

1 **Spatial relationships between entomopathogenic nematodes and nematophagous fungi in**  
2 **Florida citrus orchards**

3

4 Ekta Pathak<sup>a,b</sup>, Raquel Campos–Herrera<sup>a,c,d</sup>, Fahiem E. El–Borai<sup>a,e</sup>, Larry W. Duncan<sup>a,\*</sup>

5

6 <sup>a</sup> *Citrus Research and Education Center (CREC), University of Florida (UF), 700 Experiment Station*  
7 *Road, Fl 33850, USA*

8 <sup>b</sup> *Sam Higginbottom Institute of Agriculture, Technology & Sciences, Allahabad, U.P., India*

9 <sup>c</sup> *Instituto de Ciencias Agrarias, CSIC, Serrano 115 Dpdo, Madrid 28006, Spain*

10 <sup>d</sup> *Current address: Centro para os Recursos Biológicos e Alimentos Mediterrânicos (MeditBio), Campus*  
11 *Gambelas FCT Ed. 8 room 2.36.5, Universidade do Algarve, Faro 8005–139 (Portugal)*

12 <sup>e</sup> *Plant Protection Department, Faculty of Agriculture, Zagazig University, Egypt*

13

14 \* Corresponding author: Larry W. Duncan ([lwduncan@ufl.edu](mailto:lwduncan@ufl.edu))

15 *Citrus Research and Education Center (CREC), University of Florida (UF), 700 Experiment Station*  
16 *Road, Fl 33850, USA*

17 Phone: +1 863 9561111; Fax: +1 863 9564631

18 **Abstract**

19 Relationships between entomopathogenic nematodes (EPNs), nematophagous fungi (NF) and soil  
20 physical and chemical properties were studied in a survey of 53 citrus orchards in central ridge and  
21 flatwoods ecoregions of Florida. Seven species of NF associated with nematodes were quantified directly  
22 using a real time qPCR assay. All nematophagous fungi studied except *Arthrobotrys musiformis*  
23 and *Hirsutella rhossiliensis* were frequently detected (24-56%) in both regions. *Paecilomyces*  
24 *lilacinus* and *Gamsylella gephyropagum* were encountered more frequently in the flatwoods ( $P = 0.03$ )  
25 and on the ridge ( $P = 0.02$ ), respectively. Redundancy analysis revealed seven abiotic and biotic factors as  
26 significantly related to the NF occurrence. Multiple regression of fungi on these variables explained 78%,  
27 66%, 48%, 36%, 23% and 4% of the variation in *Catenaria* sp., *A. musiformis*, *A. dactyloides*, *P.*  
28 *lilacinus*, *A. oligospora* and *G. gephyropagum*, respectively. When the data from citrus were pooled with  
29 those reported previously from natural areas and subjected to principle component analysis, the first two  
30 principle components explained 43% of the variation in NF communities. The surveys (citrus vs natural  
31 areas) were discriminated by PC2 ( $P < 0.001$ ) and the ecoregion by PC1 ( $P < 0.002$ ), and all but one NF  
32 species were related ( $P < 0.01$ ) to one or both components. NF communities tended to have more species  
33 and greater diversity in the flatwoods, where EPN richness and diversity were the least. However, the  
34 strength of associations between individual EPN and NF species as measured by SADIE reflected the  
35 associations between each species and ground water depth, suggesting that ecoregion preferences affected  
36 the species associations. Within each ecoregion, significant relationships between the individual NF and  
37 EPN species measured by stepwise regression tended to be positive. The results did not support the  
38 hypothesis that NF modulate the spatial patterns of EPN species between or within these two ecoregions.

39  
40

41 *Key words:* Nematophagous fungi; quantitative real-time PCR; soil ecology; species-specific primers and  
42 probe; entomopathogenic nematodes; multivariate analysis

## 43 1. Introduction

44

45 Entomopathogenic nematodes (EPNs) have been studied for more than a quarter century in  
46 Florida citrus orchards for their potential as components of root weevil IPM programs. Following the  
47 deregistration of soil-applied organochlorine pesticides in the mid-1970s, the invasive pest *Diaprepes*  
48 *abbreviatus* and other root weevils such as *Pachnaeus litus* became the most economically damaging  
49 arthropods in Florida citrus due to a lack of adequate control measures (Dolinski et al., 2012). Disrupting  
50 the weevil's long life cycle, which comprises both arboreal and subterranean stages, is difficult using non-  
51 persistent chemical and biological insecticides. The introduction of the bacterial disease huanglongbing  
52 (HLB) into the citrus industry, first detected in 2005 (Halbert, 2005), intensified the need for root weevil  
53 management because HLB destroys more than half of the fibrous roots on trees before aboveground  
54 symptoms become evident (Johnson et al., 2014). As a consequence, orchards damaged by both weevils  
55 and HLB decline much more rapidly than those affected by just the bacterium or weevil (Duncan, 2014) .

56 The two primary genera of EPNs, *Steinernema* and *Heterorhabditis* (Dillman et al., 2012), are  
57 each represented by several native species in Florida. A survey of orchards (Campos-Herrera et al., 2013)  
58 and another of natural areas in the citrus growing regions of the state (Campos-Herrera et al., 2016),  
59 revealed similar associations between EPN communities and two ecoregions, the “central ridge” running  
60 north to south in the middle of the peninsula and the “flatwoods” regions surrounding the central ridge  
61 and extending to the coasts. Orchards on the higher elevation, well drained sandy soils of the central ridge  
62 were almost always inhabited by both *H. indica* and *S. diaprepesi* and frequently also by *H. zealandica*.  
63 By contrast, EPN communities in orchards on lower elevation, shallow, less well drained soils of the  
64 flatwoods were almost always composed of *H. indica*, but only occasionally in combination with *S.*  
65 *diaprepesi* and/or an undescribed *Steinernema* species in the *S. glaseri* group proposed by Spiridonov et  
66 al. (2004). The population density of *D. abbreviatus* is larger in flatwoods orchards than in those on the  
67 central ridge (Futch et al., 2005), perhaps due in part to lower rates of biological control by EPNs in the  
68 flatwoods (Duncan et al., 2003). These patterns and their relationships suggest that natural control of

69 weevils can be increased by discovering properties of habitats that can be modified in ways that conserve  
70 EPNs and enhance their services.

71 Soil properties that affect soil water potential explained the greatest amount of EPN variation in  
72 each of the two surveys by Campos Herrera et al. (2013, 2016) and results of subsequent experiments  
73 supported the possibility that different EPN spatial patterns in Florida result partly from differential  
74 species adaptation to soil moisture (Duncan et al., 2013; El-Borai et al., 2016). Additionally, natural  
75 enemies such as nematophagous fungi (NF) and micro-arthropods regulate EPN numbers locally (Ekmen  
76 et al., 2010; Hodson et al., 2012) and could also modulate EPN spatial patterns in the landscape if their  
77 effects differ by region. NF comprise a guild of diverse fungi that include saprophyte/facultative  
78 predators (most notably the ‘trapping fungi’) and obligate predators of nematodes and other soil fauna  
79 such as Chytridiomycete species that produce infective zoospores. Because many members of the guild  
80 can function at different trophic levels, conventional methods of quantifying these fungi in soil samples  
81 do not discriminate propagules derived from nematode prey from those derived from other nutrient  
82 sources. The development of molecular tools that can quantify NF associations with nematodes in nature,  
83 without the artifacts introduced by culturing methods, provides a means to study the importance of habitat  
84 properties on those associations (Pathak et al., 2012). Campos-Herrera et al. (2016) detected primarily  
85 positive associations between NF and either free-living bacterivorous nematodes or EPNs from nematode  
86 samples taken at 91 natural (non-agricultural) sites. Those relationships suggested that r-selected, rapidly  
87 increasing nematode species provided local resources for NF populations. The relative lack of negative  
88 associations might mean that NF predation of nematodes did not vary much across regions and did not  
89 affect EPN spatial patterns at the landscape scale. Jaffee et al., (1996) similarly concluded that NF were  
90 unrelated to the spatial pattern of *Heterorhabditus marelatus* that regulate subterranean ghost moth larvae  
91 and the damage they do to the roots of bush lupines in coastal California. Relationships between EPNs  
92 and NF in different habitats were also reported from field plots of citrus trees planted into holes filled  
93 either with sand or the native loam soil, thus creating well-drained or poorly drained conditions (Duncan  
94 et al., 2013). EPNs that were introduced into the plots persisted and killed sentinel weevil larvae at higher

95 rates in the sand than the loam during four years. However, real time PCR assays of DNA from  
96 nematodes isolated from soil samples showed that the NF species *Purpureocillium lilacinus* and  
97 *Catenaria* sp. consistently infected nematodes at higher rates in sand than loam, whereas no NF species  
98 appeared to be favored by conditions in the loam soil. Consequently, there is little evidence to date that  
99 the spatial patterns of EPN species at a broad, landscape scale are modulated by variation in susceptibility  
100 to NF across habitats.

101 The DNA of seven NF species recovered from samples of nematodes was identified and  
102 quantified, but not reported, in a survey of EPNs in Florida citrus orchards (Campos-Herrera et al., 2013).  
103 The agricultural inputs in those orchards created soil conditions very different than those of natural areas  
104 in the same region reported by Campos-Herrera et al. (2016). Here we report the relationships between  
105 NF and soil properties in the citrus orchards and compare them with those detected in the natural areas.  
106 Results from both surveys were also combined to explore spatial relationships between NF, member of  
107 the EPNs soil food web and abiotic factors. Our hypotheses were that 1) the occurrence frequency of  
108 some NF species would differ between orchards and natural areas due to differences in soil chemistry and  
109 management, 2) some spatial associations between species of EPNs and NF would be due to similar  
110 habitat adaptations and therefore, 3) associations between NF and EPNs within a habitat might provide  
111 better insights into the role played by NF in modulating EPN occurrence.

112

## 113 **2. Materials and methods**

114

### 115 *2.1. General survey design, sampling and soil processing*

116

117 The survey area extended nearly 200 km east to west (from the gulf coast to the Atlantic coast)  
118 and more than 300 km north to south (from Umatilla to Immokalee). The orchards were selected on the  
119 basis that they were known to be infested by *Diaprepes abbreviatus* and that representative numbers were  
120 located in each of the two ecoregions (23 sites in the central ridge and 30 in the flatwoods) in which

121 Florida's orchards occur. Sampling and sample processing methods are described in detail by Campos-  
122 Herrera et al. (2013). Briefly, two samples (30 cores, 2.5-cm diam. x 30 cm deep per sample) were  
123 obtained systematically at each 2-3 ha site during summer-early autumn 2009-2010. Samples were mixed  
124 and nematodes extracted (Jenkins, 1964) from 2, 500-cm<sup>3</sup> subsamples (i.e., 2 liters of soil per site). DNA  
125 was extracted using the UltraClean Soil DNA Extraction Kit (MoBio), quality was ensured (Nanodrop  
126 system 1000 v.3.3.0, Thermo Scientific, Wilmington, DE, USA), and the quantity adjusted for further  
127 molecular analysis. Following previous optimized protocols, seven species of NF were identified and  
128 quantified using real-time qPCR approaches (Atkins et al., 2005; Zhang et al., 2006; Pathak et al., 2012).  
129 Selected organisms previously reported in this survey by Campos-Herrera et al. (2013) were also  
130 combined in the evaluation, which included the species specific qPCR quantification of EPNs (13  
131 species), and several other nematodes, bacteria and oomycete phytopathogens, all of which are known to  
132 interact with EPNs (Huang et al., 2010; Campos-Herrera et al., 2011a, 2011b, 2012; 2013)  
133 (Supplementary data 1). In all the cases, primers and probes were synthesized by Integrated DNA  
134 Technologies Inc. (IDT, San Diego, CA), with all TaqMan® PCR probes labelled with the fluorogenic  
135 reporter dye FAM, the quencher Iowa Black™ FG and the ZEN molecule in the middle of the probe. Real  
136 time qPCR runs were performed in ABI Prism 7000 (Applied Biosystem), in optical-well reaction plates  
137 (USA Scientific, Orlando, FL, USA) with a final volume of 20 µL, and employing those reactive  
138 materials and primers/probe concentrations, annealing temperature and number of cycles described  
139 previously (Supplementary data 1). For each of the species, the identification and quantification was  
140 performed using positive (standard curve), negative (sterile de-ionized water) controls and the  
141 corresponding unknowns, with two technical replicates in each run.

142 Soil characteristics were described for the citrus survey (Campos-Herrera et al., 2013) and the  
143 natural areas study (Campos-Herrera et al., 2016). Briefly, remaining soil was air dried and sand fractions  
144 were determined by wet sieving (53 mm) followed by re-drying and processing on stacked sieves on a  
145 mechanical shaker (W.S. Tyler Co., Mentor, OH). Soil pH, organic matter (OM), percentage sand, silt and  
146 clay, electric conductivity (EC), cation-exchange capacity (CEC) and soil nutrient elements (P, K, Mg,

147 Ca, and B) were measured in a commercial laboratory (Waters Agricultural Laboratories, Camilla, GA).  
148 In addition to these studies, common for both surveys (citrus and natural areas), additional measurements  
149 were performed in the citrus surveys as follow: soil field capacity (FC) at pF 2.7, wilting point (WP) at pF  
150 4.2 and water holding capacity by Richards' method (Duchaufour, 1975); soil metal content (Cd, Cr, Cu,  
151 Fe, Mn, Ni, Pb, Zn) and others (As and Se) by Optical Emission Spectrometry (IPC) with a Plasma  
152 Spectrometer; and pesticide residues such as organochlorines, BHCs, Cyclodienes and DDT compounds  
153 following ISO 10382 (2002), in the Chromatographic service of the National Museum of Natural  
154 Sciences (Madrid, Spain) (Campos-Herrera et al., 2013).

155

## 156 *2.2. Analysis of fungal distribution in the citrus survey*

157

158 NF DNA quantity was divided by the total DNA in a sample to estimate the infection rate. The  
159 infection rate of each species was then normalized (0-1) to permit comparison of the relative effects of  
160 each species in an NF community (Pathak et al., 2012; Duncan et al., 2013). The occurrence and mean  
161 infection rate of NF was calculated for each eco-region (central ridge and flatwoods). To compare species  
162 abundance, the quantities of NF were square root transformed prior to statistical analyses. The ecological  
163 indices (i) species richness (number of species, S), (ii) the Shannon Wiener diversity index  $H' = -\sum p_i \ln$   
164  $p_i$ , where "p<sub>i</sub>" is the proportion of individuals of the i<sup>th</sup> species and (iii) the evenness, a measure of the  
165 equilibrium of species abundance ( $J' = H'/H_{\max}$ , where  $H_{\max}$  is  $\ln S$ ) were estimated for all samples and  
166 within regions. Data are presented as mean  $\pm$  SEM.

167 Regional differences in species occurrence, abundance, and ecological indices were evaluated by  
168 T-test (20.0 IBM SPSS Inc., Chicago, IL, USA). Redundancy analysis of NF species and selected  
169 organisms and soil properties were performed using the software CANOCO 5 (Ter Braak, 2009; Smilauer  
170 and Lepš, 2014). The biotic and abiotic properties used as explanatory variables were reported by  
171 Campos-Herrera et al. (2013). In particular, those derived from principal component analysis (PCA) as

172 contributing significantly to the total spatial variability (data not shown) together with GWD. The  
173 graphical results of the RDA were presented with bi-plot scaling (CANOCO 5). Stepwise multiple  
174 regression analysis (Minitab 15, State College, PA) of each NF species on the independent variables  
175 found to be significant by RDA was used to assess the relative explanatory contributions of those  
176 variables.

177

### 178 *2.3. Analysis of fungal distribution and relationships in the combined citrus and natural area surveys*

179

180 The biotic and abiotic data from natural areas and citrus orchards reported by Campos-Herrera et  
181 al. (2013, 2016) were combined with the NF data from citrus reported here for more comprehensive  
182 analyses. NF communities were characterized by principal component analysis (PCA) of the square root-  
183 transformed infection rates and subjected to ANOVA (GLM Minitab 15) to assess differences in the first  
184 two principal components between 1) surveys and 2) ecoregions. For comparison, Spatial Analysis by  
185 Distance Indices (SADIE; [http://www.rothamsted.ac.uk/pie/sadie/SADIE-home\\_page\\_1.htm](http://www.rothamsted.ac.uk/pie/sadie/SADIE-home_page_1.htm)) was also  
186 used to assess the degree of association between each NF species and each EPN species as well as NF  
187 species and GWD (Spiridonov et al., 2007). The pooled data from the citrus and natural area surveys were  
188 also subjected to RDA as described previously. Pooled data from within each ecoregion were used to  
189 determine whether NF species occurrence differed (T-test, Minitab 15) in sites with or without a given  
190 EPN species. Stepwise multiple regression of the pooled data within each ecoregion was used to assess  
191 relationships between EPNs and NF.

192

## 193 **3. Results**

194

### 195 *3.1. Analysis of fungal distribution in the citrus survey*

196



197 Few NF species varied in abundance or detection frequency across the two regions. Two  
198 exceptions were *P. lilacinus*, which was detected more frequently and at higher abundance in the  
199 flatwoods and *M. gephyropagum*, which was more frequently detected on the central ridge (Table 1). The  
200 number of species did not differ between the central ridge ( $2.3 \pm 0.24$ ) or the flatwoods ( $2.70 \pm 0.21$ ), but  
201 the species diversity ( $H' = 0.30 \pm 0.073$  vs  $0.56 \pm 0.08$ ;  $t = 2.32$ ,  $P = 0.02$ ) and evenness ( $J' = 0.34 \pm$   
202  $0.066$  vs  $0.55 \pm 0.063$ ;  $t = 2.25$ ,  $P = 0.03$ ) were highest in the flatwoods.

203 Seven environmental variables explained significant NF species variation in the redundancy  
204 analysis (Fig. 1). Stepwise multiple regression of each NF on those seven factors as independent  
205 variables revealed potassium and electrical conductivity to be positively and inversely, respectively,  
206 associated with five of the six species, AWC to be directly related to three species, pH to be positively  
207 related to two species and inversely related to one, boron to be inversely related to two species and  
208 directly related to one and phosphorus to be positively related to two species (Table 2). These variables  
209 explained 78% of the variation in *Catenaria* sp. and 66% of variation in *A. musiformis*, 48% in *A.*  
210 *dactyloides*, 36% in *P. lilacinus*, 23% in *A. oligospora* and 4% in *G. gephyropagum*.

211

### 212 3.2. Analysis of fungal distribution and relationships in the combined citrus and natural area surveys

213

214 Two way analysis of variance of data from citrus orchards and those reported previously from  
215 nearby natural areas (Campos-Herrera et al., 2016) revealed that *A. oligospora*, *A. dactyloides*, *M.*  
216 *gephyropagum* and *Catenaria* sp. were detected more frequently (all species  $P < 0.001$ ) in citrus orchards  
217 than in natural areas (Fig. 2). The pooled data confirmed that *P. lilacinus* and *G. gephyropagum* were  
218 detected more frequently in flatwoods and central ridge ecoregions, respectively. There was no significant  
219 interaction between the botanical habitat (orchard vs natural area) and the ecoregion for detection  
220 frequency of any NF species.

221 The first two principal components of the PCA accounted for 40% of the variation in the NF  
222 species composition associated with nematodes in the two ecoregions of the combined surveys (Fig. 3).

223 The first principal component discriminated the two surveys ( $P < 0.001$ ) along the x-axis with *A.*  
224 *oligospora*, *A. dactyloides*, *G. gephyrophagum*, and *Catenaria* sp., each significantly contributing ( $P <$   
225  $0.001$ ) to the difference. NF communities on the central ridge were distinguished from those in the  
226 flatwoods ( $P = 0.002$ ) by PC2 on the y-axis, with *P. lilacinus*, *A. dactyloides*, *A. musiformis*, and *G.*  
227 *gephyrophagum* (each  $P < 0.001$ ) and *A. oligospora* ( $P = 0.01$ ) being related to the components. There  
228 were no significant interactions between the survey and ecoregion effects on NF community structure.

229 The relative richness, diversity and evenness of the NF and EPN communities in the two  
230 ecoregions differed from each other consistently in both surveys (Fig. 4). NF communities tended to have  
231 more species and greater diversity in the flatwoods, where EPN richness and diversity were the least.

232 A redundancy analysis using the pooled data from both the orchard and natural area survey again  
233 revealed pH, potassium, phosphorus and *S. diaprepesi* to be significant descriptors of the overall variation  
234 in NF species (data not shown). The other significant predictive variables for citrus orchards (electrical  
235 conductivity, soil water holding capacity and boron, Fig. 1) were not measured in the natural area survey  
236 and therefore, stepwise regression using data from both surveys was not conducted.

237 SADIE association indices (non-parametric) between NF species and groundwater depth (GWD)  
238 were positively related to association indices between NF species and *S. diaprepesi* ( $P = 0.01$ ) and *H.*  
239 *zealandica* ( $P < 0.05$ ) and inversely related to those for *H. indica* ( $P = 0.02$ ) and *Steinernema* sp. ( $P =$   
240  $0.03$ ) (Fig. 5). Because *S. diaprepesi* and *H. zealandica* were reported to be directly associated with GWD  
241 and *H. indica* and *Steinernema* sp. inversely related to GWD (Campos-Herrera et al. 2013), the  
242 associations between EPN and NF species may have resulted from varying responses by each species to  
243 GWD which is greater on the central ridge than in the flatwoods. Therefore, we examined relationships  
244 between NF and EPNs separately for each ecoregion. In the flatwoods sites, the detection frequencies of  
245 six out of seven NF species in both the orchards and natural areas were greater in sites where *Steinernema*  
246 spp. was not detected (Fig. 6). There were no apparent differences in NF and EPN occurrence for any  
247 other EPN species in either region (not shown for central ridge). For central ridge sites, stepwise multiple  
248 regression of each EPN species against NF and all other measured variables for those EPN species

249 commonly detected on the central ridge (*S. diaprepesi*, *H. indica* and *H. zealandica*) revealed significant  
250 positive relationships between EPNs and citrus sites, potassium, phosphorus, *Acrobeloides*-group, *H.*  
251 *rhossiliensis*, *P. lilacinus*, *Catenaria* sp. and *P. nicotianae*, and negative relationships between EPNs and  
252 sand content and *A. musiformis* (Table 4). Subsets of those independent variables explained 55%, 73%  
253 and 46% of the variation in *S. diaprepesi*, *H. indica* and *H. zealandica*, respectively. For flatwoods sites,  
254 regression of EPN species commonly encountered in the flatwoods (*H. indica*, *S. diaprepesi* and  
255 *Steinernema* sp.) showed positive relationships between EPNs and natural areas, GWD, calcium, boron,  
256 magnesium, copper, cation exchange capacity, *G. gephyropagum*, *P. lilacinus*, *Catenaria* sp., *P.*  
257 *nicotianae* and *Acrobeloides*-group, and inverse relationships between EPNs and silt and clay content,  
258 elevation, boron, manganese, iron and between *H. indica* and both of the steinernematid species (Table 3).

259

#### 260 **4. Discussion**

261

262 As predicted, several NF species were detected at different frequency in nematode samples from  
263 the citrus orchards than were reported from natural areas nearby these orchards (Campos-Herrera et al.,  
264 2016). Each of those species (*A. oligospora*, *A. dactyloides*, *G. gephyropagum*, *Catenaria* sp.) tended to  
265 be detected in nearly half of the samples from orchards and fewer than 10% of natural sites. It is unknown  
266 whether these differences were due to the different survey times, sample processing (operator) variability  
267 or orchard *versus* natural area properties. A remarkably similar trend was reported from a Mediterranean  
268 region in which trapping fungi (primarily *A. oligospora*) and the endoparasitic *Catenaria anguliluae* were  
269 detected (by sprinkling soil on agar and baiting with nematodes) in nearly all samples from seven citrus  
270 orchards, but fewer than half of the samples from 19 nearby forests (Olivares-Bernabeu et al., 2003). If  
271 the occurrence of NF is generally greater in orchards, the potential to develop conservation biological  
272 control tactics for managing plant or animal parasitic nematodes with these fungi is evident. Several  
273 edaphic properties measured here that reflect the use of irrigation and fertilizers were significantly related  
274 to the abundance of each NF species both in the orchards and when orchard and natural area data were

275 pooled, and, therefore, merit controlled studies of their effects on these fungi (discussed below).  
276 Additionally, our orchard samples were gotten only where *Diaprepes* root weevil infestations were  
277 documented, whereas the natural sites were chosen without knowledge of soil arthropod host status.  
278 Campos-Herrera et al., (2012, 2016) reported close associations between EPNs, free-living nematodes and  
279 several species of NF, and noted that insect cadavers provide abundant resources to r-selected nematode  
280 species and their natural enemies. The lack of overall species diversity in monoculture orchards compared  
281 to natural areas may also have resulted in more EPN hosts in the orchards. Efron et al. (2001) reported a  
282 strong temporal correlation between seasonal abundance (host availability) of the grub *Maladera matrida*  
283 and steinernematids and heterorhabditids in citrus orchards in Israel.

284 We also confirmed that some spatial associations between EPNs and NF may have resulted from  
285 the ecoregion preferences of each species. SADIE association indices derived from georeferenced spatial  
286 patterns revealed consistently that the degree to which NF and EPN species were associated or  
287 disassociated was predictable by whether each tends to occupy habitats with deep or shallow water table.  
288 Nevertheless, in contrast to EPNs (Campos-Herrera et al., 2013, 2016), the regional differences between  
289 NF communities measured by either SADIE or PCA were very subtle. Only *P. lilacinus* and *G.*  
290 *gephyropagum* occurred significantly more frequently in flatwoods and central ridge, respectively.  
291 Neither species was inversely related to any EPN species, whether analyzed within or across ecoregions.  
292 Thus, whereas the community diversity of EPNs tended to be greatest on the central ridge where diversity  
293 of NF communities was the least, we found little additional evidence that NF infection rates influenced  
294 the regional occurrence of EPN species. Indeed, even within each ecoregion, the positive relationships  
295 between nematodes and fungi measured by stepwise regression suggest that EPNs may be an important  
296 resource for NF population growth, even if the NF measured here have little effect on the regional spatial  
297 distribution of the nematodes. Although *Steinernema* sp. tended to occur in sites with low incidence of  
298 most NF species, there were no significant correlations between the nematode and any NF species nor  
299 with the total NF communities in the sites. The dominant EPN *H. indica* was predictably, inversely  
300 related to its competitors *S. diaprepesi* and *Steinernema* sp. in the flatwoods, but significant relationships

301 between EPN species and NF species in flatwoods were all positive. A notable exception to this pattern  
302 occurred for an infrequently detected NF, *A. musiformis*, which was inversely related to the two dominant  
303 EPN species on the central ridge, *S. diaprepesi* and *H. indica*. However, *Catenaria* sp., *H. rhossiliensis*  
304 and *P. lilacinus* on the central ridge were positively associated with four EPN species between them. Put  
305 simply, the evidence from citrus orchards and their environs suggests that NF as a guild prey on EPNs at  
306 similar rates everywhere.

307 *Arthrobotrys oligospora* was the most frequently detected nematode-trapping species in both  
308 regions, consistent with numerous other reports from a broad range of habitats (Duddington, 1954;  
309 Shepherd, 1955; Persmark et al., 1996; Koppenhöfer et al., 1997; Durand et al., 2005; Farrell et al., 2006;  
310 Wachira et al., 2008). Similarly, Mitsui (1985) and Gray (1987) noted that net formers in general were the  
311 most frequently encountered species in numerous surveys on the occurrence of nematode trapping fungi.  
312 However, the constricting ring former *A. dactyloides*, as well as *P. lilacinus* and *Catenaria* species were  
313 equally abundant in nematode samples from these citrus groves. The more even detection of various NF  
314 guilds than reported previously may reflect the sampling method designed to measure predation as  
315 distinct from saprophagy (Pathak et al., 2012).

316 Other studies have shown positive relationships between nematodes and the endoparasitic, as well  
317 as the more parasitic nematode-trapping fungi, but only non-significant correlations with the more  
318 saprophytic nematode-trapping fungi (Jaffee et al., 1993; Persmark and Nordbring-Hertz, 1997). As  
319 noted previously, sampling to detect predation rather than saprophytic behavior in these fungi may be  
320 why positive relationships were evenly divided between obligate and facultative NF species in this study.  
321 A number of studies have also linked endoparasitic fungi to the regulation of some invertebrate  
322 communities in freshwater lakes and streams: mosquito larvae by *Coelomyces* (Whisler et al., 1974,  
323 1975), nematodes by *Catenaria anguillule* (Gray, 1985), eggs of Chironomidae by various *Catenaria*  
324 species (Martin, 1981), and *Daphnia* sp. by the chitrid *Polycaryum* leave (Johnson et al., 2006). Martin  
325 (1984) showed that the population size of midges was controlled largely by *Catenaria* sp. and other  
326 factors in littoral communities. In contrast to those reports, our findings are consistent with those of Jaffee

327 et al. (1996) who found no relationship between spatial patterns of NF and those of *H. marelatus* which  
328 regulate ghost moths (*Hepialus californicus*) and the damage they cause in bush lupines. Jaffee et al  
329 (2007) also reported that resident nematodes and trapping fungi increased when dead moth larvae were  
330 added to the soil but this increase in fungus did not affect the probability of nematode mortality.

331 Our study implicated some abiotic factors as potentially affecting the composition of NF  
332 communities in the citrus orchards. Gray (1988) studied relationships between soil chemistry and NF in  
333 agricultural fields and noted that the effects of nutrient elements on NF are likely often indirect, mediated  
334 by relationships between nutrients and numbers of nematode prey. None of the variables that explained  
335 significant variation in NF species (AWC, EC, pH, P, K, or B) occurred at especially different levels in  
336 either ecoregion (Campos-Herrera et al., 2013), consistent with the lack of regionality in most NF species.  
337 Available water capacity was strongly associated with *Catenaria* sp., which requires moisture for  
338 zoospore function. Freeman et al. (2009) found chytrids to be the dominant group in high elevation fungal  
339 communities where soils are saturated for long periods due to snow cover. The positive association of *P.*  
340 *lilacinus* and AWC was consistent with the greater occurrence of that species in flatwoods orchards,  
341 whereas *A. oligospora* was also associated with AWC, but not with either region. Electrical conductivity  
342 (EC) and potassium (K) were significantly related (inversely and directly, respectively) to 5 of the 6  
343 fungi. Salinity may have caused some of the relationships between NF and EC, but EC was also inversely  
344 correlated with pH and fungi often function better in acidic environments. Gray (1985) reported that the  
345 ring and knob-forming species were associated with soils with a low pH and the species with unmodified  
346 adhesive hyphae were isolated more frequently from soils with a higher pH. Therefore, low detection of  
347 NF at high EC may have been due to preference for low pH as was seen in *A. musiformis* and *G.*  
348 *gephyropagum*. This would unlikely be the case for *Catenaria* sp., however. Sayre and Keeley (1969)  
349 reported that in *C. anguillulae* infection rates of nematodes were highest at pH 9 and maximum growth  
350 rates for *C. anguillulae* in culture were between 8 and 9 (Nolan, 1985).

351 Phosphorus (P) content could have affected the *Catenaria* sp. and *A. oligospora* infection rates.  
352 Phosphorus is an essential element for plant health that is vastly scarce in utilizable form (mainly

353 orthophosphates). The limited availability of P is due to the binding of phosphate anions with other  
354 elements. Several studies revealed that chytrids can convert inorganic phosphorous (phosphate ion) into  
355 organic compounds (Willoughby 1962; Murray and Lovett 1966; Nolan 1970; Hassan and Catapane  
356 2000; Gleason et al. 2006). It was also reported that available soil phosphorus may be a determinant of  
357 chytrid community composition and diversity (Letcher et al. 2004). Midgley et al. (2006) showed that in  
358 addition to insoluble phosphorous, chytrids can use DNA as a source of phosphorous and they speculated  
359 that limited availability of phosphorus may influence the chytrids found in any habitat.

360 Lauber et al. (2008) reported that phylogenetic distance between fungal communities was  
361 correlated with soil P concentrations and the C:N ratio. Their analyses of the relative abundances of  
362 specific fungal taxa indicated that P-rich soils contained more Ascomycota and fewer Basidiomycota. In  
363 our results, *A. oligospora* was the only ascomycete that correlated strongly with P content. The other four  
364 ascomycetes showed little or no correlation with phosphorus content but were significantly positively  
365 associated with potassium. Kumar et al (2005) reported that application of fertilizers (urea, di-ammonium  
366 phosphate (DAP) and muriate of potash) reduced the population of nematode trapping fungi in the soil. In  
367 Florida citrus orchards, phosphorus is supplied in form of ammonium phosphates (MAP or DAP),  
368 superphosphate and NPK compound fertilizers that might be detrimental to sensitive fungi like *A.*  
369 *dactyloides*. *Purpureocillium lilacinus* enhanced the availability of soil phosphorus in the presence of  
370 calcium phosphates *in vitro* (Hernández-Leal et al, 2011). The lack of reported relationships between  
371 phosphorus and other ascomycetes in our study may also reflect the sampling method.

372 The association of *A. dactyloides* with potassium is in accordance with Gray (1988) where  
373 constricting ring forming species were correlated with soils containing high levels of K. In general,  
374 nematophagous fungi in this study (except *G. gephyropagum*) are associated with major soil nutrients (P  
375 and K) that also support large number of nematode hosts for the fungi (Gray, 1988). Eayre et al. (1987)  
376 and Eayre et al. (1983) showed that K ions stimulated germination of *H. rhossiliensis* conidia and  
377 infection of nematodes, but were unable to show a dose response to K or to increase nematode infection  
378 by addition of KCl in field trials.

379  
380  
381  
382  
383  
384  
385  
386  
387  
388  
389  
390  
391  
392  
393  
394  
395  
396  
397  
398  
399  
400  
401  
402  
403  
404

## 5. Conclusion

Few naturally occurring EPNs have been shown to be significant regulators of pests, and variables that modulate EPN services are generally poorly understood (Stuart et al., 2015; Lewis et al., 2015). By contrast, the EPNs that inhabit Florida citrus orchards are good targets for conservation tactics because they are especially virulent to *Diaprepes* root weevils on the central ridge where damage to citrus by the insect is least (Duncan et al., 2003; Futch et al., 2005). Moreover, controlled studies (Duncan et al., 2013; El-Borai et al., 2016) confirmed that relationships between abiotic soil properties and EPNs detected in citrus orchards and natural areas (Campos-Herrera et al., 2013, 2016) are potentially important in modulating the species composition of EPN communities. However, the possibility that NF communities extend a trophic cascade affecting citrus trees (NF→EPN→*Diaprepes* root weevil→citrus) differently in different ecoregions was not supported by this or a previous survey (Campos-Herrera et al., 2013). *Purpureocillium lilacinus*, the only species that predominates in the region where weevil damage is greatest, was not significantly related to EPNs across or within ecoregions. The observation that *Steinernema* sp. tended to occur in sites depauperate of most NF species may have the most relevance to the question of EPN conservation. That species and *S. diaprepesi* are the most virulent to *Diaprepes* root weevil (El-Borai et al., 2012) and *Steinernema* sp. is the best adapted of the two to flatwoods conditions (Campos-Herrera et al., 2013, 2016; El-Borai et al., 2016). However, unlike *S. diaprepesi* which is virtually ubiquitous in orchards on the central ridge, *Steinernema* sp. occurs only rarely in flatwoods. Cultural practices that mitigate the biotic or abiotic constraints to *Steinernema* sp. could represent a profitable tactic for *Diaprepes* root weevil management.



405 **Acknowledgments**

406

407 This study is supported by a USDA–CSREES Special Grant (TSTAR), and U.S–Egypt Science  
408 and Technology Joint Fund (338). R. Campos–Herrera was awarded with postdoctoral fellowships by the  
409 Ramón Areces Spanish Foundation (Natural Sciences Program 2008) and was supported by a Marie Curie  
410 International Outgoing Fellowship (FP7-PEOPLE-2009-IOF-252980) within the 7th European  
411 Community Framework Programme.

412

413 **References**

414 Atkins, S.D., Clark, I., Pande, S., Hirsch, P.R., Kerry, B.R., 2005. The use of real-time PCR and species-  
415 specific primers for the identification and monitoring of *Paecilomyces lilacinus*. *FEMS*  
416 *Microbiology Ecology* 51, 257-264.

417 Campos–Herrera, R., Johnson, E.G., El-Borai, F. E., Graham, J.H., Duncan, L.W., 2011a. Long-term  
418 stability of entomopathogenic nematode spatial patterns in soil as measured by sentinel insects  
419 and real-time PCR assays. *Annals of Applied Biology* 158, 55-68.

420 Campos–Herrera, R., El-Borai, F.E., Stuart, R.J., Graham, J.H., Duncan, L.W., 2011b. Entomopathogenic  
421 nematodes, phoretic *Paenibacillus spp.*, and the use of real time quantitative PCR to explore soil  
422 food webs in Florida citrus groves. *Journal of Invertebrate Pathology* 108, 30-39.

423 Campos–Herrera, R., El-Borai, F.E., Duncan, L.W., 2012. Wide interguild relationships among  
424 entomopathogenic and free-living nematodes in soil as measured by real time qPCR. *Journal of*  
425 *Invertebrate Pathology* 111, 126-135.

426 Campos–Herrera, R., Pathak, E., El-Borai, F.E., Stuart, R.J., Gutiérrez, C., Rodríguez-Martín, J.A.,  
427 Graham, J.H., Duncan, L.W., 2013. Geospatial patterns of soil properties and the biological  
428 control potential of entomopathogenic nematodes in Florida citrus groves. *Soil Biology and*  
429 *Biochemistry* 66, 163-174.

430 Campos-Herrera R., El-Borai, F.E., Rodríguez Martín, J.A., Duncan, L.W., 2016. Entomopathogenic  
431 nematode food web assemblages in Florida natural areas. *Soil Biology and Biochemistry* 93, 105-  
432 114.

433 Dillman, A.R., Chaston, J.M., Adams, B.J., Ciche, T.A., Goodrich-Blair, H., Stock, S.P., Sternberg,  
434 P.W., 2012. An entomopathogenic nematode by any other name. *PLoS Pathogens* 8 (3),  
435 e1002527.

436 Dolinski, C., Choo, H.Y., Duncan, L.W., 2012. Grower acceptance of entomopathogenic nematodes: case  
437 studies on three continents. *Journal of Nematology* 44, 226-235.

438 Duchaufour, P., 1975. Manual de Edafología. Toray-Masson, Barcelona, Spain (in Spanish).

439 Duddington, C.L., 1954. Nematode destroying fungi from agricultural soils. *Nature, Lond.* 173: 500.

440 Duncan, L.W. 2014. Can Diaprepes weevil be managed with physical barriers? *Citrus Industry Magazine*,  
441 June, pp. 14-16..

442

443 Duncan, L.W., Graham, J.H., Dunn, D.C., Zellers, J., McCoy, C.W., Nguyen, K., 2003. Incidence of  
444 endemic entomopathogenic nematodes following application of *Steinernema riobrave* for control  
445 of *Diaprepes abbreviatus*. *Journal of Nematology* 35,178-186.

446 Duncan, L.W., Stuart, R.J., El-Borai, F.E., Campos-Herrera, R., Pathak, E., Giurcanu, M., Graham, J.H.,  
447 2013. Modifying orchard planting sites conserves entomopathogenic nematodes, reduces weevil  
448 herbivory and increases citrus tree growth, survival and fruit yield. *Biological Control* 64, 26-36.

449 Durand, D.T., Boshoff, H.M., Michael, L.M., Krecek, R.C., 2005. Survey of nematophagous fungi in  
450 South Africa. Onderstepoort *Journal of Veterinary Research* 72, 185-187.

451 Eayre, C.G., Jaffee, B.A., Zehr., E.I., 1983. Suppression of *Criconemella xenoplax* by the fungus  
452 *Hirsutella rhossiliensis*. *Phytopathology* 73, 500.

453 Eayre, C.G., Jaffee, B.A., Zehr, E.I., 1987. Suppression of *Criconemella xenoplax* by the nematophagous  
454 fungus *Hirsutella rhossiliensis*. *Plant Disease* 71, 832-834.

455 Efron, D., Nestel, D., Glazera, I., 2001. Spatial analysis of entomopathogenic nematodes and insect hosts  
456 in a citrus grove in a semi-arid region in Israel. *Environmental Entomology* 30 (2), 254-261.

457 Ekmen, Z.I., Selcuk, H., Cakmak, I., Ozer, N., Karagoz, M., Kaya, H.K., 2010. Potential negative effects  
458 on biological control by *Sancassania polyphyllae* (Acari: Acaridae) on an entomopathogenic  
459 nematode species. *Biological Control* 54, 166-171.

460 El-Borai, F.E., Stuart, R.J., Campos-Herrera, R., Pathak, E., Duncan, L.W., 2012. Entomopathogenic  
461 nematodes, root weevil larvae, and dynamic interactions among soil texture, plant growth,  
462 herbivory, and predation. *Journal of Invertebrate Pathology* 109, 134-142.

463 El-Borai, F.E., Killiny, N., Duncan, L.W., 2016. Concilience in entomopathogenic nematode responses to  
464 water potential and their geospatial patterns in Florida. *Frontiers in Microbiology*  
465 <http://journal.frontiersin.org/article/10.3389/fmicb.2016.00356>

466 Farrell, F.C., Jaffee, B.A., Strong, D.R., 2006. The nematode-trapping fungus *Arthrobotrys oligospora* in  
467 soil of the Bodega marine reserve: distribution and dependence on nematode-parasitized moth  
468 larvae. *Soil Biology and Biochemistry* 38, 1422-1429.

469 Freeman, K.R., Martin, A.P., Karki, D., Lynch, R.C., Mitter, M.S., Meyer, M.S., Longcore, J.E.,  
470 Simmons, D.R., Schmidt, S.K., 2009. Evidence that chytrids dominate fungal communities at  
471 high-elevation soils. *Proceedings of the National Academy of Sciences* 106, 18315-18320.

472 Futch, S.H., Duncan, L.W., Zekri, M., 2005. Validation of an area-wide extension program to estimate the  
473 seasonal abundance of adult citrus root weevils with un-baited pyramidal traps. *Proceedings of*  
474 *the Florida State Horticultural Society* 117, 143-147.

475 Gleason, F.H., Midgley, D.J., Letcher, P.M., McGee, P.A., 2006. Can soil chytridiomycota survive and  
476 grow in different osmotic potentials? *Mycological Research* 110, 869-875.

477 Gray N.F., 1985. Ecology of nematophagous fungi: Effect of soil moisture, organic matter, pH and nematode  
478 density on distribution. *Soil Biology and Biochemistry* 17, 499-507.

479 Gray, N.F., 1987. Nematophagous fungi with particular reference to their ecology. *Biological Reviews* 62,  
480 245-304.

481 Gray, N.F., 1988. Ecology of nematophagous fungi: Effect of the soil nutrients N, P and K, and seven  
482 major metals on distribution. *Plant Soil* 108, 286-90.

483 Gray, N.F., 1988. Fungi attacking vermiform nematodes. pp. 3-38. In: Poinar GO, Jansson HB (eds),  
484 Diseases of Nematodes Vol. II, CRC Press, Boca Raton.

485 Halbert, S.E., 2005. The discovery of huanglongbing in Florida. In: *Proceedings of the 2nd International*  
486 *Citrus Canker and Huanglongbing Research Workshop*, no. H-3, p. 50.

487 Hassan, M.W., Catapane, E.J., 2000. Physiology of two monocentric chytrids: comparative nutritional  
488 studies of *Entophlycti spp.* and *Entophlyctis aureus* (Chytridiales). *Canadian Journal of Botany*  
489 78, 105-109.

490 Hernández-Leal, T.I., Carrión, G., Heredia, G., 2011. In vitro phosphate solubilization by a strain of  
491 *Paecilomyces lilacinus* (thom) Samson. *Agrociencia* vol.45 no.8 México.

492 Hodson, A.K., Siegel, J.P., Lewis, E.E., 2012. Ecological influence of the entomopathogenic nematode,  
493 *Steinernema carpocapsae*, on pistachio orchard soil arthropods. *Pedobiologia* 55, 51-58.

494 Huang, J., Li, C., Xiao, C., Wang, G., 2010. Detection of *Phytophthora nicotianae* in soil with realtime  
495 quantitative PCR. *Journal of Phytopathology* 158, 15-21.

496 ISO 10382, 2002. Soil quality-determination of organochlorine insecticide chlorine pesticides and  
497 polychlorinated biphenyls-gas-chromatographic method with electron capture detection.  
498 American

499 Jaffee, B.A., Tedford, E.C. Muldoon, A.E., 1993 Tests for density-dependent parasitism of nematodes by  
500 nematode-trapping and endoparasitic fungi. *Biological Control* 3, 329-336.

501 Jaffee, B.A., Strong, D.R., Muldoon, A.E., 1996. Nematode trapping fungi of a natural shrubland: Tests  
502 for food chain involvement. *Mycologia* 88, 554-564.

503 Jaffee, B.A., Bastow, J.L. Strong, D.R., 2007. Suppression of nematodes in a coastal grassland soil.  
504 *Biology and Fertility of Soils* 44, 19-26.

505 Jenkins, W.R., 1964. A rapid centrifugal-flotation technique for separating nematodes from soil. *Plant*  
506 *Disease Report* 48, 492.

507 Johnson, E.G., Wu, J., Bright, D.B., Graham, J.H., 2014. Association of 'Candidatus Liberibacter  
508 asiaticus' root infection, but not phloem plugging with root loss on huanglongbing-affected trees  
509 prior to appearance of foliar symptoms. *Plant Pathology* 63, 290-298.

510 Johnson, P., Longcore, J., Stanton, D., Carnegie, R., Shields, J., Preu, E., 2006. Chytrid infections of  
511 *Daphnia pulicaria*: development, ecology, pathology and phylogeny of *Polycaryum laeve*.  
512 *Freshwater Biology* 51(4), 634-648.

513 Koppenhofer, A.M., Jaffee, B.A., Muldoon, A.E., Strong, D.R., 1997. Suppression of an  
514 entomopathogenic nematode by the nematode-trapping fungi *Geniculifera paucispora* and  
515 *Monacrosporium eudermatum* as affected by the fungus *Arthrobotrys oligospora*. *Mycologia* 89,  
516 220-227.

517 Kumar, D., Singh, K.P., Jaiswal, R.K., 2005. Effect of fertilizers and neem cake amendment in soil on  
518 spore germination of *Arthrobotrys dactyloides*. *Mycobiology* 33, 194-199.

519 Lauber, C.L., Strickland, M.S., Bradford, M.A., Fierer, N., 2008. The influence of soil properties on the  
520 structure of bacterial and fungal communities across land-use types. *Soil Biology and*  
521 *Biochemistry* 40, 2407-2415.

522 Letcher, P.M., McGee, P.A., Powell, M.J., 2004. Distribution and diversity of zoosporic fungi from soils  
523 of four vegetation types in New South Wales, Australia. *Canadian Journal of Botany* 82, 1490-  
524 1500.

525 Lewis, E.E., Hazir, S., Hodson, A., Gulcu, B., 2015. Trophic relationships of entomopathogenic  
526 nematodes in agricultural habitats. In: R. Campos-Herrera (Ed.), *Nematode pathogenesis of*  
527 *insects and other pests - ecology and applied technologies for sustainable plant and crop*  
528 *protection*. Series: Sustainability in Plant and Crop Protection., A. Ciancio (series Ed.). Vol 1,  
529 Springer International Publishing Switzerland, pp. 139-163.

530 Martin, W.W., 1981. The natural regulation of midge populations by aquatic fungi in Virginia. *Journal of*  
531 *the Elisha Mitchell Scientific Society* 97, 162-170.

532 Martin, W.W., 1984. The dynamics of aquatic fungi parasitic in a stream population of the midge,  
533 *Chironomus attenuatus*. *Journal of Invertebrate Pathology* 44, 36–45.

534 Midgley, D.J., McGee, P.A., Letcher, P.M., 2006. Access to organic and insoluble sources of phosphorus  
535 varies among soil Chytridiomycota. *Archives of Microbiology*. 186, 211-217.

536 Mitsui, Y. 1985. Distribution and ecology of nematode-trapping fungi in Japan. *Japan Agricultural*  
537 *Research Quarterly* 18, 182-193.

538 Murray, C.L., Lovett, J.S., 1966. Nutritional requirements of the chytrid *Karlingia asterocysta*, an  
539 obligate chitinophile. *American Journal of Botany* 53, 469-476.

540 Nolan, R.A., 1970. The phycomycete *Catenaria anguillulae*: growth requirements. *The Journal of*  
541 *General and Applied Microbiology* 60, 167-180.

542 Nolan, R.A., 1985. Physiology and biochemistry. In: Couch, J.N., Bland, C.E. (Eds), *The Genus*  
543 *Coelomomyces*. Academic Press Inc, New York, pp. 321-348.

544 Olivares-Bernabeu, C., Lopez-Llorca, L.V., Boag, B., 2003. Nematodes and nematophagous fungi  
545 associated with citrus fields and *Pinus halepensis-Quercus rotundifolia* forest soils. *Nematologia*  
546 *Mediterranea* 31, 3-9.

547 Pathak, E., EL-Borai, F.E., Campos-Herrera, R., Johnson, E.G., Stuart, R.J., Graham, J.H., Duncan, L.W.,  
548 2012. Use of real-time PCR to discriminate parasitic and saprophagous behaviour by  
549 nematophagous fungi. *Fungal Biology* 116, 563-573.

550 Persmark, L., Banck, A., Jansson, H-B., 1996. Population dynamics of nematophagous fungi and  
551 nematodes in an arable soil: vertical and seasonal fluctuations. *Soil Biology and Biochemistry* 28,  
552 1005-1014

553 Persmark, L., Nordbring-Hertz, B., 1997. Conidial trap formation of nematode trapping fungi in soil and  
554 soil extracts. *FEMS Microbiology Ecology* 22, 313-323.

555 Sayre, R.M., Keeley, L.S., 1969. Factors influencing *Catenaria anguillulae* infections in a free-living and  
556 a plant parasitic nematode. *Nematologica* 15, 492-502.

557 Šmilauer, P., Lepš, J., 2014. *Multivariate Analysis of Ecological Data using Canoco 5*. Second Edition.  
558 Cambridge University Press, Cambridge, UK.

559 Shepherd, A.M., 1955. Some observations on the distribution and biology of fungi predaceous on  
560 nematodes. Ph. D. Thesis, University of London.

561 Spiridonov, S.E., Reid, A.P., Podrunka, K., Subbotin, S.A., Moens, M., 2004. Phylogenetic relationships  
562 within the genus *Steinernema* (Nematoda: Rhabditida) as inferred from analyses of sequences of  
563 the ITS1-5.8S-ITS2 region of rDNA and morphological features. *Nematology* 6, 547–566.

564 Spiridonov, S.E., Moens, M., Wilson, M.J., 2007. Fine scale spatial distributions of two  
565 entomopathogenic nematodes in a grassland soil. *Applied Soil Ecology* 37, 192-201.

566 Stuart, R.J., Barbercheck, M.E., Grewal, P.S., 2015. Entomopathogenic nematodes in the soil  
567 environment: Distributions, interactions and the influence of the biotic and abiotic factors. *In*: R.  
568 Campos-Herrera (Ed.), *Nematode pathogenesis of insects and other pests - ecology and applied*  
569 *technologies for sustainable plant and crop protection*. Series: Sustainability in Plant and Crop  
570 Protection., A. Ciancio (series Ed.). Vol 1, Springer International Publishing Switzerland, pp. 97-  
571 137.

572 Ter Braak, C.J.F., 2009. *Biometris-quantitative methods in the life and earth sciences*. Plant Research  
573 International, Wageningen University and Research Centre, Box 100, 6700 AC Wageningen (The  
574 Netherlands).

575 Wachira, P.M., Kimenju, J.W., Okoth, S., Mibey, R.K., Mungatu, J., 2008. Effect of land use on  
576 occurrence and diversity of nematode destroying fungi in TaitaTaveta, Kenya. *Asian Journal of*  
577 *Plant Sciences* 7, 447-453.

578 Whisler, H.C., Zebold, S.L., Shemanchuk, J.A., 1974. Alternative host for mosquito parasite  
579 *Coelomomyces*. *Nature* 251, 715-716.

580 Whisler, H.C., Zebold, S.L., Shemanchuk, J.A., 1975. Life history of *Coelomomyces psorophorae*.  
581 *Proceedings of the National Academy of Sciences* 72, 693-696

582 Willoughby, L.G., 1962. The fruiting behaviour and nutrition of *Cladochytrium replicatum* Karling.  
583 *Annals of Botany* 26, 13-36

584 Zhang, L.M., Liu., X.Z., Zhu, S.F., Chen, S.Y., 2006. Detection of the nematophagous fungus *Hirsutella*  
585 *rhossiliensis* in soil by real-time PCR and parasitism bioassay. *Biological Control* 36, 316-323.

586

## 587 **Figure legends**

588 **Figure 1.** Results from Redundancy analysis (RDA) for the citrus regional survey (n = 53), considering  
589 abiotic variables and entomopathogenic nematodes as explanatory variables. Together the first two  
590 axes explain 50.9% of the variation in nematophagous fungi composition. The NF species are  
591 represented by solid red arrows and are abbreviated as follows: *Arthrobotrys dactyoides* (Ad), *A.*  
592 *oligospora* (Ao), *A. musiformis* (Am), *Gamsylella gephyropagum* (Gg), *Catenaria* sp. (Cat),  
593 *Purpureocillium lilacinus* (Pl). The explanatory variables represented by solid black arrows ( $P \leq$   
594 0.05) are as follows: Sd (*Steinernema diaprepesi*), EC (electrical conductivity), P (Phosphorus), K  
595 (Potassium), B (Boron), AWC and pH. The explanatory variables represented by dotted blue  
596 arrows ( $0.05 \geq P \geq 0.09$ ) are as follows GWD (ground water depth), OM (Organic matter), Cu  
597 (Copper), Hi (*Heterorhabditis indica*) and sand.

598

599 **Figure 2.** Detection frequency (proportion positive sites) of the seven nematophagous fungi (NF) species  
600 in both surveys involving natural areas (n = 91) and citrus orchards (n = 53), discriminating  
601 between central ridge (CR) and flatwoods (FW) eco-regions. The species are abbreviated as  
602 follows: *Arthrobotrys dactyoides* (Ad), *A. oligospora* (Ao), *A. musiformis* (Am), *Gamsylella*  
603 *gephyropagum* (Gg), *Catenaria* sp. (Cat), *Purpureocillium lilacinus* (Pl), and *Hirsutella*  
604 *rhossiliensis* (Hr). Values represent average  $\pm$  SEM.

605



606 **Figure 3.** Principal components analysis (PCA) of the nematophagous fungi (NF) DNA recovered from  
607 nematode samples from this survey of citrus orchards and from natural areas that were reported by  
608 Campos-Herrera et al. (2016). Data from both surveys were pooled for analysis. Symbols are mean  
609 and standard errors of square root transformed raw data. CR = central ridge and FW = flatwoods.

610

611 **Figure 4.** Ecological indices for the entomopathogenic nematodes (EPN) and nematophagous fungi (NF)  
612 associated with nematodes in two ecoregions of the survey of citrus orchards and of natural areas  
613 reported from by Campos-Herrera et al. (2013, 2016). Indices include species richness (S),  
614 diversity ( $H'$ ) and evenness ( $J'$ ). Values above and below dashed line are highest in central ridge  
615 and flatwoods regions, respectively.

616

617 **Figure 5.** Linear relationships between the SADIE-derived association indices (Xa) for nematophagous  
618 fungi (NF) species and ground water depth and Xa for NF species and EPN species. Abbreviations  
619 for EPN are *Heterorhabditis indica* (Hi), *H. zealandica* (Hz), *S. diaprepesi* (Sd), and *Steinernema*  
620 sp. (Sx), and for NF are *Arthrobotrys dactyoides* (Ad), *A. oligospora* (Ao), *A. musiformis* (Am),  
621 *Gamsylella gephyropagum* (Gg), *Catenaria* sp. (Cat), *Purpureocillium lilacinus* (Pl), and  
622 *Hirsutella rhossiliensis* (Hr).

623

624 **Figure 6.** Mean and standard error of nematophagous fungi (NF) infection rates (normalized between 0-  
625 1) in flatwoods sites with and without the entomopathogenic nematodes (EPN) species  
626 *Heterorhabditis indica* (Hi), *Steinernema diaprepesi* (Sd) and *Steinernema* sp. (Sx). Panels A, C, E  
627 are from the orchard survey and panels B, D, F are from the survey of natural areas.

628

Figures  
Figure 1

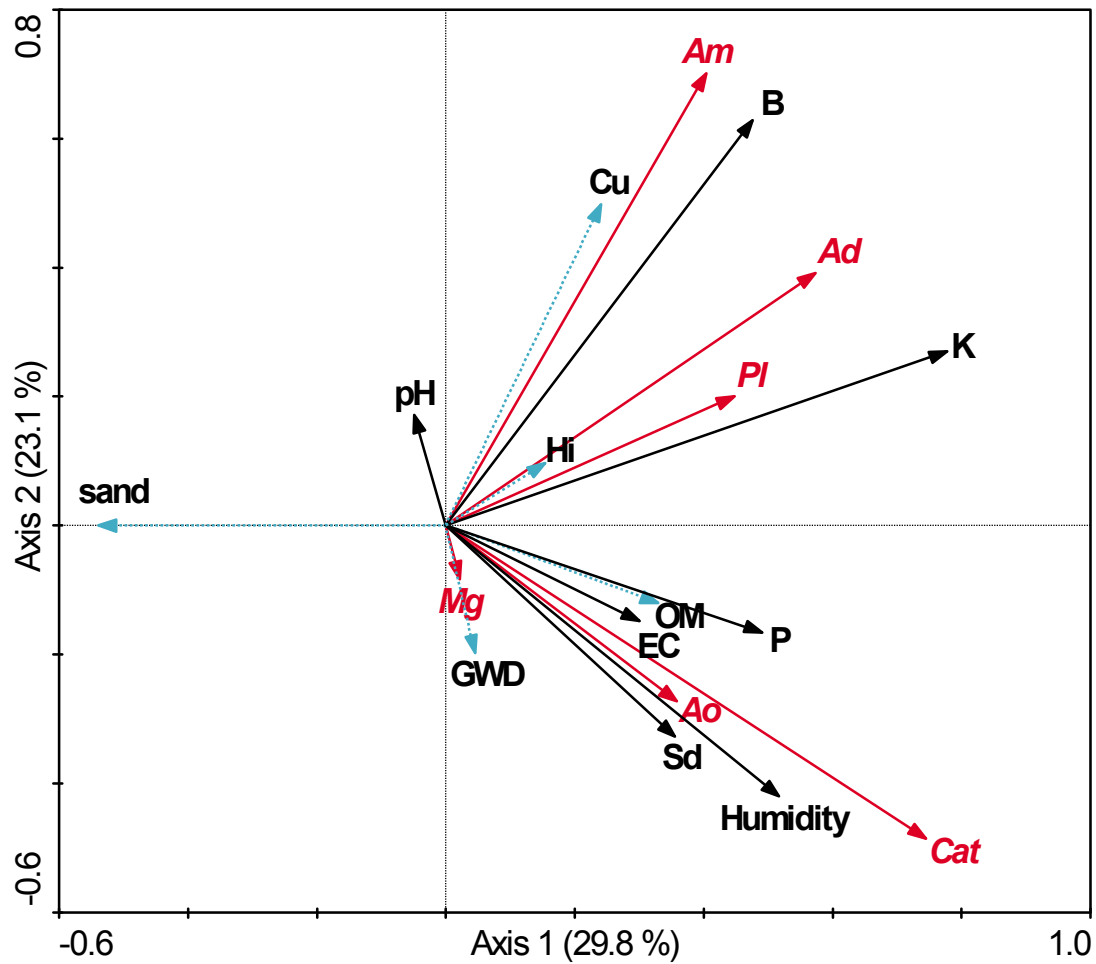


Figure 2

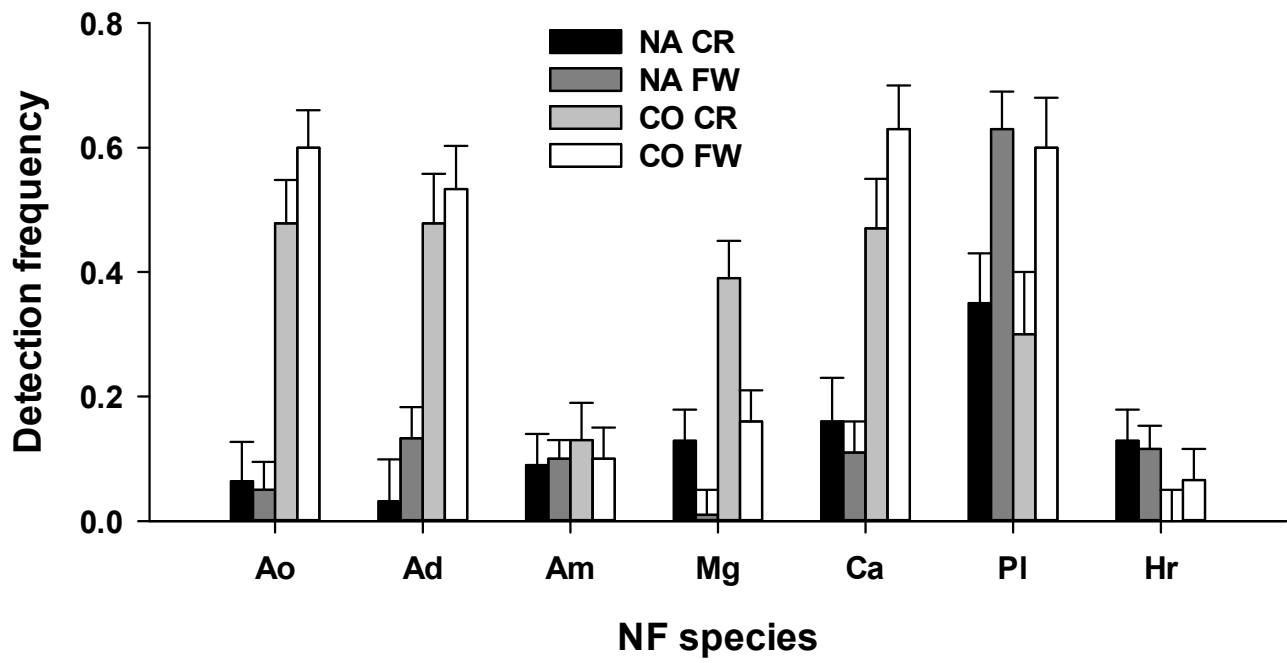


Figure 3

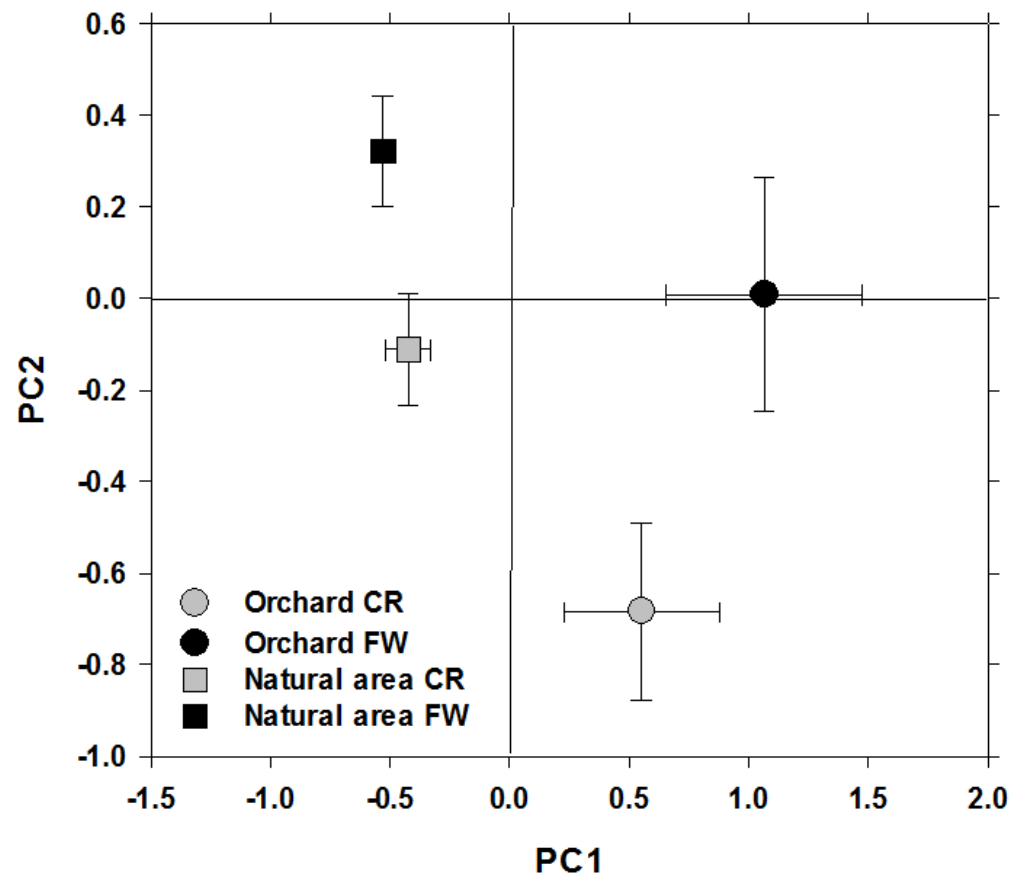


Figure 4

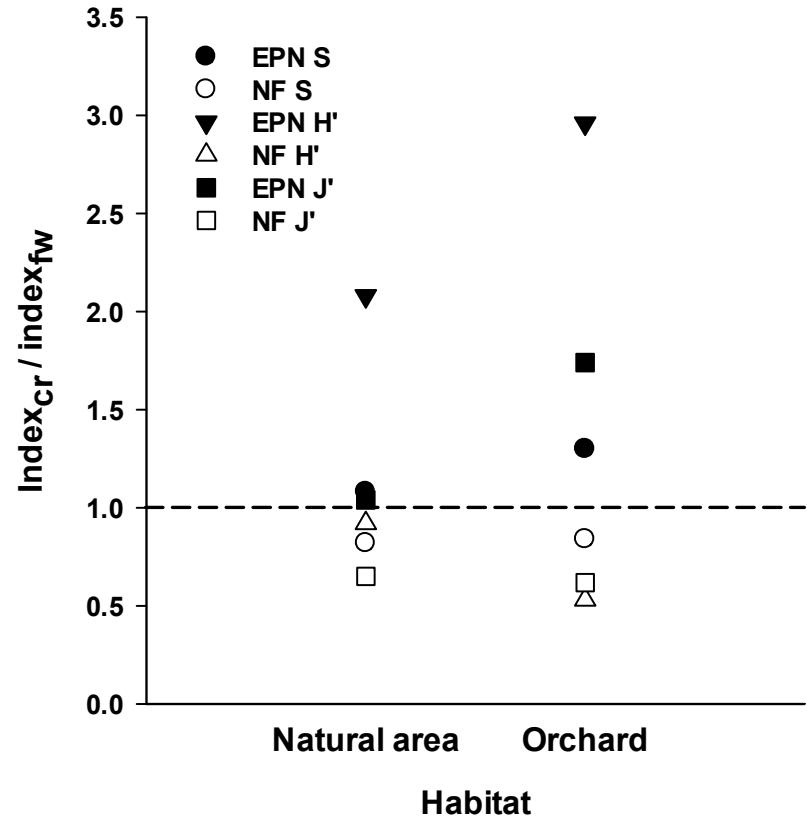
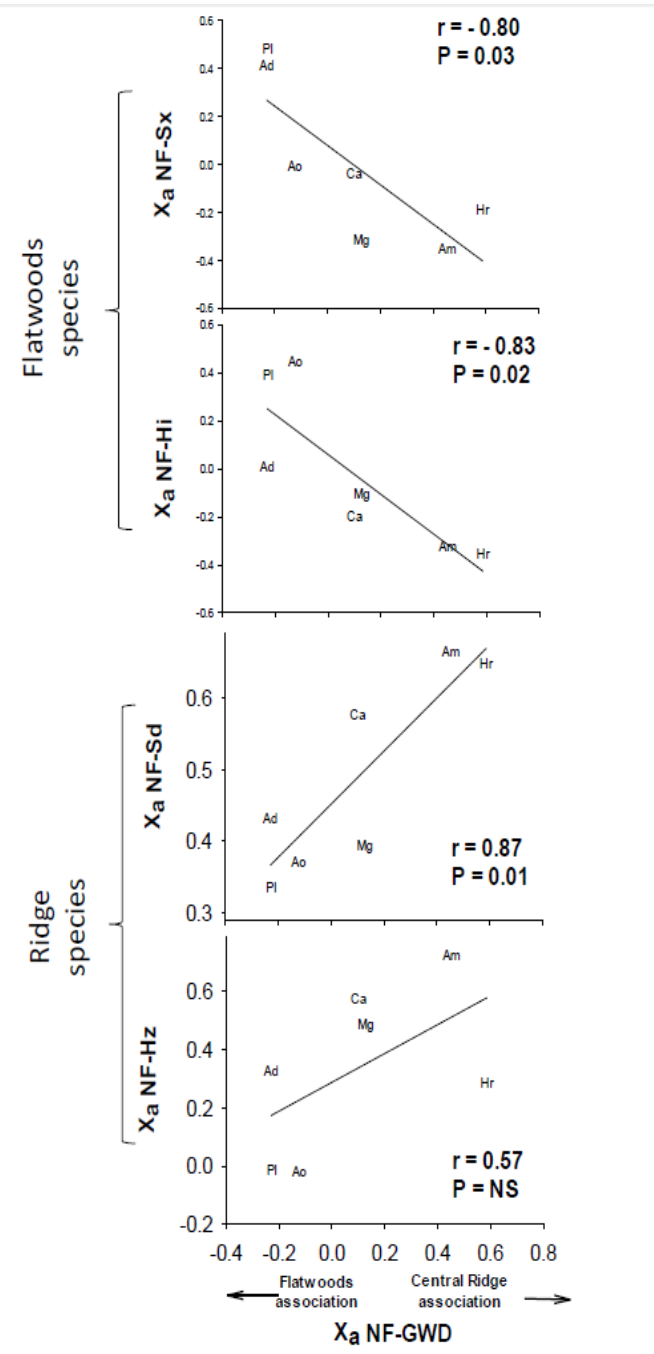


Figure 5





**Table 1.** Frequency of detection of seven species of nematophagous fungi (NF) for the two region of the citrus production in Florida. Results of the T-test analysis (df = 51) for assessing differences between regions ( $P \leq 0.05$ ).

NF species	Frequency (%)			Abundance (ng/uL)		
	Central ridge	Flatwoods	( <i>P</i> )	Central ridge	Flatwoods	( <i>P</i> )
<i>A. dactyloides</i>	50.0	51.6	n.s.	0.0332 ± 0.0225	0.0551 ± 0.0278	n.s.
<i>A. musiformis</i>	13.6	9.7	n.s.	0.00003 ± 0.00002	0.00725 ± 0.00725	n.s.
<i>A. oligospora</i>	50.0	58.0	n.s.	0.00085 ± 0.00044	0.00041 ± 0.000265	n.s.
<i>G</i>	41.0	16.1	0.02	2.65 ± 1.30	1.065 ± 0.563	n.s.
<i>gephyropagum</i>						
<i>Catenaria sp.</i>	50.0	61.3	n.s.	0.00007 ± 0.00006	0.00037 ± 0.00019	n.s.
<i>H. rhosiliensis</i>	0.0	6.5	n.s.	0.0	0.0018 ± 0.00165	n.s.
<i>P. lilacinus</i>	32.0	58	0.03	0.00043 ± 0.00024	0.00115 ± 0.000378	< 0.01



**Table 2.** Ecological indices estimated for the nematophagous fungi (NF) occurrence in citrus orchards in two ecoregions of Florida. Results of the T-test analysis (df = 51) for assessing differences between regions ( $P \leq 0.05$ ).

	Central Ridge	Flatwoods	T value ( <i>P</i> )
Richness, S	2.6 ± 0.244	3.0 ± 0.226	n.s.
Diversity, H'	0.30 ± 0.073	0.56 ± 0.08	2.32 (0.02)
Evenness, J'	0.34 ± 0.066	0.55 ± 0.063	2.25 (0.03)

**Table 3.** T-values from step-wise regressions of nematophagous fungi (dependent variable) on environmental and biotic variables found significant in redundancy analysis (RDA). Significance is denoted by \* ( $P < 0.05$ ), \*\* ( $P < 0.01$ ) and \*\*\* ( $P < 0.001$ ). Fungi abbreviated as follows: *Arthrobotrys dactyoides* (Ad), *A. oligospora* (Ao), *A. musiformis* (Am), *Gamsylella gephyropagum* (Gg), *Catenaria* sp. (Ca), *Purpureocillium lilacinus* (Pl), and *Hirsutella rhossiliensis* (Hr).

	Ad	Am	Ao	Gg	Ca	Pl
AWC	-0.08	-0.93	2.14**	0.35	9.34***	2.29**
EC	-3.47***	-1.69*	-2.62**	-0.65	-3.27***	-4.10***
pH	0.48	-2.71***	0.91	-1.84*	2.59**	1.32
P	0.22	-0.73	2.80***	-0.28	1.99**	-1.08
K	5.34***	1.74*	1.72*	0.90	5.34***	5.59***
B	-0.21	6.95***	-1.96**	-0.92	-2.24**	-2.87***
Sd	1.39	-1.04	-0.23	-0.40	2.15**	0.00
Variation explained	48 %	66 %	23 %	4 %	78 %	36 %

**Table 4.**

Results of stepwise multiple regression of entomopathogenic nematodes (EPNs) on abiotic and biotic edaphic factors using data pooled from surveys of citrus orchards and adjacent natural areas. Significance is denoted by \* ( $P < 0.05$ ), \*\* ( $P < 0.01$ ) and \*\*\* ( $P < 0.001$ ).

Independent variables	Dependent variables					
	Central Ridge			Flatwoods		
	<i>S. diaprepesi</i>	<i>H. indica</i>	<i>H. zealandica</i>	<i>S. diaprepesi</i>	<i>H. indica</i>	<i>Steinernema</i> sp. <i>glaseri</i> -group
R <sup>2</sup>	55.5	43.3	45.8	84.2	57.5	23.5
<i>Abiotic factors</i>				-	-	-
Citrus vs Natural	-6.97***	-	-	3.62**	-	-
Elevation (masl)		-	-	-	-2.52*	-
GWD (m)		-	-	4.42***	-	-
Sand content (%)	-3.31**	-	-	-	-	-
Silt content (%)	-	-	-	-	-	-2.66**
Clay content (%)	-	-	-	-2.63**	-	-
CEC	-	-	-	-	2.56*	-
pH	-	-	-	-	6.44***	-
K	-	2.83**	-	-	-	-
P	-	-	2.9**	-	-	-

B	-	-	-	-4.16***	-	2.4*
Ca	-	-	-		-4.11***	-
Mg	-	-	-	10.63***	-	-
Cu	-	-	-	4.05***	-	-
Mn	-	-	-	-2.45*	-	-
Fe	-	-	-	-2.42*	-	-
<i>Biotic factors</i>						
EPN and FLN						
<i>H. indica</i>	-	-	-	-2.11*	-	-2.33*
<i>Acroboloides</i> -group	-	-	2.11*	-	-	3.19**
Members of the citrus-EPN soil food web						
<i>P. nicotianae</i>	-	4.81***	-	7.04***	-	-
<i>A. musiformis</i>	-2.71**	-2.0*	-	-	-	-
<i>A. oligospora</i>	-	-	-	7.94***	-	-
<i>Catenaria</i> sp.	3.42**	-	-	-	-	-
<i>H. rhossiliensis</i>	-	3.36**	5.29***	8.01***	-	-
<i>P. lilacinus</i>	-	2.32**	-	-	-	-
<i>G. gephirophagum</i>	-	-	-	-	2.43*	-