A COMPARISON OF THE EXTERNAL MICROBIAL ASSEMBLAGES BETWEEN NATIVE SOUTHERN STRAIN AND WILD NORTHERN BROOK TROUT, SALVELINUS FONTINALIS, OF HATCHERY ANCESTRY

A thesis submitted to the faculty of the Graduate School of Western Carolina University in partial fulfillment of the requirements for the degree of Master of Science in Biology

By

Alex Tanner Edwards

Director: Dr. Thomas H. Martin Associate Professor of Biology Biology Department

Committee Members: Dr. Joseph Pechmann, Biology Dr. Seán O'Connell, Biology

November 2012

ACKNOWLEDGEMENTS

I would like to thank my committee for support and guidance. I would also like to thank the Biology Department for providing the materials needed to complete my research. Many thanks to Daniel Brewer, Jeffrey Drummond, Mitch Mincey, Corey Green, Daniel Sollenberger, and Kyle Stowe for help during field collection. Additional thanks to Ryan Simmons and Brandon Smith for their help in editing and formatting my thesis.

TABLE OF CONTENTS

Page	
List of Tables	iv
List of Figures	v
Abstract	vi
Introduction	7
Methods	
Results	19
Discussion	
Conclusions	
Literature Cited	
Appendix	

LIST OF TABLES

Tał	ble	Page
1.	Field sites showing locations and characteristics of streams	18
2.	Water parameters taken at each stream reach prior to sampling	22
3.	Individual and total capture lengths (in cm) of brook trout sampled at each	
	field site	23
4.	Median CFU abundance of fish and water samples at each field site	24
5.	Statistical summaries of ANOVA and Tukey's pairwise comparisons among	
	median estimates of CFU abundance (per ml) in fish samples	25
6.	Statistical summaries of ANOVA and Tukey's pairwise comparisons among	
	median estimates of CFU abundance (per ml) in water samples	26
7.	Percent of private CFUs per sample based on total abundance	27
8.	Statistical summaries of ANOVA and Tukey's pairwise comparisons for	
	percent private CFUs based on total abundance	28
9.	Percent of private CFU richness per sample based on occurrence of unique	
	CFU types	29
10.	Statistical summaries of ANOVA and Tukey's pairwise comparisons for	
	percent private CFU richness based on occurrence	30
11.	Frequencies of shared CFU types among the different samples at each field	
	site	31

APPENDICES

1.	Characterization data of identified CFUs	.62
2.	Total CFU counts in each plate in the different dilution series for each sample.	.82
3.	Presence and absence of unique CFU types within each sample	. 84

LIST OF FIGURES

Fig	gure	Page
1.	Box plot showing ranges of median CFU abundance for each grouped	
	sample	32
2.	Box plot showing ranges of percent private CFUs based on total	
	abundance for each grouped sample	33
3.	Box plot showing ranges of percent private CFUs based on richness of	
	individual CFU types for each grouped sample	34

APPENDIX

1.	Photos of selected	plate cultures	19	9
----	--------------------	----------------	----	---

ABSTRACT

A COMPARISON OF THE EXTERNAL MICROBIAL ASSEMBLAGES BETWEEN NATIVE SOUTHERN STRAIN AND WILD NORTHERN BROOK TROUT, SALVELINUS FONTINALIS, OF HATCHERY ANCESTRY

Alex Tanner Edwards, M.S.

Western Carolina University (November 2012)

Director: Dr. Thomas H. Martin

Sean O'Connell, Joseph Pechmann

Hatchery reared, northern strain brook trout have been stocked in streams within the home range of southern strain brook trout in an effort to restore or enhance native trout populations since the late 1800s. But, brook trout native to the southern Appalachians are genetically distinct; raising ecological and ethical concerns regarding the impact of the past stockings. In this study, the external microbial assemblages on native southern and wild fish of hatchery ancestry were compared by characterizing colony morphologies and estimating densities of colony forming units. The hatcheryancestry fish had significantly higher densities, and assemblages were more similar to that of the surrounding water than those of the southern strain fish. These results suggest that the native southern strain fish exhibit a greater ability to inhibit microbial growth in their epidermal mucus than do the fish with hatchery ancestry.

INTRODUCTION

The brook trout (Salvelinus fontinalis) is the only native salmonid of the southern Appalachian Mountains (MacCrimmon and Campbell 1969). Brook trout have been stocked to restore or enhance depleted native trout populations (Lennon 1967, Jones 1975, Wilson 2011). However, the brook trout reared in these early hatcheries were not derived from the southern strain, but were of northern ancestry (Lennon 1967) and have distinct genetic differences (McCracken et al. 1993, Kreigler et al. 1995, Hayes et al. 1996, Galbreath et al. 2001). Northern strain brook trout are generally considered to be those found north of the New River drainage in Virginia, while the southern strains include the New River drainage and all waters south (Hayes et al. 1996). Using allozyme analysis, McCracken et al. (1993) found genetic differences between hatchery strain brook trout of northern descent and native southern strain populations. Wild northern strain fish of hatchery origin (NBKT) were found to have less genetic diversity than native southern strain brook trout (SBKT; Hayes et al. 1996). NBKT had only 4 mtDNA haplotypes, while SBKT populations had 12 haplotypes and exhibited almost as much intra-strain variation as they did with NBKT (Hayes et al. 1996).

Hayes *et al.* (1996) argued that the low diversity seen in the hatchery fish could be explained by the bottlenecking effect of the hatchery. However, northern strains also exhibited low genetic diversity in their native streams; it was hypothesized that this is due to the contraction and subsequent re-expansion of the northern brook trout's range after the mass glaciations of the Pleistocene (Hayes *et al.* 1996). In contrast, the genetic diversity of SBKT was shown to be much greater than that of the far northern populations and those of hatchery ancestry (Hayes *et al.* 1996). The difference between the two

strains may be explained by two hypotheses: 1) unlike the NBKT, who were forced into refugia during the Pleistocene glaciation, SBKT flourished in the South and when the glaciations ended they subsequently did not have a bottlenecking effect of reestablishment; and 2) after the Pleistocene glaciation, as the climate began warming and the lower reaches of major rivers became too warm, they had to seek refuge in the cooler headwater streams. Thus they became isolated into distinct populations increasing the chances of diversification across their southern range (Hayes *et al.* 1996).

Many concerns have been raised about introducing nonnative trout into native populations (Allendorf and Phelps 1980, Ferguson 1990, Krueger and May 1991). In particular, the loss of native genetic diversity through introgression of NBKT alleles is a possibility (Krueger and May 1991, Hayes *et al.* 1996). The process of hatchery rearing may have selected for traits or behaviors that are maladapted for natural streams (Hindar *et al.* 1991). Though the genetic differentiation between the strains has been known for some time, only recently have the ecological differences been quantified. Wesner *et al.* (2011) found differences between NBKT and SBKT in growth, behavior, and survival under experimental conditions. Hybridization with hatchery stock can swamp the genetic makeup of native trout (Hindar *et al.* 1991, Hansen and Loeschcke 1994). A loss of genetic integrity could disrupt unique naturally selected ecological and physiological responses, putting the genetic diversity and fitness of native fish populations at risk (Allendorf and Phelps 1980, Ferguson 1990).

Another potential ecological difference between the strains is how fish react to the microbial communities in their habitat. Fish have slow reacting specific immune responses that are affected by temperature (Ellis 1982, Bly and Clem 1991) and must

therefore rely heavily on their innate immune response (Subramanian *et al.* 2008). The epidermal mucosal layer of fish (slime) serves as an integral part of innate immunity (Ellis 1974, Ingram 1980) and is considered the fish's first line of defense against microorganisms (Hjelmeland *et al.* 1983, Austin and McIntosh 1988, Grinde *et al.* 1988, Fouz *et al.* 1990, Nagashima *et al.* 2001, Sarmaşik 2002). The process in which slime protects fish from harmful pathogens works in three layers: first the slime acts as a physical barrier between the fish and the environment; second, the slime is continually replenished and sloughed off, removing microbes that have attached (Pickering 1974, Alexander and Ingram 1992, Rombout *et al.* 1993, Aranishi and Nakane 1997, Ellis 2001); and third, the presence of broad-spectrum, defensive agents within the slime prevent or destroy growth of foreign invaders (Austin and McIntosh 1988, Ellis 2001, Hellio *et al.* 2002, Subramanian *et al.* 2008; reviewed in: Bols *et al.* 2001, Ellis 2001).

Subramanian *et al.* (2008) attempted to identify and describe the defensive agents within the slime using aqueous, organic, and acidic extracts of concentrated mucus from different fish species, including brook trout. The slime of brook trout was found to exhibit among the strongest antimicrobial properties of the tested fish. The agent which seemed to have the greatest antimicrobial properties was the small peptide molecules found in the acidic extracts. Fish become highly susceptible to infection (bacterial and fungal) after slime removal (Wedemeyer 1996, Madetoja *et al.* 2000). When Madetoja *et al.* (2000) challenged rainbow trout (*Oncorhynchus mykiss*) with intact mucal layers to immersion in baths containing known fish pathogens, no mortalities occurred. Fish in which the mucus had been removed resulted in mortalities of 27% of the sample, while fish with removed mucus and skin abrasions resulted in an average of 95% mortalities.

The antimicrobial activity of mucus differs among fish species as do the cells producing the mucus, which could lead to differences in mucus composition and thus variation in antimicrobial effectiveness (Shephard 1993, Subramanian *et al.* 2008). Mucus composition has also been shown to vary due to ecological and physiological conditions such as water quality and induced stress (Agarwal *et al.* 1979, Zuchelkowski *et al.* 1981, Blackstock and Pickering 1982, Pottinger *et al.* 1984, Lebedeva 1999). Isolation and differing selective pressures between these two strains of brook trout could have provided different trajectories of innate immune responses.

Though some antimicrobial agents have been identified and described from fish slime, much is still unknown (Subramanian et al. 2008). One fairly unexplored possibility is that of associative microbes living within the slime (Subramanian et al. 2008). Microbial species that have been identified in the mucus of fish have been shown to exhibit their own antimicrobial components (Ebran et al. 1999, Parret et al. 2005). Microorganisms associated with the exterior of host organisms can be beneficial in protecting the host from deleterious pathogens, forming a mutualistic relationship (Wingender et al. 1999). Studies of the external microbial assemblages of amphibians have shown that different species held different assemblages of microorganisms on their skin (Culp *et al.* 2007). Many of these microorganisms have been found to be unique to the host and were not constituents of the aquatic environment, suggesting a symbiotic relationship (Gilbert 1944, Culp et al. 2007). Specific analyses have shown that mutualistic relationships do exist between some salamanders and their skin flora (Lauer et al. 2007, Lauer et al. 2008). Thus the possibility of beneficial skin or mucus flora on brook trout acting as an antimicrobial agent should not be discounted.

Since much of the antimicrobial action of fish slime is believed to be broadspectrum, reduced overall growth and especially that of environmental microorganisms within the slime could indicate a greater antimicrobial action and thus a greater ability to fight off potential pathogens (Ellis 2001, Hellio *et al.* 2002, Magnadóttir 2004, Balasubramanian *et al.* 2011). Results of differing antimicrobial activity would then potentially define selective differences of innate immunity in hatchery reared fish. Being that these differences in innate immune responses are genetically inherited (Secombes and Olivier 1997), the changes endured prior to the hatchery or selected for by hatchery pressures could persist through generations of stocked fish, raising concerns that potential hybridization between native and nonnative strains which may result in a loss of fitness by contaminating the genetic makeup of native fish (i.e. replacing their naturally selected immune responses for a maladaptive artificially selected response) (Allendorf and Phelps 1980, Currens *et al.* 1997, Lynch and O'Hely 2001, Davis 2006).

I extracted slime from NBKT and SBKT and cultured it in the lab to assess quantity and composition of the external microbial assemblages of these. There were three possible outcomes. The first was more microbial growth in the slime and/or an increased presence of environmental microbes in the microbial slime assemblage of NBKT. This could be indicative of either a lack of adaptation to the streams they have colonized. An alternative explanation is reduced mucal activity due to selection or lack thereof in the hatchery, perhaps because of the use of antibiotics and antimicrobials to control disease (Kirkan *et al.* 2003). The second possibility was that SBKT would exhibit a higher microbial count and a less endemic assemblage. This would result if either NBKT's historical ancestry in northern environments selected for fish with more advanced immune systems based on environmental factors or if harsh conditions common to hatchery rearing (Tomasso *et al.* 1981, Piper *et al.* 1989, Winfree *et al.* 1998, Ellis *et al.* 2002) selected for more effective innate immunity. The final possibility was that negligible variation in mucosal immune responses exists in these sub-populations, suggesting that isolation (either hatchery or historic geographic) has not resulted in selection of differences in mucal activity. Based on the endemicity of the native strain and potentially mal-selective pressures of fish hatcheries, I hypothesized that SBKT would exhibit reduced density and diversity of colony forming units.

METHODS

The study sites were two tributaries of each of three separate major rivers in western North Carolina (French Broad, Pigeon, and Tuckasegee). Study sites were chosen on the basis of being distinct, isolated watersheds so as to control for any effect of the water itself, and to look at the microbial diversity of isolated populations. Each pair of streams consisted of one stream previously identified by the NC Wildlife Resource Commission to be populated with SBKT, and one occupied by NBKT (Table 1). The streams chosen for the French Broad were Sawmill Creek and Shoal Creek, containing SBKT and NBKT respectively. From the Pigeon I sampled Scapecat Creek – hybridized population originally believed to be SBKT - and Flat Laurel Creek -NBKT. From the Tuckasegee I sampled Mull Creek -SBKT - and Beechflat Creek - NBKT. The samples from Scapecat Creek were later removed from the SBKT category when I learned that fish of mixed native and hatchery ancestry (hybrid brook trout - HBKT) had been identified from this stream (Galbreath et al. 2001). Both the mucus and water samples were used in the statistical analyses; however, they were placed into their own category of HBKT and HBKT stream.

Two rounds of sampling were undertaken. The first sampling was conducted in 2011 between October 8th and November 19th and the second during July 2012 (17th-19th). Shoal Creek was not sampled during 2012 due to the difficulty of sampling and low capture rate, and thus its paired stream Sawmill Creek was also omitted that year.

Upon arrival at a sample site, a grab-sample of stream water for microbial analysis was taken by immersing a sterilized 50 ml conical tube in non-turbulent but swift flowing water. Temperature, pH, % DO, and conductivity of the stream were then measured using a dissolved oxygen meter (model YSI 650, YSI Inc., Yellow Springs, OH, USA). Before fish collection all submersible capture gear was sterilized in 15% house-hold bleach. Resealable polyethylene bags were sterilized overnight using 70% isopropyl alcohol then rinsed and filled with 500 ml of 0.85% saline solution and placed on ice.

I collected brook trout were via electro-fishing or hook and line from each stream. Hook and line was used when shocking was not effective either due to extremely low conductivity or unmanageable terrain making capture after shocking difficult or dangerous). Because the amount of slime on the fish is relative to the surface area (size) of the fish, the collection was not based on numbers of fish but by a cumulative total fish length of 50-100 cm per sample site. Collected fish were placed in a sterilized resealable plastic bag containing 500 ml of saline solution and lightly shaken for 15 seconds for slime extraction. Fish were removed from the bag by hand using a fresh nitrile glove after each catch. The slime was pooled for each stream reach, reusing the same bag of saline solution for each new capture. After slime extraction, the bag was placed on ice. The total length of the fish was measured and then they were immediately released.

Negative controls were used to test for the occurrence of outside contamination. Using the same sterile saline solution and plastic-bag-setup procedure (but without fish), the process of the slime extraction was mimicked. All samples of water, extracted slime, and negative controls were stored on ice while transported back to Western Carolina University and refrigerated at 4°C to minimize growth and preserve spec imines until culture.

The microbiota assemblage of the slime, water, and controls were examined by plating samples (either the slime from the saline solution or the water) on an R2A nutrient agar medium. A dilution series using a 0.85% saline solution was conducted using the undiluted sample, a 1:10 dilution, and a 1:100 dilution. 100 μ l of each concentration from each sample were plated on three different T100x15 mm Petri dishes. Dishes were plated and evenly spread under a sterile hood using a blower with laminar flow. After plating, the dishes were stored upside down in the dark at ambient temperature (approximately 20°C), undisturbed for one week. After one week the dishes were observed and plates with few to no colonies were marked and were then placed in refrigeration at 4°C to minimize further colony growth. This would indicate if further growth was occurring after refrigeration. Colony forming units (CFUs) were counted on each plate. Individual CFUs were observed and described using a six-characteristic microbe check list similar to the protocol described by Breakwell et al. (2007). While CFUs were counted from samples taken during both sample periods, only samples from the first sample season were used for the colony morphology descriptions. Thus analyses that used the morphology data only reflect a total of three samples from each grouped origin.

The plates used to determine the counts were the three plates from the dilution series for each sample that fell within the 20-200 CFU range. If no plate's count for a sample fell within the range, the nearest appropriate plates were used. If the CFU count for all plates was less than 20, then the plates with the highest values were used, and if all CFU counts were greater than 200, then the plates with the lowest values were used. If there were more than three plates within the optimal range the three closest to the middle of the 20-200 range (i.e. nearest 110) were used. The median CFU density (number per mL of sample) from the three subsamples was used as the CFU density for the sample.

Each sample was analyzed as independent even though some samples were from the same locations but at different times. Independence was assumed based on the belief that the microbial assemblages would represent new populations due to the continuous renewal of the mucal layer and the rapid frequency in which microbes reproduce. The data were transformed (log(x)) to meet the assumptions of normality and homoscedasticity. To measure differences in the microbial abundance among the samples data were analyzed by ANOVA. Analysis of variance was followed by pairwise comparisons of means using Tukey's correction to maintain an experiment-wise error rate of 0.05. For abundance analysis water and mucus samples were analyzed separately due to the differences in the initial dilution of samples.

To measure the uniqueness of CFU composition, the abundance of morphological colony types uniquely represented in a particular sample and shared by no other samples (referred to in this paper as "private CFUs", based on my terminology and not related to other scientific literature) was totaled for each sample and a proportion was calculated using the number of private colonies divided by the total number of colonies for the sample "percent private CFU abundance" (Appendix Tables 1 and 2). This is interpreted as a test for potential endemicity of microbes in different populations and as a screen for antimicrobial activity, based on the premise that absence of environmental microbes suggests inhibition. An ANOVA followed by Tukey's pairwise comparisons was performed using arcsine transformed data. The raw count/classification data (Appendix Table 1) was summarized into a table of occurrence, based on the presence or absence of

specific CFU types within the different sample locations (Appendix Table 3). A similar private CFU analysis based on the richness of private CFU types was performed "percent private CFU richness". Richness in this context refers to the number of distinct CFU types for each sample. The ratios of private to total CFU types were calculated and analyzed in the same manner as the previous private CFU analysis. This test of private CFUs was conducted to remove the bias from abundant CFU types. For example, a sample that exhibits high abundance of a particular private CFU type and low private CFU richness would result in the previous analysis "percent private CFU abundance" showing a high percent of private CFUs, whereas the "percent private CFU richness" analysis would show a low percent of private CFUs.

Table 1. Field sites showing locations and characteristics of streams containing northern strain brook trout derived from hatchery ancestry (N), native, southern strain brook trout (S), and fish of mixed genetic origin (H).

Stream	Drainage	Strain	Latitude	Longitude	Elevation (m)
Sawmill Creek - above falls	French Broad	S	35.191101	-82.82089	824
Shoal Creek	French Broad	Ν	35.260136	-82.849281	925
Scapecat Creek	Pigeon	Н	35.380413	-82.892151	1065
Flat Laurel Creek	Pigeon	Ν	35.327363	-82.901912	1521
Mull Creek	Tuckasegee	S	35.365794	-83.020409	1055
Beechflat Creek - above falls	Tuckasegee	Ν	35.352388	-83.015501	1082

RESULTS

Water conditions were similar among the streams (Table 2). Temperature differed between the two seasons with the July 2012 sampling season running warmer (15.0-16.9°C) than the October through November 2011 season (7.9-12.0°C). Conductivities were similar and typical of high elevation mountain streams (5-18 μ S). Dissolved oxygen had little variation (84.3-91.4 sat%) as did pH (typical range 6.14-7.30 pH), except for a single outlying data point, the second sampling of Flat Laurel Creek exhibited a much higher level of acidity than the other streams (4.98 pH) and also than itself the previous season (7.30 pH).

All fish capture totals (Table 3) were within the desired range of 50-100 cm except for Shoal Creek from which I was only able to sample two fish (summed length of 42 cm). Fish capture was markedly easier and quicker in the streams inhabited by SBKT, and in general it took much longer to acquire fish from northern strain streams. Though the total length of trout collected differed among streams, I kept the paired streams (NBKT and SBKT of the same river system) within similar total fish length ranges.

The control plates yielded no CFUs, with the exception of one replicate. A small growth was seen in along the edge in a single 1:100 dilution. The frequency of microbes per plate was proportionate to the dilution except for one Mull Creek fish sample. One plate from the 1:100 dilution (2 CFUs) yielded more CFUs than the 1:10 dilutions (0 CFUs); however, in this case the CFU count was extremely low throughout all the plates and during later scrutiny it was observed that there were four 1:100 plates labeled and only two 1:10 plates labeled. This error went unnoticed because most of the plates from both dilutions yielded no CFUs. In addition, the total occurrence of CFUs

from Mull Creek fish was so low that it could be realistically considered as no growth. However, I took a broad approach and the plate was evaluated despite having fewer than 20 CFUs. Most samples' dilution series produced at least three plates in the 20-200 CFU range. Most of the locations that yielded no plates reaching the optimal range belonged to SBKT samples as a result of their low yields. One stream (Shoal Creek) produced very high CFU counts (Table 4, Appendix Table 2), but samples appeared uncontaminated due to the fact that the dilutions still produced the correct proportions and microbes described occurred in most of the plates from that stream and even appeared in plates from other sample locations.

The undiluted HBKT samples from Scapecat Creek were different from the SBKT samples in appearance, due to the presence of spreading CFUs (irregular colonies that had no uniformity and established themselves over a large area of the plate). Though "spreaders" were not uncommon within the water and NBKT samples they were relatively small or absent from the SBKT samples. All "spreaders" were counted as a single colony, unless completely divided by edged space. The filmy, layered appearance within plates with heavy "spreaders" made it difficult to characterize some CFU forms and discern whether they were individual colonies or of the same colony. CFUs that were formed within "spreaders" were counted for abundance data but were not characterized to avoid possible misidentification. Selected pictures (replicate "A" from each dilution of each sample) are provided in the Appendix (Figure 1).

SBKT had significantly lower median CFU density than NBKT (p = 0.016) (Table 5, Figure 1). The water samples exhibited similar abundance values with no significant differences (Table 6, Figure 1). HBKT exhibited CFU counts between SBKT and NBKT and thus was not significantly different from either of the two strains. The water samples, regardless of occupying strain, and NBKT samples all had very similar CFU densities.

The presence of potentially fungal-like colonies was also noted by features of dullness, opaqueness, and fibrous or rhizoid projections. These fungal-like colonies were not infrequent within the water, NBKT, and HBKT samples, but were virtually absent from the SBKT samples (Appendix table 1).

Based on abundance, SBKT possessed a significantly higher percent of total private CFUs in comparison to NBKT (p = 0.023), SBKT streams (p = 0.030), and NBKT streams (p = 0.015; Tables 7 and 8, Figure 2). The majority of the microbial composition of the native trout was CFUs that were unique to each sample of SBKT (Appendix table 3). The other samples exhibited much higher abundances of shared colonies. Based on the richness analysis, when compared to NBKT and all of the stream samples, SBKT exhibited a significantly higher percent of private CFU types than NBKT (p = <0.001) and the water samples (SBKT.W p = 0.002, NBKT.W p = <0.001, HBKT.W p = 0.006), which exhibited similarly insignificant differences among each other (Tables 9 and 10, Figure 3). However, HBKT exhibited a significantly greater percent of private CFU forms in comparison to NBKT streams (p = 0.046) and a higher mean percent than NBKT (p = 0.057) but not significant at the 95% confidence level. Only a single CUF type cultured from samples of SBKT was shared by one other sample (Table 11) But CFUs cultured from the water in which the brook trout were living often shared many types with other stream samples and with NBKT samples. HBKT exhibited shared CFUs with the environment and with NBKT but do a lesser degree than NBKT.

21

Table 2. Water parameters taken at each stream reach prior to sampling.

Abbreviations: temperature (Temp), specific conductivity (SP Cond), dissolved oxygen saturation (DO sat%).

Location	Strain	Date	Time	Temp (°C)	SP Cond (µS)	DO (sat%)	рН
Beechflat Creek	Ν	10/8/2011	12:37	12.0	18	86.7	6.14
Mull Creek	S	10/8/2011	16:06	12.0	15	87.2	6.81
Scapecat Creek	Н	10/23/2011	12:46	9.1	11	88.2	7.14
Flat Laurel Creek	Ν	10/25/2011	16:22	7.9	5	86.3	7.30
Sawmill Creek	S	11/3/2011	10:35	9.6	9	91.4	7.13
Shoal Creek	Ν	11/3/2011	12:45	9.1	11	85.3	6.92
Beechflat Creek	Ν	7/17/2012	10:00	15.0	10	89.6	6.39
Mull Creek	S	7/18/2012	8:41	15.2	15	89.9	6.97
Scapecat Creek	Н	7/19/2012	8:34	16.7	14	87.5	7.07
Flat Laurel Creek	Ν	7/19/2012	15.10	169	6	84 3	4 98

Table 3. Individual and total capture lengths (in cm) of brook trout sampled at each field site. The number "2" refers to the sample taken during the second season.

SBKT			NBKT					HBKT	
Mull	Sawmill	Mull 2	Beechflat	Flat Laurel	Shoal	Beechflat 2	Flat Laurel 2	Scapecat	Scapecat 2
11	15	10	15	15	19	20	20	9	14
8	15	13	17	14	23	25	15	7	12
6	14	15	15	18		19	18	10	15
16	9	13	9	19			20	10	13
9	9	12		17				7	6
13								15	
								22	
63	62	63	56	83	42	64	73	80	60

Origin	Strain	Median abundance
Fish Samples		
Mull	S	2.00E+01
Sawmill	S	1.10E+02
Mull 2	S	5.00E+01
Beechflat	Ν	8.40E+02
Flat Laurel	Ν	2.20E+02
Shoal	Ν	1.15E+05
Beechflat 2	Ν	5.90E+03
Flat Laurel 2	Ν	1.07E+04
Scapecat	Н	1.60E+02
Scapecat 2	Н	4.20E+02
Water Samples		
Mull	S	1.21E+03
Sawmill	S	1.16E+03
Mull 2	S	1.10E+04
Beechflat	Ν	6.00E+02
Flat Laurel	Ν	4.30E+02
Shoal	Ν	7.20E+03
Beechflat 2	Ν	6.20E+03
Flat Laurel 2	Ν	2.30E+03
Scapecat	Н	8.40E+02
Scapecat 2	Н	1.10E+03

Table 4. Median CFU abundance of fish and water samples at each field site. CFU totals are an average of the three plates closest to the 20-200 range.

Table 5. Statistical summaries of ANOVA and Tukey's pairwise comparisons among median estimates of CFU abundance (per ml) in fish samples. Abbreviations: degrees of freedom (df), sum of squares (SS), mean squared (MS), f value (F), p value (P), standard error (SE), t value (t), fish (F), water (W).

ANOVA					
Source	df	SS	MS	F	Р
Sample Origin	2	39.58	19.792	5.508	0.0366
Error	7	25.15	3.594		
Tukey Pair-wise Com	parisons:				_
Comparison	Estimate	SE	t	Р	
NBKT.F-HBKT.F=0	2.79	1.586	1.759	0.2488	_
SBKT.F-HBKT.F=0	-1.688	1.73	-0.976	0.6119	
SBKT.F-NBKT.F=0	-4.479	1.384	-3.235	0.0334	_

Table 6. Statistical summaries of ANOVA and Tukey's pairwise comparisons among median estimates of CFU abundance (per ml) in water samples. Abbreviations: degrees of freedom (df), sum of squares (SS), mean squared (MS), f value (F), p value (P), standard error (SE), t value (t), fish (F),

water (W).

ANOVA						
Source	df	SS	MS	F	Р	
Sample Origin	2	1.125	0.5627	0.39	0.691	
Error	7	10.093	1.4419			
Tukey Pair-wise Comparisons:						
Comparison	Estimate	SE	t	Р		
NBKT.W-HBKT.W=0	0.6949	1.0047	0.692	0.774	_	
SBKT.W-HBKT.W=0	0.9518	1.0962	0.868	0.674		
SBKT.W-NBKT.W=0	0.257	0.8769	0.293	0.954	_	

Table 7. Percent of private CFUs per sample based on total abundance. FromOctober/November samples only.

Sample Location	Strain	Total CFUs	Total Private CFUs	%Private CFUs
Fish Samples				
Mull	S	4.00E+00	4.00E+00	100.00
Sawmill	S	3.90E+01	3.10E+01	79.48
Beechflat	Ν	1.87E+02	3.70E+01	19.78
Flat Laurel	Ν	6.70E+01	2.10E+01	31.34
Shoal	Ν	3.33E+04	2.00E+00	0.0059
Scapecat	Н	6.00E+01	4.30E+01	71.66
Water Samples				
Mull	S	4.44E+02	3.40E+01	7.65
Sawmill	S	3.29E+02	4.20E+01	12.76
Beechflat	Ν	3.45E+02	2.60E+01	7.53
Flat Laurel	Ν	4.23E+02	1.60E+01	3.78
Shoal	Ν	6.62E+02	7.00E+01	10.57
Scapecat	Н	1.82E+02	4.60E+01	25.27

Table 8. Statistical summaries of ANOVA and Tukey's pairwise comparisons for percent private CFUs based on total abundance. Abbreviations: degrees-of-freedom (df), sum-of-squares (SS), mean-squared (MS), F-value (F), p-value (P), standard error (SE), tvalue (t).

ANOVA									
Source	df	SS	MS	F	Р				
Sample Origin	5	1.9014	0.3803	7.303	0.0156				
Error	6	0.3124	0.0521						
Tukey Pair-wise Comparisons:									
Comparison	Estimate	SE	t	Р	-				
HBKT.W-HBKT.F=0	-0.48273	0.32271	-1.496	0.6734	-				
NBKT.F-HBKT.F=0	-0.65519	0.26349	-2.487	0.2552					
NBKT.W-HBKT.F=0	-0.74115	0.26349	-2.813	0.1774					
SBKT.F-HBKT.F=0	0.32629	0.27948	1.168	0.8336					
SBKT.W-HBKT.F=0	-0.68662	0.27948	-2.457	0.2636					
NBKT.F-HBKT.W=0	-0.17246	0.26349	-0.655	0.9805					
NBKT.W-HBKT.W=0	-0.25842	0.26349	-0.981	0.9061					
SBKT.F-HBKT.W=0	0.80902	0.27948	2.895	0.1617					
SBKT.W-HBKT.W=0	-0.20389	0.27948	-0.73	0.9694					
NBKT.W-NBKT.F=0	-0.08596	0.18632	-0.461	0.9958					
SBKT.F-NBKT.F=0	0.98148	0.20831	4.712	0.0232					
SBKT.W-NBKT.F=0	-0.03143	0.20831	-0.151	1.0000					
SBKT.F-NBKT.W=0	1.06744	0.20831	5.124	0.0155					
SBKT.W-NBKT.W=0	0.05454	0.20831	0.262	0.9997					
SBKT.W-SBKT.F=0	-1.01291	0.22819	-4.439	0.0303					

Table 9. Percent of private CFU richness per sample based on occurrence of unique CFUtypes. From October/November samples only.

Sample Location	Strain	Total CFUs	Total Private CFUs	% Private CFUs		
Fish Samples						
Mull	S	4.00E+00	4.00E+00	100.00		
Sawmill	S	1.40E+01	1.30E+01	92.85		
Beechflat	Ν	8.50E+01	3.30E+01	38.82		
Flat Laurel	Ν	5.00E+01	1.30E+01	26.00		
Shoal	Ν	1.50E+01	3.00E+00	20.00		
Scapecat	Н	3.00E+01	2.20E+01	73.33		
Water Samples						
Mull	S	8.80E+01	3.00E+01	34.09		
Sawmill	S	7.00E+01	2.30E+01	32.85		
Beechflat	Ν	8.10E+01	1.90E+01	23.45		
Flat Laurel	Ν	6.40E+01	1.40E+01	21.87		
Shoal	Ν	1.13E+02	3.70E+01	32.74		
Scapecat	Н	5.80E+01	2.00E+01	34.48		

Table 10. Statistical summaries of ANOVA and Tukey's pairwise comparisons for percent private CFU richness based on occurrence. Abbreviations: degrees of freedom (df), sum of squares (SS), mean squared (MS), f value (F), p value (P), standard error (SE), t value (t).

Source	df	SS	MS	F	Р				
Sample Origin	5	1.319	0.26386	23.3	0.000729				
Error	6	0.068	0.01133						
Tukey Pair-wise Comparisons:									
Comparison	Estimate	SE	t	Р	_				
HBKT.W-HBKT.F=0	-0.40054	0.15051	-2.661	0.21019	-				
NBKT.F-HBKT.F=0	-0.47102	0.12289	-3.833	0.05719					
NBKT.W-HBKT.F=0	-0.49433	0.12289	-4.022	0.04682					
SBKT.F-HBKT.F=0	0.40736	0.13035	3.125	0.12444					
SBKT.W-HBKT.F=0	-0.4112	0.13035	-3.155	0.12049					
NBKT.F-HBKT.W=0	-0.07049	0.12289	-0.574	0.98895					
NBKT.W-HBKT.W=0	-0.09379	0.12289	-0.763	0.96336					
SBKT.F-HBKT.W=0	0.8079	0.13035	6.198	0.00610					
SBKT.W-HBKT.W=0	-0.01066	0.13035	-0.082	1.00000					
NBKT.W-NBKT.F=0	-0.0233	0.0869	-0.268	0.99968					
SBKT.F-NBKT.F=0	0.87839	0.09716	9.041	< 0.001					
SBKT.W-NBKT.F=0	0.05982	0.09716	0.616	0.98497					
SBKT.F-NBKT.W=0	0.90169	0.09716	9.281	< 0.001					
SBKT.W-NBKT.W=0	0.08313	0.09716	0.856	0.94285					
SBKT.W-SBKT.F=0	-0.81857	0.10643	-7.691	0.00197	_				

Table 11. Frequencies of shared CFU types among the different samples at each field site. The diagonal indicates the total discrete CFU type occurring in a sample and the off-diagonal contains the number of shared CFU type between the two samples indicated by column and row. Abbreviations: Mull Creek (ML), Beechflat Creek (BF), Scapecat Creek (SC), Flat Laurel Creek (FL), Sawmill Creek (SM), Shoal Creek (SH), suffix designation for fish sample (F), water sample (W).

Origin	Strain	ML.F	ML.W	BF.F	BF.W	SC.F	SC.W	FL.F	FL.W	SM.F	SM.W	SH.F	SH.W
ML.F	S	4											
ML.W	S	0	88										
BF.F	Ν	0	31	85									
BF.W	Ν	0	38	32	81								
SC.F	Н	0	2	2	2	30							
SC.W	Н	0	14	14	14	7	58						
FL.F	Ν	0	18	13	14	3	15	50					
FL.W	Ν	0	19	15	18	3	13	22	64				
SM.F	S	0	0	0	0	0	0	0	1	14			
SM.W	S	0	17	15	17	3	11	9	19	0	70		
SH.F	Ν	0	3	3	3	0	1	1	2	0	6	15	
SH.W	Ν	0	29	27	36	3	15	16	24	0	32	10	113



Figure 1: Box plot showing ranges of median CFU abundance for each grouped sample. Letters above plots represent the results of pair-wise comparisons. Origins with the same letter were not found to be significantly different. "F" refers to fish mucus samples while "W" refers to water samples. Water and fish samples were collected in different dilutions and should not be compared for relatedness or differences in abundance.



Figure 2. Box plot showing ranges of percent private CFUs based on total abundance for each grouped sample. Letters above plots represent the results of pair-wise comparisons. Origins with the same letter were not found to be significantly different. SBKT show significantly higher percentage of private colonies; however, this describes each individual sample and not the grouped location.



Figure 3. Box plot showing ranges of percent private CFUs based on richness of individual CFU types for each grouped sample. Origins with the same letter were not found to be significantly different.

DISCUSSION

Differences in microbial growth (both in quantity and composition) were observed between native southern and hatchery derived northern strain brook trout (Tables 5, 8, and 10; Figures 1, 2, and 3). The differences seen between the two strains were most likely due to their physiological differences rather than due to differences in their environments. Environmental parameters were mostly consistent among streams (Table 2). The high acidity of the second sample of Flat Laurel Creek may have been due to a rain event the night before in addition to the frequent rain experienced that week. No significant differences were observed in the CFU quantity or composition of water samples from the streams based on the strain of the resident brook trout (Tables 6, 8, and 10; Figures 1, 2, and 3).

SBKT had significantly lower CFU density than did NBKT (Table 5, Figure 1). The native southern brook trout could either have increased broad-spectrum antimicrobial activity, in either intensity or effectiveness. An alternative hypothesis, not mutually exclusive from the previous, is that the innate antimicrobial action is specialized and more effective against the local microbial community (Ellis 1999, Magnadóttir 2004). Not only did the SBKT exhibit reduced growth in terms of reduced CFU density, but they also seemed to exclude the number of CFU types found in the water as evidenced by their high percentage of private CFUs (Tables 8 and 10, Figures 2 and 3). The extent of private CFUs within SBKT appears to extend to the population level, with samples showing uniqueness among the different sample sites. This could be an indication of endemic microbes specifically associated with the SBKT, but due to their differentiation among the populations and their low abundance transiently associated CFUs should not be ruled out. NBKT showed a much lower percent of private CFUs based on both total abundance and richness and contained several repeated representatives from the water samples. This indicates that not only is there higher occurrence of environmental microbes within NBKT but that a broader spectrum of microbial species is uninhibited. Even though the test did not specifically screen for pathogens, non-pathogenic opportunistic microbes may pose a significant and more omnipresent threat to fishes' health (Ellis 2001, Magnadóttir 2004). Common water microbes such as *Saprolegnia* readily infect fish after injury, mucal removal, or other stressors (Pickering and Willoughby 1982).

Due to the inhibitive properties of fish slime, the majority of the cultured microbes may have only been transiently associated with the outermost layer and therefore not part of a stable population established within the mucosal layers (Cahill 1990, Ellis 2001). The data could be representative of a short window of what was present on the exterior of the fish at the moment of extraction (Cahill 1990). This effect of ephemeral microbial association may be responsible for the high percent of private CFUs observed in the SBKT samples (Tables 8 and 10, Figures 2 and 3). In essence, some of the observed growth may not be representative of a stable or established population of microbes that could grow in the mucus uninhibited. However, nonpathogenic microbes are more readily inhibited than pathogenic species which have developed strategies to allow for their penetration of the slime (Jung *et al.* 2000, Ellis 1999, 2001, Magnadóttir 2004). Thus, the CFUs cultured from slime samples represents a sample of the river water microbes the slime was exposed to and/or a sample of associated species of organisms that penetrated the mucus and are established or both.
Regardless of how the microbes came to be and whether they represent pathogenic microbes or not, a clear trend in microbial quantity and composition was observed within the samples. Considering the broad-spectrum activity of the slime (Hellio *et al.* 2002, Subramanian *et al.* 2008), the data represent a significant difference in the number of CFUs present in two strains of fish from the same species. The analysis of possible reasons and pressures attributing to the differences in presumed antimicrobial effectiveness and microbial composition could help us understand how hatchery environments and/or historic isolating factors can affect the reaction of fishes' mucosalbased innate immune systems to the microbial environment.

Overall, it appears that the SBKT in their natural habitat show far greater antimicrobial abilities than do the NBKT in nonnative waters. This is most likely due to either artificial selection from a hatchery environment or an artifact of historic isolation in their native habitat. The latter option would suggest a difference based on evolution to combat endemic microbes (Ellis 1999, Magnadóttir 2004) or different historic environmental factors altering the innate immune response (Blackstock and Pickering 1982, Lebedeva 1999), and thus an increased susceptibility to microbial establishment may be seen when they are subjected to alien microbes in new environments.

The SBKT have been genetically isolated, not only from NBKT, but from other populations of SBKT since the Pleistocene glaciations (Hayes *et al.* 1996). Their isolation and island population distribution could lead to selection for specialized immune responses limiting establishment and possible infection from local microbes (Agarwal *et al.* 1979, Zuchelkowski *et al.* 1981, Blackstock and Pickering 1982, Pottinger *et al.* 1984, Ellis 1999, Lebedeva 1999, Magnadóttir 2004). Thus the NBKT

may not be as well adapted to the habitats of southern Appalachia (Lennon 1967). Though the brook trout strains being sampled historically evolved in allopatry (Hayes *et al.* 1996) the physiological requirements of brook trout are fairly similar between the strains. There are differences in their traditional environmental habitats, but it is likely that they would seek similar physiological optimums (e.g. temperature, pH, dissolved oxygen, etc.) regardless of geographic location (MacCrimmon and Campbell 1969, Rashleigh *et al.* 2005, Ficke *et al.* 2009). Thus similar environmental pressures, including microbial load, would be expected.

Due to the broad-spectrum antimicrobial action of the slime identified in brook trout (Subramanian et al. 2008) and the relative similarities between natural northern and southern strain environments (Rashleigh et al. 2005, Hudy and Thieling 2008), I argue that it is more likely that the majority of differences in antimicrobial activity found in this study arose due to hatchery rearing. A hatchery environment in which natural stream microbes have been removed may result in a loss of resistance in to common stream microorganisms over time (Davis 2006). Hatchery origin rainbow trout, historically unexposed the myxosporean parasite *Ceratomyxa shasta*, exhibited significantly higher mortality rates after exposure than native rainbow trout from streams in which the parasite resides (Currens et al. 1997). Shrimpton et al. (1994) found that coho salmon (Oncorhynchus kisutch) raised in hatcheries for the first part of their lives showed increased susceptibility to pathogens when placed in a natural setting in comparison to wild fish. This may be a result of anthropogenic conditioning occurring in hatcheries (Maynard et al. 1994, Reisenbichler and Rubin 1999, Hansen 2002, Álvarez and Nicieza 2003, Davis 2006). Genetic, behavioral, and physiological differences selected by

domestication in hatcheries are well documented (Hindar *et al.* 1991, Maynard *et al.* 1994, Reisenbichler and Rubin 1999, Ellis *et al.* 2002, Hansen 2002, Álvarez and Nicieza 2003) and aspects of domestication have been shown to reduce survival of fish when stocked in natural systems (Shrimpton *et al.* 1994, Currens *et al.* 1997, Reisenbichler and Rubin 1999, Hansen 2002, Davis 2006). A degree of the reduced survival may be attributed to the low genetic variation commonly seen in hatchery fish (Hindar *et al.* 1991, Kreigler *et al.* 1995, Hayes *et al.* 1996, Hansen *et al.* 2001) which has been shown to have a negative effect on the fish's immune system (Allendorf and Phelps 1980).

The hatcheries could be providing an environment that allows for reduced antimicrobial activity. However, some researchers have concluded that hatchery rearing may produce fish that are more resistant to pathogens because of the intense selection due to generally poorer environmental conditions (Ruzzante 1994, Davis 2006). Water in hatcheries is generally of poorer quality than would be found in a natural environment (Tomasso *et al.* 1981) due to waste accumulation by fish in unnaturally high densities (Piper *et al.* 1989) as well as decomposing material from uneaten food (Cho *et al.* 1994, Conte 2004). High densities can also lead to greater rates of infection due to unnaturally close proximity and frequently compromised mucosal layer due to physical abrasion from aggressive fish interactions (Pickering 1989, Winfree *et al.* 1998, Ellis *et al.* 2002). In addition to the direct health impacts, chronic stress, which can result from the previous conditions (reviewed in: Conte 2004, Davis 2006), can have a negative impact on general health as well (Pickering and Duston 1983, Dhabhar and McEwen 1996, 1997, Barton 2002, Ellis *et al.* 2002, Magnadóttir 2004, Davis 2006). These conditions would lead one

to suspect a selective pressure for a more responsive and stronger immune system in hatchery reared fish. However, other aspects of hatchery rearing could be reducing selection for broad-spectrum innate resistance.

Though no longer an accepted practice in modern aquaculture, the heavy and unregulated use of antibiotics was once common (Watts et al. 2001, Benbrook 2002, Thurman et al. 2002, Anand et al. 2011), due to the increased potential for fish infection from the typically poorer water conditions and the overcrowding of fish (Klinger and Floyd 1998, Ellis et al. 2002). Broad-spectrum antibiotics are known to compromise innate mucosal defense in humans (Brandl et al. 2008). The mucosal innate immune system is fairly analogous among vertebrates, but out of necessity fish have higher concentrations of mucus forming cells and a larger arsenal of defensive agents (Alexander and Ingram 1992, Ellis 2001). This is because fish have a more primitive acquired immune system and are intimately in touch with the microbial environment, (Ellis 2001, Magnadóttir 2004). A history of persistent antibiotic use could lead to immunosuppression (Anand et al. 2011), as has been shown to occur in common carp (*Cyprinus carpio* L.; Rijkers *et al.* 1980). Heavy antibiotic use leading to immunosuppression may remove the pressure for individuals to have adequate immune responses. However, there is also the potential that antibiotics may select for immunosuppressed individuals. Because sperm cells are non-self, inflammation and infection can cause them to be targeted by defensive agents. Immunosuppressed fish therefore have more viable sperm which would suggest greater reproductive success (Måsvær *et al.* 2004). In a natural environment this is a positive selective pressure. Fish that are resistant to foreign-invaders which could cause inflammation would experience

greater reproductive success, while those that were immunosuppressed but susceptible would suffer reduced fitness from infection (Måsvær *et al.* 2004). However, in a hatchery with antibiotics causing the immunosuppressive tendencies, the selection towards the more immunocompromised individual would not be based on increased pathogen resistance, which could lead to the selection of fish with the greatest immunosuppression, without the benefit of resistance.

In conjunction with the use of antibiotics, other factors of hatchery life could lead to a reduction of inherited immune responses (Bosakowski and Wagner 1994, Carballo et al. 1995). The application of antibiotics in the presence of continuously stressful environments may lead to a selective pressure against natural stress responses for indication of infection. In a hatchery setting where stress is omnipresent, chronic stress responses tend to be detrimental to fish health (Dhabhar and McEwen 1996, 1997, Barton 2002, Conte 2004, Davis 2006). Chronic stressors may result in energetically taxing responses (Davis et al. 1985, Pickering 1990). Prolonged stress is generally seen to compromise the immune system (Maule et al. 1989, Pickering 1989, Dhabhar and McEwen 1996, 1997, Ortuño et al. 2001, Davis 2006), including reduced levels of mucal secretion (Barton 1987) and inhibition of defensive agents within the mucus (Hjelmeland et al. 1983). However, stress is not entirely detrimental (Barton 2002). Stress responses are evolutionary adaptations to signal potential threats and to initiate the appropriate reaction (Barton 2002, Volpato et al. 2007). Responding to stress in the appropriate way is beneficial and exposure to acute stress enhances the immune system (Pickering and Pottinger 1989, Dhabhar and McEwen 1996, 1997, Chrousos 1998, Davis 2006) and initially leads to an increase in mucal production (Barton 1987).

Due to these persistent stressors, hatchery fish might benefit from a reduced stress response and higher tolerances, and thus the selection of fish with higher stress tolerances has likely occurred in hatcheries (Woodward and Strange 1987, Davis 2006). This could promote higher survival and success in hatchery systems due to the removal of the deleterious effects of chronic stressors. Under hatchery conditions negative reactions to chronic stress may supersede the benefits of responding to acute stressors, thus the beneficial adaptation for hatchery conditions may result in the general suppression of stress and loss of an adaption for natural habitats (Davis 2006). This may result in hatchery fish responding inappropriately to potential stressors in natural settings (Reisenbichler and McIntyre 1977, Chilcote *et al.* 1986, Pickering and Pottinger 1989, Hindar *et al.* 1991, Barton 2002, Davis 2006,). Hatchery derived traits have been shown to be maladaptive and to reduce the success of fish in the wild, in addition these traits may share genetic inheritance and lead to generations of poorly adaptive fish in natural environments (Reisenbichler and Rubin 1999, Hansen 2002).

The water quality of hatcheries can impact mucosal effectiveness (Lang *et al.* 1987, Grinde 1989, Mock and Peters 1990), due to waste accumulation from high fish densities (Piper *et al.* 1989). Hatcheries have higher ammonia levels than would occur in natural environments (Piper *et al.* 1989) and ammonia is known to interfere with mucus renewal (Lang *et al.* 1987) and reduce the presence of defensive agents in the slime of rainbow trout (Grinde 1989, Mock and Peters 1990). Other toxicants also interfere with mucal production and suppress the innate immune system (reviewed in: Carballo *et al.* 1995, Bols *et al.* 2001). Hatchery substrate has a significant effect on the disease susceptibility and welfare of trout (Bosakowski and Wagner 1994) and other fish based

on abrasiveness (Mahoney *et al.* 1973) (i.e. steel or concrete enclosures lead to greater instances of fin, scale, and mucus removal). Other common hatchery practices attributing to epidermal abrasions include handling, transport, and overcrowding (Abbott and Dill 1985). In conjunction with the treatment of antibiotics (Benbrook 2002, Thurman *et al.* 2002), the reduction of mucal effectiveness through abrasions and toxicants may result in the repeated replacement of slime as a pointless and expensive process. This could create a pressure within hatchery settings selecting for less slime production and less investment of defensive agents within the slime.

Since the environmental microbes were largely inhibited in SBKT, the possibility of endemic microbes specific to the fish may explain the presence of the private CFUs (Tables 8 and 10, Figures 2 and 3; Austin 1982, 1983). Several microbes have been identified within the mucus of fish that exhibit their own antimicrobial properties (Ebran *et al.* 1999, Parret *et al.* 2005). In addition, if uninhibited, the integument of fish could present itself as a beneficial environment for the microbe. Living in a microbially hostile environment would present little competition for the microbe and relatively homeostatic conditions as well as a stable carbon source (Bordas *et al.* 1996). Evidence of microbial mutualisms associated with the integument of red-backed salamanders (*Plethodon cinereus*) has been shown (Lauer *et al.* 2007) and similar relationships may exist within the mucus of fish (Subramanian *et al.* 2008).

Because the sampling of the hybridized stream Scapecat Creek was accidental, the sample of HBKT was from only a single stream. However, it is interesting to note that CFU abundance (Figure 1) and the percent of private CFUs were intermittent between the two pure strains (Figures 2 and 3). Aspects of innate immune response are known to be

heritable (Secombes and Olivier 1997); and though the NBKT studied in this experiment were of hatchery descent, these fish represent wild populations separated by their hatchery predecessors by many generations. If the selective pressures of ecological isolation or hatchery conditioning led to these differences in innate immune activity, then these characteristics have persisted through several generations and have strong basis for genetic inheritance. This creates a problematic situation in terms of introgression between the strains. It is well documented that in many occasions hatchery-reared brook trout and other salmonids experience lower survival than wild fish after being stocked in natural settings (Greene 1952, Miller 1952, Salo and Bayliff 1958, Reimers 1963, Reisenbichler and McIntyre 1977, Fraser 1981, Webster and Flick 1981,). Though there have been several identified causes for this trend, the possibility of a more effective innate immune system in native fish may be an additional factor attributing to the reduced survivability commonly seen in transplanted hatchery fish. A concern over the loss of genetic integrity in native fish through the introgression of introduced hatchery stock has been a well discussed topic of concern (Allendorf and Phelps 1980, Ferguson 1990, Krueger and May 1991). Galbreath et al. (2001) demonstrated that introgression has frequently occurred between SBKT and NBKT. Though the degree to which wild trout hybridize with cultivated fish is only partially known and understood (Galbreath *et al.*) 2001, Hansen 2002) empirical evidence shows that at least in some situations it does occur in spite of selective forces acting against hatchery raised fish (reviewed in: Hansen 2002). Hansen (2002) showed that introgression between native and hatchery brown trout (Salmo trutta) does not occur as frequently as expected if both strains were equally contributing to new progenies of fish. It is hypothesized that the lack of introgression of

the hatchery stock into the gene pool of native trout may be due to the poor survival and maladaptedeness of the introduced strain (Hansen 2002). Though maladaptive traits will be selected against in a natural environment, introgression still occurs and may have detrimental effects on native populations (Lynch and O'Hely 2001, Hansen 2002). Negative effects on fitness may also occur long after stocking has ceased (Lynch and O'Hely 2001). However, introgression may also explain some trends favoring heterosis in wild X hatchery hybrids. Webster and Flick (1981) showed reduced survival of hatchery ancestry brook trout reared in a wild setting when compared to wild fish but found the survival of hybrids to be as great as or greater than the wild fish. However, the presence of poor alleles is still potentially detrimental. Though a degree of initial heterosis may be exhibited in early generations, the continuous introgression of fish with impaired innate immune systems could swamp the adaptive genetics of native fish and lead to an overall population of fish with maladapted innate immune responses (Allendorf and Phelps 1980, Hindar et al. 1991, Hansen and Loeschcke 1994). Though not conclusive due to small sample size, it appears likely that the introgression of hatchery brook trout into native populations may reduce the activity of the innate immune system, potentially threatening the fitness of wild populations.

CONCLUSIONS

The inherited immunities or microbes associated with an organism's ability to protect itself from potentially harmful invaders may be shaped by the historic environment of the organism. Propagation of many generations of fish in unnatural hatchery settings may also apply evolutionary pressures, making them less fit in natural settings. The fish in the streams sampled represent artifacts of historical hatcheries and stocking practices and do not necessarily reflect the artificial pressures of hatcheries today. Antibiotics, immunization, and chronic anthropogenic stress may play major roles in the suppression of the immune system in hatchery fish. These changes could become problematic if these fish hybridize with native fish. These results suggest further reasons for caution when stocking fertile fish in areas where they could interbreed with native fish species. While in North Carolina, hatchery fish are no longer stocked into streams with naturally reproducing populations, and in general, hatcheries are moving toward production of non-fertile fish for recreational stocking (Davis 2006, Wilson 2011), selective pressures of modern hatcheries may still have a negative effect on the innate immune response. Thus, hatcheries rearing fish intended for supplementation, should consider rearing fish in more natural settings with lower densities promoting better water quality, reducing the need of antimicrobial additives. Additionally hatchery fish may benefit from exposure to acute stressors while minimizing chronic stressors, promoting a more natural and robust stress response. The mimicking of natural settings and reduction of anthropogenic influences should more appropriately create natural conditions for hatchery fish intend for supplementation, and may reduce the potential for genetic or physiological changes due to hatchery selection.

46

- Abbott, J. C. and L. M. Dill, 1985. Patterns of aggressive attack in juvenile steelhead trout (*Salmo gairdneri*). *Canadian Journal of Fisheries and Aquatic Sciences* 42: 1702-1706.
- Agarwal, S. K., T. K. Banerjee, and A. K. Mittal, 1979. Physiological adaptation in relation to hyperosmotic stress in the epidermis of a fresh-water teleost Barbus sophor (*Cypriniformes, Cyprinidae*): a histochemical study. *Zeitschrift fur Mikroskopisch-Anatomische Forschung* **93**: 51–64.
- Alexander J. B. and G. A. Ingram, 1992. Noncellular nonspecific defense mechanisms of fish. Annual Review of Fish Diseases 2: 249-279.
- Allendorf, F. W. and S. R. Phelps, 1980. Loss of genetic variation in a hatchery stock of cutthroat trout. *Transactions of the American Fisheries Society* **109**: 5: 537-543.
- Álvarez, D. and A. G. Nicieza, 2003. Predator avoidance behavior in wild and hatcheryreared brown trout: the role of experience and domestication. *Journal of Fish Biology* **63**: 1565–1577.
- Anand, J., A. A. Thomas, and M. P. Salini, 2011. Fluctuations in the microbial load at certain nonspecific immune sites of *Macrobrachium rosenbergii* supplemented with *Ocimum sanctum*. *The Ecoscan*, Special Issue 1: 389-392.
- Aranishi F. and M. Nakane., 1997. Epidermal proteases of the Japanese eel. Fish Physiology and Biochemistry 16: 471-478.
- Austin, B., 1982. Taxonomy of bacteria isolated from a coastal, marine fishrearing unit. *Journal of Applied Microbiology* **53**: 253-268.

- Austin, B., 1983. Bacterial microflora associated with a coastal, marine fish-rearing unit. *Journal of the Marine Biological Association of the United Kingdom* **63**: 585-592.
- Austin, B. and D. McIntosh, 1988. Natural antibacterial compounds on the surface of rainbow trout, *Salmo gairdneri* Richardson. *Journal of Fish Diseases* 11: 275– 277.

Balasubramanian S., R. P. Baby, P. A. Arul, M. Prakash, P. Senthilraja, and G.
Gunasekaran , 2011. Antimicrobial properties of skin mucus from four freshwater cultivable Fishes (*Catla catla, Hypophthalmichthys molitrix, Labeo rohita* and *Ctenopharyngodon idella*). African Journal of Microbiology Research 6: 24: 5110-5120.

- Barton, B. A., 1987. Effects of chronic cortisol administration and daily acute stress on growth, physiological conditions, and stress responses in juvenile rainbow trout. *Diseases of Aquatic Organisms* 2: 173-185.
- Barton, B. A., 2002. Stress in Fishes: A diversity of responses with particular reference to changes in circulating corticosteroids. *Integrative and Comparative Biology* 42: 517–525.
- Benbrook, C. M., 2002. Antibiotic drug use in U.S. aquaculture. The Northwest Science and Environmental Policy Center Sandpoint, Idaho. *Integrated Agriculture Training Program* Report: 1-18.
- Blackstock, N. and A. D. Pickering, 1982. Changes in the concentration and histochemistry of epidermal mucous cells during the alevin and fry stages of the brown trout *Salmo trutta*. *Journal of Zoology* **197**: 463–471.

- Bly, J. and W. Clem, 1991. Temperature-mediated processes in teleost immunity: in vitro immunosupression induced by in vivo low temperature in channel catfish. *Veterinary Immunology and Immunopathology* 28: 365–377.
- Bols, N. C., J. L. Brubacher, R. C. Ganassin, and L. E. J. Lee, 2001. Ecotoxicology and innate immunity in fish. *Developmental and Comparative Immunology* 25: 853-873.
- Bordas, M. A., M. C. Balebona, I. Zorilla, J. J. Borrego, and M. A. Morinigo,
 1996. Kinetics of adhesion of selected fish-pathogenic *Vibro* strains of
 skin mucus of gilt-head sea bream (*Sparus aurata* L.). *Applied and Environmental Microbiology* 62: 3650-654.
- Bosakowski, T. and E. J. Wagner, 1994. A survey of trout fin erosion, water quality, and rearing conditions at state fish hatcheries in Utah. *Journal of the World Aquaculture Society* **25**: 2: 308-316.
- Brandl, K., G. Plitas, C. N. Mihu, C. Ubeda, T. Jia, M. Fleisher, B. Schnabl, R. P. DeMatteo, and E. G. Pamer, 2008. Vancomycin-resistant enterococci exploit antibiotic-induced innate immune deficits. *Nature* 455: 804-808.
- Breakwell, D., C. Woolverton, B. MacDonald, K. Smith, and R. Robison. 2007. Colony morphology protocol. The Microbe Library (<u>http://www.microbelibrary.org/library/laboratory-test/3136-colony-morphologyprotocol</u>). Accessed 30 October 2012.
- Cahill, M. M., 1990. Bacterial flora of fishes: A review. *Microbial Ecology* 10: 21-41.
- Carballo, M., M. J. Muñoz, M. Cuellar, and J. V. Tarazona, 1995. Effects of waterborne copper, cyanide, ammonia, and nitrite on stress parameters and changes in

susceptibility to Saprolegniosis in rainbow trout (*Oncorhynchus mykiss*). Applied and Environmental Microbiology **61**: 2108–2112.

- Chilcote, M. W., S. A. Leider, and J. J. Loch. 1986. Differential reproductive success of hatchery and wild summer-run steelhead under natural conditions. *Transactions* of the American Fisheries Society 115: 726–735.
- Cho, C. Y., J. D. Hynes, K. R. Wood, and H. K. Yoshida, 1994. Development of highnutrient-dense, low-population diets and prediction of aquaculture wastes using biological approaches. *Aquaculture* 124: 293-305.
- Chrousos, G. P., 1998. Stressors, stress, and neuroendocrine integration of the adaptive response. *Annals of the New York Academy of Sciences* **851**: 311–335.
- Conte, F. S., 2004. Stress and the welfare of cultured fish. *Applied Animal Behaviour Science* **86**: 205–223.
- Culp, C. E., J. O. Falkinham III, and L. K. Belden, 2007. Identification of the natural bacterial microflora on the skin of Eastern newts, bullfrog tadpoles and Redback salamanders. *Herpetologica* 63: 66–71.
- Currens, K.P., A. R. Hemmingsen, R. A. French, D. V. Buchanan, C. B. Schreck, and H. W. Li, 1997. Introgression and susceptibility to disease in a wild population of rainbow trout. *North American Journal of Fisheries Management* 17: 1065-1078.
- Davis, K. B., 2006. Management of physiological stress in finfish aquaculture. North American. *Journal of Aquaculture* **68**: 116–121.
- Davis, K. B., P. Torrance, N. C. Parker, and M. A. Suttle. 1985. Growth, body composition, and hepatic tyrosine aminotransferase activity in cortisol-fed

channel catfish, *Ictalurus punctatus* Rafinesque. *Journal of Fish Biology* **27**: 177-184.

- Dhabhar, F. S. and B. S. McEwen, 1996. Stress-induced enhancement of antigen-specific cell mediated immunity. *Journal of Immunology* **156**: 2608–2615.
- Dhabhar, F. S. and B. S. McEwen, 1997. Acute stress enhances while chronic stress suppresses immune function in vivo: a potential role for leucocyte trafficking. *Brain Behavior and Immunology* **11**: 286–306.
- Ebran, N., S. Julien, N. Orange, P. Saglio, C. Lemaître, and G. Molle, 1999. Poreforming properties and antibacterial activity of proteins extracted from epidermal mucus of fish. *Comparative Biochemistry and Physiology* **122**: 181–189.
- Ellis, A. E., 1974. Non-specific defense mechanisms in fish and their role in disease processes. *Developments in Biological Standardization* **49**: 337–352.
- Ellis, A. E., 1982. Differences between the immune mechanisms of fish and higher vertebrates. In: Roberts RJ (ed) *Microbial diseases of fish*. Academic Press, New York, London: 1-29.
- Ellis, A. E., 1999. Immunity to bacteria in fish. *Fish and Shellfish Immunology* **9**: 291-308.
- Ellis, A. E., 2001. Innate host defense mechanisms of fish against viruses and bacteria. *Developmental and Comparative Immunology* **25**: 827–839.
- Ellis, T., B. North, A. P. Scott, N. R. Bromage, M. Porter, and D. Gadd, 2002. The relationships between stocking density and welfare in farmed rainbow trout. *Journal of Fish Biology* **61**: 493–531.

- Fraser, J. M., 1981. Comparative survival and growth of planted wild, hybrid and domestic strains of brook trout (*Salvelinus fontinalis*) in Ontario lakes.
 Canadian Journal of Fisheries and Aquatic Sciences 38: 1672-1684.
- Ferguson, M. M., 1990. The genetic impact of introduced fishes on native species. Canadian Journal of Zoology 68: 1053-1057.
- Ficke, A. D., D. P. Peterson, and B. Janowsky, 2009. Brook trout (*Salvelinus fontinalis*): a technical conservation assessment: 1-58.
- Fouz, B., S. Devesa, K. Gravningen, J. L. Barja, and A. E. Toranzo, 1990. Antibacterial action of the mucus of turbot. Bulletin: *European Association of Fish Pathologists* 10: 56–59.
- Galbreath, P. F., N. D. Adams, S. Z. Guffey, C. J. Moore, and J. L. West. 2001.
 Persistence of native southern Appalachian brook trout populations in the Pigeon river system, North Carolina. *North American Journal of Fisheries Management* 21: 927–934.
- Gilbert, P. W., 1944. The alga-egg relationship in *Ambystoma maculatum*, a case of symbiosis. Ecological Society of America. *Ecology* **25**: 3: 366-369.
- Grinde, B., 1989. Lysozyme from rainbow trout, *Salmo gairdneri*, as an antibacterial agent against fish pathogens. *Journal of Fish Diseases* **12**: 95–104.
- Grinde, B., J. Jolles, and P. Jolles, 1988. Purification and characterization of two lysozymes from rainbow trout (*Salmo gairdneri*). European Journal of Biochemistry **173**: 269–273.
- Greene, C. W., 1952. Results from stocking brook trout of wild and hatchery strains at Stillwater Pond. *Transactions of the American Fisheries Society* **81**: 43-52.

- Hansen, M. M., 2002. Estimating the long-term effects of stocking domesticated trout into wild brown trout (*Salmo trutta*) populations: an approach using microsatellite DNA analysis of historical and contemporary samples. *Molecular Ecology* 11: 1003–1015.
- Hansen, M. M., D. E. Ruzzante, E. E. Nielsen, and K. D. Mensberg, 2001. Brown trout (Salmo Trutta) stocking impact assessment using microsatellite DNA markers. *Ecological Applications* 11: 1: 148-160.
- Hansen, M. M. and V. Loeschcke, 1994. Effects if releasing hatchery-reared brown trout to wild trout populations. In *Conservation Genetics*, Edited by V. Loeschcke, J. Tomiuk, and S. K. Jain. Boston: Berlin: Birkhäuser: 273-290.
- Hayes, J. P., S. Z.Guffey, F. J. Kriegler, G. F. McCracken, and C. R. Parker, 1996. The genetic diversity of native, stocked, and hybrid populations of brook trout in the southern Appalachians. *Conservation Biology* **10**: 1403–1412.
- Hellio, C., A. M. Pons, C. Beaupoil, N. Bourgougnon, and Y. L. Gal, 2002. Antibacterial, antifungal and cytotoxic activities of extracts from fish epidermis and epidermal mucus. *International Journal of Antimicrobial Agents* 20: 214–219.
- Hindar, K., N. Ryman, and F. Utter, 1991. Genetic effects of cultured fish on natural fish populations. *Canadian Journal of Fisheries and Aquatic Sciences* **48**: 945–957.
- Hjelmeland, K., M. Chrisite, and J. Raa, 1983. Skin mucus protease from rainbow trout, Salmo gairdneri Richardson, and its biological significance. Journal of Fish Biology 23: 13–22.

- Hudy, M. and T. M. Thieling, 2008. Distribution, status, and land use characteristics of subwatersheds within the native range of brook trout in the eastern United States.
 North American Journal of Fisheries Management 28: 1069–1085.
- Ingram, G.A., 1980. Substances involved in the natural resistance of fish to infection a review. *Journal of Fish Biology* **16**: 23–60.
- Jones, R. D., 1975. Regional distribution trends of the trout resource. Proceedings of the southeastern trout resource: ecology and management symposium. Virginia Polytechnic Institute. Blacksburg.
- Jung, T. S., K. Thompson, and A. Adams, 2000. Identification of sialic acid on *Photobacterium damsela* subspecies *pisicida* - possible role in cell adhesion and survival in the fish host. *Fish and Shellfish Immunology* **10**: 285.
- Kırkan, S., E. Ö. Göksoy, and O. Kaya, 2003. Isolation and antimicrobial susceptibility of *Aeromonas salmonicida* in rainbow trout (*Oncorhynchus mykiss*) in Turkey Hatchery Farms. *Journal of Veterinary Medicine* **50**: 339–342.
- Klinger, R.E. and R. F. Floyd, 1998. Introduction to freshwater fish parasites. University of Florida, Institute of Food and Agricultural Sciences, Extension: 1-13.
- Kreigler, F. J., G. F. McCracken, J. W. Habera, and R. J. Strange, 1995. Genetic characterization of Tennessee brook trout populations and associated management implications. *North American Journal of Fisheries Management* **15**: 804–813.
- Krueger, C. C. and B. May, 1991. Ecological and genetic effects of salmonid introductions in North America. *Canadian Journal of Fish Aquatic Science* 1: 66-77.

- Lang, T., G. Peters, R. Hoffmann, and E. Heyer, 1987. Experimental investigation on the toxicity of ammonia: effects on ventilation Frequency, growth, epidermical mucous cells and gill structure of rainbow trout, *Salmo gairdneri*. *Diseases of Aquatic Organisms* **3**: 159–165.
- Lauer, A., M. A. Simon, J. L. Banning, B. A. Lam, and R. N. Harris, 2008. Diversity of cutaneous bacteria with antifungal activity isolated from female four-toed salamanders. *The International Society of Microbial Ecology Journal* 2: 145–157.
- Lauer, A., M. A. Simon, J. L. Banning, E. André, and K. Duncan, 2007. Common cutaneous bacteria from the eastern red-backed salamander can inhibit pathogenic fungi. *Copeia* 3: 630-640.
- Lebedeva, N.Y., 1999. Skin and superficial mucus of fish: biochemical structure and functional role. In: Saksena, D.N. (Ed.), Ichthyology: *Recent Research Advances*. Science publishers, New Hampshire: 179–193.
- Lennon, R. E. 1967. Brook trout of the Great Smoky Mountains National Park. U.S.Bureau of Sport Fish Technical Paper 15. Department of the Interior, Washington, D.C.
- Lynch, M. and M. O'Hely, 2001. Captive breeding and the genetic fitness of natural populations. *Conservation Genetics* **2**: 363–378.
- MacCrimmon, H. R. and J. S. Campbell, 1969. World distribution of brook trout, Salvelinus fontinalis. Journal of the Fisheries Research Board of Canada 26: 1699–1725.

- Madetoja, J., P. Nyman, and T. Wiklund, 2000. Flavobacterium psychrophilum, invasion into and shedding by rainbow trout Oncorhynchus mykiss. Diseases of Aquatic Organisms 43: 27–38.
- Magnadóttir, B., 2004. Innate immunity of fish (overview). *Fish & Shellfish Immunology* **20**: 137-151.
- Mahoney, J. B., F. H. Midlege, and D. G. Deuel, 1973. A fin rot disease of marine and euryhaline fishes in the New York Bight. *Transactions of the American Fisheries Society* 102: 596-605.
- Måsvær, M., S. Liljedal, and I. Folstad, 2004. Are secondary sex traits, parasites and immunity related to variation in primary sex traits in the Arctic charr? *Proceedings of the Royal Society B*, London **271**: S40-S42.
- Maule, A. G., R. A. Tripp, S. L. Kaattari, and C. B. Schreck, 1989. Stress alters immune function and disease resistance in chinook salmon (*Oncorhynchus tshawytscha*) *Journal of Endocrinology* **120**: 135-142.
- Maynard, D. J., T. A. Flagg, and C. V. W. Mahnken, 1994. A review of seminatural culture strategies for enhancing the postrelease survival of anadromous salmonids. Coastal Zone and Estuarine Studies Division., Northwest Fisheries Science Center, National Marine Fisheries Service, and National Oceanic and Atmospheric Administration, Montlake Boulevard E. Seattle, Washington.
- McCracken, G. F., C. R. Parker, and S. Z. Guffey, 1993. Genetic differentiation and hybridization between stocked hatchery and native brook trout in Great Smoky

Mountains National Park. *Transactions of the American Fisheries Society* **122**: 533–542.

- Miller, R. B., 1952. Survival of hatchery-reared cutthroat trout in an Alberta stream. *Transactions of the American Fisheries Society* **81**: 35-42.
- Mock, A. and G. Peters, 1990. Lysozyme activity in rainbow trout, *Oncorhynchus mykiss*, stressed by handling, transport and water pollution. *Journal of Fish Biology* 37: 873–885.
- Nagashima, Y., A. Sendo, K. Shimakura, T. Kobayashi, Kimura, and T. Fujii, 2001. Antibacterial factors in skin-mucus of rabbitfishes. *Journal of Fish Biology* 58: 1761–1765.
- Ortuño J., M. A. Esteban, and J. Meseguer 2001. Effects of short-term crowding stress on the gilthead seabream (*Sparus aurata* L.) innate immune response. *Fish & Shellfish Immunology* **11:** 187–197.
- Parret, A. H. A., K. Temmerma, and R. D. Mot, 2005. Novel lectin-like bacteriocins of biocontrol strain *Pseudomonas fluorescens* Pf-5. Applied *and Environmental Microbiology* 71: 9: 5197–5207.
- Pickering, A. D., 1974. The distribution of mucus cells in the epidermis of the brown trout Salmo trutta (L.) and the char Salvelinus alpinus (L). Journal of Fish Biology 6: 111–118.
- Pickering, A. D., 1989. Factors affecting the susceptibility of salmonid fish to disease.In: Fifty-seventh annual report for the year. Ambleside, UK, *Freshwater Biological Association:* 61-80.

- Pickering, A. D. 1990. Stress and the suppression of somatic growth in teleost fish. *Progress in Comparative Endocrinology*. Edited by A. Epple, C.G. Scanes and M.H. Stetson. Wiley-Liss, New York: 473-479.
- Pickering, A. D. and D. J. Duston, 1983. Administration of cortisol to brown trout, Salmo trutta L., and its effects on the susceptibility to Saprulegnia infection and furunculosis. Journal of Fish Biology 23: 163-175.
- Pickering, A. D. and L. G. Willoughby, 1982. Saprolegnia infections of salmonid fish. In: Annual Report, Freshwater Biological Association, Ambleside. Fiftieth annual report: 38-48.
- Pickering, A. D. and T. G. Pottinger, 1989. Stress responses and disease resistance in salmonid fish: Effects of chronic elevation of plasma cortisol. *Fish Physiology* and Biochemistry 7: 1-4: 253-258.
- Piper, R. G., I. B. McElwain, L. E. Orme, J. P. McCraren, L. J. Fowler, and J. R. Leonard, 1989. *Fish hatchery management*, 4th edition. U.S. Fish and Wildlife Service, Washington, D.C.
- Pottinger, T. G, A. D. Pickering, and N. Blackstock, 1984. Ectoparasite induced changes in epidermal mucification of the brown trout, *Salmo trutta* L. *Journal of Fish Biology* 25: 1: 123-128.
- Rashleigh, B., R. P. Parmar, J. M. Johnston, and M. C. Barber, 2005. Predictive habitat models for the occurrence of stream fishes in the mid-Atlantic Highlands. *North American Journal of Fisheries Management* 25: 1353–1336.

- Reimers, N., 1963. Body condition, water temperature, and overwinter survival of hatchery reared trout in Convict Creek, California. *Transactions of the American Fisheries Society* **92**: 39-46.
- Reisenbichler, R. R. and J. D. McIntyre, 1977. Genetic differences in growth and survival of juvenile hatchery and wild steelhead trout, *Salmo gairdneri*. *Journal of the Fisheries Research Board of Canada* **34**: 123–128.
- Reisenbichler, R. R. and S. P. Rubin, 1999. Genetic changes from artificial propagation of Pacific salmon affect the productivity and viability of supplemented populations. ICES *Journal of Marine Science* 56: 459–466.
- Rijkers, G. T., A. G. Teunissen, R. Van Oosterom, and W. B. Van Muiswinkel, 1980.
 The immune system of cyprinid fish. The immunosuppressive effect of the antibiotic oxytetracycline in carp (*Cyprinus carpio* L.). *Aquaculture* 19: 2: 177–189.
- Rombout J., N. Taverne, M. van de Kamp, and A. J. Taverne-Thiele, 1993. Differences in mucus and serum immunoglobulin of carp (*Cyprinus carpio* L.). *Developmental* and Comparative Immunology 17: 309-317.
- Ruzzante, D. E., 1994. Domestication effects on aggressive and schooling behavior in fish. *Aquaculture* **120**: 1–24.
- Salo, E. O. and W. H. Bayliff, 1958. Artificial and natural production of silver salmon, Oncorhynchus kisutch, at Minter Creek, Washington. Washington Department of Fisheries Research Bulletin 4.
- Sarmaşik, A., 2002. Antimicrobial peptides: a potential therapeutic alternative for the treatment of fish diseases. Turkey Journal Biology **26**: 201–207.

- Secombes, C. J. and G. Olivier, 1997. Host-pathogen interactions in salmonids. In:
 Bernoth, E. M., A. E. Ellis, P. J. Midtlyng, G. Olivier, and P. Smith, editors.
 Furunculosis. Multidisciplinary fish disease research. *Academic Press* 1997: 269-296.
- Shephard, K. L., 1993. Mucus on the epidermis of fish and its influence on drug delivery. *Advanced Drug Delivery Reviews* **11**: 403–417.
- Shrimpton, J. M., N. J. Bernier, G. K. Iwama, and D. J. Randall, 1994. Differences in measurements of smolt development between wild and hatchery-reared juvenile coho salmon (*Oncorhynchus kisutch*) before and after saltwater exposure.
 Canadian Journal of Fisheries and Aquatic Sciences 51: 2170–2178.
- Subramanian, S., N. W. Ross, and S. L. MacKinnon, 2008. Comparison of antimicrobial activity in the epidermal mucus extracts of fish. *Comparative Biochemistry and Physiology*. Part B **150**: 85–92.
- Tomasso, J. R., K. B. Davis, and N. C. Parker, 1981. Plasma corticosteroid dynamics in channel catfish (*Ictalurus punctatus*) during and following oxygen depletion. *Journal of Fish Biology* 18: 519–526.
- Thurman, E., J. Dietze, and E. Scribner, 2002. Occurrence of antibiotics in water from fish hatcheries. U.S. Dept. of Interior, U.S. Geological Survey Toxic Substances Hydrology Program, Fact Sheet 120: 02: 1-4.
- Volpato, G. L., E. Goncalves-de-Freitas, and M. Fernades-de-Castilho, 2007. Insights into the concept of fish welfare. *Diseases of Aquatic Organisms* **75**: 165-171.
- Watts, M., B. L. Munday, and C. M. Burke, 2001. Immune responses of teleost fish. Australian Veterinary Journal 79: 8: 570-574.

- Webster D. A. and W. A. Flick, 1981. Performance of indigenous, exotic, and hybrid strains of brook trout (*Salvelinus fontinalis*) in waters of the Adirondack Mountains, New York. *Canadian Journal of Fisheries and Aquatic Sciences* 38: 12: 1701-1707.
- Wedemeyer, G. A., 1996. Physiology of fish in intensive culture systems. Springer: 1-232.
- Wesner, J. S., J. W. Cornelison, C. D. Dankmeyer, P. F. Galbreath, and T. H. Martin, 2011. Growth, pH tolerance, survival, and diet of introduced northern-strain and native southern-strain appalachian brook trout. *Transactions of the American Fisheries Society.* 140: 1: 37 -44.
- Wilson, K. and illustrated by Tim Lee, 2011. Fish by the millions. Wildlife in North Carolina WINC. Part 1 of 3:20-24.
- Winfree, R. A., G. A. Kindschi, and H. T. Shaw, 1998. Elevated water temperature, crowding, and food deprivation accelerate fin erosion in juvenile steelhead. *Progressive Fish-Culturist* 60: 192–199.
- Wingender, J., T. R. Neu, and H. C. Flemming, 1999. What are bacterial extracellular polymeric substances? *Microbial Extracellular Polymeric Substances: Characterization, Structure and Function*: 1-19.
- Woodward, C. C. and R. J. Strange, 1987. Physiological stress responses in wild and hatchery-reared rainbow trout. *Transactions of the American Fisheries* 116: 4: 574-579.
- Zuchelkowski, E. M., R. C. Lantz, and D. E. Hinton, 1981. Effects of acid-stress on epidermal mucous cells of the brown bullhead *Ictalurus nebulosus* (Leseur): a morphometric study. *The Anatomical Record* 200: 33–39.

APPENDIX

Table 1. Characterization data of identified CFUs.

Chart heading abbreviations: CFU identification (ID), CFU count (Cnt), opaque (OPQ), translucent (TRL), shiny (SHY), dull (DUL), circular (CRC), irregular (IRR), filamentous (FIL), rhizomous (RHZ), entire (ENT), undulated (UND), lobate (LOB), erose (ERO), curled (CRL), flat (FLT), raised (RAI), convex (CVX), and umbro (UMB). In chart abbreviations: check (+), repeated CFU type ('), center (Cn), edge (Eg), pale (Pl), light (Li), heavy (Hv), very (Vy), mostly (Mo), large (Lg), small (Sm),different (Dif), similar (Sim), particles (Pts), ring (Rg), fringe (Fg), texture (Tex), possible repeat (PR), red (Rd), yellow (Yl), orange (Og), green (Gn), pink (Pk), peach (Ph), black (Blk), purple (Pp), gray (Gry), brown (Bwn), beige (Bg), tangerine (Tng), clear (Cl), white (Wht), milky (Mlk).

Sample	ID	Cnt	t Size i	n mn	n Color	Consi	stency		Color	y Appe	earance	;		Edge						Elev	/ation			Notes
		#	<1 1	-22-	7	OPQ	TRL	SHY	DUL	CRC	IRR	FIL	RHZ	ENT	UND	LOB	ERO	FIL	CRL	FLT	RAI	CVX	UMB	
M.F 1:10A	Α	1			Og, Pl, small translucent Eg	+		+		+				+								+		
M.F 1:10A	В	1		+	- Yl Cn, Li Cn, Lg Wht Eg	+		+		Cn	Eg				+								+	Fried Egg look
M.F 1:X A	C	1		+	- Og, Dk Og in Cn		Li	+		+				+					+		+			More translucent than A
M.F 1:X A	D	1		+	Ph (salMon)	+		+		+				+								Hv		Li Dif in Eg color
M.W 1:100A	B*	1		+	- Y1 Cn, Cl/Bg Eg		+	+		Cn	Eg					+							+	Sim but Dif to B and R
M.W 1:100A	50	1		+	- Pastel Yl	+		Li		+				+							+	+		
M.W 1:100A	51	1			Gn/Dk/Blk	+			+	+		+	+					+					+	Volcano shape, Fungal-like
M.W 1:100A	51'	1																						
M.W 1:100A	52	1			Cl Pl Yl/Bg		+	+		+						+			+		+			RAI w/ CVX Rgs, FLT Cn, Pts in Cn
M.W 1:100A	K	1		+	- Bg Eg, Li Rg, Yl Pts in Cn	Cn Pts	Eg	+		+					+								+	Pts make Y1 Cn
M.W 1:100B	54	1			Wht	Cn	Eg	+		+					+						+			Crater-shape, no ridges
M.W 1:100B	55	1			Cl/Bg Eg, Bwn Pts in Cn	Pts	Mo	+		+					+								+	Sim to others, Bwn Cn w/ Dk Bwn Pts
M.W 1:100B	56	1			Og w/ Dkr Pts		+	+		Mo					+							Vy Li		CRC but w/ 2 side runs, Dk Og Pts inside
M.W 1:100C	25'	1																						
M.W 1:100C	7'	1		+	-																			
M.W 1:100C	57	1		+	Li Bwn Rg, Cl Fib Eg, Dk Bwn		+	+		+	Eg			Cn	+			+				Li	+	
M.W 1:100C	58	1			Cl Eg. Li Yl Rg. Dk Yl Cn		+	+		Cn	Eg	+		Cn	+						+			
M.W 1:100C	59	1			Cl Eg, Pl Yl Cn		+	+		Cn	Eg				+								Li	Tex Cn
M.W 1:100C	60	1			Cl Eg, Bg Rg, Dkr Bg/Bwn Cn		+	+		Cn	Eg				+								Li	
M.W 1:100C	AAA	' 1																						sLight waviness (possibly countour behind)
M.W 1:10A	DD'	1	+																					
M.W 1:10A	TTT	• 1		+	 Cl w/ Pts inside 	Pts	Mo	+		+				Mo									+	Sim but Dif to TTT
M.W 1:10A	15'	1																						
M.W 1:10A	HH'	1		+	-																			
M.W 1:10A	X'	1	+																					
M.W 1:10A	G'	1																						
M.W 1:10A	61'	1		+																				
M.W 1:10A	FF'	1		+	-																			
M.W 1:10A	30'	1		+	-																			
M.W 1:10A	ZZ	1																						
M.W 1:10A	ZZZ	1		+	-																			
M.W 1:10A	62'	1		+	-																			
M.W 1:10A	64'	1	+																					
M.W 1:10A	20'	2	+	+					1	1			1					1		1		1		
M.W 1:10A	7'	4		+ +	-																			SLightly CRL
M.W 1:10A	7*'	1	11	+	-	1		1	1	1		1	1			1		1		1	1	1	1	
M.W 1:10A	65	1		+	Li Bg Eg, Dkr Cn	Li Cn	Mo Es	+		+					+				+		Li		Li	SLightly Crater-Like

Sample	ID	Cnt	: Size	in n	nm (Color	Consi	istency		Colon	y Appe	earance	.		Edge						Elev	ation			Notes
I		#	<1	1-2	2-7		OPO	TRL	SHY	DUL	CRC	IRR	FIL	RHZ	ENT	UND	LOB	ERO	FIL	CRL	FLT	RAI	CVX	UMB	
M W 1.10A	VV'	1	1 1			Pri VI CIEg						1		1	Mo		T i		1	1			1	1	IDD Eibroug Da just before Ea
M W 1.10A	67	1			+	VI/Of Sm Cl Eq	1	+	+	-	+			-	Mo	T i	LI			T ;	+		LL.	<u>ц</u> .,	IKK FIDIOUS Kg just before Eg
M.W 1.10A	67	1		+				+	+		+				WIO				_	LI			nv	Πv	TOP
M.W 1:10A	68	1		_	+	PI YI, LI Eg, Nearly CI		+	+	-	+			-		+			-	Li	+	C L			
M.W 1:10A	69	1		_	-	Bg/Li Bwn Cn, Cl Eg, Mik Fg		+	+	-	CnEg			-		+			-			CnLi			Smooth Sm Ch, Lg grainy Eg
M.W 1:10A	69'	1															+								Sim to 69 but heavier UND and coarser Tex
M.W 1:10B	70	1			+	Bri Yl Eg, Bri Og Cn		+	+		+				+							Cn			Dk Sm Pts inside
M.W 1:10B	71	1			+	Gry Dkr in Cn, Dkr Ph Rg		Mo	+		+				+					Hv			Li		Each Rg towards Cn increasingly Dkr
M.W 1:10B	72	1				Cl Eg, Bg Cn	Cn Li	+	+		+					+				Hv				Li	Frilled Eg, patterned Lines in from CRL
M.W 1:10B	73	1				Cl Eg, Cloudy Bg Cn		+	+		+				+								Li		Boardered by 72 otherwise would be CRC
M.W 1:10B	74	1			+ 1	Bg Cn, Mlk Eg		+	+		+					Li						Li			Hv dimpled golfball Tex
M.W 1:10B	75	1			+	Cl Eg, Gry Cn		+	+		+						+					+	Li		Lumpy and Crater-Like
M.W 1:10B	76	1		+		Bg Cn, Lg Cl Eg		Eg	+		+					+							Li		
M.W 1:10B	77	1]	Pl Li Yl Cn, Cl Eg	Cn	+	+		Cn	Eg				+	+						Cn		Cn Circle off centered, Tex Eg Dif from 69
M.W 1:10B	78	1			+			+	+		+					+								+	Tex Eg, CVX Cn
M.W 1:10B	77'	1			+																				
M.W 1:10B	7'	1			+																				
M.W 1:10B	7*'	1			+																				
M.W 1:10B	G'	1																							
M.W 1:10B	DD'	1		+																					
M.W 1:10B	29'	1		- I	+																				
M W 1.10B	32'	1			+																				
M W 1:10B	11'	1	1 1	-	+																				
M W 1:10B	14'	1	+ +	-	+														-	-			-		
M W 1:10D	62'	1	+ +	-	+														-	-			-		
MW 1:10D	70	1			+	Cl Eq. Wht Bg & Cn	1			-				-											
M.W 1.10C	19	1		+	- 6	Dr. Dh.Cr. Dh.Er.Er.	1	+	+	-	+ C=	Ea		-	÷	11							C _n L :		
M.W 1:10C	15	1	+ +	_	+ .	Bg, DK CII, DK Fg Eg		+	+		Cn	Eg				пv	+			-		т.:	CILL		
M.W 1:10C	81	1	+ +	_	+			LI	+		C	+			+		+			-		LI			D 1 1 F T 1 1 C C C (1 4 001 4 D)C
M.W 1:10C	82	1		_	+ .	Bg/Og broken up Cn, CI Eg,		+	+		Cn	EgLi				Hv						CnLi		6	Raised Fg, Tex space before Cn, Sim to 80 but Dif
M.W 1:10C	83	1		_		Og/YI biofilm, Dkr Cn		+	+		Li					+						~		Cn	Bio film across Most of plate, fine Tex, raised Dkr Cn point
M.W 1:10C	84	1			-	Bg Cn, Mlk Rg, Cl Eg		+	+		Cn	Li Eg				Hv				+		CnLı			a
M.W 1:10C	85	1			+	Pl Og Cn, Li Eg		+	+		+					+								+	Crater-Like
M.W 1:10C	7'	2			+																				
M.W 1:10C	11'	1			+																				
M.W 1:10C	20'	2		+																					
M.W 1:10C	59'	1			+																				
M.W 1:10C	54'	1			$^+$																				
M.W 1:10C	FF'	1			+																				
M.W 1:10C	25'	1																							
M.W 1:10C	86	1			+	Mlk Eg, Cloudy Gry Cn	Cn	Mo	+		+				Mo								+		OPQ Gry swirls in Cn
M.W 1:10C	87	1		+		Cl		+	+		+				+					+		Li			
M.W 1:10C	88	1]	Pl Yl Cn, Mlk Cl Eg		+	+		Cn	Eg				+	+			Cn			Cn		Tree-Like Rgs in Eg
M.W 1:10C	89	1				Sm Li Bg Cn. Lg Cl Eg		+	+		Cn					+								+	
M.W 1:10C	90	1			+ 1	CI		+	+		+		+					+	+					+	Crater-Like. Hy Tex Eg
M.W 1:10C	91	1				Sm Li Yl Cn. Lø Cl Eø		+	+		Cn	Eg			Cn	+							Cn		Very fine Tex Eg
M W I X A	CCC'	1		+	1																				
M.W.1:X A	EE'	1 i	+ +	+			1			1			1	1				1		1			<u> </u>		
M W I·X A	W'	1	++	+			1			1			1	1				1		1			<u> </u>		
MW1·XA	14'	1	+	+	-		1	-		1	-		1	1			-	1	1	1		-	1		
MW1.YA	A A 2'	1	1_1	г	-		-						+					+		+					
MW1.Y	BBP'	1	+	-	+		+			+				+					1	+					
MW1.XA	20'	1	++	-	+		+			+				+					-	+					
WI.W I:X A	39	1	+	_	+						<u> </u>	<u> </u>					<u> </u>			+					
M.W IXA	GGG	2	++	_	+						<u> </u>	<u> </u>	<u> </u>				<u> </u>	<u> </u>					<u> </u>		
M.W I:X A	RR3	1	1	+			I	<u> </u>		I	I	<u> </u>	I	I	I		I	I	<u> </u>		<u> </u>	<u> </u>	<u> </u>	<u> </u>	
M.W 1:X A	HH'	2	1		+		1	Î.		Î.	1	1	1	Î.	Ì		1	1	1	1	1	Î.	1	1	

Image: Constraint of the second sec	Sample	ID	Cnt	Size	in m	m Color	Const	istency		Colon	y Appe	earance			Edge						Elev	/ation			Notes
MW 1/X OV 1 0 <t< td=""><td>· ·</td><td></td><td>#</td><td><1</td><td>-2 2</td><td>-7</td><td>OPQ</td><td>TRL</td><td>SHY</td><td>DUL</td><td>CRC</td><td>IRR</td><td>FIL</td><td>RHZ</td><td>ENT</td><td>UND</td><td>LOB</td><td>ERO</td><td>FIL</td><td>CRL</td><td>FLT</td><td>RAI</td><td>CVX</td><td>UMB</td><td></td></t<>	· ·		#	<1	-2 2	-7	OPQ	TRL	SHY	DUL	CRC	IRR	FIL	RHZ	ENT	UND	LOB	ERO	FIL	CRL	FLT	RAI	CVX	UMB	
Nu Yi Xa Ya	MW1.VA	WWW	1 2	T			1								1				1		1				
NAME NA Late Late <thlate< th=""> Late Late <</thlate<>	MW1.XA	CCC'	1		+																				
NM VIA NM VIA<	M.W I:A A	LUL 17	1		_	+	-																		
M. W. A.	M.W I:X A	47	1			+																			
M. W. X.	M.W I:X A	DDD	1																						
M.W.X.A. 228 1 + -	M.W 1:X A	25'	1			+																			
M.W.IX.A 88 1 1 4 - <	M.W 1:X A	22B'	1		+																				
M.W.IXA 40 1 4	M.W 1:X A	88'	1			+																			
M.W.IXA 36 1 4	M.W 1:X A	40'	1			+																			
M.W.IXA 7 7 <td< td=""><td>M.W 1:X A</td><td>34'</td><td>1</td><td></td><td></td><td>+</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></td<>	M.W 1:X A	34'	1			+																			
MW 1XA DP 6 1 0 <t< td=""><td>M.W 1:X A</td><td>7'</td><td>7</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></t<>	M.W 1:X A	7'	7																						
MW IXA 27 9 1 4 <t< td=""><td>M.W 1:X A</td><td>DD'</td><td>6</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></t<>	M.W 1:X A	DD'	6																						
NW IXB P I <td>M W I X A</td> <td>20'</td> <td>9</td> <td></td> <td>1</td> <td></td> <td>1</td> <td></td> <td></td> <td></td> <td></td>	M W I X A	20'	9																1		1				
Div D	MW1.YB	- 20 T	1		-	+															1				
Xi Wi XB Y 1<	MW1.XB	57'	1		-1-	T	-	-		-										-		-			
NY 18 2 1 <td>MW LV D</td> <td>77</td> <td>1</td> <td></td> <td>+</td> <td></td> <td>-</td> <td></td> <td>-</td> <td></td> <td>-</td> <td></td> <td></td> <td></td> <td></td>	MW LV D	77	1		+		-												-		-				
a. W. I. B. JO. J. I.	WI.W IIA B		1	+	-+-			<u> </u>		<u> </u>	<u> </u>				l		<u> </u>	<u> </u>	<u> </u>		<u> </u>				
M.W. 18, B S8 1 i <td< td=""><td>M.W I:X B</td><td>0</td><td>2</td><td></td><td>+</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></td<>	M.W I:X B	0	2		+																				
MW 128 KK 1 + + - </td <td>M.W 1:X B</td> <td>35B'</td> <td>2</td> <td></td> <td>+</td> <td></td>	M.W 1:X B	35B'	2		+																				
MW 1X B 40° 1 4 -	M.W 1:X B	KK'	1			+																			
MW LXB FP' 3 + - - - - - - - - - MW LXB X 1 + - - - - - - - - - MW LXB X 1 + - - - - - - - - - MW LXB X 1 + - - - - - - - - - MW LXB MT - + - - - - - - - - - MW LXB RR ² 1 + - <	M.W 1:X B	40'	1			+													1		1				
MW 1XB 1* 1 + - </td <td>M.W 1:X B</td> <td>FF*'</td> <td>3</td> <td></td> <td></td> <td>+</td> <td></td>	M.W 1:X B	FF*'	3			+																			
M.W.IXB X 1 + - <t< td=""><td>M.W 1:X B</td><td>1*'</td><td>1</td><td></td><td>+</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></t<>	M.W 1:X B	1*'	1		+																				
M.W.1XB 87 1 + + - <	M.W 1:X B	X'	1	+																					
M.W.IXB HHF I	M.W 1:X B	87'	1			+																			
MW IX B 27 1 4 - 0 <	M.W 1:X B	HH'	1			+																			
MW 12 B RR3 1 + Image: Constraint of the second seco	M.W 1:X B	27'	1			+																			
MW IXB NNN 1 I	M.W 1:X B	RR3'	1		+																				
MW 11X B EEE 1 + -	M.W 1:X B	NNN'	1																1		1				
MW 1X C 88' 1 + -	M W 1·X B	EEE'	1			+													1		1				
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	M W I X C	88'	1			+																			
MW 1/X C GGG 1 + + +	MW1.XC	34'	1			+																			
M.W. IXC 3BB 1 + -	MW1XC	54	1		-	4																			
M.W. LXC J.D. I <td< td=""><td>MW LVC</td><td>25P'</td><td>1</td><td></td><td>+</td><td>T</td><td>-</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td>-</td><td></td><td>-</td><td></td><td></td><td></td><td></td></td<>	MW LVC	25P'	1		+	T	-												-		-				
M.W. HAC 22 1 + -	MW LXC	250	1		+		-												-		-				
M.W. HAC 22 1 + -	MW LXC	23	1		_	+																			
M.W. IXC 0'2' 1 + - <th< td=""><td>MWIXC</td><td>22</td><td>1</td><td></td><td>_</td><td>+</td><td>-</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></th<>	MWIXC	22	1		_	+	-																		
M.W. 12, C RKS 1 + - <t< td=""><td>M.W IXC</td><td>62</td><td>1</td><td></td><td></td><td>+</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></t<>	M.W IXC	62	1			+																			
M.W. I:XC YY I	M.W I:X C	RR3	1		+																				
M.W. I:XC 13' 1 + + - <td< td=""><td>M.W 1:X C</td><td>YYY</td><td>1</td><td></td><td></td><td>+</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></td<>	M.W 1:X C	YYY	1			+																			
M.W. I:XC I/O I <th< td=""><td>M.W 1:X C</td><td>13'</td><td>1</td><td></td><td></td><td>+</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></th<>	M.W 1:X C	13'	1			+																			
M.W.IXCCCCI++	M.W 1:X C	10'	1			+	1	1		1									1	1	1	1			
M.W.IXCUUIII<	M.W 1:X C	CCC'	1			+																			
BF.F 1:100AEI+++ <th< td=""><td>M.W 1:X C</td><td>UU'</td><td>1</td><td></td><td></td><td>+</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></th<>	M.W 1:X C	UU'	1			+																			
BF.F 1:100FI+YICn, Clear Eg++++++++II Overly defined pointed lobeBF.F 1:10AMI+PI By_LLEg++<	BF.F 1:100A	E	1			+ Wht/Bg	+		+		+					+					+				
BFF 1:100 L 1 + Pl Bg, LiEg + + + + - + -	BF.F 1:100B	F	1			+ Yl Cn, Clear Eg		+	+		+					+					+				1 Overly defined pointed lobe
BF.F 1:10A M 1 Pl YI/Og, Dk Eg + </td <td>BF.F 1:100C</td> <td>L</td> <td>1</td> <td></td> <td>+</td> <td>Pl Bg, Li Eg</td> <td></td> <td>+</td> <td></td> <td>+</td> <td>+</td> <td></td> <td></td> <td></td> <td></td> <td>+</td> <td></td> <td></td> <td></td> <td></td> <td>+</td> <td></td> <td></td> <td></td> <td></td>	BF.F 1:100C	L	1		+	Pl Bg, Li Eg		+		+	+					+					+				
BF.F 1:10A N 1 + Li Yl/Gn, Li Eg + + + + Mo Within M BF.F 1:10A O 1 + Wht Cn, Cl Eg Cn Eg + + + + Mo Within M BF.F 1:10A O' 2 + Wht Cn, Cl Eg Cn Eg + + + Mo + Within M BF.F 1:10A P 1 Cl Yl, Dk at Eg Blo Mo + + Mo + Workprecessore BF.F 1:10A Q 1 + Pure Wht Cn, Li Eg Cn Eg + + + Mo + Mostly Cl and FLT w/ Dk OPQ CVX YL Blotches of CVX areas BF.F 1:10A Q' 1 + Pure Wht Cn, Li Eg Cn Eg + + + Hv Hv Hv BF.F 1:10A Q' 1 + YICn, Plog Eg Cn Eg + + + Hv Hv BF.F 1:10A K' 1 + YICn, Plog Eg Cn Eg +	BF.F 1:10A	М	1			Pl Yl/Og, Dk Eg		+	+			+				+	+						Li	Li	Scattered CVX/UMB areas within
BFF 1:10A O I + Wht Cn, Cl Eg Cn Eg + + + - + Within M BF.F 1:10A O' 2 + + C F + + Within M BF.F 1:10A O' 2 + + C F + Within M BF.F 1:10A O' 2 + + C F H Mo + Mostly Cl and FLT w/ Dk OPQ CVX YL Blotches of CVX areas BF.F 1:10A Q 1 + Pure Wht Cn, LiEg Cn Eg + + + Hv BF.F 1:10A Q' 1 + Pure Wht Cn, LiEg Cn Eg + + H Hv BF.F 1:10A Q' 1 + Within M Hv Hv Hv Hv Hv BF.F 1:10A Q' 1 + Pure Wht Cn, LiEg Cn Eg + + Hv Hv Hv BF.F 1:10A K' 1 + Y1Cn, Plog Eg Cn Eg Hv	BF.F 1:10A	N	1			+ Li Yl/Gn, Li Eg	1	+	+	1	+				+		1	1	1	1	Mo	1			Within M
BF.F 1:10A O' 2 + F C <th< td=""><td>BF.F 1:10A</td><td>0</td><td>1</td><td></td><td>+</td><td>Wht Cn, Cl Eg</td><td>Cn</td><td>Eg</td><td>+</td><td>1</td><td>+</td><td></td><td></td><td></td><td>+</td><td></td><td>1</td><td>1</td><td>1</td><td>1</td><td></td><td>1</td><td>+</td><td></td><td>Within M</td></th<>	BF.F 1:10A	0	1		+	Wht Cn, Cl Eg	Cn	Eg	+	1	+				+		1	1	1	1		1	+		Within M
BF.F 1:10A P 1 Cl YI, Dk at Eg Blo Mo + Li + Mo + Mostly Cl and FLT w/ Dk OPQ CVX YL Blotches of CVX areas BF.F 1:10A Q 1 + Pure Wht Cn, Li Eg Cn Eg + + + Hv Hv BF.F 1:10A Q' 1 + Pure Wht Cn, Li Eg Cn Eg + + Hv Hv BF.F 1:10A Q' 1 + Pure Wht Cn, Plog Eg Cn Eg + + Hv Hv	BF.F 1:10A	O'	2	+	+	The second se	1			1	1				1		1	1	1	1	1	1			
BFF 1:10A Q 1 + Pure Wht Cn, Li Eg Cn Eg + + + Main Hv BFF 1:10A Q' 1 + -	BF.F 1:10A	Р	1			Cl Yl, Dk at Eg	Blo	Mo	+	1	1	+			1	Li	+	1	1	1	Mo	1	+		Mostly Cl and FLT w/ Dk OPQ CVX YL Blotches of CVX areas
BFF 1:10A Q' 1 + - <th< td=""><td>BF.F 1:10A</td><td>0</td><td>1</td><td></td><td>+</td><td>Pure Wht Cn. Li Eg</td><td>Cn</td><td>Eg</td><td>+</td><td>1</td><td>+</td><td></td><td></td><td></td><td>+</td><td></td><td></td><td></td><td>1</td><td>1</td><td>1</td><td></td><td>Hv</td><td></td><td></td></th<>	BF.F 1:10A	0	1		+	Pure Wht Cn. Li Eg	Cn	Eg	+	1	+				+				1	1	1		Hv		
BFF 1:104 K' 1 + Y1Cn, Pl Og Eg Cn Eg + + + + + + + + + + + + Sim to B but smaller and Dif	BF.F 1:10A	O'	1		+		1			1					1				1	1	1				
	BE E 1.10A	K'	1	+	+	YI Cn Pl Og Eg	Cr	Eσ	+	1	+					+			1	<u> </u>	1	<u> </u>		+	Sim to B but smaller and Dif
REF1:10B G 1 PLYURg Nearly CL + + Ob + 2 Li Slight ("Depression mild Tex	BE E 1:10B	G	1	+	<u> </u>	Pl Yl/Bg Nearly Cl	Cil	+	+	1		Ob			<u> </u>	+	2	1	+	1	+	Li			Slight Cn Depression mild Tex
	BEE 1:10B	н	1	+	+	VI/Bg	+		+	1	+	00			Me	-	-	1	+	1	+				Sugar on Depression, mild Tex

$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	Sample	ID	Cnt	Size	in m	m Color	Cons	istency		Color	y Appe	earance	;		Edge						Elev	ration			Notes
BIF 1000 1 <th1< th=""> 1<!--</th--><th>-</th><th></th><th>#</th><th><1 1</th><th>-2 2</th><th>2-7</th><th>OPQ</th><th>TRL</th><th>SHY</th><th>DUL</th><th>CRC</th><th>IRR</th><th>FIL</th><th>RHZ</th><th>ENT</th><th>UND</th><th>LOB</th><th>ERO</th><th>FIL</th><th>CRL</th><th>FLT</th><th>RAI</th><th>CVX</th><th>UMB</th><th></th></th1<>	-		#	<1 1	-2 2	2-7	OPQ	TRL	SHY	DUL	CRC	IRR	FIL	RHZ	ENT	UND	LOB	ERO	FIL	CRL	FLT	RAI	CVX	UMB	
Diff 100 J I<	BF.F 1:10B	Ĭ	1			+ Dk Yl/Og		Li	+		+				+						Mo				
BIT P1 00 K I Bit MULTING Co. Fig. Model Li Co. Distance	BF F 1:10B	Ĭ	1		-	+ off Wht	+		+		+					+							Li		Light inner depression
Diff File Diff File <thdiff< th=""> Diff File Diff</thdiff<>	BF F 1:10B	ĸ	1		-	Pastel VI Li Eg	Cn	Eσ	+							Ti						Τi			Depression in Cn. Textured Cn w Gn Pts
Diff Line Diff Line <thdiff line<="" th=""> <thdiff line<="" th=""> <thd< td=""><td>BEE 1:10C</td><td>п</td><td>1</td><td></td><td>-</td><td>Dk Yl Gn/Blk Pts</td><td>Ro</td><td>Eg/Cr</td><td>Rσ</td><td>Eg/Cr</td><td></td><td></td><td>+</td><td></td><td></td><td>1.1</td><td></td><td></td><td>+</td><td></td><td>+</td><td>1.1</td><td></td><td></td><td>Multi colored Fil w/ growths and Blk dots Mold Fungal-like</td></thd<></thdiff></thdiff>	BEE 1:10C	п	1		-	Dk Yl Gn/Blk Pts	Ro	Eg/Cr	Rσ	Eg/Cr			+			1.1			+		+	1.1			Multi colored Fil w/ growths and Blk dots Mold Fungal-like
Diff P LOC KK 1 <th< td=""><td>BF F 1:10C</td><td>п</td><td>1</td><td></td><td></td><td>Dk Cn Gn/Bwn Eg</td><td>Eg/Cr</td><td>R a</td><td>K5</td><td>±</td><td>+</td><td></td><td>+</td><td></td><td></td><td></td><td></td><td></td><td>+</td><td></td><td>+</td><td></td><td></td><td></td><td>Fil w/ color gradient Mold, Fungal-like</td></th<>	BF F 1:10C	п	1			Dk Cn Gn/Bwn Eg	Eg/Cr	R a	K5	±	+		+						+		+				Fil w/ color gradient Mold, Fungal-like
Bit Find Li I	BEE 1:10C	KK	1		-	+ Bright V1	Lg/CI	- Kg			-				-					-				-1-	Cux Eq and Cn
Diff 1 Co MM 1 N N 1 0 1 1 1 1 1 1 1	BF F 1:10C	II	1	+	-	Physic 11	+	т	+		Τ -				+ +					т			Hv	т	eva Eg and en
Diff 10C NN 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1 0 1	BEE 1:10C	MM	1		+	Wht	-		+		-				-								117	Hv	
Bir Filloc Do L Dinger With Column Dinger With	BF F 1:10C	NN	1		Ŧ	Dull Bwn VI/Og Dkr Eg	т	-	Ŧ	-	- T				т	+	-		-		+			110	Dkr. Lightly raised Fil Eg
Diff Ling PP L P L P L P P P P L P L P L P L P L P L P L P L P L P P L P P L P P L P P L P P L P P L P	DF.F 1:10C	00	1		-	Bright V1		т		Ŧ	- T					T	т		Ŧ		т				DKi Eighty faised Hi Eg
Diff E find O D <thd< th=""> D <thd< th=""> <thd< th=""> <thd< <="" td=""><td>BF.F 1.10C</td><td>DD</td><td>1</td><td>+</td><td>-</td><td>Dull VI</td><td>+</td><td></td><td>+</td><td>1</td><td>+</td><td></td><td></td><td></td><td>+</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td>÷</td><td>T :</td><td>Light $\mathbf{P}_{\mathbf{\sigma}}/\mathbf{C}_{\mathbf{p}}$ at ightly raised</td></thd<></thd<></thd<></thd<>	BF.F 1.10C	DD	1	+	-	Dull VI	+		+	1	+				+								÷	T :	Light $\mathbf{P}_{\mathbf{\sigma}}/\mathbf{C}_{\mathbf{p}}$ at ightly raised
Diff IXA OC I I P N La brok Dy YUCh Ch I I I I I I I P N La bord Age DEF IXA T I I P N La bord Age I	DF.F 1:10C	00	1		-	Dun 11	-	т	Ŧ		Ŧ				т									Li	Too Dificult to identify
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	DEE LV A	<u>v</u> v	1		-	DIVIER DEORVICE	C.	Ea		1						11			т.:						Flower Libra
DEF 1XA V I V </td <td>DF.F I:A A</td> <td><u>з</u></td> <td>1</td> <td></td> <td></td> <td>+ PI II Eg, DK Og/ II Cli</td> <td>Cn</td> <td>Eg</td> <td>+</td> <td></td> <td>+</td> <td></td> <td></td> <td></td> <td></td> <td>пv</td> <td></td> <td></td> <td>LI</td> <td></td> <td></td> <td></td> <td></td> <td>+</td> <td>Flower Like</td>	DF.F I:A A	<u>з</u>	1			+ PI II Eg, DK Og/ II Cli	Cn	Eg	+		+					пv			LI					+	Flower Like
DEF ISA V I </td <td>DEE LV A</td> <td>T</td> <td>1</td> <td></td> <td>-</td> <td>+ FITLIOUEI Kg</td> <td>-</td> <td>÷</td> <td>+</td> <td>1</td> <td>÷</td> <td></td> <td></td> <td></td> <td>÷</td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td>+</td> <td></td>	DEE LV A	T	1		-	+ FITLIOUEI Kg	-	÷	+	1	÷				÷									+	
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	DF.F I:A A	1	1		_	+ DI V1	-														Ma				Sim but Differen T.I. intertaine
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	BF.F I:X A	U	1		_	+ PI YI	6	+	+	-	+					+					MO		¥ .		Sim but Dif from 1 Light depression
BF FIXA W I I Prode_scale Ca Eg I I I Into V but Dim BF FIXA X I I I Prode_scale F Into V but Dim Into V but Dim BF FIXA X I I I Into V but Dim Into V but Dim BF FIXA X I I Into V but Dim Into V but Dim Into V but Dim BF FIXA X I I Into V but Dim Into V but Dim Into V but Dim BF FIXA X I I Into V but Dim Into V but Dim Into V but Dim BF FIXA X I Into V but Dim Into V but Dim Into V but Dim BF FIXA BA Into V but Dim Into V but Dim Into V but Dim Into V but Dim BF FIXA BA Into V but Dim Into V but Dim Into V but Dim Into V but Dim BF FIXA BA Into V but Dim Into V but Dim Into V but Dim Into V but Dim BF FIXA BA Into V but Dim Into V but Dim Into V but Dim Into V b	BF.F I:X A	V	1		_	+ PLOg	Cn	Eg	+		+				+								Li		61 - 111 - D16
By Fi XA X 1 + F +	BF.F I:X A	W	1		+	PLOg, CLEg	Cn	Eg	+		+				+				Lı				Lı		Sim to V but Dif
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	BF.F I:X A	X	1		+	YI/Og sm CI Eg	?	+	+		+				+								+		
Br.F. 1X.A. Y 1 0 Councy Char Print +	BF.F I:X A	X	1	+	_		-				1:0														P1 1.14
BFF IXA Y 1<	BF.F 1:X A	Y	1			Cloudy Cl w/ Pl Yl	_	+		+	Lı Cn	Eg							+		+				Fungal-like
BFF IXA Y 1 - Pastel Y1 C E C - - - - - Pastel Y1 BFF IXA AA 1 + Clav(bk R4, Pp Pis) Pis + + + - + - Pis and branches within BFF IXA AA 1 + Clav(bk R4, Pp Pis) Pis + + + + - Pis and branches within BFF IXA ABF 1 + Pis and branches within +	BF.F 1:X A	Y'	1																						
BFF IXA Z I I I Perplay Hendel Markers Perplay Ber BixA Markers Perplay Ber BixA Markers Perplay Ber BixA Ber BixA Ber BixA Ber BixA Ber BixA H H C D H Perplay Dec Markers Perplay Dec Markers <thdec markers<="" th=""> Dec Markers <th< td=""><td>BF.F 1:X A</td><td>Y"</td><td>1</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></th<></thdec>	BF.F 1:X A	Y"	1																						
BFF IXA AA I + CW / DR AD, PP S Ps + + + - - + - Pe and branches within BFF IXA BB I + Pe - <td>BF.F 1:X A</td> <td>Z</td> <td>1</td> <td></td> <td></td> <td>+ Pastel Yl</td> <td>Cn</td> <td>Eg</td> <td>Cn</td> <td>Eg</td> <td></td> <td></td> <td></td> <td></td> <td></td> <td>+</td> <td></td> <td></td> <td>Li</td> <td></td> <td></td> <td></td> <td></td> <td>+</td> <td>Fungal-like</td>	BF.F 1:X A	Z	1			+ Pastel Yl	Cn	Eg	Cn	Eg						+			Li					+	Fungal-like
BF F1XA BB 1 I + Ph LiCo Eg + + - - - Li Wordjoining colonies BFF 1XA CC I + Ph Li I - Li - - - Li - - - Li - - -	BF.F 1:X A	AA	1			+ Cl w/ Dk Rd, Pp Pts	Pts	+	+		+					+						+			Pts and branches within
BF: F1:XA BB': I I	BF.F 1:X A	BB	1			+ Ph	Li Cn	Eg	+		+				+					+				Li	two adjoining colonies
BFF IXA CC 1 + P_{0} wD kPs + + + + - <td>BF.F 1:X A</td> <td>BB'</td> <td>1</td> <td></td> <td>+</td> <td></td>	BF.F 1:X A	BB'	1		+																				
BEF I:XA Q 1 + -	BF.F 1:X A	CC	1			+ Og w/ Dk Pts		+	+		+					+				?			Li		Hv Tex and embedded w/ inner circles Like wood
BEF F1XA DD I + Whit Ch, Cleg Cn Eg + + - - Li - - Li - BFF F1XA FF I + BFT (XA FF I - C1 HV + + Mo I I I - - - Li - Li <td>BF.F 1:X A</td> <td>Q'</td> <td>1</td> <td></td> <td>+</td> <td></td>	BF.F 1:X A	Q'	1		+																				
BFF IXA EE 1 + Horizon Li + Horizon	BF.F 1:X A	DD	1		+	Wht Cn, Cl Eg	Cn	Eg	+		+				+								Li		
BFF IXA FF 1 C1 HV + + - + - Li BFF IXA GG 1 + YL LiEg HV + + - HV + - - Mo - Li Tex w/Rgs BFF IXA HH 1 - + C1 HV + + - Mo - Li Tex w/Rgs BFF IXA HH 1 - + C1 HV + + - Mo - Li Tex m/Rgs BFF IXA UUU 1 + YL, BwnOg, CHEg LiC - + - - - Li - - Li - - Li - - Li - - -	BF.F 1:X A	EE	1			+ Bri Yl Cn, Cl Eg		Li	+		+				Mo	Li						Li			
BF.F I:XA GG I I YI, Li Eg HV I I I I I C w Rgs Mo I Li Tex w Rgs Mo I Fex w Rgs BF.F I:XA SS I I + C1 HV + + Mo I I Tex w Rgs fmc ray and the ray of the ra	BF.F 1:X A	FF	1			Cl		Hv	+		+				+					+				Li	
BF.F. IX.A HH I + + Cl HV + + Mo - - - + fine Tex in Cn BF.F. IX.A UUU I + YI, BwnOg, CIEg Li Cn + - Li - Li - Li - Li rater Like, nutliced edges BF.F. IX.A VUV I + + CLE, g., Bg/Bvn Cn + + Mo Li + + Li - Li - Li rater Like, nutliced edges BF.F. IX.A VVV I I + CLE, g., Bg/Bvn Cn + + Mo Li - + + Li - Li - Li Taree Like, nutliced edges BF.F. IX.A VVV I I 00, C.O. CTex Eg + + Mo - + + - Li - HV	BF.F 1:X A	GG	1			+ Yl, Li Eg		Hv	+		+				+						Mo			Li	Tex w/ Rgs
BFF I:XA UUU VN to Cn, Cleg Cn Eg + Cn Lie + Li Li caracterize Li Li Li caracterize Li Li Li Caracterize Li Li Caracterize Li Li Caracterize Li Li Caracterize Li Li Li Caracterize Li Li <thli< th=""> Li <thli< th=""></thli<></thli<>	BF.F 1:X A	HH	1			+ Cl		Hv	+		+				Mo									+	fine Tex in Cn
BFF I:XA UUU I I YL BwnOg, ClEg LiCn I	BF.F 1:X A	SS	1			Wht Cn, Cl Eg	Cn	Eg	+		Cn	Li Eg				+								Li	crater Like, ribbed edges
BFF 1:XA VVV 4 + CL Eg, Bg/Bwn Cn + + Mo Li + + Li Fibros, outlined Eg BFF 1:XA WWW 1 0 Og Cn, Cl Tex Eg + + Cn Eg Eg - + + 0 Li Fibros, outlined Eg BFF 1:XA XXX 1 + Tk Mo + + Mo - - + + 0 Li Fibros, outlined Eg BFF.1:XA XXX 1 + Tk Mo + + Mo - - Li + Hv	BF.F 1:X A	UUU	1			+ Yl, Bwn/Og, Cl Eg		Li Cn	+		+					Li							Li		
BF.F. 1:X.A WWW 1 × Og Cn, Cl Tex Eg + + Cn Eg Eg × +	BF.F 1:X A	VVV	4			+ CL Eg, Bg/Bwn Cn		+	+		Mo	Li	+					+	+			Li			Fibrous, outLined Eg
BFF 1:XA XXX 1 + Tk Mo + Mo - Li - No - Hv Hv bump, Lightly inrEg BFF 1:XA 20 1 + CL + + + + + - + + - - Hv Hv bump, Lightly inrEg BFF 1:XA 400 - + + + + + + + + + + - - Hv Hv bump, Lightly inrEg BFF 1:XA 400 - + <t< td=""><td>BF.F 1:X A</td><td>WWW</td><td>1</td><td></td><td></td><td>Og Cn, Cl Tex Eg</td><td></td><td>+</td><td>+</td><td></td><td>Cn</td><td>Eg</td><td></td><td>Eg</td><td></td><td></td><td></td><td>+</td><td>+</td><td></td><td></td><td></td><td></td><td>Li</td><td>Tex Eg, SMooth Cn</td></t<>	BF.F 1:X A	WWW	1			Og Cn, Cl Tex Eg		+	+		Cn	Eg		Eg				+	+					Li	Tex Eg, SMooth Cn
BF.F.1X.A 20 1 + Cl + + + + + + - - - - - - Original BF.F.1X.A ZZZ 1 + Ph/Pk + + + + + + + - + - - + Original BF.F.1X.A ZZZ 1 + Hyhpkr Cn + + + + - + -	BF.F 1:X A	XXX	1	+		Tk	Mo		+		Mo					Li								Hv	Hv bump, Lightly irrEg
BFF 1:XA 40 1 + Ph/Pk +	BF.F 1:X A	20	1	+		Cl		+	+		+				+								+		Original
BFF 1:XA ZZZ I I H <th< td=""><td>BF.F 1:X A</td><td>40</td><td>1</td><td></td><td></td><td>+ Ph/Pk</td><td></td><td>+</td><td>+</td><td></td><td>+</td><td></td><td></td><td></td><td>+</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td>+</td><td></td><td>Original</td></th<>	BF.F 1:X A	40	1			+ Ph/Pk		+	+		+				+								+		Original
BEF.F1:XA57'1+Li Bwn Rg, Cl Fib Eg, Dk Bwn+++EgCn++Not CVX at EgBF.F1:XA632+Y1 Dkr Cn++++MoLiLiLiLi Li Lightly Crater LikeBF.F1:XA641+++++MoLiLiLiLi Li Lightly Crater LikeBF.F1:XAZZ'1+++++-+-+-BF.F1:XAZZ'1++BF.F1:XAWW1BF.F1:XABBF, 1:XABF.F1:XBAAABF.F1:XBBBB1+Pk/Ph, Dk Cn, Cl Eg+++-+BF.F1:XBDDD1+Li Li Q/YI Cn, Cl Eg+++-+-+ </td <td>BF.F 1:X A</td> <td>ZZZ</td> <td>1</td> <td></td> <td></td> <td>+ Wht, Dkr Cn</td> <td>+</td> <td></td> <td>+</td> <td></td> <td>+</td> <td></td> <td></td> <td></td> <td>Mo</td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td></td> <td>Li</td> <td></td> <td></td>	BF.F 1:X A	ZZZ	1			+ Wht, Dkr Cn	+		+		+				Mo								Li		
BFF 1:XA 63 2 + + Y1Dkr Cn + + + + Mo Li	BF.F 1:X A	57'	1			+ Li Bwn Rg, Cl Fib Eg, Dk Bwn	1	+	+		+	Eg			Cn	+			+				+		Not CVX at Eg
BFF 1:XA 64 1 + Ph Dkr Cn + + + + - + -	BF.F 1:X A	63	2			+ Yl Dkr Cn		+	+		+				Mo	Li				Li				Li	Lightly Crater Like
BFF 1:XA ZZ 1 + + -	BF.F 1:X A	64	1	+		Ph Dkr Cn		+	+		+				+								+		
BF.F 1:X A WW 1 Image: Clystem of the second	BF.F 1:X A	ZZ	1			+	1			1															
BF.F 1:X B AAA 1 Cl/Y1 + + + Li Mo Bio-film, takes up 1/2 of plate, dimpled Like a golf ball BF.F 1:X B VV 1 -	BF.F 1:X A	WW'	1							1	1						1		1	1	Ì		1		
BF.F 1:X B VV 1	BF.F 1:X B	AAA	1			CI/Y1	1	+	+	1		+				Li					Mo				Bio-film, takes up 1/2 of plate, dimpled Like a golf ball
BFF 1:X B BBB 1 + Pk/Ph, Dk Cn, Cl Eg + + + Li Li Li BF.F 1:X B CCC 1 Wht/B Cn, Cl Eg + + Cn Eg + Li Li Li Li BF.F 1:X B DDD 1 + Li Og/Y1 Cn, Cl Eg + + Cn Eg + + Sm Cn, Lg Eg BF.F 1:X B DDD 1 + Li Og/Y1 Cn, Cl Eg + + + Ii Li Li Li Lg Cn, Sm Eg BF.F 1:X B DDD 1 + LiOg/Y1 Cn, Cl Eg + + + Iii Li Lg Cn, Sm Eg BF.F 1:X B DDD 1 + HPL & Assey + + H	BF.F 1:X B	VV'	1				1			1											Ĺ				
BF.F 1:X B DCC 1 1 1 1 1 1 1 1 1 BF.F 1:X B DDD 1 + Li Og/Y1 Cn, C1Eg + + + + + Sm Cn, Lg Eg BF.F 1:X B DDD 1 + Li Og/Y1 Cn, C1Eg + + + + Li Li Lg Cn, Sm Eg BF.F 1:X B DDD' 1 + + + + + + + BF.F 1:X B DDD' 1 + + + + + + +	BF.F 1:X B	BBB	1		Ť	+ Pk/Ph. Dk Cn. Cl Eg	1	+	+	1	+					+	1		1	Li	1		1	Li	
BF.F 1:X B DDD 1 + + + + BF.F 1:X B DDD 1 + H H H BF.F 1:X B FFE 1 + H H H	BF.F 1:X B	CCC	1		Ť	Wht/Bg Cn. Cl Eg	1	+	+	1	Cn	Eg				+	1		1	1	+		1		Sm Cn. Lø Eø
BF.F I:X B FFE I + PIPk Rosev + + + + + + + + + + + + + + + + + + +	BFF1·XB	DDD	1 î	\vdash	-	+ Li Og/YI Cn Cl Eg	1	+	+	1	+					<u> </u>		+						Li	L o Cn Sm Eo
BFF1XB FFF 1 + PIPk Rosev + + + + +	BEELVE	DDD'	1	\vdash	+	+	1		-	+					-		<u> </u>		<u> </u>	<u> </u>	<u> </u>		<u> </u>		25 0m, 0m 25
	BFF1·XB	EEE	1 î	\vdash	-	+ Pl Pk Rosev	1	+	+	1	+						+					+			

Sample	ID	Cnt	Size i	n m	m Color	Consi	istency		Colon	у Арре	arance	;		Edge						Elev	/ation			Notes
-		#	<1 1	-22	2-7	OPQ	TRL	SHY	DUL	CRC	IRR	FIL	RHZ	ENT	UND	LOB	ERO	FIL	CRL	FLT	RAI	CVX	UMB	
BEE1 X B	FFF	1			Wht	Cn	Eσ	+			+				+	+							+	
BF F 1 X B	GGG	1		+	Pastel YI Cn Cl Wht Eg	Cn	Eσ	+		+				+							+		Li	
BEE1:XB	ннн	1		<u> </u>	+ PLLiOg/XLCn_CLEg	C.	 	+		Cn	Fσ					+					-			
BF F 1 X B	III	1			$+ \Omega \sigma w/Dk Pts$		+	+		+	25				+				+				+	Split open causing scattered Pts. Sim to DD
BEE1:XB	III	1	+	-	Bri VI		+	+		+	Li			+	· ·	Li						Hv		Spin open causing seattered 1 (s, bin to DD
BEELVE	KKK	1	-	-	+ VICn Cl Wht Eq		-	+		-	Li			Mo	T i	1.1						11,	-	Uncooked Eag look round Eg
DEE LV D	LLI	1		-	+ ITCH, CI Whit Eg	-	+	+		+				WIO	LI								+	Er More OPO Eurgel like
DEE LV D	NOOI	1			V1/On w/ Dla Dta		т		Ŧ	т		Ŧ						Ŧ		Ŧ				Eg More Of Q, I uligal-like
DEE LV D	NNN	1		+	Vory Li PLOg Eg. Dkr Cn	-	+	+		Cn	+ Ea				+							÷		Egg Look
DEE LV D	INININ 0000	1		-	Very Li, Frog Eg, DKi Cli	- · ·	÷	+		- Cli	Eg						Ŧ			÷				Egg Look
DF.F I.A D	DDD	1		_	+ win	+		+		+				+				т.:				+		Sim to 77
DEELVD	PPP	1		_	+ CLOg		+	+		+								LI		+			¥ ·	
BF.F I:X B	QQQ	1		_	+ PILIYIPh	+		+		+					+	+							Li	Tex Eg
BF.F I:X B	RRR	1		_	+ YI/Og, Dk Cn and Eg, Li Rg	_	+	+		+				+									+	Lighter Rg
BF.F I:X B	555	1		+	YI/Og, Dkr Eg	~	+	+		+							+		+		+			Tex Dkr Eg w/ scattered Pts
BF.F 1:X B	TIT	1		_	+ Dk Yl Cn, Li Eg	Cn	+	+		+					+								Lı	Cl w/ Flecks of Wht, Dkr Eg
BF.F 1:X C	RR	1		_	Cl w/ Wht Pts		+		+	+		+						+		+				Nearly whole plate covered, Wht FIL growth, Fungal-like
BF.F 1:X C	V2'	1			+																			
BF.F 1:X C	TT	1			+ Wht, Bwn/Yl Cn	+			+		+				+							+		Scattered about plate, raised but w/o uniformity, Fungal-like
BF.F 1:X C	TT'	1			+																			
BF.F 1:X C	UU	1			+ Yl Cn, Cl Eg		+	Cn	Eg	+					Hv			+					+	
BF.F 1:X C	VV	1			+ Blk/Gn Cn, Cl Eg	Cn	Eg		+	Li		+						+		Mo			Li	Fungal-like
BF.F 1:X C	WW	1			Wht	Mo	Li Eg	+			+				+					+				
BF.F 1:X C	XX	1		+	Yl Cn, Li Wht Eg		+	+		+				Cn	Li					+				
BF.F 1:X C	DD'	1		+																				
BF.F 1:X C	YY	1			Cl, Pl Yl Cn, Dk Eg		+	+			+						+			+				Raised Eg
BF.F 1:X C	ZZ	1			+ Cl, Dk Eg		+	+		+							+			+				Finely UND
BF.F 1:X C	ZZ	2			+																			
BF.F 1:X C	Y'	1			+																			
BF.F 1:X C	CCC'	1																						
BF.F 1:X C	7	1			+ Og		+	+		+														Originals Clustered
BF.F 1:X C	YYY	1			+ C1		+	+			+				+						Li			Amaboid shaped w/ wrinkled Tex
BF.F 1:X C	46	1			+ Wht. Mo Og/Y1 Cn	Li	+	+		+				+								+		
BF.F 1:X C	61	1			Y1 Bg Cn. Cl Bg rest. Dk Bg		+	+		Mo	Eg			CRC	Mo					+				Tex in non CRC area
BF.F 1:X C	7*'	1			+																			
BF.W 1:100A	1	1			Y1/Og. Dkr Cn&Rg. Cl Eg		+	+	Eg	Cn Li	Mo				+				LI	+				Hy dimples/bumps all over More in Cn
BF.W 1:100A	2	1			Cl/Bg		+	+		+				+								Li		May be Sim to Fish
BF W 1.100B	SS'	1																						
BF W 1:100B	DD'	1		+																				
BF W 1:100B	II'	1	+																					
BF W 1:100B	3	1			Pl Ph Past		+	+		+				+					+		+			Dif from BBB
BF W 1:100B	4	1		4	Ta	+	Ŧ	Ť		т	+			Ŧ	-			-	Ŧ		т	-	-	Vy lumpy IPP
DF.W 1:100D	5	1		T .	Pd/Og	- T		T i			- T				т Ц.,			-			-	- T	- T	vy lumpy iKK
BF.W 1.100B	5	1		+	Og Sm Eg	+	Sm E				+				nv	Ŧ						+	+	Marka Sim to Fish
DF.W 1.100C	WW	1		-	Og, Shi Eg	+	SIILE	+		+				÷								÷		Maybe Shin to Fish
DF.W 1:10A	WWW	1	\vdash	_	0-04																			SLinkly Direction AAA and Finan Jamela (human
BF.W 1:10A	AAA [*]	1		_	Og/YI	-	**															¥ ·	¥ ·	SLignly Dkr than AAA, and Finer dimples/bumps
DF.W 1:10A	7	1	\vdash	_	Ug, LI UI Eg	+	HV	+		+				+								LI	LI	very connvion
BF.W 1:10A	7	4		_	+ . L: VI.C. CI.F.	+	<u> </u>	<u> </u>		<u> </u>							.	<u> </u>	.		<u> </u>	<u> </u>	· · ·	
BF.W 1:10A	8	1		_	+ LI YI Ch, CI Eg	+	+	+		+							Lı	<u> </u>	Li		<u> </u>	<u> </u>	Li	
BF.W 1:10A	8.	2		_	+	-	I		I							I			I	I				
BF.W 1:10A	9	1		+	Ph Cn, Wht Eg, Dk Bwn Rg	+	I		+	+					Lı	I			I	I		+		
BF.W 1:10A	10	1		_	+ YI Cn, Cl/Wht Eg	-	+	+	L	+					+	L		<u> </u>		L	<u> </u>	<u> </u>	Li	
BF.W 1:10A	11	1			+ Yl Cn, Cl Eg	Li Cn	+	+		+				+									+	Relatively Sm Eg
BF.W 1:10A	10'	1			+																			
BF.W 1:10A	12	1			+ Yl Cn, Cl Eg		+	+		+					Li								Li	Cn Tex

Sample	ID	Cnt	Size in	mn	n Color	Consi	istency		Colon	у Арре	arance	;		Edge						Elev	ration			Notes
-		#	<1 1-	22-	-7	OPQ	TRL	SHY	DUL	CRC	IRR	FIL	RHZ	ENT	UND	LOB	ERO	FIL	CRL	FLT	RAI	CVX	UMB	
BF W 1.10A	13	1		+	+ Wht Cn Mo Cl Eg Wht outer		+	+		Cn					Hv								+	Wht Cn Mo Cl Eg w/ Wht outer fRge
BF W 1.10A	14	1		+	+ Li Og/Y1 C1 Eg		+	+		+				+					+		+			More entire color not Egg-Like
BEW 1:104	15	1		T.	Li VI Cn Cl Eg			+		Cn	Cn			<u> </u>	+					+				I g Eg Sim to NNN
BEW 1:10A	16	1		1	Br VI Cn Cl/Wht Eg		+	+		Cn	Cn						Hv						Li	PR
BEW 1:10A	17	1		1	CI/VI Eg Li Og Cp		-	-		Ch	+						+			-			1.1	Dimpled Like golfball_sMooth Cn
BEW 1:10A	20'	1	-		Mo Cl Li Og/Wht Cn		-	-		4				-								-		Might be Sim to HH
BF W 1:10A	20'	1	- T	-	Mo Ci, Li Og/ Witt Ch		т	Ŧ		т				Ŧ								Ŧ		Might be Shift of HIT
DE W1.10P	20 A A A *'	1	т	-									-					-				-		
DF.W1.10D	DDD'	1		+	+																			Dividing
DF.W1.10D	DDD	1		+	+																			Dividing
DF.W1:10D	7'	1		-														-						
DF.W1:10D	7"	1		-														-						
BF.W1:10B	7	1		-																				
BF.W1:10B	-7"	1		+	-																			
BF.W1:10B	DD'	1		30	<u>m</u>																			Eg Li UND due to overlap
BF.W1:10B	FF'	1			4																			
BF.W1:10B	17'	1			4																			
BF.W1:10B	14'	1		+																				
BF.W1:10B	18	1			Cl Lg Eg, Li Og Cn		+	+		Mo					Li					+				Sim to 17 but not dimpled
BF.W1:10B	19	1		+	- Og/Ph		+	+		+					+						+			Uniflorm UND, Crater-Like
BF.W1:10B	CCC'	1																						Dk flecks within, due to overlap, probably a Dif colony
BF.W1:10B	21	1			Cl Lg Eg, Grad to Yl Cn		+	+		Cn	+			Mo	Li					+				PR
BF.W1:10B	22	1		+	Pl Yl, Sm Clearer Eg		+	+		+				+					+				+	PR
BF.W1:10B	20'	3	+		1																			
BF.W1:10B	HH'	1	+																					
BF.W1:10B	39'	1		+	+																			
BF.W1:10B	23	1		+	+ Y1 C1 Eg		+	+		+				+					+				+	
BF.W 1:10C	AAA'	1																						
BF.W 1:10C	CCC'	1		+	÷																			
BF W 1.10C	7'	1		T.																				
BEW 1:10C	7"	2		1																				
BEW 1:10C	7'''	2		4	+																			
BF W 1:10C	w'	2	+ +		+																			
BEW 1:10C	22'	1																						
BF W 1:10C	24	1			Cl w/ yery Sm Bg Cn		-	-			T i		-	Mo	T i			-		+		-		Mo straight ENT Eq. w/ Light breaks. Mo IPP but round shape
DF.W 1.10C	14'	1		۰.	La Vary Li Og/VI Cn. Cl Ea		- T	-			Li		-	IVIO	LI			-	т;	т		T i		Mo straight ENT Eg, w/ Eight breaks, wo tick but found shape
BF.W 1:10C	14	1		+	DI WH/O-		+	+		+				+				-	LI			LI		
DF.W 1.10C	20	1		+			+	+		+				+					- L1		Ŧ		т.:	
BF.W 1:10C	29	1		+	· LI Ug Ch, CI Eg		+	+		+				+				-	+				LI	
BF.W 1:10C	24	1		_	01 F 11/0 F					G	F											G		
BF.W 1:10C	25	1		-	CIEg, YI/Og Eg		+	+		Cn	Eg				+							Cn		
BF.W 1:10C	26	1		+	Sherbert Og	+		+		+				+								+		
BF.W 1:10C	92	1	+		PI YI, Dkr Cn		+	+		+				+								Lı		
BF.W 1:10C	1*	1	+		Cl Eg, Yl Cn		+	+		+					+								+	Tex Eg
BF.W 1:X A	62	1		+	· Cl		+	+		+				Mo	Li				Cn				+	Tex Eg & Li Cn Sand paper
BF.W 1:X A	NNN'	1		+	-																			
BF.W 1:X A	7'	10		+	-																			
BF.W 1:X A	RR'	1																						
BF.W 1:X A	WWW'	3		+																				
BF.W 1:X A	ZZ'	1		+	-																			
BF.W 1:X A	AAA*'	2																						
BF.W 1:X A	FF*	1		+	- Cl		+	+		+					Li						+			Raised and dips in sLightly
BF.W 1:X A	FF'	1		+	r																			
BF.W 1:X A	30	1		+	Pr, Dkr Eg & Cn		+	+	+	+							+		+				Li	Cn and outer Eg Dkr
BF.W 1:X A	X'	2	+		1	1	1	1	1				1	1		1			1	1		1		
BF.W 1:X A	W'	1		+	- Lg Og Cn, Cl/Wht Eg	1	+	+	1	+			1	1	Li	1			+	1	+	1		Sim to W

Sample	ID	Cnt	Size	in n	nm	Color	Consi	istency		Color	y Appe	earance	e		Edge						Elev	/ation			Notes
,		#	<1	1-2	2-7		OPQ	TRL	SHY	DUL	CRC	IRR	FIL	RHZ	ENT	UND	LOB	ERO	FIL	CRL	FLT	RAI	CVX	UMB	
BEW 1.YA	32	1		T	+	Pl Og		4	-	T		Тi		1		-				1	T	+	T		Crater shaped
BEW 1.X A	7'	16	-1-	+	Ŧ	ITOg		т	т		-	Li	-		-	т	-		-			т			Crater-snaped
DEW LV A	20'	10	+	-							-		-		-	-	-		-						
DEW LV A	20 DDD'	15	+	+						1											-		-		
DEW LV A	DDD'	1			+					1											-		-		
DF.W IXA	DD	4		+																					
BF.W IXA	HH	1		_	+	W1 (/P)	L'C	Г		-						¥7 ¥ ·					-		-		DD.
BF.W I:X A	35	1		_	+	wnt/Bg	LICN	Eg	+	-	+					VYLI					-		-	+	PR E
BF.W I:X A	34	1		_	+	Ph Cn, Cl Eg		+	+		+				Mo	Lı								Li	Tex Eg
BF.W I:X A	BBB.	2		_	+	4																			-
BF.W 1:X A	35	1			+	Cl		+	+		+				+								+		Tex
BF.W 1:X A	36	1			+	Cl Eg, Yl/Og/Bg Cn		+	+		+							+			+				Sim to 14 but Dif, Lg Cl Eg & Tex Eg
BF.W 1:X A	37	1			+	Cl Eg, Cl/Wht Cn		+	+		+					+								Hv	Highly Tex, nipple shaped
BF.W 1:X A	XXX	1	+																						
BF.W 1:X A	EE'	1	+			Yl		+	+		+				+								+		
BF.W 1:X A	39	1		+		Cl		+	+		+				+								+		Crater-shaped, patterned Lines from Cn
BF.W 1:X A	40'	1			+	Bri Ph/Pk		+	+		+				+								+		
BF.W 1:X B	VV'	1		+																					
BF.W 1:X B	37'	1			+																				
BF.W 1:X B	7'	14		+	+																				
BF.W 1:X B	ZZ'	1			+																				
BF.W 1:X B	20'	11	+	+																					
BF.W 1:X B	40'	1			+																				
BF.W 1:X B	CCC'	1			+																				SLightly Dif due to background film
BF.W 1:X B	41	1				Mo Cl. Li spots of Wht/Yl		+	+			+				+	+				+				Bio film, non uniform wayes, splotches of Li color
BF.W 1:X B	22'	1			+																				
BF.W 1:X B	41*	1			+	Cl Cn & Eg Wht Rg		+	+		Mo							+			+				
BF.W 1:X B	42	1		+	÷	Cl Eg. Yl Cn & Dkr Rg		+	+		+				+									Hv	Cn nipple
BF W 1·X B	FF*'	1		- I	+																				
BF W 1·X B	YYY	2		-	+																				
BEW 1.X B	43	1		-	+	Cl		+	+		+				Mo					+	+				Tex and Sim to others but Dif
BEW 1:Y B	30'	1		-		61									1010										Tex and Shiri to balers but Dir
BEW 1:X B	DD'	1		+	-																				
BEW 1:Y B	44	1		+	-	Cl Eg. Cl/Burn Cn. Dk Pts			-		-1-				-1-								-		Colored Ptsicles inside
BFW 1.X B	44	1		+		CI LIG, CI/DWITCH, DKTIS		- - τ	- T		- T		-		- T	-	-		-	-			+		bas scattered hits arround
DEW LV D	45	1		Ŧ		ei		т	т	-	т		-		т	-	-		-	Ŧ			Ŧ		has scattered bits arround
DEW LV D	40 M	1		-	+					1											-		-		
DF.W LAD	IVI XV/	1		-	_					1											-		-		
DEW LV D	40		+	-				 .	<u> </u> .		<u>+ .</u>				<u>+ .</u>								T :	т:	Sim to 7 but m/ Dum Dt inside
DF.W IX B	48	4	+	_	+	CI Eg, Og Ch, Bwh Pts in Ch		+	+	+	+	<u> </u>		<u> </u>	+	<u> </u>	<u> </u>					<u> </u>	LI	LI	SIII to / but w/ bwn Pts inside
DF.W IX B	/*	1	+	_		Og/ 11		+	+	+		+		<u> </u>		+	+					+			
BF.W I:X C	M	1		_																					LOB w/ gaps throught dish
BF.W I:X C	W'	1	+	_			<u> </u>	<u> </u>		<u> </u>	<u> </u>		<u> </u>	<u> </u>	<u> </u>	<u> </u>	<u> </u>		<u> </u>			<u> </u>			
BF.W 1:X C	CCC'	1	\square		+			I	I	<u> </u>				I						I	 		 		
BF.W 1:X C	7	21	\square	+	+			I	I	<u> </u>				I						I	 		 		
BF.W 1:X C	20'	17	+	+			Ļ			<u> </u>	<u> </u>		Ļ		<u> </u>	<u> </u>	<u> </u>		Ļ		<u> </u>	<u> </u>	<u> </u>		
BF.W 1:X C	XX'	1	+					I	L	I				I						L	L		L		
BF.W 1:X C	00'	1	+					I	L	I				I						L	L		L		
BF.W 1:X C	47	1			+	Cl		+	+		Mo							+			L		L	Li	Tex in Cn sMooth on outside
BF.W 1:X C	37'	1			+																L		L		
BF.W 1:X C	TT'	2		+																					very Sim, less developed
BF.W 1:X C	92	1	LT		+																				
BF.W 1:X C	48'	1	LT		+																				
BF.W 1:X C	49	1	+			Ph/Bwn Cn, Dk Pts	Cn Pts	+	+		+				+								+		Original on 1:a F
BF.W 1:X C	ZZ'	2			+																				
BF.W 1:X C	AAA*	3																							

Sample	ID	Cnt	Size i	in mr	n Color	Const	istency	r	Colon	y Appe	earance			Edge						Elev	/ation			Notes
<u>^</u>		#	<1 1	-2 2	7	OPQ	TRL	SHY	DUL	CRC	IRR	FIL	RHZ	ENT	UND	LOB	ERO	FIL	CRL	FLT	RAI	CVX	UMB	
BEW 1.X C	41*'	1		-	-																			
BEW 1:X C	FF'	1		+		1																		
BEW IXC	GGG	1		+		-																		
BFW 1XC	000	1	+	Ŧ		-	-	-	-					-				-	-	-	-	-		
DEW LX C	49'	1	Ŧ	-		-	-	-	-					-				-	-	-	-	-		
SCE 1.10A	40	1																						Element ite actions however
SC.F 1:10A	A3	1			- BII II	+		+		+					+							+		Flower-Like pattern, lumpy
SC.F 1:10A	A5 D2	1		+	WILLE G. C.			T ·																
SC.F 1:10A	B3	1		+	whiteg, Gry Ch	+	¥ *	Li		+		**			+						+	+	¥ ·	
SC.F 1:10B	0.5	1		_	PI Bg/wht	-	Lì	+		+		HV						+	¥ ·				Li	Tightiy FiL outside Linear ridges, Fungai-like
SC.F 1:10B	D3	1		_		_	+	+		+					+				Lı	Mo	6			
SC.F 1:10B	E3	1		_	Lg Bwn Og Cn, Cl Eg	_	+	+		+					+			+			Cn			Cloudy blobs w/ Cl Eg, RAI in Rgs around CRL
SC.F 1:10B	F3	1			Mlk Bg, Cl Eg	_	+	+			+				+	+					Lı			
SC.F 1:10B	F3'	1																						Rgs of CRL
SC.F 1:10B	G3	1			Cl Rg, Bwn/Og Cn, Mlk/Cl Eg		+	+		Li					+	+		+	+				Li	FIL Lightly branching from Cn to Eg
SC.F 1:10B	H3	1			Li Bg arms, Cl otherwise		+	+		Mo			+		+					Mo				Dk Pts within
SC.F 1:10C	13	1		+	 Og/Bwn/Ph, Dk Cn, Li Rg 		+	+		+					+	+						Li		Bri bump Og film
SC.F 1:10C	J3	1			Og		+	+			+				+					+				
SC.F 1:10C	K3	1		+	Pl Dull Yl		Li	+		+				Mo								Li		
SC.F 1:10C	I3'	1		4	-																			
SC.F 1:X A	L3	1		4	Cl, Cloudy		+	+		Li					+	Li					+			Lumpy RAI and Indentions
SC.F 1:X A	M3	1		÷	 Cloudy Eg, Ph Cn 	Cn	+	+		Cn	Eg				+	+						Cn		Sm Cn
SC.F 1:X A	N3	1		Ŧ	- Cl		+	+		+					+	+						+		RAI Cn, CRL RAI, Pts at Eg
SC.F 1:X A	J3'	1																						
SC.F 1:X A	F3'	7		-	-																			
SC.F 1:X A	42A'	1																						
SC.F 1:X A	53	1		-	- Cl		+	+		Mo				Mo			Li			+				Sim to othe ones like ZZ
SC.F 1:X A	ZZ	4		-																				
SC F 1:X B	03	1		-	- C1		+	+		+					+						+			More CRC than Sim ones, Crater-shaped
SC F 1·X B	03'	1		4																				
SC F 1:X B	P3	1			Cl Sm Li Dkr Cn		+	+		+							+						+	Sim to others Rough Tex
SC F 1:X B	03	1		+	Bwn Cn Cl Eg	Cn	Fσ	+						Mo								+		onn to oulois, Hough Tex
SC F 1:X B	13*	1		-	Cl	Ch	105	+			+			+						+				very Sim to others ?
SC F 1:X B	P3	1			Cl Eg. Mlk Wht/Bg Cn	1	-	-		-					-								T i	Fan from Cn
SC F I Y B	T3	1			CI Lg, with with bg cir	-	- T	- T	-	- T				-	- T			-	-	-	-	-	Li	Li Tay in Cn
SC.F LV P	96*	1			Cl/Bg_Milt Wht Clouds	-	- T	- T	-	- Cn	Ea			Cn	- T			-	-	Ea	-	Cn		Cloudy not in Page Elt Mile Eilm & Eg IDP
SC.F.I.X.B	111	1		-	CI/Bg, Mik will Clouds	-	+	÷		Cli	Eg			Cli	+	÷				Eg		Cli		Cloudy not in Kgs, Fit Wik Film & Eg IKK
SC.F LX B	42 A	1			-	-																		
SC.F I:A B	42A	1		-		-																		
SC.F I:X B	F5	2			-	-																		
SC.F I:X B		1	\vdash		MILWILLE C	6	Г	<u> </u>		<u> </u>			<u> </u>		<u> </u>						. .			
SC.F I:X C	83	1		-	Mlk Wht/Bg Cn	Cn	Eg	+		+					+						Li			Fan from Cn
SC.F I:X C	P3'	2		_		_																		
SC.F 1:X C	\$3	1		+	-	_																		
SC.F 1:X C	13	2	\vdash			_				I		I	I		I	I								
SC.F 1:X C	J3'	1	\square		Og	_	+	+	<u> </u>	L	+			Ļ	+	+		<u> </u>	Ļ	<u> </u>	+	<u> </u>		Lgr Tex and More bubbly
SC.F 1:X C	27A'	1		+		_				L		L	I		L	L								
SC.F 1:X C	FF*'	2		+		_				L		L	I		L	L								
SC.F 1:X C	31A'	1																						
SC.F 1:X C	15B'	1		н	-																			
SC.F 1:X C	F3'	3		4	-																			
SC.F 1:X C	ZZ	1		4	-																			
SC.W 1:100A	25A	1			Tng, Dk Eg	Eg	+	+		+					+	+					Eg			Raised Eg, dipped Cn, pattern from Cn, thin lobed Eg
SC.W 1:100A	KK2'	1		4	-																			
SC.W 1:100A	26A	1		+	Bg Eg, Bwn Cn	+		+		+					+							+		
SC.W 1:100B	27A	1		+			+			Li					+	+					LiEg			

Sample	ID	Cnt	Size in	mm	1 Color	Consi	istency		Colon	y Appe	arance			Edge						Elev	ation			Notes
-		#	<1 1-	2 2-'	7	OPQ	TRL	SHY	DUL	CRC	IRR	FIL	RHZ	ENT	UND	LOB	ERO	FIL	CRL	FLT	RAI	CVX	UMB	
SC W 1.100B	٥٨'	1		T	V1/Og	Ì	1	1		1		1	1					I		I		1		
SC W 1:100B	28 1	1		+	11/0g		-	-	-	-				-								Тį		PP too crowded to tell
SC W 1:100D	204	1			Mile Whet Eq. Dg/Dh Cn	Cn	Fali	-	-	- T				т	T ;							Li		r K, too crowded to ten
SC.W 1:100C	29A	1		+	Mik will Eg, Bg/Pfi Cli	Cn	EgLi	+		+					LI						T :	LI		Sim to 20 A but Dif
SC.W 1:100C	50A	1		+	Bg, Dkr Cli	Cn	EgLi	+		+					+						LI			Sill to 29A but Dil
SC.W 1:100C	15	1		-																				
SC.W 1:100C	31A	1		_	Cl/Li Wht		+	+			+				+	+				+				PR, spaced grainy Tex
SC.W 1:100C	20'	1	+	_																				PR, possibly ph/og, possible infulence by J3
SC.W 1:10A	32A	1		+	Pk/Ph Cn, Cl Eg		+	+		+					+					Eg		+		
SC.W 1:10A	33A	1			Pl Yl, Li Eg		+	+		Li					+	+			+	+				
SC.W 1:10A	13'	1																						
SC.W 1:10A	30A'	2	+	+																				
SC.W 1:10A	25'	1																						
SC.W 1:10A	J3'	1																						
SC.W 1:10A	FF*'	1		+																				
SC.W 1:10A	J3"	1																						
SC.W 1:10A	J3""	1			1 1																			
SC W 1.10A	34A	1	+	1	Βσ/ΥΙ Οσ		Li	+		+					+							+		PR
SC W 1:10A	0'	1		+	bymog																			
SC W 1:10R	35.4	1		-	PLV1 Dkr Eg		T i			-				-					+		EaCn		T i	Cadia
SC W 1:10B	261	1		+	PI VI/Og		Li			- T				- T					Ŧ		LgCI	T i	Li	Simila to 25 A
SC.W 1.10B	27 A	1		+	CLI MILEE DIS DE DE LICE		+			÷				÷							D.e.	ы		Deired De die in Ce
SC.W 1:10B	3/A	2			CI LI MIK Eg, DKF Bg Kg, LI CI		+	+			+				+	+					ĸg			Kaiseu Kg, uip in Ch
SC.W 1:10B	38A	2		+	Bg/Og		+	+		+					+						+			
SC.W 1:10B	39A	1		+	Mik Ci Eg, Og Cn		+	+		+					+				+		CRL			
SC.W 1:10B	40A	1			Cl, Li Mlk Cn		+	+		Li							+			+				Sim to ZZ, but sMooth Cn, Tex Eg
SC.W 1:10B	31A'	1		+																				PR
SC.W 1:10B	NN3'	1		+																				
SC.W 1:10B	J3'	2																						
SC.W 1:10B	63'	1		+																				
SC.W 1:10B	V'	1		+																				
SC.W 1:10B	7'	1		+	-																			
SC.W 1:10C	41A	1			Cl,		+	+		+					Hv						Li Eg			Crater-Like, Lg hole in Cn, Li Tex Eg, Eg ridge frilled
SC.W 1:10C	42A	2		+	Cl w/ Gray Pts	Pts	+	+		+					+							Hv		Volcano-shaped
SC.W 1:10C	43A	1		+	Bri Yl		+	+		+					Hv						Li			Raised Eg. dipped Rg. Li raised Cn
SC.W 1:10C	44A	2		+	- Cl Og		+	+						+								+		a, H. a,
SC W 1.10C	45A	1			Cl		+	+		Cn	Fσ				+						Cn			Crater-Like Cn. Smooth flat Eg
SC W 1:10C	464	1		+			+				15				Ţ.				+		Ch		+	Sim to FF
SC.W 1:10C	FE2'	1		Ŧ	Ci		Ŧ		-	Ŧ					LI				т				Ŧ	500 1011
SC.W 1.10C	121	4	+	-	4																			
SC.W 1:10C	12	4	<u> </u>	-																				
SC.W 1:10C	0	1	+	-																				
SC.W 1:10C	30A	3		+	D (0																			T 1 0 01 - 004
SC.W 1:10C	4/A	2		+	Bg/Og		+	+		+					+					+				Tex in Cn, Sim to 38A
SC.W 1:10C	21A'	1	+																					
SC.W 1:10C	YYY'	1		+	·																			
SC.W 1:10C	48A	3		+	Dull Wl/Bg		Li	+		+				Mo								+		PR
SC.W 1:X A	J3'	1																						
SC.W 1:X A	32A'	1		+																				
SC.W 1:X A	31A'	1		+																				
SC.W 1:X A	25'	1		+																				
SC.W 1:X A	PP2'	1		+																				
SC.W 1:X A	35'	4	+	+					1									1		1				
SC.W 1:X A	44B'	1		+	1				1									1		1				
SC W I X A	HH3'	1		1.	1 1		<u> </u>	<u> </u>	1	<u> </u>		<u> </u>	<u> </u>		<u> </u>			1		1		<u> </u>		
SC W LX A	40'	10	<u>н</u> н	+ -					1			-										-		
SC W 1:X A	W'	1		+	1		<u> </u>	<u> </u>	1	<u> </u>		1	<u> </u>	-		-		<u> </u>		<u> </u>		<u> </u>		

Sample	ID	Cnt	Size	n mr	n Color	Cons	istency	r	Colon	y Appe	earance			Edge						Elev	/ation			Notes
		#	<1 1	-2 2-	7	OPO	TRL	SHY	DUL	CRC	IRR	FIL	RHZ	ENT	UND	LOB	ERO	FIL	CRL	FLT	RAI	CVX	UMB	
COW LV A	40.4	1		Ŧ.	Mile Whet Dies Ca		T :		1			1		1				1		Ea		Ca		I - C- S- E-
SC.W I:A A	49A	1		4	Mik whi, Dkr Ch	_	LI	+		+				X	+	+		-		Eg		Cn		Lg Cli, Sili Eg
SC.W I:X A	003	1		-			+	+	MO					MO								+		PR
SC.W 1:X A	51A	3		+	- Cl		+	+			+				+					Mo				PR
SC.W 1:X A	FF*'	1		-																				
SC.W 1:X A	UU3'	7	+	+	Li Yl		+	+		+				+								+		Sim to X, PR but Lighter
SC.W 1:X B	LLL'	1																						
SC.W 1:X B	35'	4		+																				
SC.W 1:X B	1*'	1		+																				
SC.W 1:X B	YYY'	1		+																				
SC.W 1:X B	HH3'	1																						
SC.W 1:X B	EEE'	3		-				1																
SC W 1·X B	13'	1																				1		
SC W 1:X B	32 4'	5																						
SC W 1:X B	40'	7			-																			
SC.WI.XD	22 41	1		T 7	-	_			-									-		-		-		
SC.W I:A B	21A	1		-	-	_																		
SC.W I:A B	JIA	1		_		_	-	-	-									-						
SC.W I:X B	KK3	1		_																				
SC.W I:X B	88	1		-	-	_																		
SC.W 1:X B	41*'	1		4	-																			
SC.W 1:X B	49A'	2		4	-																			
SC.W 1:X B	UU3'	1		+																				
SC.W 1:X B	51A'	3		4	-																			
SC.W 1:X B	W'	3	+	+																				
SC.W 1:X B	XXX'	1	+																					
SC.W 1:X B	UU3'	6	+	+																				
SC.W 1:X C	86'	1		+																				
SC.W 1:X C	32A'	1		+																				
SC.W 1:X C	40'	1		+																				
SC.W 1:X C	41A'	1		+																				
SC.W 1:X C	40A'	1																						
SC.W 1:X C	J3'	1																						
SC W 1:X C	UU3'	4																				1		
SC W 1·X C	77'	1		-																				
SC W 1:X C	35'	3		+ .																				
SC W 1:X C	W'	7	+																	1		1		
SC W I:X C	TITI3.	3	-	+	1	-		+						+					+		+			
EL E 1:100A	112	2		T	CIPa	_	· .		-			T i		-										Fundal lika
ELE 1.100A	V2	1	+		Wht	-	+	+	1	+		LI		Mc	+			+	T ÷	+		+	T i	rungar-nko
FL.F 1:100A	V 3 W2	1	\vdash	+	W IIL XX71-4	+	<u> </u>	+		+		<u> </u>	<u> </u>	1V10 M.	<u> </u>	<u> </u>	<u> </u>				+	<u> </u>	LI	Directown in Co. annuned DI- Bto in Co.
FL.F 1:100A	W 3	1	\vdash		Will DI C	+	<u> </u>	+		+		<u> </u>	<u> </u>	IVIO	<u> </u>	<u> </u>	<u> </u>		+			 		Dips down in Cit, arranged DK PIS in Cit
FL.F 1:10B	X3	1	\vdash		TI/Gn, Dk Cn	+	<u> </u>	+		+			<u> </u>	+	·		<u> </u>		+	<u> </u>	<u> </u>	+	.	
FL.F 1:10B	¥ 3	1	\vdash		PI MIK Bg, Dk Cn	-	+	+		+		<u> </u>	<u> </u>	<u> </u>	Li	<u> </u>	I		I	I		I	Li	Crater-Like, but not uniform Eg
FL.F 1:10B	Z3	1	\square	+	YI, PI Cn	Eg	Cn	+	<u> </u>	Li				<u> </u>	+			<u> </u>	<u> </u>	<u> </u>	Hv	<u> </u>		
FL.F 1:10B	AA3	1	\square	+	CI	_	+	+	<u> </u>	+				+				<u> </u>	<u> </u>	<u> </u>	<u> </u>	+		
FL.F 1:10B	BB3	1			Cl		+	+			+				+	+			+	+		1		Looks Like contour Lines
FL.F 1:X A	CC3	1		4	Lg Bln Cn, Cl Eg		+	+		+				Mo	Li						Li			Fine UND Eg w/ other UND, PR
FL.F 1:X A	DD3	2			Cl		+	+		Li	+						+			+				PR
FL.F 1:X A	EE3	1		+	Cl Eg, Li Yl Cn		+	+		+				+									+	Lg UMB Cn, Sm FLT UND Eg, PR
FL.F 1:X A	FF3	3		+	Cl		+	+		+					Hv								+	
FL.F 1:X A	AA3'	1	+																					
FL.F 1:X A	20'	1		+																				
FL.F 1:X A	92'	1		+																				
FL.F 1:X A	39'	1		4	-		1	1	1	1		1	1	1	1	1	1	1	1	1	1	1		
FL.F 1:X A	GG3	1		1	Cl/Li Bg		+	+	1	1	+	1	1	1	1	1	1	1	1	+	1	1		Grainy fine Tex, Sim to some
FL.F 1:X B	HH3	1			Cl Eg, Mlk Wht/Bg		+	+		1	+	1	1	1	+	+	1		1	1	Li	1		Sim to M. Li RAI at Eg, Frilled Eg

Sample	ID	Cnt	Size i	in mr	n Color	Cons	istency	r	Colon	y Appe	earance	;		Edge						Elev	/ation			Notes
-		#	<1 1	-2 2	7	OPQ	TRL	SHY	DUL	CRC	IRR	FIL	RHZ	ENT	UND	LOB	ERO	FIL	CRL	, FLT	RAI	CVX	UMB	
FL F 1 X B	113	1		4	- Cl Eg Wht Rg Grav Cn	Cn	Eσ	+		Mo				+				+				+		Dkr towards Cn. In sol id Ros. Fungal-like
FL F 1 X B	113	2			Cl	- Cii	+	+			+				+	+		<u> </u>		+		<u> </u>		Hy frilled Eg
FL F 1:X B	KK3	2		-	Cl			+			+				+	<u> </u>				+				Scaled Tex
FL F 1:X B	II 3	1		-	Cl	-	+	+			+				+	+								Sim to II3 Light Eg Eine grainy Tex PR
FL F 1:X B	MM3	1			Pl VI/Bg Cn_Grad To Cl		+	+	1	+		+			+			1					Li	Sin to 355, Eight 1 g 1 no grainy 10x, 1 K
FL F 1:Y B	NN3	1		4	Ph/SalMon	Fσ	Cn	-			+				-						Εσ		1.1	Hy DALEG Lumpy
FL F 1:X B	51'	1		Ŧ	1 Il Salvioli	Lg	CII	т			т				т						Lg			IIV KAI Eg, Eulipy
FL F 1:X B	51"	1		-																				
FL F 1.X B	51	1			-	_	-		1							1		1		-				
FL E LV P	11'	2			-		-	-					-	-						-	-	-		
FLF LAD	12'	2			-	-			-							-		-						
FL.F LA D	15	1			-	-			-							-		-						
FL.F I:A B	251	1			-	-																		
FL.F I:X B	35	1			-	-			-							-		-						
FL.F I:A B	111	1			-	-																		
FL.F I:X B		1			-	-			-							-		-						
FL.F I:X B	63	1		-	-	-			-							-		-						
FL.F I:X B	88	1		-	-																			
FL.F I:X B	64	1	+	_		~	_														~			
FL.F 1:X C	003	1			Wht Cn, Bg Eg	Cn	Eg	+		+					Hv	+					Cn			
FL.F 1:X C	PP3	1			Mo Cl, Bg Blotches inside	_	+	+			+				+						?			Hv frilled Eg (on one side) grainy Tex inside, RAI blotches
FL.F 1:X C	QQ3	2			Li Bwn/Wl/Og Cn, Cl Eg		+	+		+					+									
FL.F 1:X C	RR3	1			Bg Cn, Grad Rg, Cl Eg	CnLi	+	+		+							+		Li	+	LiCn			Frilled Fg Tex Rg and Cn, Cloudy Cn
FL.F 1:X C	DD3'	2																						
FL.F 1:X C	SS3	1		+	 Og/Tng, Dkr Pts 	Pts	Li			Li					+							+		Non uniform CVX, lumpy Dkr OPQ Pts within
FL.F 1:X C	TT3	1		+	Wht Cn, Cl Eg	Cn	Eg	+		+				+									+	PR
FL.F 1:X C	UU3	1		+	Cl		+	+		+					Li							+		PR
FL.F 1:X C	VV3	1		4	- Cl, Li Mlk		+	+		+					+	+			+				+	CRL Cn. RAI dotted Lg Tex Eg, No defined Eg
FL.F 1:X C	WW3	1			Gn/Algae-Like		+	+												+				Tex, Li grainy
FL.F 1:X C	LLL'	1																						
FL.F 1:X C	TT'	1		4	-																			
FL.F 1:X C	DD'	1		+																				
FL.F 1:X C	20'	3	+	+																				
FL.F 1:X C	FF'	1		÷	-																			
FL.F 1:X C	W'	1		÷	-																			
FL.F 1:X C	X'	1		Ŧ	-																			
FL.F 1:X C	LL3'	1																						
FL.F 1:X C	DD3'	2																						
FL.W 1:100A	34'	1			Pk, Dk Near-Rd Cn		+	+		+						Li			+	Mo				Sim to BBB
FL.W 1:100A	DD'	1		+			1	1	1	1		1	1	1	1	1	1	1	1		1	1		
FL.W 1:100A	AA3'	1	+																					
FL.W 1:100A	FF3'	1		+			1	1	1	1		1	1	1	1	1	1	1	1	1	1	1		
FL.W 1:100B	2A	1			Dull Yl/Bg	Mo		+		+					+							Li		Single Rg inside Eg
FL.W 1:100B	3A	1		4	- Dull Bg/Wht	Li		+		+					+	Li					Li			IrrEgular Te, lumpy top
FL.W 1:10A	34'	1																						
FL.W 1:10A	DD'	1		+			1	1	1	1		1	1	1	1	1	1	1	1	1	1	1		
FL W 1.10A	FF*'	1																						
FL W 1:10A	EE3'	1		+			1	1	1				1	1		1	1	1	1		1	1		
FL W 1.10A	GG3'	1	\vdash	·	-	-	1	1	+	<u> </u>		<u> </u>	1	1	<u> </u>	+	1	+	1	1	1	1		
FL W 1.10A	94	1	\vdash	-	PLOg	-	+	+	+	<u> </u>	+	<u> </u>	1	1	+	+	1	+	1	+	1	1		Sim to SCCK PR
FL W 1.10A	104	1	\vdash	+	Pl Wht Li Cn Near Cl	-	+	+	+	Me		<u> </u>	1	Me	Li	+	1	+	1	+	1	1		PR
FL W 1.10A	111	1	+		CIEg VICn	Cr	Fα	т +					1	1410	- 51		1		1	-	1	1	يلى ا	PR
ELW 1.10A	124	1	+		Li Dull WI Ca. Cl/Pa E-	Ch	Eg	+	+	+				+	<u> </u>	+		+					+	DD
FL W 1.10A	12A 20'	3	-	-	- Li Dull WICH, CI/Bg Eg	-	+	+	+	+					+	+		+		+				1 K
ELW 1.10A	20	1	+	т	1	_			+							+		+		-				
I.F.W 1110B	DDJ	1			1		1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	
Sample	ID	Cnt	Size	in mr	n Color	Consi	istency	1	Color	iy Appe	earance	•		Edge						Elev	ation			Notes
------------	-----------------	-----	-----------	-------	--------------------------------	----------	----------	----------	----------	------------	---------	----------	----------	----------	--------	-----	-----	----------	-----	------	--------	-----	-----	----------------------------------
		#	<1 1	-2 2	7	OPQ	TRL	SHY	DUL	CRC	IRR	FIL	RHZ	ENT	UND	LOB	ERO	FIL	CRL	FLT	RAI	CVX	UMB	
FL W 1.10B	ľ	1		-	_																			
FL W 1.10B	34'	1																						
FL W 1:10B	134	1			Patched Cloudy, VI/Cl		+	+			+				Li	+					Τi			
FL W 1:10B	144	1		-	Bg Og Cn Grad Dkr Cn Cl Eg		+	+		Li					+				+		Li			
FL W 1:10B	15.4	1		-	Li Bg/VI Cn Cl Eg		+	-		-					-				T i		1.1		T i	Hy Detal Frills on Eg
FL W 1:10B	164	1		-	Li Bg/VI Cn Miky Cl Eg		т +	τ +		- τ - ⊥					τ +				LI				Li	IIV I chai I filis oli Eg
FL W 1:10B	174	1			PI VI Cn Cl Eg	Cn	Fσ	- T	-	- T		-		-	- T			-					Li	
FL W 1:10B	194	1			Tng/Dh	Cli	Lg	- T	-	- T		-			т			-				T i	Li	PB Sim to 21
FL W 1:10B	DD2	1		+	Pum/Cn Cn Sm Cl Eq	-	+	+		+				+								LI		FK, Shii to 51
FL W 1:10B	204	1			Cl Eq. Mik Pa Ca	1	+	+	-	+				+	T i							LI	T i	DP but Light UND and no Frills
FL W 1:10B	20A	1		1	VICa CIEg	-	+	+		+ Cn	Ea				L1								LI	PR, but Light UND and no Filits
FL.W 1.10B	21A	1		+	Cl/Da asttarr. Dia Dta in Ca	-	+	+		- Ch	Eg				+	÷						т.:	ы	FK Design inside DI-Dis in Ca
FL.W 1:10B	22A	1			- CI/Bg pattern, DKr Pis in Ch		+	+		+				+		¥ ·								Design inside, DK Pts III Cli
FL.W 1:10B	23A	1	<u> </u>	-	- Bg/ whit Ch, Cl Eg		+	+		+				MO		LI						LI		PK DD
FL.W 1:10B	20	2	+	+	C1															14				PD 6: + 20
FL.W 1:10B	24A	1		+	CI	-	+	+		+				+						MO				PK Sim to 20
FL.W 1:10C	54 DD21	1		_						+														
FL.W 1:10C	BB3	1		-		-		-																
FL.W 1:10C	J.	1		-		Di													¥ .		E C	L		
FL.W 1:10C	3A*	1		-	Og/Ing	Pts	MO	+		+					+						Eg, Cr	1		
FL.W 1:10C	4A	1			CIEg, Grad to Bg/Gray Ch	LICI	MO	+		Ovai				+					пv		LI			
FL.W 1:10C	5A	1		-	- CIEg, Grad to Bg Cn	Cn	MO	+		+					+							Lì		Grainy Tex
FL.W 1:10C	6A	1		-	- CIEg, Tng Cn	LiCn	Eg	+		+				+								+		PR
FL.W 1:10C	555	1		-	-	-		-														**		branched Pts inside Dkr
FL.W 1:10C	8A	1		+	Tng	ļ	+	+		+					+							Hv		Volcano-shaped, hollow Cn
FL.W 1:10C	19'	2	+	+															¥ ·					
FL.W I: A	9A	1		_											+				Li					
FL.W I: A	FF [*]	2		-	-	-		-							+									
FL.W I: A	34	8		-	-										+							• •		
FL.W I: A	30	1		-	-	-		-						+								Li		
FL.W I: A	GG3	1		_		-		-						+	* -							Lì		
FL.W I: A	KK 72	1				-		-							Li									
FL.W I: A	Z3	1		+		-		-							+	+								
FL.W I: A	LLS	1		-		-		-						+		¥ ·								
FL.W I: A	YY	1		-	-	-		-						MO		Lì								
FL.W I: A	48A	1		+																м				
FL.W I: A	EE2	1		-	-									+						Mo				
FL.W I:X B	34	22		+ -	-																			
FL.W 1:X B	RR3	1		+		ļ																		
FL.W I:X B	11'	3	\vdash		-	<u> </u>	<u> </u>	<u> </u>		<u> </u>		<u> </u>	<u> </u>	<u> </u>				<u> </u>						
FL.W 1:X B	FF*'	2		+	-																			
FL.W 1:X B	15A'	2		-		ļ																		
FL.W 1:X B	16A'	1		+	-																			
FL.W 1:X B	17A'	1		+	-																			
FL.W 1:X B	KK'	1		_																				
FL.W 1:X B	QQ3'	1	\square		+	<u> </u>		Ļ		Ļ				Ļ		L		L						
FL.W 1:X B	EE3'	1	\vdash	+		 		<u> </u>	<u> </u>				I		I	I			I	I		I		
FL.W 1:X B	42A'	1	\square	+	+	<u> </u>		Ļ		Ļ				Ļ		L		L						
FL.W 1:X B	24A'	1	\square	+	+	I		ļ	 				I	I	I	I		I	I	ļ		ļ		
FL.W 1:X B	35'	5	\square	4	-	<u> </u>		<u> </u>		Ļ				Ļ		L		L						
FL.W 1:X B	22B'	3	\square	+	+	<u> </u>		<u> </u>		Ļ				Ļ		L		L						
FL.W 1:X B	ZZ'	6	\square	4	-	L	<u> </u>	L	<u> </u>	<u> </u>		<u> </u>	L	<u> </u>	L	L		<u> </u>	L	L		L		
FL.W 1:X B	X'	1	+		4	<u> </u>	<u> </u>	L	<u> </u>	<u> </u>		<u> </u>	L	<u> </u>	L	L		<u> </u>	L	L		L		
FL.W 1:X B	37'	1		-	-	L		I	I				I		L	L			L	L		L		
FL.W 1:X B	FF'	1 1	1		-	Î.	1	1	1	1		1	1	1	1	1	1	1	1	1		1		

Sample	ID	Cnt	Size i	n mr	n Color	Cons	istency		Colon	y Appe	earance	;		Edge						Elev	/ation			Notes
,		#	<1 1	-2 2-	7	OPQ	TRL	SHY	DUL	CRC	IRR	FIL	RHZ	ENT	UND	LOB	ERO) FIL	CRL	FLT	RAI	CVX	UMB	
EL W LV D	42'	1								<u> </u>	1	1			1	1		1	1	1				
FL.W LAD	42 WW	1		+												1		-		1				
FL.WI.XD	W W	1		1	-	-												-						
FL.W IXB	E2	1		-	-	_												_						
FL.W I:X B	20B	1		+		_												_						
FL.W 1:X C	34'	9		-	-																			
FL.W 1:X C	42'	3		+																				
FL.W 1:X C	22A'	1		+																				
FL.W 1:X C	FF3'	1		+																				
FL.W 1:X C	15A'	3		4	-																			
FL.W 1:X C	24A'	2		+																				
FL.W 1:X C	HH'	1		+	-																			
FL.W 1:X C	J'	1		+	-																			
FL W 1:X C	63'	1		4																				
FL W 1·X C	SS3'	3		+ +	-																			
FLW 1:X C	BB3	1																						
FLW 1:X C	35'	5				-																		
FLW LXC	55	1	-		-	-										-		-		-				
ELW LVC	EE F	1		τ.	+	+												+	1					
FL.W IAC	1	1			-		<u> </u>		<u> </u>			<u> </u>	+				<u> </u>							
FL.W I:X C	12	1		+	-		<u> </u>			<u> </u>		<u> </u>			<u> </u>	<u> </u>								
FL.W I:X C	15	1		+																				
FL.W 1:X C	46A'	1		4	-																			
FL.W 1:X C	RR'	1																						
FL.W 1:X C	42A'	2		+																				
FL.W 1:X C	U3'	1																						
FL.W 1:X C	EE3'	1	+																					
FL.W 1:X C	QQ3'	1																						
SM.F 1:10B	A2	1			Off Wht		+	+		Cn Li	+				+	+					Li			Sim to G but Dif, Eg LOB instead of FIB/FIL, Crater-Like
SM.F 1:10C	B2	1		+	Y1	+				+				+								Hv		over CVX, balloon-shaped
SM F 1:10C	C2	1		-	- Y1	+			+	Li					+							Hy		Tex. Irregular CVX
SM F 1·X A	D2	1		-	Mlk Wht/C1 Hy Rg	CnI	i Mo	+		+					Hy								+	Erilled I g Eg. Sm Pk Cn
SME LY A	D2'	4	-	-	Mill Mill Of, IT Rg	Cn L										1				1				Third Dg Dg, om TR On
SMELY A	E2	1		-	Off Wht/Ba	Mo	-				-		-	-	+	-	-	-	-		-	lumpe		IPP massas w/ OPO CVX lumps within
SMLP LA A	E2	1	_	-	Sama and Dia Dia ingida	WIO				т.:	+				÷	Ŧ		-		1		iumps		IKK masses w/ OFQ CVX tumps within
SMLF IXA	E2	4		4	- Some w/ Dk Pis inside	+	· · ·			LI							1	-	_					lumps not connected by Sm
SM.F I:X A	F2	1		+	CIEg, PI YICh	_	Li	+		+	_			+				_					+	
SM.F 1:X A	G2	1		-	- Bg, Dk Yl Cn		+	+		Cn	Eg				+	+							Lı	CVX Cn, IRR Eg
SM.F 1:X A	H2	1	+	_	Ph/Pk, Dk patterned Eg		+	+		+				+								+		Dk Patterned
SM.F 1:X B	D2'	5																						
SM.F 1:X B	E2'	1															I	1						
SM.F 1:X B	I2	1		+	- Off Wht/Bg, Cl Eg		+	+		+					Li							+		Color from Spoke-Like Cn protrusions
SM.F 1:X B	J2	1	+	T	Ph/Pk		+	+		+				+								+		
SM.F 1:X B	H2	3		+	Ph/Pk, Dk patterned Eg		+	+		+					Li							+		W/ Lighter UND and Lg Cn
SM.F 1:X C	E2'	2																						
SM.F 1:X C	D2'	4				1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1			
SM F 1:X C	J2'	2	+		T	1	1						1	1	1	1	1	1	1	1	1			
SM F 1·X C	K2	1			- C1	1	+	+	1	+	1	1	1	1	+	1	1	1	+	+	1	1		
SM F 1.X C	1.2	1			- Bo Dkr Eo	+	+	+	<u> </u>	Li	<u> </u>	1	1	1	Hy	+	1	+		+	1	<u> </u>		
SMELYC	1.2'	1				+		<u> </u>				-	1	1	117	1	1	+	1		1	-		
SMLF LAC	M2	1		+	Cl/Bg Cn Mo Bwn Eg	+	-	-		-				+		+	+	+	1	+			Li	
SWLF I:A C	NI2	1		÷	VIC: MILVIE	E	+ M.	+		+				+		<u> </u>		+	+ .		т:		ы	
SNI.W 1:100A	IN2	1			11 CII, MIK/ 11 Eg	Eg	MO	+		+				<u> </u>		+		+	+		LI			
SM.W 1:100B	02	1			- TI/Bg		+	+		+		<u> </u>	<u> </u>	+	<u> </u>	.	<u> </u>		+	<u> </u>	+			
SM.W 1:100B	P2	1			- Sm Cl Eg, Tng Cn	+	L	+		Mo			<u> </u>	+	<u> </u>	Li	<u> </u>	-	<u> </u>	<u> </u>	L	L		Sm LOB Eg
SM.W 1:100B	Q2	1		+	- Tng/Rd		Li	L	L	Li	L	I			+	I	I			I	Hv	Hv		Hv&IRR RAI Crater, Rough Tex
SM.W 1:100B	R2	1		4	- Pl Tng	+		+		Li					+	L		1	1	L	Hv			Hv&IRR RAI Crater, Smooth Tex
SM.W 1:100B	BB3'	1																						Uniform RAI

Sample	ID	Cnt	Size in	mm	Color	Consi	stency		Colon	y Appe	arance			Edge						Elev	/ation			Notes
		#	<1 1-	2 2-7	1	OPQ	TRL	SHY	DUL	CRC	IRR	FIL	RHZ	ENT	UND	LOB	ERO	FIL	CRL	FLT	RAI	CVX	UMB	
SM W 1-100P	EE'	1		1															1				-	
SM W 1.100B	EE 62	1	+		Terr		т.:			т.:											11			
SM.W 1:100C	32	1	- I .	+	Thg			+							+						ПV			
SM.W 1:100C	Z2 T2	1	+		Ing		+	+		Li					+						HV			F' T F I'
SM.W 1:100C	12	1		+	Ing, LIEg		+	+		+												+		Fine Tex on Eg, Li
SM.W 1:100C	02	1		+	Off wht	+		+		+				+					+			Lı		Sm CRC colonies making Lg IRR mass
SM.W 1:100C	V2	1		+	Off Wht/Cl		+	+		Lı				+		Lı					Lı			
SM.W 1:100C	W2	1		_	Sm Yl Cn, Lg Cl Eg,		+	+		+		Li			+			Li					Li	
SM.W 1:10A	X2	1			Wht	+		+		Cn	Mass				Li							+		Crater-Like Mostly CRC but IRR mass
SM.W 1:10A	Y2	1		+	Li Og/Yl		+	+		+				+						+				
SM.W 1:10A	AA2	1		+	Bri Yl		Li	+			+				+						+			
SM.W 1:10A	BB2	1		+	Mlk Wht, Li Cl Cn	Eg	Cn	+		+					+						+			
SM.W 1:10A	AA2'	1	+																					
SM.W 1:10A	CC2	1		+	Cloudy, Vy Li Og/Bwn		+	+		+				Mo						+				Course Tex
SM.W 1:10B	DD2	1			Wht, Cl Eg	Mo		+			+				+	+					Eg	Eg		Sim to X2 but sMoother & w/o as many distinct colonies and Eg
SM.W 1:10B	EE2	1		+	Li Wht, Mo Cl		+	+		+				+								+		
SM.W 1:10B	FF2	1	+		Yl/Og Cn, Li Eg		+	+		+				+								Li		
SM.W 1:10B	GG2	1		+	Li Pastel Og/Yl		Li	+		+				+								Li		
SM.W 1:10B	HH2	11	+		Yl. Li Cn	Eg	Cn	+		+					+						+			
SM.W 1:10B	BB2'	1		+																				
SM.W 1:10C	112	1		1	Wht	+		+			+				+						+	Eg		Sin to X2 and DD2, sMooth Like DD2 but w/o distinct Eg
SM W 1.10C	72'	1	+																					
SM W 1:10C	BB2'	1		+												+								
SM W 1:10C	112	1	-		Bri Pastel VI	+		+		+				+								+		
SM W 1:10C	KK2	1			Tng		4	-			+				-						-1-			Limpy
SM.W 1:10C	LL2	1		-	Pastal VI Li Eg	Mo	т	- T	-	-	Ŧ				Ti			-	-	-	т		-	Lumpy
SM.W 1.10C	MM2	1			Critica Li Da Cl Ea	IVIO		τ		Ŧ											τ.:		Ŧ	Center Libr
SM.W 1.10C	NN12	1		+	Gry Dk Eg. Li Cn	Ea	+ Cn	+		+					+						Ea			Claici-Like
SM.W 1.10C	002	1		+	OIY, DK Eg, LI CII	Eg	Cli	+			+				+						Eg	¥ ·		
SM.W 1:10C	002	1	+	-	ŶÌ		+	+		+				+								Li		
SM.W I:X A	60	1		_																				
SM.W I:X A	62	2		+																				
SM.W 1:X A	FF*'	3		+																				
SM.W 1:X A	22B'	3	+																					
SM.W 1:X A	36B'	1		_																				
SM.W 1:X A	BB2'	1		+																				
SM.W 1:X A	HH'	1		+																				
SM.W 1:X A	45B'	1																						
SM.W 1:X A	42A'	1		+																				
SM.W 1:X A	86'	1		+																				
SM.W 1:X A	QQ2'	1		+																				
SM.W 1:X A	SS'	1																						
SM.W 1:X A	44B'	1																						
SM.W 1:X A	RR2'	4		+																				
SM.W 1:X A	FF2'	1	+																					
SM.W 1:X A	30'	1		+																				
SM.W 1:X A	TT'	1	+	1				1	1							1		1	1	1	1	1	1	
SM.W 1:X A	PP2'	1		+														1	1	1	1			
SM.W 1:X A	41B'	1		1					1									1	1	1	1			
SM.W 1:X A	12A'	1		+				1	1							1		1	1	1	1	1	1	
SM W 1·X A	XXX'	1	+	†÷					<u> </u>									1	<u> </u>	1	1			
SM W 1·X A	40'	3		+				<u> </u>								<u> </u>		1	1	1	1	1	<u> </u>	
SM W 1·X A	7*'	4		1				<u> </u>								<u> </u>		1	1	1	1	1	<u> </u>	
SMLW LAA	/ · DD2'	4		+																				
SIVLW I:A B	KK2 02'	2	<u> </u>	+																				
SIVI. W TA B	02	2	+	+				<u> </u>								<u> </u>		<u> </u>		<u> </u>	<u> </u>	<u> </u>	<u> </u>	
SIVLW TAB	1	1		1 +	1			1	1							1		1	1	1	1	1	1	

Sample	ID	Cnt	Size i	n mr	1 Color	Consi	istency		Colon	y App	earance			Edge						Elev	ation			Notes
-		#	<1 1	-2 2-'	7	OPQ	TRL	SHY	DUL	CRC	IRR	FIL	RHZ	ENT	UND	LOB	ERO	FIL	CRL	FLT	RAI	CVX	UMB	
SM W 1·X B	35B'	1		+																				
SM W 1 X B	WW'	1		·																				
SM W 1:X B	FF2'	1		L.																				
SM W 1:X B	13'	1		<u> </u>																				
SMW 1:XB	MM2'	1																						
SMW 1:XB	BB2	1		1																				
SMW 1.X B	124'	3		+			-		-					-		-		-	-	-		-		
SMI.W LX D	42A	1		+			-		-					-		-		-	-	-		-		
SMI.W LAD	22B	1		-	1	-							1											
SMI.W LAD	DD2	1			1	-							1											
SM.WIAD	PP2 97	1		+																				
SMI.W LAD	44D'	1		+	1	-							1											
SM.WIXD	44D	1		+																				
SM.W IXB	W	1		+		-							-											
SM.W IXB	62	2		+		-							-											
SM.W I:X B	10A'	1		÷																				
SM.W I:X B	VV'	1		+																				
SM.W 1:X B	34'	1		+																				
SM.W 1:X B	19'	5	+	+																				
SM.W 1:X B	J'	1		+																				
SM.W 1:X B	16A'	1		+																				
SM.W 1:X C	WW2'	2	+			Mo		+		+					+							+		
SM.W 1:X C	FF2'	1		÷																				
SM.W 1:X C	41B'	1																						
SM.W 1:X C	86'	1		+																				
SM.W 1:X C	11'	1		+																				
SM.W 1:X C	GG3'	1																						
SM.W 1:X C	34'	1		+																				
SM.W 1:X C	12A'	1		+																				
SM.W 1:X C	GGG'	1		+																				
SM.W 1:X C	35'	1		+																				
SM.W 1:X C	J'	1		+																				
SM.W 1:X C	EE'	1		+																				
SM.W 1:X C	35B'	1		÷																				
SM.W 1:X C	64'	1	+																					
SM.W 1:X C	J3'	1																						
SM.W 1:X C	44B'	1		+																				
SM.W 1:X C	TT2'	1		+																				
SM W1:X C	16B'	1																						
SM.W 1:X C	42A'	1		+																				
SH.F 1:100A	PP2	62		+	Bg Cn. Mlk Wht Eg	1	+	+	1	+			1	1	+	1		1	1	1		1	+	Sim to D2 but Dif, not as OPO, no Rg, under 1 cm, More CRC
SH.F 1:100A	002	24		+	Bg Cn. Mlk Wht Eg	LiCn	Mo	+							+								Li	Larger Cn. CRC, less pointed UMB, Dk Cn
SH F 1.100A	RR2	5		+	Lo Bo Cn Sm Li Bo Fo	Cn	Eσ	+							Li							+		
SH.F 1:100A	SS2	9	\vdash	. +	Dk Bg Cn. Mlk Wht Eg	LiCn	+	+	1	Li	+		1	<u> </u>	Hy	1		1	<u> </u>	1		<u> </u>	Li	Larger Cn. More IRR but smaller Cn than RR2
SH F 1:100A	TT2	15		ь <u>т</u>	Off Wht	Mo	Li	+									+			Mo				Earger end more inter our sindler en daar inter
SH F 1:100A	PP2'	40		· · · ·	on one	1410			1	1	<u> </u>	1	+		<u> </u>	1	<u> </u>	1	1	1010	<u> </u>	1	-	
SH F 1.100A	002	30	\vdash	+			+		+				-	+		+		+		+		+		PR 2 one w/ More VI Cn other w/ Wht and less OPO
SH F 1:100A	RR2'	2		-		+			1	1	<u> </u>	1	+		<u> </u>	1	1	1	1	1	<u> </u>	1	-	The 2, one with those The cit, other with the and less of Q
SH F 1.100A	\$\$2'	8	\vdash	-			+		+					+		+		+		+		+		
SHE 1.100B	DD'	2	\vdash	-			+		+					+		+		+		+		+		
SHE 1.100B	DD'	<u> <u> </u></u>	\vdash		1	+							+											
SHE 1:100D	002	26	\vdash	+	1	+							+											
SH.F 1:100C	QQ2'	20	\vdash	+																				
SH.F 1:100C	KKZ'	2		÷						<u> </u>		<u> </u>	+				<u> </u>							
SH.F 1:100C	552	10	\vdash			ł	-							-										DD.
SH.F 1:100C	DD'	5	1 1			1	1	1	1	1	1		1	1	1	1	1	1	1	1	1	1	1	PK

$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	Sample	ID	Cnt	Size	in m	m Color	Const	istency		Colon	y Appe	earance	;		Edge						Elev	vation			Notes
SHP 10A PC P<	<u> </u>		#	<1	1-2 2	-7	OPQ	TRL	SHY	DUL	CRC	IRR	FIL	RHZ	ENT	UND	LOB	ERO) FIL	CRL	FLT	RAI	CVX	UMB	
Diff 10. COL OP I I I	SH E 1-10A	DD2'	SD								<u> </u>	1		1						1		1	1	1	
Diff Dial No. Display Display <thdisplay< th=""> <thdisplay< th=""> <thdis< td=""><td>SH.F 1.10A</td><td>002</td><td>SF</td><td>+</td><td>+</td><td></td><td>-</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td>-</td><td></td><td></td><td></td><td></td><td></td><td></td></thdis<></thdisplay<></thdisplay<>	SH.F 1.10A	002	SF	+	+		-												-						
Diff Find OCC Diff Find Diff	SH.F 1:10A	QQ2	SP	+	+		-											-	-						
Number 1 1<	SH.F 1:10A	SS2 [*]	SP	+	+																				
Bill 10 Bill 1 I <t< td=""><td>SH.F 1:10A</td><td>CCC</td><td>1</td><td></td><td></td><td>+</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></t<>	SH.F 1:10A	CCC	1			+																			
SHF 1:0A DC I	SH.F 1:10A	HH'	1			+																			
SHE 160 VQ I YI I	SH.F 1:10A	DD'	1	+	+																				
SHF 1:100 YP SP I <th< td=""><td>SH.F 1:10A</td><td>UU2</td><td>1</td><td></td><td>+</td><td>Yl</td><td>+</td><td></td><td>+</td><td></td><td>+</td><td></td><td></td><td></td><td></td><td>+</td><td></td><td></td><td></td><td></td><td></td><td></td><td>+</td><td></td><td>Shadowed Eg</td></th<>	SH.F 1:10A	UU2	1		+	Yl	+		+		+					+							+		Shadowed Eg
SHF 1:00 QC SP I <thi< td=""><td>SH.F 1:10B</td><td>PP2'</td><td>SP</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></thi<>	SH.F 1:10B	PP2'	SP																						
SHF 1:00 SS: SP A <	SH.F 1:10B	QQ2'	SP																						
SHF 1:00 CC 1 6 7	SH.F 1:10B	SS2'	SP																						
SHF 1:08 CCC 1	SH.F 1:10B	RR2'	SP																1			1			
Silf Fill No I	SH F 1.10B	CCC'	1																1			1			
Bit Fillor Dir Image: Series of the state of the sta	SH E 1:10B	DD'	1		-															1		1			
Diff File Core	SHE 1:10D	DD'	CD 1	+	-		-												-	-		-			
BATE JUNC CX3 BST Image: CX3	SHE 1.10C	002	CD	+	-		-												-	-		-			
Bith Fillow Skill Sign Image of the state of the sta	SH.F 1:10C	002	SP		_		-											-	-						
Mit Fillor Mit Fillor <td>SH.F 1:10C</td> <td>552</td> <td>SP</td> <td></td> <td>_</td> <td></td> <td>-</td> <td></td> <td>-</td> <td></td> <td></td> <td></td>	SH.F 1:10C	552	SP		_															-		-			
SHF 1:102 CCC 1 <td< td=""><td>SH.F 1:10C</td><td>RR2</td><td>SP</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></td<>	SH.F 1:10C	RR2	SP																						
SHP E10C DV 1 - VIOg. Sm Li Eg + + + + - <td>SH.F 1:10C</td> <td>CCC</td> <td>1</td> <td></td>	SH.F 1:10C	CCC	1																						
SHP i:A VV2 1 + VV2 1 + Wo Li + Dkrblocks inside SHP i:A QQ2 SP -	SH.F 1:10C	DD'	1																						
SHF 1XA QP2 SP I Image: SHF 1XA QQ2 SP Image: SHF 1XA SS2 SP Image: SHF 1XA SS2 SF Image: S	SH.F 1:10C	VV2	1	+		Yl/Og, Sm Li Eg	+		+		+				Mo		Li					+			Dkr blotches inside
SHF IXA QQ2 SP I Image: Second constraints of the second consecond constraints	SH.F 1:X A	PP2'	SP																						
SHF 1XA SS2 SP I Image: ShF 1XA SS2 SP Image: ShF 1XA SS2 SP Image: ShF 1XA SS2 SF	SH.F 1:X A	QQ2'	SP																						
SHF 1X A RE2 SP	SH.F 1:X A	SS2'	SP																						
SHF 1X A WW2 2 Mo Grainy Tex SHF 1X A UU2 2 + H	SH.F 1:X A	RR2'	SP																						
SHE FLX A DD2 1 + - C N <th< td=""><td>SH.F 1:X A</td><td>WW2</td><td>2</td><td>Mo</td><td></td><td>Gry/Wht</td><td>+</td><td></td><td>+</td><td></td><td>+</td><td></td><td></td><td></td><td></td><td>+</td><td></td><td></td><td></td><td></td><td></td><td></td><td>+</td><td></td><td>Grainy Tex</td></th<>	SH.F 1:X A	WW2	2	Mo		Gry/Wht	+		+		+					+							+		Grainy Tex
SHF F1X A UU2 2 + Image: Constraint of the cons	SH.F 1:X A	DD2	1		+																				Several that apperar Sim
HIF IX B PPP SP I Image: SP Imag	SH.F 1:X A	UU2'	2		+																				
EHF 1X B OQ2 SP Image: SP	SH.F 1:X B	PP2'	SP																						
SHE F iX B SS2 SP I I I I I I I I I I SHE F XB RR2' SP I I I I I I I I SHF F XB RR2' SP I I I I I I I I I I SHF F XB XX2 I I Clew solid PS. Pis + + + + + + + I H SHF F XB XX2 I Clew solid PS. Pis + + + + + + + + H SHF F XC PP2 SP I Clew solid PS. Pis + + + + + + + H SHF F XC PP2 SP I I Clew solid PS. Pis I I I I I IRC n SHF F XC PP2 SP I I I I I I I I I IRC n SHF F XC U22 I I I I I I I I I	SH.F 1:X B	002'	SP																						
Shifix B RR2 SP Image: Shifix B RA R	SH F 1·X B	\$\$2'	SP																						
Shifi X.B UU2 2 + + + + + + + + + All in connected lumps SHF IX B XX2 1 PlogPh + + + + + + RAI in connected lumps SHF IX B YX2 1 Cl.few solid Ps. Pts + + + + + RAI in connected lumps SHF IX B YX2 1 Cl.few solid Ps. Pts + + + + + RAI in connected lumps SHF IX C PV2 SP -	SHE 1:X B	RR2'	SP																						
Diff. 1X.B CX2 1 Plog/Ph + + + + + + + ARI in connected lumps SH.F 1X.B YY2 1 Cl, few solid Pts. Pts + + HV Mo IRR Cn SH.F 1X.C QQ2 SP Image: Close Solid Pts. Pts + + HV Mo IRR Cn SH.F 1X.C QQ2 SP Image: Close Solid Pts. Fts Image: Close Solid Pts. Fts Image: Close Solid Pts. Fts	SHE 1:X B	LILL2'	2		+																				
Initial Direction Pice Pi	SHE 1:Y B	XX2	1		<u> </u>	Pl Og/Ph		-1-	-			-				-									PAL in connected lumps
Init ND	SHELV B	VV2	1		-	Cl. faw soLid Pts	Dtc	- T	- T	-	-	т				H _V	т				Mo	т		-	IPP Cn
SHLF 1:XC QQ2 SP Image: Constraint of the second sec	SHE LVC	DD2'	CD 1		-	CI, ICW SOLID I IS.	115	т	т	-	т					117	-				WIO			-	ikk ch
Sh.F 1:XC QQ2 SF I <t< td=""><td>SHELVC</td><td>002</td><td>SF</td><td></td><td>-</td><td></td><td>-</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td>-</td><td></td><td></td><td></td><td></td><td></td><td></td></t<>	SHELVC	002	SF		-		-												-						
SHF 11XC SK2 SP Image: SP Iman	SHFIXC	002	SP		-		-												-						
Sh.F. 1:X.C KR2 SP Image: SP I	SH.F I:X C	552	SP		_															-		-			
Sh.F. 1:X.C UO2 3 + - <	SH.F 1:X C	RR2	SP		_		_												_						
SH.F.1:X C YY2'' 1 I I Hylg, DkrY1Cn Mo I Hv Li Image: Constraint of the second s	SH.F I:X C	002	3	+	+		I	<u> </u>	<u> </u>	<u> </u>	I	<u> </u>		<u> </u>	<u> </u>	<u> </u>	<u> </u>	I		I	<u> </u>	I	<u> </u>	I	
SH.F.1XCZZZI++YHg, Dkr Y1CnMo+LiHvLiLiLiLimpy but Li CVX shapeSH.W.1100A021++<	SH.F 1:X C	YY2'	1	\square			1	<u> </u>	I	<u> </u>	L	I		I	L	L	L		ļ	I	<u> </u>	I	I		
SH.W 1:100A 02° 1	SH.F 1:X C	ZZ2	1			+ Yl/Bg, Dkr Yl Cn	Mo		+		Li					Hv	Li						Li		Lumpy but Li CVX shape
SH.W 1:100A1B1PK/PH++++VyLi+MoLiEnd but not complete CRC, Li RAI on CRLSH.W 1:100A3B1+Tng+LiEg+MoLiLiHvSMooth but not quite CRCSH.W 1:100A3B1+HH+HLiLiHvSMooth but not quite CRCSH.W 1:100BYY'1+H+HHHHHvSMooth but not quite CRCSH.W 1:100B46'1+HHHHHHHHSH.W 1:100B40'1+HHHHHHSH.W 1:100B19'1+HHHHHHSH.W 1:100B19'1HHHHHHHHSH.W 1:100B19'1HHHHHHHHSH.W 1:100B19'1HHHHHHHHSH.W 1:100B19'1HHHHHHHHHSH.W 1:100B5B1HHHHHHHHHSH.W 1:100B5B1HHHHHHHHHSH.W 1:100B5B1HHHHHHHHH	SH.W 1:100A	O2'	1			+																L			
SH.W 1:100A2B1+Tng+LEg+MoLiLiLiHvSMooth but not quite CRCSH.W 1:100B3B1+BwnOg++++LiLiLiHvSMooth but not quite CRCSH.W 1:100BYY1-BwnOg+++-LiLiHvHvSH.W 1:100BS6'1+LiLi-HvHvSH.W 1:100B46'1+LiHvHvSH.W 1:100B19'1+SH.W 1:100B19'1+SH.W 1:100B14'-PIYlog, Sm ClEg+++MoLi-+CRLPR, RAI at CRL, CL FLT EgSH.W 1:100B5B1-PIYlog+++Hv+CRLLiFg at Eg outside of CRL, RAI at CRLSH.W 1:100B5B1-+Tmg+++Hv++PRSH.W 1:100B5B1-PINCG Gradue ClEg+++Hv+PR+PRSH.W 1:100B6B1+Tmg++Hv+LiHv+PRSH.W 1:100B7B1+	SH.W 1:100A	1B	1			PK/PH		+	+		+				+	VyLi				+	Mo	Li			Ent but not complete CRC, Li RAI on CRL
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	SH.W 1:100A	2B	1			+ Tng	+	LiEg	+		Mo					Li	Li						Hv		SMooth but not quite CRC
SH.W 1:100B YY' 1 Image: Constraint of the state of t	SH.W 1:100A	3B	1		+	Bwn/Og	+		+			+					Li						Hv		
SH.W 1:100B 86' 1 +	SH.W 1:100B	YY'	1																						
SH.W 1:100B 40' 1 + <td< td=""><td>SH.W 1:100B</td><td>86'</td><td>1</td><td></td><td></td><td>+</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></td<>	SH.W 1:100B	86'	1			+																			
SH.W 1:100B 19' 1 + SH.W 1:100B 19' 1 PI YI/0g. Sm Cl Eg + + Mo Li + CRL PR, RAI at CRL, CL FLT Eg SH.W 1:100B JT I I PI YI/0g. Sm Cl Eg + + Mo Li + CRL PR, RAI at CRL, CL FLT Eg SH.W 1:100B JS 1 PI YI/0g + + Hv + CRL Li Fa at Eg outside of CRL, RAI at CRL SH.W 1:100B GB 1 + Tmg + + Hv + CRL Li Fa at Eg outside of CRL, RAI at CRL SH.W 1:100B GB 1 + Tmg + + Hv + Li + PR SH.W 1:100B 7B 1 - - - - Li - + PR SH.W 1:100B 7B 1 - + + + - + Li - + PR SH.W 1:100B 7B 1 - - - - Li - + PR SH.W 1:100B 7B 1 - + + - + Li - <td>SH.W 1:100B</td> <td>40'</td> <td>1</td> <td></td> <td></td> <td>+</td> <td>1</td> <td>1</td> <td>1</td> <td>1</td> <td>1</td> <td>1</td> <td></td> <td>1</td> <td></td>	SH.W 1:100B	40'	1			+	1	1	1	1	1	1		1	1	1	1	1	1	1	1	1	1	1	
SH.W 1:100B 4B 1 PI Y/Vg, Sm Cl Eg + + Mo Li + CRL PR, RAI at CRL, CL FL Eg SH.W 1:100B J' 1 -	SH.W 1:100B	19'	1			+	1	1	1	1	1	1		1	1	1	1	1	1	1	1	1	1	1	
SH.W 1:100B J I I I I I I SH.W 1:100B 5B 1 I I I I I I SH.W 1:100B 5B 1 I I I I I I SH.W 1:100B 6B 1 I I I I I I SH.W 1:100B 6B 1 I I I I I I SH.W 1:100B 7B 1 I I I I I I SH.W 1:100B 7B 1 I I I I I I	SH.W 1:100B	4B	1	t t		Pl Yl/Og, Sm Cl Eg	1	+	+	1	+				Mo	Li	1		1	+	1	CRL			PR. RAI at CRL, CL FLT Eg
SH.W 1:100B SB I PI Y/V0g + + Hv + CRL Li Fg at Eg outside of CRL, RAI at CRL SH.W 1:100B 6B I + Tmg + + + CRL Li Fg at Eg outside of CRL, RAI at CRL SH.W 1:100B 7B I + + + + Li + PR SH.W 1:100B 7B I + + + Li + PR	SH.W 1:100B	J.	1	t t			1	1		1							1		1	1	1				· · · · · ·
SH.W 1:100B 6B 1 + Tng + + + + + Li + PR SH.W 1:100B 7B 1 - P Og Cn Grad to CLE q + + + + + Li - + R	SH W 1.100B	5B	1	+		ΡΙ ΥΙ/Οσ	1	+	+	1	+					Hv	<u> </u>	1	1	+	<u> </u>	CRI		Li	Eg at Eg outside of CRL, RAL at CRL
HW 11008 78 1 Plog Cn Grad to ClEg	SH W 1.100B	6B	1	+		+ Tng	1	+	+	1	+	<u> </u>		<u> </u>	<u> </u>	+	Li		+		1	CILL	+		PR
	SH W 1.100B	7B	1	+	-+	PLOG Cn Grad to Cl Eg	1	+	+	1	+	<u> </u>		<u> </u>	<u> </u>	Li	Li		+	Li	Me	+			Li RALon Cn & CRL Pts in pattern toward Eg

Sample	ID	Cnt	Size	in m	n Color	Consi	istency	r	Color	y Appe	earance	e		Edge						Elev	ation			Notes
		#	<1 1	1-2.2	-7	OPQ	TRL	SHY	DUL	CRC	IRR	FIL	RHZ	ENT	UND	LOB	ERO	FIL	CRL	FLT	RAI	CVX	UMB	
SH W 1.100B	XXX'	1	+																					
SH W 1:100C	1B'	1	-																					
SH W 1:100C	XXX'	1	+																					
SH W 1:100C	FF'	1	-	-	L .																			
SH W 1:100C	8B	2		-	PLV1 DkrCn		-	-		-					-1-		тi				-			Frilled F.g. Lumpy Tex. Dkr. More OPO Cn
SH W 1:100C	OR	1		-	Bwn Cn Dkr Pg Bg/Bwn Eg	Cn	Fσ	т +		- T - ⊥					τ +	т 	Li		T i		- T - ⊥			Trinca Eg, Eampy Tex, DK More of Q En
SH W 1:100C	10B	1		-	Bwn Cn, Lg Pl Vl Eg	Cii	Lg	- T		- T		-		-	Ti	т	T i		Li	Mo	Ŧ	-		Sim to 0B but Dif
SH W 1:100C	11D	1		-	Dk Pg	-	T ;	-		- T		-		-		-	Li		-	WIO		-	T i	
SH.W 1:100C	11D	1		-	+ DK Bg		LI	÷	1	÷					+		LI	-		-			Li	
SH.W 1.100C	110	1		-					1									-	т.:	-			т:	Sim to L & 14 but Dif
SH.W 1:100C	12D DD2'	1			+ bg, ci bg		÷	÷	1	÷				÷			÷	-	LI	-			Li	Siin to 1 & 14 but Di
SH.W 1.100C	12D	1		+	W7b4			т:	1		т:				т.:			-		-			T.	DD.
SH.W 1:100C	130	1		+	WIII	+	C	LI		+	LI				LI						F		пv	PK
SH.W 1:100C	14D	2		+	Dk Ing	Eg	Cn	+		+					пv						Eg			Crater-Like, PK
SH.W 1:100C	200	3		_		-																		
SH.W 1:10A	HH	1		_	+				-									-		-				
SH.W 1:10A	35	1		_	+				-									-		-				
SH.W 1:10A	RR3	1		+		_																		
SH.W 1:10A	003	1		_					-									-		-				
SH.W 1:10A	62	1		_	+																			
SH.W 1:10A	25	1		_	+				-									-		-				
SH.W 1:10A	14	1		_	+	_																		
SH.W 1:10A	19'	1		_	+																			
SH.W 1:10A	20	2		+		_																		
SH.W 1:10A	24A'	2		+																				
SH.W 1:10A	FF'	1		+		-																		
SH.W 1:10A	DD'	4			D. C. MILWEID, CL.D.	6								**										
SH.W 1:10A	15B	1		_	Bg Cn, Mlk Wht Rg, CL Eg	Cn	Mo	+		+				Hv			+			Mo				Pr But Dif, Multi Rg, ridged/frilled inner Eg, Fit Tex outer Eg
SH.W 1:10A	16B	1		_	Li Bg Cn, CL Eg		+	+		+				Hv					Lı				Lı	Sim to 72, Hv frills on Eg
SH.W 1:10A	16B'	1				-																		
SH.W 1:10A	17B	1		_	+ Cloudy Og/YI, Grad to CI Eg		+	+		+				Mo		Lı						+		
SH.W 1:10A	22B'	1		+																				Original
SH.W 1:10A	18B	1			Pl Mlk Wht/Ph		+	+		+				Mo						Mo		Li		Not Entirely CRC
SH.W 1:10A	19B	1			+ Bwn/Rd, Dk Pts		+	+		+				+							Li	Li		Small OPQ Pts
SH.W 1:10A	20B	1		_	+ Li Bg/Yl Cn, Cl Eg		+	+		+					+								+	PR, totally FLT Eg, Hv CVX Cn
SH.W 1:10A	21B	1			Cl, Li Bg Cn		+	+			+				+	+	Cn			Mo				
SH.W 1:10A	22B	1		+	Cl		+	+		+				+					+				+	PR, Sim to small FF
SH.W 1:10A	23B	1			+ Yl, Li Eg		+	+		+					+	+	Cn					Li		
SH.W 1:10A	24B	1			Cl		+	+		Cn	+				+	+			Li	Mo				Sim but Dif from 21B
SH.W 1:10A	25B	1			Mlk Bg/Wht Cn, Cl/Bg Eg	LiCn	Mo	+			+				+	+							Hv	Small frilled Eg, Eg bulbous cloudy Cn
SH.W 1:10A	26B	2			Li Bg Cn, Cl Eg		+	+		+					+		HvCr	1					+	PR, Sim to 62
SH.W 1:10B	14'	1			+																			
SH.W 1:10B	SS2'	1			+																			
SH.W 1:10B	YY2'	1			+																			
SH.W 1:10B	SS'	1																						
SH.W 1:10B	J'	1																						
SH.W 1:10B	W'	4		+	+																			
SH.W 1:10B	8A'	3		+	+																			
SH.W 1:10B	RR2'	2																						
SH.W 1:10B	72'	1			+																			
SH.W 1:10B	45'	1		+																				
SH.W 1:10B	42'	1	+																					
SH.W 1:10B	QQ2'	3			+																			
SH.W 1:10B	Γ	2			÷																			
SH.W 1:10B	X'	2		+	+																			

Sample	ID	Cnt	Size	in m	n Color	Cons	istency	r	Color	y Appe	earance			Edge						Elev	/ation			Notes
· ·		#	<1 1	-22	-7	OPQ	TRL	SHY	DUL	CRC	IRR	FIL	RHZ	ENT	UND	LOB	ERO	FIL	CRL	FLT	RAI	CVX	UMB	
SH W 1.10B	VV'	1																						
SH W 1:10B	DD'	5	+	+ .																				
SH W 1:10D	20'	6		<u> </u>																				
SH.W 1.10B	24 4'	1		-	1				1							-				-				
SH.W 1.10D	24A EE*!	1		+					1							-				-				
SH.W 1:10B		0			F	-												_						
SH.W 1:10B	25B	1			-	_												_	¥ ·					DD.
SH.W 1:10B	2/B	1			F PI YI/Og	_	+	+		+				+					Lı		Li		Li	PR
SH.W 1:10B	26B'	1																						
SH.W 1:10B	28B	1			Lg Cloudy Bg Cn, Mlk Wht Eg	LiCn	Mo	+		+					+								+	Hv frills on Sm Eg
SH.W 1:10B	28B'	1																						
SH.W 1:10B	29B	1			Sm Bg Cn, CL Eg		+	+		+					+				+				+	Lg Frills
SH.W 1:10B	30B	1			Bwn/Og Cn, Wht Eg	+		+		+					+	+						+		
SH.W 1:10B	30B'	2			F																			
SH.W 1:10B	31B	1			Bwn/Og Cn, Cl Eg	Cn	Eg	+		+					+								+	
SH.W 1:10B	32B	1			 Yl/Bwn Cn, Cl/Wht Eg 		Li	+		+					+						+			
SH.W 1:10B	32B'	2			+																			
SH.W 1:10B	33B	1			Bg/Mlk Wht, Dkr Cn		Li			+					+								Li	PR frilled Eg
SH.W 1:10B	34B	1			+ C1		+	+		+					+		+					+		PR !!!!
SH.W 1:10B	35B	1			- Og/Bwn Cn. Cl Eg		+	+		+					+				Li			+		not quite CRC. Dkr Pts
SH.W 1:10B	36B	1			YI/Og		+	+		Mo				Mo	Li							Li		Hy frills on Eg. Sim to OO2
SH W 1.10B	37B	1			Bwn/Og Cn Cl Eg		+	+		+					+								Li	
SH W 1:10C	30'	1			Dwill Og Cil, Ci Eg																		Li	
SH W 1:10C	10'	1		+		-	-	-		-				-	-			-			-	-		
SH W 1:10C	1) V'	1		T .		-	-	-		-				-	-			-			-	-		
SH W 1:10C	1 1 2'	1		-	F				1							-				-				
SILW 1.10C	10	1		-					1							-				-				
SH.W 1:10C	002	1			F	-												_						
SH.W 1:10C	00	1			+	-			-							-				-				
SH.W 1:10C	II'	1			+	_																		
SH.W 1:10C	RR2	6			+	_																		
SH.W 1:10C	86'	2			+																			
SH.W 1:10C	SS2'	1																						
SH.W 1:10C	35'	1			+																			
SH.W 1:10C	VV'	3		+																				
SH.W 1:10C	U2'	1			F																			
SH.W 1:10C	DD'	3	+		F																			
SH.W 1:10C	37B'	2			F																			
SH.W 1:10C	32B'	2																						
SH.W 1:10C	26B'	3																						
SH.W 1:10C	31B'	1			÷																			
SH.W 1:10C	FF*'	1			F																			
SH.W 1:10C	W'	2			E .																			
SH.W 1:10C	38B	1			Dull YI. Dkr Cn	1	+	+	1	+				1	+	1	+	1	1	1	1	1	+	
SH.W 1:10C	39B	1 î	+		Pl Yl. Dkr Pts	1	+	+	1	Li				1	+	1	t ·		1	1	<u> </u>	Li	<u> </u>	Sim to 36 but More UND and More Pts
SH W 1:10C	39B'	1 î	+	-		1	<u> </u>	<u> </u>	1			<u> </u>	<u> </u>	1	<u> </u>	1		1		1			<u> </u>	
SH W 1:10C	40P	1	+	-	Bg/Og Dkr Cloudy Cr	1	Тi	+	1	-			-	+	1	1	1	1	1	1	1	Тi		DD
SH W 1.10C	41P	1	++	-		1	- LI 	+	+	Ť			 	+	+	+	Cr	+	-	+		LI		
SH.W 1.10C	410	1	++	_		+	+	+	+	LI					+	+	Cil	1		+				
SH.W 1:10C	40	1	\vdash			+	<u> </u>	<u> </u>			<u> </u>				<u> </u>	<u> </u>	т;			<u> </u>				
SH.W 1:10C	42B	1	\vdash				+	+		<u> </u>	+	<u> </u>	<u> </u>	<u> </u>	+	+	니		<u> </u>	+	¥ ·	<u> </u>	¥ ·	E'11 E
SH.W 1:10C	43B		\vdash		HIOCI, LI YI Ch	+	+	+	<u> </u>	+			<u> </u>	<u> </u>	+		<u> </u>		+		Li	<u> </u>	Li	Frills on Eg
SH.W 1:10C	HH'	1	\vdash	+					<u> </u>			I	I			 		<u> </u>		 				
SH.W 1:10C	TT'	1	\square	+			<u> </u>	<u> </u>	<u> </u>	<u> </u>				<u> </u>										
SH.W 1:10C	44B	1			+ Wht/Gry Cn		+	+		Oval				+		L				L		Li		
SH.W 1:10C	O'	1		+												I				I				
SH.W 1:10C	1*'	1	ΙΓ	+			1								1	1 -	1	1 -	1	1 -	1	1		

Sample	ID	Cnt	Size	in m	m Color	Cons	istency	r	Color	iy Appe	earance			Edge						Elev	/ation			Notes
<u>,</u>		#	<1 1	-2.2	-7	OPQ	TRL	SHY	DUL	CRC	IRR	FIL	RHZ	ENT	UND	LOB	ERO	FIL	CRL	FLT	RAI	CVX	UMB	
SH W LV A	46D	1	Π		Bg Dkr Cn. Grad out				1											1			T i	
SH.W LXA	40D	1		-	Bg DKi Cii, Glad out	_	Ŧ	Ŧ	-	т					Ŧ		т			-			LI	
SH.W LAA	40D	1			-	-			1											-				
SH.W I:X A	KK2	1		+																				
SH.W I:X A	VV2	3	+	_																				
SH.W 1:X A	RKK'	1			+																			
SH.W 1:X A	PP2'	8			+																			
SH.W 1:X A	44'	5		+	+																			
SH.W 1:X A	FF*'	4		+	+																			
SH.W 1:X A	45B	1			+ Pp Dkr Rgs		Li	+		+				+								+		Sim to 30 but w/ CVX
SH.W 1:X A	45B'	3			+																			
SH.W 1:X A	QQ2'	3			+																			
SH.W 1:X A	40'	2			+																			
SH.W 1:X A	30'	1			+																			
SH.W 1:X A	11'	2			+															1				
SH W 1·X A	HH'	5		+	+																			
SH W 1:X A	RR'	1		<u>.</u>																				
SH W LV A	86'	1		-			-	-		-				-		-		-			-			
SH.W IXA	80 11112	1	<u> </u>	_	+	-																		
SH.W EA A	002	1	+	_		_	-																	
SH.W I:X A	K	1		_	+	_																		
SH.W 1:X A	LL'	1	+	_		_																		
SH.W 1:X A	62'	4			+																			
SH.W 1:X A	26B'	3			+																			
SH.W 1:X A	Y'	1																						
SH.W 1:X B	Y3'	1			+																			
SH.W 1:X B	44A'	1			+																			
SH.W 1:X B	75'	1			+																			
SH.W 1:X B	30'	2			+																			
SH.W 1:X B	45B'	3			+																			
SH.W 1:X B	002'	7			+																			
SH W 1.X B	40'	, 			+																			
SH W 1.Y B	777'	2		-																				
SH.W LAB	62'	5		-	+	-			1											-				
SH.W LAB	02 26D	2		-	. 1	-			1											-				
SH.W LA D	200	2		_	+	-																		
SH.W IX B	29	1		_	+	_	-																	
SH.W I:X B	FF*'	3		+	+	_																		
SH.W I:X B	4	1		+		_																		
SH.W 1:X B	PP2'	6			+																			
SH.W 1:X B	UU2'	1			+		L		L											1				
SH.W 1:X B	11'	2			+															I				
SH.W 1:X B	8'	1			+																			
SH.W 1:X B	V'	1																						
SH.W 1:X B	42A'	1			+																			
SH.W 1:X B	FF'	4			+	1																		
SH.W 1:X B	22B'	5		+	+																			
SH.W 1:X B	44'	6			+	1	1	1	1	1		1	1	1	1	1		1	1	1	1			
SH W 1 X B	RR2'	Ĭ			+	1	1		1			<u> </u>			<u> </u>				<u> </u>	1	1			
SHW I-Y P	HH'	5		-			1	1	1	1		-	1	1	-	1		1	-	1	1			
SHW 1.Y P	S'	2	+	-		+	<u> </u>					<u> </u>		+	<u> </u>	+		+	<u> </u>	+				
SH.W IAB	5 15 A '	4	++	_	+																			
SILW LX C	15A	4	++		+									-		-		-		ł		I		
SH.W I:X C	39	2			+	_	<u> </u>	<u> </u>	I	<u> </u>		I	I	I	<u> </u>	I		I	I	I	<u> </u>			
SH.W 1:X C	QQ2'	7			+	_	I	<u> </u>	I	<u> </u>		I	I		L				I	I	<u> </u>			
SH.W 1:X C	GGG'	1			+							I							I	I				
SH.W 1:X C	44A'	6			+																			
SH.W 1:X C	SS2'	1		T	+			1	1	1				1	1	1		1		1	1			

Sample	ID	Cn	i Size	in mn	n Color	Consi	istency	r	Colon	у Арре	earance			Edge						Elev	ration			Notes
		#	<1 1	-2 2-	7	OPQ	TRL	SHY	DUL	CRC	IRR	FIL	RHZ	ENT	UND	LOB	ERO	FIL	CRL	FLT	RAI	CVX	UMB	
SH.W 1:X C	2'	1		+	-																			
SH.W 1:X C	39'	1		+	-																			
SH.W 1:X C	40'	6		+	-																			
SH.W 1:X C	45B	1		+	-																			
SH.W 1:X C	22B	1		+																				
SH.W 1:X C	HH	4		+	-																			
SH.W 1:X C	FF'	3		+	-																			
SH.W 1:X C	RR2	5		+ +	-																			
SH.W 1:X C	WW2	2' 1		+	-																			
SH.W 1:X C	62'	1		+	-																			
SH.W 1:X C	26B	3		+	-																			
SH.W 1:X C	15A	' 4		+	-																			
SH.W 1:X C	88'	1		+	-																			
SH.W 1:X C	Y'	1																						

Table 2. Total CFU counts in each plate in the different dilution series for each sample.

Selected median values in "bold".

Sample	Count	Sample	Count	Sample	Count
M F 1:100 A	2	SC F 1:100 A	0	SM F 1:100 A	0
M F 1:100 B	0	SC F 1:100 B	0	SM F 1:100 B	0
M F 1:100 C	0	SC F 1:100 C	0	SM F 1:100 C	0
M F 1:10 A	0	SC F 1:10 A	3	SM F 1:10 A	0
M F 1:10 B	0	SC F 1:10 B	7	SM F 1:10 B	1
M F 1:10 C	0	SC F 1:10 C	4	SM F 1:10 C	2
M F 1:X A	<u>2</u>	SC F 1:X A	17	SM F 1:X A	13
M F 1:X B	0	SC F 1:X B	13	SM F 1:X B	<u>11</u>
M F 1:X C	0	SC F 1:X C	16	SM F 1:X C	12
M W 1:100 A	6	SC W 1:100 A	3	SM W 1:100 A	1
M W 1:100 B	3	SC W 1:100 B	3	SM W 1:100 B	6
M W 1:100 C	7	SC W 1:100 C	5	SM W 1:100 C	6
M W 1:10 A	26	SC W 1:10 A	12	SM W 1:10 A	12
M W 1:10 B	22	SC W 1:10 B	15	SM W 1:10 B	25
M W 1:10 C	24	SC W 1:10 C	33	SM W 1:10 C	16
M W 1:X A	130	SC W 1:X A	80	SM W 1:X A	104
M W 1:X B	<u>121</u>	SC W 1:X B	94	SM W 1:X B	137
M W 1:X C	105	SC W 1:X C	84	SM W 1:X C	<u>116</u>
BF F 1:100 A	2	FL F 1:100 A	4	SH F 1:100 A	<u>115</u>
BF F 1:100 B	6	FL F 1:100 B	0	SH F 1:100 B	91
BF F 1:100 C	1	FL F 1:100 C	0	SH F 1:100 C	143
BF F 1:10 A	27	FL F 1:10 A	0	SH F 1:10 A	1000+
BF F 1:10 B	21	FL F 1:10 B	5	SH F 1:10 B	1000+
BF F 1:10 C	14	FL F 1:10 C	1	SH F 1:10 C	1000+
BF F 1:X A	106	FL F 1:X A	12	SH F 1:X A	10000+
BF F 1:X B	74	FL F 1:X B	<u>22</u>	SH F 1:X B	10000+
BF F 1:X C	<u>84</u>	FL F 1:X C	24	SH F 1:X C	10000+
BF W 1:100 A	2	FL W 1:100 A	4	SH W 1:100 A	4
BF W 1:100 B	6	FL W 1:100 B	2	SH W 1:100 B	15
BF W 1:100 C	1	FL W 1:100 C	0	SH W 1:100 C	16
BF W 1:10 A	22	FL W 1:10 A	12	SH W 1:10 A	37
BF W1:10 B	21	FL W 1:10 B	17	SH W 1:10 B	66
BF W 1:10 C	19	FL W 1:10 C	9	SH W 1:10 C	<u>72</u>
BF W 1:X A	74	FL W 1:X A	35	SH W 1:X A	142
BF W 1:X B	51	FL W 1:X B	60	SH W 1:X B	186
BF W 1:X C	<u>60</u>	FL W 1:X C	<u>43</u>	SH W 1:X C	224

Table 2. Continued.

Sample	Count	Sample	Count
M2 F 1:100 A	0	SC2 F 1:100 A	0
M2 F 1:100 B	0	SC2 F 1:100 B	1
M2 F 1:100 C	1	SC2 F 1:100 C	1
M2 F 1:10 A	1	SC2 F 1:10 A	9
M2 F 1:10 B	1	SC2 F 1:10 B	9
M2 F 1:10 C	1	SC2 F 1:10 C	8
M2 F 1:X A	4	SC2 F 1:X A	46
M2 F 1:X B	6	SC2 F 1:X B	55
M2 F 1:X C	<u>5</u>	SC2 F 1:X C	<u>42</u>
M2 W 1:100 A	12	SC2 W 1:100 A	5
M2 W 1:100 B	13	SC2 W 1:100 B	3
M2 W 1:100 C	24	SC2 W 1:100 C	0
M2 W 1:10 A	121	SC2 W 1:10 A	30
M2 W 1:10 B	<u>110</u>	SC2 W 1:10 B	17
M2 W 1:10 C	83	SC2 W 1:10 C	17
M2 W 1:X A	326	SC2 W 1:X A	114
M2 W 1:X B	306	SC2 W 1:X B	98
M2 W 1:X C	288	SC2 W 1:X C	<u>110</u>
BF2 F 1:100 A	13*	FL2 F 1:100 A	19
BF2 F 1:100 B	9	FL2 F 1:100 B	23
BF2 F 1:100 C	8*	FL2 F 1:100 C	10
BF2 F 1:10 A	<u>59</u>	FL2 F 1:10 A	123
BF2 F 1:10 B	71	FL2 F 1:10 B	<u>107</u>
BF2 F 1:10 C	58	FL2 F 1:10 C	90
BF2 F 1:X A	328	FL2 F 1:X A	448
BF2 F 1:X B	287	FL2 F 1:X B	522
BF2 F 1:X C	353	FL2 F 1:X C	337
BF2 W 1:100 A	15	FL2 W 1:100 A	5
BF2 W 1:100 B	21	FL2 W 1:100 B	4
BF2 W 1:100 C	21	FL2 W 1:100 C	5
BF2 W 1:10 A	56	FL2 W 1:10 A	27
BF2 W 1:10 B	68	FL2 W 1:10 B	21
BF2 W 1:10 C	<u>62</u>	FL2 W 1:10 C	<u>23</u>
BF2 W 1:X A	281	FL2 W 1:X A	333
BF2 W 1:X B	232	FL2 W 1:X B	312
BF2 W 1:X C	286	FL2 W 1:X C	299

* 13 and 8 colonies within "normal" recordable size ranges occurring with pin-point colonies that were too numerous to count.

Table 3. Presence or absence of unique CFU types within each sample. Abbreviations: presence indicated by (1),

absolute by (0), et o species type (5pm), lower eon type corresponds to the designation in (Appendix 1a

		~	~ ~ ~	~ ~	~ .	~ ~		~ -			
	CFU Type	Sp 1	SP 2	Sp 3	Sp 4	Sp 5	Sp 6	Sp 7	Sp 8	Sp 9	Sp 10
Sample Origin		А	В	С	D	B*	50	51	52	Κ	54
M F		1	1	1	1	0	0	0	0	0	0
MW		0	0	0	0	1	1	1	1	1	1
BF F		0	0	0	0	0	0	0	0	1	0
BF W		0	0	0	0	0	0	0	0	0	0
SC F		0	0	0	0	0	0	0	0	0	0
SC W		0	0	0	0	0	0	0	0	0	0
FL F		0	0	0	0	0	0	1	0	0	0
FL W		0	0	0	0	0	0	0	0	0	0
SM F		0	0	0	0	0	0	0	0	0	0
SM W		0	0	0	0	0	0	0	0	0	0
SH F		0	0	0	0	0	0	0	0	0	0
SH W		0	0	0	0	0	0	0	0	1	0

	Sp 11	Sp 12	Sp 13	Sp 14	Sp 15	Sp 16	Sp 17	Sp 18	Sp 19	Sp 20	Sp 21
Sample Origin	55	56	57	58	59	60	TTT*	65	67	68	69
MF	0	0	0	0	0	0	0	0	0	0	0
MW	1	1	1	1	1	1	1	1	1	1	1
BF F	0	0	1	0	0	0	0	0	0	0	0
BF W	0	0	0	0	0	0	0	0	0	0	0
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	0	0	0	0	0	0	0	0	0	0	0
FL F	0	0	0	0	0	0	0	0	0	0	0
FL W	0	0	0	0	0	0	0	0	0	0	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	0	0	1	0	0	0	0	0
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	0	0	0	0	0	0	0	0	0	0

	Sp 22	Sp 23	Sp 24	Sp 25	Sp 26	Sp 27	Sp 28	Sp 29	Sp 30	Sp 31	Sp 32
Sample Origin	70	71	72	73	74	75	76	77	78	79	81
MF	0	0	0	0	0	0	0	0	0	0	0
MW	1	1	1	1	1	1	1	1	1	1	1
BF F	0	0	0	0	0	0	0	0	0	0	0
BF W	0	0	0	0	0	0	0	0	0	0	0
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	0	0	0	0	0	0	0	0	0	0	0
FL F	0	0	0	0	0	0	0	0	0	0	0
FL W	0	0	0	0	0	0	0	0	0	0	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	0	0	0	0	0	0	0	0
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	0	1	0	0	1	0	0	0	0	0

	Sp 33	Sp 34	Sp 35	Sp 36	Sp 37	Sp 38	Sp 39	Sp 40	Sp 41	Sp 42	Sp 43
Sample Origin	82	83	84	85	86	87	88	89	90	91	Е
M F	0	0	0	0	0	0	0	0	0	0	0
MW	1	1	1	1	1	1	1	1	1	1	0
BF F	0	0	0	0	0	0	0	0	0	0	1
BF W	0	0	0	0	0	0	0	0	0	0	0
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	0	0	0	0	1	0	1	0	0	0	0
FL F	0	0	0	0	0	0	1	0	0	0	0
FL W	0	0	0	0	0	0	0	0	0	0	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	0	1	0	0	0	0	0	0
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	0	0	0	1	0	1	0	0	0	0

	Sp 44	Sp 45	Sp 46	Sp 47	Sp 48	Sp 49	Sp 50	Sp 51	Sp 52	Sp 53	Sp 54
Sample Origin	F	L	M	N	0	P	Q	G	H	I	J
M F	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	1	0	0	1	0	1	0
BF F	1	1	1	1	1	1	1	1	1	1	1
BF W	0	0	1	0	1	0	0	1	0	0	0
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	0	0	0	0	1	0	1	0	0	0	0
FL F	0	0	0	0	0	0	0	0	0	1	0
FL W	0	0	0	0	0	0	0	0	0	1	1
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	0	0	0	0	1	0	1	1
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	0	0	0	1	0	0	1	0	1	1

	Sp 55	Sp 56	Sp 57	Sp 58	Sp 59	Sp 60	Sp 61	Sp 62	Sp 63	Sp 64	Sp 65
Sample Origin	K	II	JJ	KK	LL	MM	NN	00	PP	QQ	S
M F	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	1	0	0	0	0	0	0	0
BF F	1	1	1	1	1	1	1	1	1	1	1
BF W	0	0	0	0	1	0	0	1	0	0	0
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	0	0	0	0	1	0	0	0	0	0	0
FL F	0	0	0	1	1	0	0	0	0	0	0
FL W	0	0	0	1	0	0	0	0	0	0	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	0	0	0	0	0	0	0	0
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	0	0	0	1	0	0	1	0	0	1

	Sp 66	Sp 67	Sp 68	Sp 69	Sp 70	Sp 71	Sp 72	Sp 73	Sp 74	Sp 75	Sp 76
Sample Origin	Т	U	V	W	X	Y	Z	AA	BB	CC	DD
MF	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	1	1	0	0	0	0	0	1
BF F	1	1	1	1	1	1	1	1	1	1	1
BF W	0	0	0	1	1	0	0	0	0	0	1
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	0	0	1	1	0	0	0	0	0	0	0
FL F	0	0	0	1	1	0	0	0	0	0	1
FL W	0	0	0	0	1	0	0	0	0	0	1
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	1	0	0	0	0	0	0	0
SH F	0	0	0	0	0	0	0	0	0	0	1
SH W	0	0	1	1	1	1	0	0	0	0	1

	Sp 77	Sp 78	Sp 79	Sp 80	Sp 81	Sp 82	Sp 83	Sp 84	Sp 85	Sp 86	Sp 87
Sample Origin	EE	FF	GG	HH	SS	UUU	VVV	WWW	XXX	20	40
MF	0	0	0	0	0	0	0	0	0	0	0
MW	1	1	0	1	0	0	0	1	0	1	1
BF F	1	1	1	1	1	1	1	1	1	1	1
BF W	1	1	0	1	1	1	0	1	1	1	1
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	0	0	0	0	0	0	0	0	1	1	1
FL F	0	1	0	0	0	0	0	0	0	1	0
FL W	1	1	0	1	0	0	0	0	0	1	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	1	1	0	0	0	1	0	1
SH F	0	0	0	1	0	0	0	0	0	0	0
SH W	1	1	0	1	1	0	0	0	1	1	1

	Sp 88	Sp 89	Sp 90	Sp 91	Sp 92	Sp 93	Sp 94	Sp 95	Sp 96	Sp 97	Sp 98
Sample Origin	ZZZ	57	63	64	AAA	BBB	CCC	DDD	EEE	FFF	GGG
MF	0	0	0	0	0	0	0	0	0	0	0
MW	1	0	1	1	1	1	1	1	1	0	1
BF F	1	1	1	1	1	1	1	1	1	1	1
BF W	0	0	0	0	1	1	1	1	0	0	1
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	0	0	1	0	0	0	0	0	1	0	0
FL F	0	0	1	1	0	0	0	0	0	0	0
FL W	0	0	1	0	0	0	0	0	0	0	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	1	0	0	0	0	0	0	1
SH F	0	0	0	0	0	0	1	0	0	0	0
SH W	1	0	0	0	0	0	0	0	0	0	1

	Sp 99	Sp 100	Sp 101	Sp 102	Sp 103	Sp 104	Sp 105	Sp 106	Sp 107	Sp 108	Sp 109
Sample Origin	HHH	III	JJJ	KKK	LLL	MMM	NNN	000	PPP	QQQ	RRR
M F	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	0	0	1	0	0	0	0
BF F	1	1	1	1	1	1	1	1	1	1	1
BF W	0	0	0	0	0	0	1	0	0	0	0
SC F	0	0	0	0	1	0	0	0	0	0	0
SC W	0	0	0	0	1	0	0	0	0	0	0
FL F	0	0	0	0	1	0	0	0	0	0	0
FL W	0	0	0	0	0	0	0	0	0	0	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	0	0	0	0	0	0	0	0
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	0	0	0	0	0	0	0	0	0	1

	Sp 110	Sp 111	Sp 112	Sp 113	Sp 114	Sp 115	Sp 116	Sp 117	Sp 118	Sp 119	Sp 120
Sample Origin	SSS	TTT	RR	TT	UU	VV	WW	XX	YY	ZZ	7
MF	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	1	0	0	0	0	1	1
BF F	1	1	1	1	1	1	1	1	1	1	1
BF W	0	0	1	1	0	1	0	1	0	1	1
SC F	0	0	0	0	0	0	0	0	0	1	0
SC W	0	0	0	0	0	0	0	0	0	1	1
FL F	0	0	0	1	0	0	0	0	0	1	0
FL W	0	0	1	0	0	0	1	0	1	1	1
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	1	0	1	1	0	0	0	0
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	0	1	1	0	1	0	0	1	0	0

	Sp 121	Sp 122	Sp 123	Sp 124	Sp 125	Sp 126	Sp 127	Sp 128	Sp 129	Sp 130	Sp 131
Sample Origin	YYY	46	61	1	2	3	4	5	6	AAA*	8
MF	0	0	0	0	0	0	0	0	0	0	0
MW	1	0	1	0	0	0	0	0	0	0	0
BF F	1	1	1	0	0	0	0	0	0	0	0
BF W	1	1	0	1	1	1	1	1	1	1	1
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	1	0	0	0	0	0	0	0	0	0	0
FL F	0	0	0	0	0	0	0	0	0	0	0
FL W	0	0	0	0	0	0	0	0	0	0	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	0	0	0	0	0	0	0	0
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	0	0	0	1	0	1	0	0	0	1

	Sp 132	Sp 133	Sp 134	Sp 135	Sp 136	Sp 137	Sp 138	Sp 139	Sp 140	Sp 141	Sp 142
Sample Origin	9	10	11	12	13	14	15	16	17	18	19
MF	0	0	0	0	0	0	0	0	0	0	0
MW	0	1	1	0	1	1	1	0	0	0	0
BF F	0	0	0	0	0	0	0	0	0	0	0
BF W	1	1	1	1	1	1	1	1	1	1	1
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	0	0	0	0	1	0	0	0	0	0	0
FL F	0	0	1	0	1	0	0	0	0	0	0
FL W	0	0	1	0	1	0	0	0	0	0	1
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	1	0	0	0	0	0	0	0	1
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	0	1	0	0	1	0	0	0	1	1

	Sp 143	Sp 144	Sp 145	Sp 146	Sp 147	Sp 148	Sp 149	Sp 150	Sp 151	Sp 152	Sp 153
Sample Origin	21	22	23	24	27	29	25	26	92	1*	62
MF	0	0	0	0	0	0	0	0	0	0	0
MW	0	1	0	0	1	1	1	0	0	1	1
BF F	0	0	0	0	0	0	0	0	0	0	0
BF W	1	1	1	1	1	1	1	1	1	1	1
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	0	0	0	0	0	0	1	0	0	0	0
FL F	0	0	0	0	0	0	0	0	1	0	0
FL W	0	0	0	0	0	0	0	0	0	0	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	0	0	0	0	0	0	0	1
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	0	1	0	0	1	0	0	0	1	1

	Sp 154	Sp 155	Sp 156	Sp 157	Sp 158	Sp 159	Sp 160	Sp 161	Sp 162	Sp 163	Sp 164
Sample Origin	FF*	30	32	33	34	35	36	37	39	41	41*
MF	0	0	0	0	0	0	0	0	0	0	0
MW	1	1	1	0	1	0	0	0	1	0	0
BF F	0	0	0	0	0	0	0	0	0	0	0
BF W	1	1	1	1	1	1	1	1	1	1	1
SC F	1	0	0	0	0	0	0	0	0	0	0
SC W	1	0	0	0	0	1	0	0	0	0	1
FL F	1	0	0	0	0	1	0	0	1	0	0
FL W	1	1	0	0	1	1	0	1	0	0	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	1	1	0	0	1	1	0	0	0	0	0
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	1	1	0	0	0	1	0	0	1	0	0

	Sp 165	Sp 166	Sp 167	Sp 168	Sp 169	Sp 170	Sp 171	Sp 172	Sp 173	Sp 174	Sp 175
Sample Origin	42	43	44	45	48	7*	47	92	49	A3	B3
MF	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	0	1	1	0	0	0	0
BF F	0	0	0	0	0	1	0	0	0	0	0
BF W	1	1	1	1	1	1	1	1	1	0	0
SC F	0	0	0	0	0	0	0	0	0	1	1
SC W	0	0	0	0	0	0	0	0	0	0	0
FL F	0	0	0	0	0	0	0	0	0	0	0
FL W	1	0	0	0	0	0	0	0	0	0	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	0	0	1	0	0	0	0	0
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	1	0	1	1	0	0	0	0	0	0	0

	Sp 176	Sp 177	Sp 178	Sp 179	Sp 180	Sp 181	Sp 182	Sp 183	Sp 184	Sp 185	Sp 186
Sample Origin	C3	D3	E3	F3	G3	Н3	I3	J3	K3	L3	M3
MF	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	0	0	0	0	0	0	0
BF F	0	0	0	0	0	0	0	0	0	0	0
BF W	0	0	0	0	0	0	0	0	0	0	0
SC F	1	1	1	1	1	1	1	1	1	1	1
SC W	0	0	0	0	0	0	0	1	0	0	0
FL F	0	0	0	0	0	0	0	0	0	0	0
FL W	0	0	0	0	0	0	0	0	0	0	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	0	0	0	0	1	0	0	0
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	0	0	0	0	0	0	0	0	0	0

	Sp 187	Sp 188	Sp 189	Sp 190	Sp 191	Sp 192	Sp 193	Sp 194	Sp 195	Sp 196	Sp 197
Sample Origin	N3	53	03	P3	Q3	J3*	R3	T3	86*	S3	25A
MF	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	0	0	0	0	0	0	0
BF F	0	0	0	0	0	0	0	0	0	0	0
BF W	0	0	0	0	0	0	0	0	0	0	0
SC F	1	1	1	1	1	1	1	1	1	1	0
SC W	0	0	0	0	0	0	0	0	0	0	1
FL F	0	0	0	0	0	0	0	0	0	0	0
FL W	0	0	0	0	0	0	0	0	0	0	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	0	0	0	0	0	0	0	0
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	0	0	0	0	0	0	0	0	0	0

	Sp 198	Sp 199	Sp 200	Sp 201	Sp 202	Sp 203	Sp 204	Sp 205	Sp 206	Sp 207	Sp 208
Sample Origin	26A	27A	28A	29A	30A	31A	32A	33A	34A	35A	36A
MF	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	0	0	0	0	0	0	0
BF F	0	0	0	0	0	0	0	0	0	0	0
BF W	0	0	0	0	0	0	0	0	0	0	0
SC F	0	1	0	0	0	1	0	0	0	0	0
SC W	1	1	1	1	1	1	1	1	1	1	1
FL F	0	0	0	0	0	0	0	0	0	0	0
FL W	0	0	0	0	0	0	0	0	0	0	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	0	0	0	0	0	0	0	0
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	0	0	0	0	0	0	0	0	0	0

	S= 200	S= 210	S= 211	S= 212	S= 212	S= 214	S= 215	S= 216	S= 217	C= 219	S= 210
	Sp 209	Sp 210	Sp 211	Sp 212	Sp 215	Sp 214	Sp 215	Sp 210	Sp 217	Sp 218	Sp 219
Sample Origin	37A	38A	39A	40A	41A	42A	43A	44A	45A	46A	47A
M F	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	0	0	0	0	0	0	0
BF F	0	0	0	0	0	0	0	0	0	0	0
BF W	0	0	0	0	0	0	0	0	0	0	0
SC F	0	0	0	0	0	1	0	0	0	0	0
SC W	1	1	1	1	1	1	1	1	1	1	1
FL F	0	0	0	0	0	0	0	0	0	0	0
FL W	0	0	0	0	0	1	0	0	0	1	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	0	0	1	0	0	0	0	0
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	0	0	0	0	1	0	1	0	0	0

	Sp 220	Sp 221	Sp 222	Sp 223	Sp 224	Sp 225	Sp 226	Sp 227	Sp 228	Sp 229	Sp 230
Sample Origin	48A	49A	51A	U3	V3	W3	X3	Y3	Z3	AA3	BB3
MF	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	0	0	0	0	0	1	0
BF F	0	0	0	0	0	0	0	0	0	0	0
BF W	0	0	0	0	0	0	0	0	0	0	0
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	1	1	1	0	0	0	0	0	0	0	0
FL F	0	0	0	1	1	1	1	1	1	1	1
FL W	1	0	0	1	0	0	0	0	1	1	1
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	0	0	0	0	0	0	0	1
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	0	0	0	0	0	0	1	0	0	0

	Sn 231	Sn 232	Sn 233	Sn 234	Sn 235	Sn 236	Sn 237	Sn 238	Sn 239	Sn 240	Sp 241
Sample Origin	CC3	DD3	EE3	FF3	GG3	НН3	II3	JJ3	KK3	LL3	MM3
MF	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	0	0	0	0	0	0	0
BF F	0	0	0	0	0	0	0	0	0	0	0
BF W	0	0	0	0	0	0	0	0	0	0	0
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	0	0	1	0	0	1	0	0	1	0	0
FL F	1	1	1	1	1	1	1	1	1	1	1
FL W	0	0	1	0	1	0	0	0	0	1	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	0	1	0	0	0	0	0	0
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	0	0	0	0	0	0	0	0	0	0

	Sp 242	Sp 243	Sp 244	Sp 245	Sp 246	Sp 247	Sp 248	Sp 249	Sp 250	Sp 251	Sp 252
Sample Origin	NN3	003	PP3	QQ3	RR3	SS3	TT3	UU3	VV3	WW3	2A
MF	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	1	0	0	0	0	0	0
BF F	0	0	0	0	0	0	0	0	0	0	0
BF W	0	0	0	0	0	0	0	0	0	0	0
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	1	0	0	0	0	0	0	1	0	0	0
FL F	1	1	1	1	1	1	1	1	1	1	0
FL W	0	0	0	1	1	1	0	0	0	0	1
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	0	0	0	0	0	0	0	0
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	1	0	0	1	0	0	0	0	0	0

	Sp 253	Sp 254	Sp 255	Sp 256	Sp 257	Sp 258	Sp 259	Sp 260	Sp 261	Sp 262	Sp 263
Sample Origin	3A	9A	10A	11A	12A	13A	14A	15A	16A	17A	18A
MF	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	0	0	0	0	0	0	0
BF F	0	0	0	0	0	0	0	0	0	0	0
BF W	0	0	0	0	0	0	0	0	0	0	0
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	0	1	0	0	0	0	0	0	0	0	0
FL F	0	0	0	0	0	0	0	0	0	0	0
FL W	1	1	1	1	1	1	1	1	1	1	1
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	1	0	1	0	0	0	1	0	0
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	0	0	0	0	0	0	1	0	0	0

	Sp 264	Sp 265	Sp 266	Sp 267	Sp 268	Sp 269	Sp 270	Sp 271	Sp 272	Sp 273	Sp 274
Sample Origin	20A	21A	22A	23A	24A	3A*	4A	5A	6A	8A	A2
M F	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	0	0	0	0	0	0	0
BF F	0	0	0	0	0	0	0	0	0	0	0
BF W	0	0	0	0	0	0	0	0	0	0	0
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	0	1	0	0	0	0	0	0	0	0	0
FL F	0	0	0	0	0	0	0	0	0	0	0
FL W	1	1	1	1	1	1	1	1	1	1	0
SM F	0	0	0	0	0	0	0	0	0	0	1
SM W	0	0	0	0	0	0	0	0	0	0	0
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	0	0	0	1	0	0	0	0	1	0
	Sp 275	Sp 276	Sp 277	Sp 278	Sp 279	Sp 280	Sp 281	Sp 282	Sp 283	Sp 284	Sp 285
---------------	--------	--------	--------	--------	--------	--------	--------	--------	--------	--------	--------
Sample Origin	B2	C2	D2	E2	F2	G2	H2	I2	J2	H2	K2
MF	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	0	0	0	0	0	0	0
BF F	0	0	0	0	0	0	0	0	0	0	0
BF W	0	0	0	0	0	0	0	0	0	0	0
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	0	0	0	0	0	0	0	0	0	0	0
FL F	0	0	0	0	0	0	0	0	0	0	0
FL W	0	0	0	1	0	0	0	0	0	0	0
SM F	1	1	1	1	1	1	1	1	1	1	1
SM W	0	0	0	0	0	0	0	0	0	0	0
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	0	0	0	0	0	0	0	0	0	0

	Sp 286	Sp 287	Sp 288	Sp 289	Sp 290	Sp 291	Sp 292	Sp 293	Sp 294	Sp 295	Sp 296
Sample Origin	L2	M2	N2	02	P2	Q2	R2	S2	<u>Z2</u>	T2	U2
MF	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	0	0	0	0	0	0	0
BF F	0	0	0	0	0	0	0	0	0	0	0
BF W	0	0	0	0	0	0	0	0	0	0	0
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	0	0	0	0	0	0	0	0	0	0	0
FL F	0	0	0	0	0	0	0	0	0	0	0
FL W	0	0	0	0	0	0	0	0	0	0	1
SM F	1	1	0	0	0	0	0	0	0	0	0
SM W	0	0	1	1	1	1	1	1	1	1	1
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	0	0	1	0	0	0	0	0	0	1

	Sp 297	Sp 298	Sp 299	Sp 300	Sp 301	Sp 302	Sp 303	Sp 304	Sp 305	Sp 306	Sp 307
Sample Origin	V2	W2	X2	Y2	AA2	BB2	CC2	DD2	EE2	FF2	GG2
MF	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	0	0	0	0	0	0	0
BF F	1	0	0	0	0	0	0	0	0	0	0
BF W	0	0	0	0	0	0	0	0	0	0	0
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	0	0	0	0	0	0	0	0	0	0	0
FL F	0	0	0	0	0	0	0	0	0	0	0
FL W	0	0	0	0	0	0	0	0	1	0	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	1	1	1	1	1	1	1	1	1	1	1
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	0	0	0	0	0	0	0	0	0	0	0

	Sp 308	Sp 309	Sp 310	Sp 311	Sp 312	Sp 313	Sp 314	Sp 315	Sp 316	Sp 317	Sp 318
Sample Origin	HH2	II2	JJ2	KK2	LL2	MM2	NN2	002	PP2	QQ2	RR2
M F	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	0	0	0	0	0	0	0
BF F	0	0	0	0	0	0	0	0	0	0	0
BF W	0	0	0	0	0	0	0	0	0	0	0
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	0	0	0	1	0	0	0	0	1	0	0
FL F	0	0	0	0	0	0	0	0	0	0	0
FL W	0	0	0	0	0	0	0	0	0	0	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	1	1	1	1	1	1	1	1	1	1	1
SH F	0	0	0	0	0	0	0	0	1	1	1
SH W	0	0	0	0	0	0	0	0	1	1	1

	Sp 319	Sp 320	Sp 321	Sp 322	Sp 323	Sp 324	Sp 325	Sp 326	Sp 327	Sp 328	Sp 329
Sample Origin	SS2	TT2	UU2	VV2	WW2	DD2	XX2	YY2	ZZ2	1B	2B
MF	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	0	0	0	0	0	0	0
BF F	0	0	0	0	0	0	0	0	0	0	0
BF W	0	0	0	0	0	0	0	0	0	0	0
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	0	0	0	0	0	0	0	0	0	0	0
FL F	0	0	0	0	0	0	0	0	0	0	0
FL W	0	0	0	0	0	0	0	0	0	0	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	1	0	0	1	0	0	0	0	0	0
SH F	1	1	1	1	1	1	1	1	1	0	0
SH W	1	0	1	1	1	0	0	1	0	1	1

	Sp 330	Sp 331	Sp 332	Sp 333	Sp 334	Sp 335	Sp 336	Sp 337	Sp 338	Sp 339	Sp 340
Sample Origin	3B	4B	5B	6B	7B	8B	9B	10B	11B	12B	13B
M F	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	0	0	0	0	0	0	0
BF F	0	0	0	0	0	0	0	0	0	0	0
BF W	0	0	0	0	0	0	0	0	0	0	0
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	0	0	0	0	0	0	0	0	0	0	0
FL F	0	0	0	0	0	0	0	0	0	0	0
FL W	0	0	0	0	0	0	0	0	0	0	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	0	0	0	0	0	0	0	0
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	1	1	1	1	1	1	1	1	1	1	1

	Sp 341	Sp 342	Sp 343	Sp 344	Sp 345	Sp 346	Sp 347	Sp 348	Sp 349	Sp 350	Sp 351
Sample Origin	14 B	15B	16B	17B	18B	19B	20B	21B	22B	23B	24B
M F	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	0	0	0	0	1	0	0
BF F	0	0	0	0	0	0	0	0	0	0	0
BF W	0	0	0	0	0	0	0	0	0	0	0
SC F	0	1	0	0	0	0	0	0	0	0	0
SC W	0	0	0	0	0	0	0	0	0	0	0
FL F	0	0	0	0	0	0	0	0	0	0	0
FL W	0	0	0	0	0	0	1	0	1	0	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	1	0	0	0	0	0	1	0	0
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	1	1	1	1	1	1	1	1	1	1	1

	Sp 352	Sp 353	Sp 354	Sp 355	Sp 356	Sp 357	Sp 358	Sp 359	Sp 360	Sp 361	Sp 362
Sample Origin	25B	26B	27B	28B	29B	30B	31B	32B	33B	34B	35B
MF	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	0	0	0	0	0	0	1
BF F	0	0	0	0	0	0	0	0	0	0	0
BF W	0	0	0	0	0	0	0	0	0	0	0
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	0	0	0	0	0	0	0	0	0	0	0
FL F	0	0	0	0	0	0	0	0	0	0	0
FL W	0	0	0	0	0	0	0	0	0	0	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	0	0	0	0	0	0	0	0	0	0	1
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	1	1	1	1	1	1	1	1	1	1	1

	Sp 363	Sp 364	Sp 365	Sp 366	Sp 367	Sp 368	Sp 369	Sp 370	Sp 371	Sp 372	Sp 373
Sample Origin	36B	37B	38B	39B	40B	41B	42B	43B	44B	46B	45B
M F	0	0	0	0	0	0	0	0	0	0	0
MW	0	0	0	0	0	0	0	0	0	0	0
BF F	0	0	0	0	0	0	0	0	0	0	0
BF W	0	0	0	0	0	0	0	0	0	0	0
SC F	0	0	0	0	0	0	0	0	0	0	0
SC W	0	0	0	0	0	0	0	0	1	0	0
FL F	0	0	0	0	0	0	0	0	0	0	0
FL W	0	0	0	0	0	0	0	0	0	0	0
SM F	0	0	0	0	0	0	0	0	0	0	0
SM W	1	0	0	0	0	1	0	0	1	0	1
SH F	0	0	0	0	0	0	0	0	0	0	0
SH W	1	1	1	1	1	1	1	1	1	1	1

Sample Origin	Total CFU Richness	Total shared CFUs
M F	4	0
MW	88	58
BF F	85	52
BF W	81	62
SC F	30	7
SC W	58	38
FL F	50	37
FL W	64	50
SM F	14	1
SM W	70	47
SH F	15	12
SH W	113	76

Figure 1. Photos of selected plate cultures to visually demonstrate growth trend within samples. Only the first replicate (A) of each dilution is depicted.

Mull Creek fish from first sample.



Beechflat Creek fish from first sample.



Figure 1. Continued.

Mull Creek fish from second sample.



Beechflat Creek fish from second sample.



Figure 1. Continued.

Scapecat Creek fish from first sample.



Sawmill Creek fish from first sample.



Figure 1. Continued.

Mull Creek water from first sample.



Beechflat Creek water from first sample.

