biomed Mater. 2011;4(2):153-161.
- 45. Uzel SG, Buehler MJ. Nanomechanical sequencing of collagen: tropocollagen features heterogeneous elastic properties at the nanoscale. Integr Biol (Camb). 2009;1(7):452-459.
- 46. Woodford-Rogers B, Cyphert L, Denegar CR. Risk factors for anterior cruciate ligament injury in high school and college athletes. J Athl Train.1994;29(4):343-346.
- 47. Wright RW, Dunn WR, Amendola A, et al. Risk of tearing the intact anterior cruciate ligament in the contralateral knee and rupturing the anterior cruciate ligament graft during

the first 2 years after anterior cruciate ligament reconstruction. Am J Sports Med. 2007;35:1131-1134.

- 48. Young BB, Zhang G, Koch M, Birk DE. The roles of types XII and XIV collagen in fibrillogenesis and matrix assembly in the developing cornea. J Cell Biochem. 2002;87(2):208-220.
- 49. Yu WD, Panossian V, Hatch JD, Liu SH, Finerman GA. Combined effects of estrogen and progesterone on the anterior cruciate ligament. Clin Ortho Rel Res. 2001;383:268-281.