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Complete or High-Quality Draft Genome Sequences of Six *Xanthomonas hortorum* Strains Sequenced with Short- and Long-Read Technologies

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Resource Announcements

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ABSTRACT We report the genome sequences of six *Xanthomonas hortorum* species-level clade members, *X. hortorum* pathovars taraxaci, pelargonii, cynarae, and gardneri (complete genome sequences) and *X. hortorum* pathovars carotae and vitians (high-quality draft genome sequences). Both short- and long-read sequencing technologies were used.

The genetic relatedness of the *Xanthomonas hortorum* species-level clade (slc) was established by Parkinson et al. using partial *gyrB* gene sequences (1). The most recently updated taxonomy of the clade (2) includes seven pathovars of *X. hortorum* (cynarae, gardneri, carotae, hederae, pelargonii, taraxaci, and vitians type B) (1–3).

We report here the whole-genome sequences of six *X*. *hortorum* strains (Table 1). The strains were isolated between 1942 and 2008 from a wide range of hosts and in various countries (Table 1). The genome sequences published in this work are either complete genome sequences or high-quality draft genome sequences, and all have five contigs or less (Table 1).

The strains were initially obtained as freeze-dried cultures in glass ampoules from two international strain collections abbreviated in the strain names as CFBP (Collection Française de Bactéries Associées aux Plantes, Beaucouzé, France) and NCPPB (National Collection of Plant Pathogenic Bacteria, York, United Kingdom). After revival on nutrient yeast extract glycerol agar plates (4) for 2 days at 28°C, an isolated colony was streaked onto the same medium and grown in the same manner. The strains were then stored in a -80° C ultrafreezer as glycerol stocks in 50% (vol/vol) nutrient yeast extract glycerol broth (4) until further use. The strains were revived as described above, and a single colony was used as starting material for downstream experiments.

Genomic DNA (gDNA) for Illumina MiSeq short-read sequencing was extracted from cells grown overnight at 28°C in nutrient yeast extract glycerol broth (4) using the NucleoSpin tissue kit (Macherey-Nagel, Düren, Germany) according to the manufacturer's protocol with the following specifications: elution buffer was heated at 70°C before use, and the gDNA was eluted with 60 μ l of this buffer. The quality of the gDNA was checked using a fragment analyzer (Advanced Analytical Technologies, Inc., Ankeny, IA) and quantified using the Quant-iT PicoGreen double-stranded DNA (dsDNA) quantification assay (Thermo Fisher Scientific, Waltham, MA). Library preparation was done using the Nextera XT DNA library prep kit (Illumina, San Diego, CA) following the manufacturer's instructions. Sequencing was performed on a MiSeq Illumina sequencer with 2 × 300-bp paired-end reads using a MiSeq reagent kit version 3 (Illumina) according to the manufacturer's instructions.

Genomic DNA for long-read sequencing was extracted from overnight-grown cells using the Gentra PureGene Yeast/Bact kit protocol (Qiagen, Hilden, Germany). The Citation Dia NC, Rezzonico F, Smits THM, Pothier JF. 2020. Complete or high-quality draft genome sequences of six *Xanthomonas hortorum* strains sequenced with short- and long-read technologies. Microbiol Resour Announc 9:e00828-20. https://doi.org/10.1128/ MRA.00828-20.

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| TABLE | : 1 Genome n | ietrics and a | ccession numl | TABLE 1 Genome metrics and accession numbers of the newly sequenced genomes within the <i>Xanthomonas hortorum</i> species-level clade ^a | ly sequ | renced genc | omes wit | thin the Xa | nthomonas <i>i</i> | hortorum | species- | level clade ^a | | | |
|---|---|---|--|---|-----------------------|---|--|--|---|------------------------------|------------|---|---------------------------|--|--|
| | | | | | | | | Illumina data: | ta: | | Oxford | Oxford Nanopore data: | ata: | | |
| Strain ⁶ | Bacterial species | Origin (yr) | Host | G+C Genome content size (bp) (%) | | Total Genome no. of status genes (N ₅₀ [bp]) | No. of contigs/ no. of plasmids | No. of contigs/ no. of 17/15 plasmids indexes | Avg read Avg Total no. length coverage of reads (bp) (×) | d Avg gth covera) (×) | BC | Total Read Avç no. of length cov reads N ₅₀ (bp) (x) | Avg coverage p) (×) | Avg coverage SRA accession no. (×) (MiSeq/MinlON) | ENA accession no. |
| CFBP 498 | X. hortorum pv. vitians | USA (1949) | Lactuca sp. | 5,678,543 63.23 | 4,976 | 4,976 HQ draft (5,365,193) | 4/3 | N703/S504 | N703/S504 1,112,420 259 | 38 | BC18 | BC18 27,882 9,898 | 58 | ERR4326259, ERR4327117 | LR828257 (chr.), LR828258 (p224), LR828259 (p47), LR828260 (p41) |
| CFBP 2044 | X. hortorum | France (1981) Cynara scolvi | Cynara scolvmus L. | 5,119,234 63.67 | 4,322 | Complete | 2/1 | N705/S517 | N705/S517 2,593,908 277 | 96 | BC16 | BC16 18,849 12,128 | 55 | ERR4326260, ERR4327118 | LR828251 (chr.), LR828252 (p40) |
| CFBP X. 2533 ^p | CFBP X. hortorum pv. New Zealand Pelargonium 2533 ^p pelargonii (1974) peltatum | v. New Zealand (1974) | Pelargonium peltatum | 5,287,542 63.72 | 4,427 | Complete | 3/2 | N702/S517 | N702/5517 3,398,636 261 | 111 | BC15 | BC15 14,695 8,396 | 20 | ERR4326261, ERR4327119 | LR828261 (chr.), LR828262 (p47), I R828263 (n41) |
| CFBP 7900 | CFBP X. hortorum 7900 ^p pv. carotae | USA (2008) | | Daucus carota 5,149,201 63.77 (seed) | 4,274 | HQ draft (2.659.169) | 5/ND | N702/S504 | N702/S504 1,721,852 264 | 65 | BC14 | BC14 7,777 8,728 | 15 | ERR4326262, ERR4327120 | CAJDKC00000000 |
| CFBP 8129 | CFBP X. <i>hortorum</i> pv. Costa Rica 8129 gardneri (1991) | /. Costa Rica (1991) | Solanum lycopersicum | 5,440,942 63.48 | 4,655 | ů | 4/3 | N705/S504 | N705/S504 1,653,590 258 | 27 | BC20 | BC20 45,436 8,073 | 80 | ERR4326263, ERR4327121 | LR828253 (chr.), LR828254 (p211), LR828255 (p46), I R828256 (n26) |
| NCPPB 940 ^p | NCPPB X. hortorum pv. USA (1942) Taraxacum 940 ^p taraxaci kok-sagh) | ۰. USA (1942) | Taraxacum kok-saghyz | 5,029,134 63.83 | 4,316 | 4,316 Complete | 2/1 | N701/S503 | N701/S503 1,197,722 269 | 43 | BC17 | BC17 9,847 7,654 | 18 | ERR4329153, ERR4327122 | LR828264 (chr.), LR828265 (p30) |
| ^{<i>a</i>} BC, bé ^{<i>b</i>} The ci Bacteri | ^a BC, barcode; HO, high-quality; ND, not determined; chr., chromosome. ^b The culture collections providing strains are abbreviated in the strain r Bacteria, York, United Kingdom). Superscript P following a strain name | quality; ND, no providing strai (ingdom). Super | t determined; ch ns are abbreviatu script P followin | BC, barcode; HQ, high-quality; ND, not determined; chr., chromosome. The culture collections providing strains are abbreviated in the strain names as CFBP (Collection Française de Bactérie: Bacteria, York, United Kingdom). Superscript P following a strain name indicates the pathotype strain for the pathovar. | nes as C dicates t | CFBP (Collection the pathotype | n Français strain for 1 | e de Bactérie: the pathovar. | s Associées aux | x Plantes, B | seaucouzé, | France) and N | VCPPB (Nati | ^σ BC, barcode; HQ, high-quality: ND, not determined; chr., chromosome. ^Φ The culture collections providing strains are abbreviated in the strain names as CFBP (Collection Française de Bactéries Associées aux Plantes, Beaucouzé, France) and NCPPB (National Collection of Plant Pathogenic Bacteria, York, United Kingdom). Superscript P following a strain name indicates the pathotype strain for the pathovar. | nt Pathogenic |

gDNA was quantified and its quality checked as described above. Library preparation and sequencing were performed with the ligation sequencing kit (catalog no. SQK-LSK109; Oxford Nanopore Technologies, Oxford, United Kingdom) and run on an R9.4.1 flow cell with a MinION sequencer. The native barcoding expansion kit (catalog no. XP-NBD114) was used for multiplexing. Base calling was performed using Guppy version 3.0.7 in the "accurate" mode implemented in the MinION release 19.06.8.

A hybrid assembly using the MiSeq and MinION reads was conducted with Unicycler version 0.4.7 (5). To check for misalignments, the MiSeq reads were mapped against the Unicycler assemblies using SeqMan Pro version 12.2 (DNAStar, Madison, WI). Contigs were reordered using the Mauve Contig Mover version 2.3.1 (6) when required (7). The scrub and align options of Unicycler were used to detect chimeras and check for high-error regions, respectively. The manual improvement of the genomes was finalized with Bandage version 0.8.1 (8). The genomes were then annotated using Prokka version 1.14.5 (9). Indels were checked using Ideel (10). All tools were run with default parameters unless otherwise specified.

The sizes of the hybrid assemblies ranged from 5,029,134 to 5,678,543 bp, a size range typically found in *Xanthomonas* genomes (Table 1). The G+C contents of the genomes varied from 63.23% to 63.83%, comparable to other *Xanthomonas* genome G+C contents. The sequenced genomes discussed here will be used for further analysis of evolution within the *X. hortorum* slc.

Data availability. The annotated genome sequences of the six *X*. *hortorum* strains have been deposited in ENA under BioProject no. PRJEB38812. The genome and raw read accession numbers for each isolate are shown in Table 1.

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