



THE UNIVERSITY *of* EDINBURGH

Edinburgh Research Explorer

Genotype and sex-based host variation in behavior and susceptibility drives population disease dynamics

Citation for published version:

White, LA, Siva-Jothy, J, Craft, ME & Vale, PFD 2020, 'Genotype and sex-based host variation in behavior and susceptibility drives population disease dynamics', *Proceedings of the Royal Society B-Biological Sciences*. <https://doi.org/10.1098/rspb.2020.1653>

Digital Object Identifier (DOI):

[10.1098/rspb.2020.1653](https://doi.org/10.1098/rspb.2020.1653)

Link:

[Link to publication record in Edinburgh Research Explorer](#)

Document Version:

Peer reviewed version

Published In:

Proceedings of the Royal Society B-Biological Sciences

General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.



1 **Genotype and sex-based host variation in behavior and**
2 **susceptibility drives population disease dynamics**

3
4 Lauren A. White^{1,2}, Jonathon A. Siva-Jothy³, Meggan E. Craft², Pedro F. Vale³

5
6 1. National Socio-Environmental Synthesis Center SESYNC, 1 Park Place, Suite 300,
7 Annapolis, MD 21401, USA.

8
9 2. Department of Veterinary Population Medicine, University of Minnesota, St Paul, MN
10 55126

11
12 3. Institute of Evolutionary Biology, School of Biological Sciences, University of Edinburgh,
13 Ashworth Labs, Charlotte Auerbach Road, EH9 3JT Edinburgh, UK.

14
15
16

17 **Abstract**

18 Host heterogeneity in pathogen transmission is widespread and presents a major hurdle to predicting
19 and minimizing disease outbreaks. Using *Drosophila melanogaster* infected with *Drosophila C* virus
20 as a model system, we integrated experimental measurements of social aggregation, virus shedding,
21 and disease-induced mortality from different genetic lines and sexes into a disease modelling
22 framework. The experimentally measured host heterogeneity produced substantial differences in
23 simulated disease outbreaks, providing evidence for genetic and sex-specific effects on disease
24 dynamics at a population level. While this was true for homogeneous populations of single
25 sex/genetic line, the genetic background or sex of the index case did not alter outbreak dynamics in
26 simulated, heterogeneous populations. Finally, to explore the relative effects of social aggregation,
27 viral shedding and mortality, we compared simulations where we allowed these traits to vary, as
28 measured experimentally, to simulations where we constrained variation in these traits to the
29 population mean. In this context, variation in infectiousness, followed by social aggregation, was the
30 most influential component of transmission. Overall, we show that host heterogeneity in three host
31 traits dramatically affects population-level transmission, but the relative impact of this variation
32 depends on both the susceptible population diversity and the distribution of population-level variation.

33

34 **Key words:** *Drosophila melanogaster*, disease transmission, social aggregation, virus shedding,
35 contact networks, disease modelling, transmission heterogeneity

36

37 **Introduction**

38 Individual heterogeneity in host traits affecting disease transmission has major consequences for the
39 predictability and severity of outbreaks of infectious disease, and in extreme cases can lead to
40 ‘superspreaders’ or ‘supershedders’ of infection [1–3]. An individual’s transmission potential can be
41 described as a function of: (1) its rate of contact with susceptible individuals, (2) the likelihood of that
42 contact resulting in infection, and (3) the length of time that individual remains infectious [4,5]. It is
43 therefore important to understand how common sources of variation, such as host genetic background
44 and sex, may contribute to the variance in these traits and how individual variation may scale up to
45 population level disease dynamics [4,6,7].

46

47 Disease dynamics may be disproportionately driven by individuals with extreme behavioural and
48 physiological traits including social aggregation, pathogen shedding, or in the host’s ability to resist
49 or tolerate the infection. For example, sex differences in immunity [8] or nutritional and thermal
50 effects on host behaviours [9,10] can lead to differences in hosts’ ability to tolerate infection and
51 consequently increase transmission rates. Similarly, there are also examples of genetic differences
52 driving the extent of pathogen shedding [11] and behaviours that mediate contact between infected
53 and susceptible individuals [12,13]. Quantifying these relevant behavioural, physiological and
54 immune traits and their interactions remains tremendously challenging, particularly in wild or natural
55 disease settings [4].

56

57 One potentially useful approach is experimentally infecting model systems under controlled
58 laboratory settings in order to quantify the roles of physiological and behavioural host heterogeneity
59 on pathogen transmission [12,14,15]. This experimental approach offers the advantage of providing
60 an experimentally tractable framework to partition the variance in individual transmission among a
61 range of behavioural, physiological and immune phenotypes [4], while minimising environmental
62 variation and allowing highly replicated measurements of individual host traits. However, such
63 studies may be limited in their ability to extrapolate the effects of measured heterogeneity at the level

64 of individual hosts to population-level epidemic dynamics. Mathematical modelling is a useful tool to
65 efficiently test different hypotheses and infer patterns across scales [16], but many theoretical studies
66 often rely on assumptions about the level of heterogeneity in host traits, in the absence of empirical
67 data [4,5]. A helpful approach is therefore to use mathematical modelling of epidemiological
68 dynamics whereas many parameters as possible are informed by experimental data measured on
69 individual hosts in controlled laboratory settings.

70

71 Here we combine experimental data and a simulation approach to test how population-level disease
72 transmission dynamics are affected by experimentally measured levels of variation in pathogen
73 shedding, lifespan following infection and social aggregation. We previously measured individual-
74 level variation in behavioural and physiological traits that are relevant to pathogen transmission in the
75 fruit fly (*Drosophila melanogaster*) when infected with its viral pathogen *Drosophila C Virus* (DCV)
76 [13,17]. DCV is a horizontally transmitted ssRNA virus of *Drosophila*. While relatively little is
77 known about DCV dynamics in the wild, it appears to be common as a low-level persistent infection
78 with apparently little pathology among several species of *Drosophila* [18,19]. Following what is
79 presumably a predominantly fecal-oral route of transmission, DCV replicates in the fly's reproductive
80 and digestive tissues leading to intestinal obstruction, lower metabolic rate and reduced locomotor
81 activity [20–22]. Some experimental work has also shown that cannibalism of infectious fly cadavers
82 is a viable route of transmission, but it is unknown how common this transmission route is in the wild
83 [23]. Previously, we observed sex-based and genetic-based variation in both locomotor activity and
84 social aggregation following DCV infection [13]. We also showed that fly genetic background, sex
85 and female mating status significantly influenced infected lifespan, viral growth, virus shedding, and
86 viral load at death [17]. These experiments leveraged genetic and sex-specific sources of variation in
87 three traits that likely affect individual transmission potential of DCV: the degree of group-level
88 social aggregation (as an indicator of potential contact rate); how much DCV each individual sheds
89 into its environment (as a proxy measure of infectiousness); and mortality rate (which defines the
90 duration of infection).

91

92 In the present study, we explore the interactions of social aggregation, viral shedding, and mortality
93 on pathogen transmission when we: (1) vary population means of these traits; (2) vary the individual
94 traits of the index case; and (3) constrain the variance of these traits in the population at large. First,
95 we asked if genetic and sex-specific variation in the population means of social aggregation, virus
96 shedding, and duration of infection – as measured in a lab setting – would result in different predicted
97 epidemics in theoretical populations. By comparing simulated epidemics in host populations
98 comprised of a single sex and one genetic background, we isolated genetic and sex-specific sources of
99 variation in disease transmission. Second, to test the relative importance of the index case vs. group
100 composition, we simulated epidemics in populations where the index case's traits were sampled from
101 a larger phenotypic distribution, including males and females from all ten genetic backgrounds. Third,
102 to test the relative importance of variation in specific host traits on epidemic dynamics, we compared
103 epidemic dynamics of populations exhibiting experimentally-measured levels of variation in social
104 aggregation, viral shedding and mortality, to populations where we constrained variation in these
105 traits to the population mean.

106

107 **Methods**

108 **Simulation model**

109 We developed an individual-based, stochastic, discrete time model that tests how
110 experimentally measured variation in host social aggregation, mortality, and viral shedding in *D.*
111 *melanogaster* translates to differences in disease dynamics. The simulated contact networks
112 underlying this model were generated from degree distributions derived from experimental
113 measurements of social aggregation specific to the sex (σ) and genetic line (g) present in the
114 simulated population. Using a susceptible-infected-removed (*SIR*) process, we simulated direct
115 transmission of DCV in a closed population with no births and where only infected individuals die
116 [24]. We did not include background (i.e. non-disease related) mortality. Note that these transmission
117 processes are consistent with other agent-based models that encompass contact heterogeneity[25].

118 Let the time step be equal to one day, $S(t)$ equal the number of susceptible hosts at time t ,
119 and $I(t)$ equal the number of infectious hosts at time t . The total number of hosts, $N(t)$, in the
120 population at time t is represented by: $N(t) = S(t) + I(t)$. The number of susceptible (S) and infected
121 (I) individuals at the next time step is given by:

$$122 \quad S(t + 1) = S(t) - \sum_{i=1}^{S(t)} \sum_{j=1}^{I(t)} \beta_{ij}(t) s_i(t) i_j(t)$$

$$123 \quad I(t + 1) = I(t) + \sum_{i=1}^{S(t)} \sum_{j=1}^{I(t)} \beta_{ij}(t) s_i(t) i_j(t) - \alpha_j(\sigma, g) i_j(t)$$

124 Here $s_i(t)$ is a vector of susceptible individuals at time t , and $i_j(t)$ is a vector of infected individuals
125 at time t . Therefore, the summations in both equations above iterate over individuals and not time
126 steps.

127

128

129 The processes of mortality $\alpha_j(\sigma, g)$ and transmission (β_{ij}) were individual-specific reflecting the

130 covariates of sex (σ) genetic line (g). More specifically, the transmission between a susceptible
131 individual (s_i) and infectious host (j) is given by:

$$132 \beta_{ij}(t) = \kappa_j(\sigma, g)\eta\tau x_{ij}(t)$$

133 where $\kappa_j(\sigma, g)$ represents the infectiousness of infectious host (j) and x_{ij} represents whether or not an
134 edge exists in the network between individuals s_i and j ($x_{ij}(t) = \begin{cases} 1 \\ 0 \end{cases}$). Because of the uncertainty
135 surrounding the DCV transmission process, we also include scaled infectiousness (η) and
136 transmission efficiency of the pathogen (τ) as components of $\beta_{ij}(t)$, which we discuss in further detail
137 below.

138 For each susceptible individual s_i at each time step, transmission was a stochastic process
139 governed by a Bernoulli draw based on the value of $\beta_{ij}(t)$. Likewise, for each infectious individual (j),
140 mortality was a stochastic process based on a Bernoulli draw for the value of
141 $\alpha_j(\sigma, g)$. Individuals removed during the mortality process no longer contributed to transmission
142 dynamics.

143

144 **Experimental data distributions: Measuring social aggregation, viral shedding and mortality** 145 **rate in infected *D. melanogaster***

146 We used experimental measurements of host social aggregation, mortality, and viral shedding from
147 *D. melanogaster* infected with DCV (Figure 1a-c; note the heterogeneity among genotypes) to test
148 how the sex-specific and genetic variation translates to differences in disease dynamics. An in-depth
149 analysis of these experimental data has been carried out previously, showing substantial genotype-by-
150 sex interactive effects on each of these traits [13,17]. Briefly, we established systemic infections with
151 DCV in males and females of ten lines (Table 1) from the *Drosophila* Genetic Resource Panel
152 (DGRP) [26] and measured a number of traits including social aggregation [20], the infected lifespan
153 and the viral shedding of each line-by-sex combination [17].

154 Here, we focus on the frequency distributions of these data for each fly line and sex (Figure
155 1), as the simulations described below were parameterized using these experimentally derived

156 distributions. Of particular note is the distribution of viral shedding (Figure 1b), which showed
157 substantial zero-inflation, due to many flies not shedding DCV in detectable quantities despite being
158 infected.

159

160 **Social aggregation and contact network degree distribution**

161 Social aggregation was measured by calculating the nearest neighbour distance (NND) from a
162 photograph of groups of ten to twelve flies of the same genetic background, sex and infection status,
163 in 55mm Petri dishes [13]. In accordance with other studies of *D. melanogaster* social aggregation
164 [45], photos were taken of fly groups in Petri dishes following 30 minutes of acclimation to ensure
165 minimal fly activity. Social aggregation was measured in $n = 14\text{--}16$ replicate groups of 12 flies for
166 every combination of genetic background and sex (580 groups of flies in total).

167 The dynamics of faecal-oral DCV transmission are poorly understood [27,28], but the virus
168 readily proliferates through laboratory stocks of *Drosophila* [29]. To account for this uncertainty in
169 transmission mode and to assess the relative importance of possible direct transmission routes, we
170 considered three threshold radii (10, 15 or 20mm) for feasible transmission. For each of these
171 thresholds, the qualifying neighbours for each focal individual was calculated using the coordinates of
172 each fly generated with the ImageJ multipoint tool.

173 To generate a simulated contact network reflecting contact rates of different phenotypes, we
174 started by creating empirical contact networks where an individual (node) shared an edge in the
175 network if they appeared within the prescribed threshold radius of the focal fly. Importantly, using
176 social aggregation as a proximate measure of contact rate assumes the likelihood of contact with DCV
177 is proportional to an individual's proximity to an infected fly. Using the number of neighbours within
178 this radius for each fly (i.e., unweighted degree centrality), we derived an empirical degree
179 distribution for each genetic line and sex combination. From this empirical degree distribution, we
180 sampled 1000 times with replacement to generate a larger degree distribution representing more
181 individuals. To produce a random graph with this given degree sequence, we then used the
182 `samp_degseq` function from the `igraph` package [30]. Note that we resampled if the degree sequence

183 summed to be odd. This produced a network where the mean degree (rather than network density)
184 was maintained between experimental and simulated populations.

185

186 **Infectiousness.** We estimated infectiousness (κ_j) for any given infected individual, j , from our
187 experimental measurements of viral shedding [17]. Viral shedding was measured by housing single
188 infected flies in 1.5ml Eppendorf tubes for 24 hours, removing the fly, washing out the tube with 50 μ l
189 of TRI-reagent to preserve viral RNA, and freezing this sample at -70°C to await RT-PCR and qPCR.
190 Each combination of sex and genetic background consisted of a minimum of 20 replicate flies, with
191 most combinations consisting of 32-38 shedding samples [17].

192 The untransformed distribution of this data was highly skewed and zero-inflated, with some
193 rare flies shedding exceedingly high viral titres (i.e. supershedders)—over two orders of magnitude
194 greater than the population mean—and others not shedding any virus at all (within the technical limit
195 of detection). To account for this disparity, we used the natural log to transform our viral load shed
196 distribution and then normalized values by the greatest amount of virus shed. This transformation
197 yielded a distribution constrained between 0 and 1 with a median value of 0 and a mean value of 0.23.
198 With this transformed distribution, only extreme supershedders at the upper end of the distribution
199 would ensure a high probability transmission, with all other individuals had a probability much less
200 than one.

201 Since the amount of virus needed to ensure DCV transmission is unclear, we also considered
202 a ‘scaled infectiousness’ (η) parameter to explore what would happen if average or non-zero shedders
203 could also shed enough to ensure infection. This scenario was implemented by multiplying our
204 measure of infectiousness (κ_j) by 2. This step expanded the range of the transformed experimental
205 distribution from 0 to 2. Note that for the Bernoulli trial determining whether a transmission event had
206 occurred, the final transmission probability (τ) was then capped at a maximum value of 1.

207 Finally, because the dosage and viability of DCV in the environment remain unclear, we
208 included a transmission efficiency (τ) parameter in our model to account for this uncertainty. The
209 three levels, $\tau = 0.1, 0.5, \text{ or } 1$, altered infectiousness and correspond to 10, 50, and 100% probability

210 of transmission given contact. Both scaled infectiousness (η) and transmission efficiency (τ) were held
211 constant in simulations unless specifically mentioned.

212

213 **Mortality rate.** DCV results in death for infected flies, making our experimental measurement of the
214 time between inoculation and death an ideal measure of mortality rate. Infected lifespan was
215 measured by housing single flies in standard Lewis medium vials following systemic DCV infection
216 and monitored daily until death. For eighteen of twenty sex and genetic background combinations, the
217 lifespan following infection was measured for $n=17-20$, two combinations consisted of $n=13$ and
218 $n=15$ flies [17]. For simulations, we calculated mortality rate, $\alpha_j(\sigma, g)$, as the inverse of
219 experimentally-measured disease-related mortality for a given sex and genetic background.

220

221 **Simulation factorial design**

222 The effects of all parameters on outbreak dynamics were tested in a full-factorial design. For each
223 parameter set, 500 simulations were conducted for a population of 1000 individuals over the course of
224 1000 time steps (Tables 1-3). A wide variety of outbreaks of infectious disease were produced by
225 different combinations of these parameters. To avoid datasets becoming predominated by fadeout, we
226 have presented the outbreaks in populations defined by a set of parameters ($r=15\text{mm}$, $\tau=1$, $\eta=2$) most
227 conducive to outbreaks of infectious disease. Key metrics to measure outbreak dynamics included:
228 fadeout probability, maximum number of infected individuals, outbreak duration, and time to
229 maximum number of infected individuals. Fadeout probability represents the probability of an
230 outbreak stochastically dying out [31]; in this case, we define it as the proportion of simulations
231 where DCV fails to spread beyond the index case. We use R_0 as a measure of the number of secondary
232 cases of infection caused by the index case for the duration of the simulation. Code to conduct these
233 simulations was written in R (Version 3.4.4) and is available at:

234 <https://github.com/whit1951/Drosophila>

235

236 **Random forest analysis**

237 Parsing out the effects of individual variables in simulation modelling can be challenging because of
238 collinear effects and sensitivity of frequentist measures of significance to sample size. To further a
239 descriptive discussion of our simulation results, we have used random forest analysis – a machine
240 learning method that can handle complex, non-linear relationships between model inputs and outputs,
241 as well as potential collinearity between covariates [32]. Random forest analysis is a recursive
242 partitioning method that combines the predictions from numerous fittings of classification or
243 regression trees to the same set of data [32,33]. A higher mean decrease in accuracy correlates with
244 higher variable importance, i.e., more predictive power is lost if this variable is excluded from the
245 analysis. For all three simulation experiments, we analysed outputs of: fadeout probability (whether
246 the infection spread beyond initially infected individual), maximum prevalence, outbreak duration,
247 and R_0 (the number of secondary cases resulting from a single infectious individual in an entirely
248 susceptible population). A detailed description of the analyses can be found in supplementary
249 information and Figures S1-S3.

250

251 **Results**

252

253 **Simulation results**

254 Overall, our findings were robust to changes in various parameter combinations (Table 1). Threshold
255 radius had strong effects on maximum prevalence but was not as strong predictor of a predictor of
256 outbreak likelihood (Figures S1-S4). Here, we present results for a threshold radius of 15mm, a
257 transmission efficiency of 1, and a scaled infectiousness of 2, which were generally representative of
258 most parameter spaces. Summary figures for every parameter combination are presented in Figures
259 S4-S15.

260

261 **Theoretical simulation #1:** We scaled-up the experimental degree distributions for males and
262 females of our ten genetic backgrounds to a theoretical population size of 1000. In each simulated
263 population, flies were of the same sex and genetic background. We allowed infectiousness, duration
264 of infection, and social aggregation to vary based on experimental measurements for each
265 combination of sex and genetic background (Table 1). For each individual simulation, we generated a
266 new network from the scaled-up degree distribution, and randomly selected an individual from the
267 network to start as the index case.

268

269 **Individual variation in host infectiousness, social aggregation, and mortality rate produced** 270 **variation in population-level, pathogen transmission dynamics.**

271 The variation in experimental treatment groups produced distinct outbreaks of infectious disease in
272 populations comprised solely of one genetic background and sex (Figure 2a-d). This finding held true
273 when comparing both genetic lines and sexes. For example, the median outbreak size for line 373
274 females was ~200 flies compared to ~1 fly for line 373 males. In contrast the median outbreak size for
275 line 818 females was ~1 fly, but approached ~500 flies for line 818 males. Random forest analysis
276 suggested that the two top predictors for outbreak likelihood were genetic and sex-specific variation
277 (Figure S1). Given a successful outbreak, host genetic and sex-specific variation also affected the

278 maximum number of infected individuals at any given time step (Figures 2c & S1) and outbreak
279 duration (Figures 2d & S1). However, host genetic background and sex were less important than the
280 threshold radius used to derive social network degree distribution for both outcomes (Figure S1) and
281 less important than transmission efficiency for predicting the maximum number of infected
282 individuals (Figure S1).

283
284 **Theoretical simulation #2:** Many natural host populations have highly variable levels of genetic
285 diversity which can significantly affect host-pathogen dynamics [34]. To test the relative importance
286 of trait differences among potential index cases, we simulated populations where males and females
287 of all ten genetic backgrounds were combined in equal proportion. More specifically, the simulated,
288 scaled-up populations of 1000 individuals were comprised of 20 sub-populations each containing 50
289 sampled individuals drawn from the larger experimental distribution for each respective line/sex
290 combination. Individuals maintained their respective experimentally measured distributions for
291 aggregation, infectiousness, and duration of infection according to their genetic background and sex
292 combination. A connected network of these sub-populations was created by sampling an expected
293 degree for each node based on its subpopulation traits and then using the `samp_degseq` function from
294 the `igraph` package to create a random graph with the given degree sequence as described in the
295 Methods [30]. Thus, flies with different covariate traits (as simulated from sampling from their
296 respective experimental data distributions) could be connected in the network. These simulated
297 populations therefore reflect a relatively diverse population. We then varied which genetic
298 background and sex combination served as the index case (Table 1). We conducted 500 replicates per
299 index case type. For each recorded replicate, the traits of the simulated population were resampled,
300 and a new network was generated.

301

302 **Effects of the index case were outweighed by heterogeneity in the susceptible population.**

303 The genetic background or sex of the index case did not alter outbreak dynamics in diverse
304 populations where 20 experimental treatment groups (all genotype by sex combinations) were equally
305 sampled to create a heterogeneous population (Figure 3 & S2). This was true for all outbreak

306 parameters (Figure 3 & S2). Based on the random forest analysis, threshold radius and transmission
307 efficiency were the top two predictors for fadeout probability, maximum number of infected
308 individuals, outbreak duration, and R_0 (Figure S2).

309

310 **Theoretical simulation #3:** To determine the relative importance of experimentally observed
311 variation in social aggregation, viral shedding, and disease-related mortality on disease transmission
312 in a heterogeneous population, we simulated heterogeneous populations derived from the variation
313 seen across all genetic backgrounds and both sexes. To determine the effect of population-level
314 variation, we iteratively constrained the variation in each three host traits to the population's mean.
315 During these simulations, the unconstrained traits were free to vary according to their experimentally
316 determined distributions (Table 1). For example, to understand at the effect of variation in social
317 aggregation in isolation, we constrained social aggregation to take on the experimentally determined
318 mean degree distribution of the entire heterogeneous population but allowed viral shedding and
319 mortality rate to vary according to their experimentally-measured distributions across all genetic
320 backgrounds and both sexes. In the case of degree of the network, we rounded this value to ensure a
321 whole number, which is essential for contact network formation (e.g., an individual cannot have 2.5
322 contacts). We also considered interactions between variability of these three traits (Table 1).

323

324 **Variation in infectiousness increased fadeout probability and decreased maximum prevalence of**
325 **successful outbreaks, but increased outbreak duration.**

326 Constraining the infectiousness of a population to the mean (0.23, 0.46 for scaled infectiousness (η)
327 levels 1 and 2, respectively) of the experimentally measured distribution increased the outbreak
328 severity (Figure 4a), made outbreaks 2-fold more likely (Figure 4b), more than doubled the maximum
329 prevalence (Figure 4a,c), and persisted in the population for longer (Figure 4a,d). Limiting variation
330 in infectiousness also made outbreaks more predictable, reducing the variance of the time taken to
331 reach the maximum number of infected individuals (Figure 4d). According to the random forest

332 analysis, variation in infectiousness was the top predictor for whether or not an outbreak spread
333 beyond the initially infected individual (Figure S3).

334

335 **Variation in social aggregation did not influence fadeout probability but made outbreaks more**
336 **severe**

337 When social network degree distribution of simulated populations was confined to the mean of the
338 experimental data (2, 3 and 4 for threshold radii of 10, 15 and 20mm respectively), outbreaks became
339 less severe (Figure 3a) compared to simulations based on the complete degree distribution. Simulated
340 DCV spread to fewer individuals (Figure 4c) and was quicker to die-out than in simulations where
341 infectiousness, social aggregation, and mortality varied freely (Figure 4d).

342

343 **Variation in disease-related mortality did not affect epidemic outcomes.**

344 When constrained to the mean of the experimental data (13.6 days), we found disease-related
345 mortality had little to no effect on any aspect of disease outbreak (Figure 4). This is supported by the
346 random forest analysis which identified variation in mortality rate as the least important predictor
347 across outbreak metrics (Figure S3).

348

349 **Variation in infectiousness, followed by social aggregation, was the most influential component**
350 **of transmission.**

351 An increase in the maximum number of infected individuals was only seen when variation in
352 infectiousness was constrained. Interestingly the same effect was seen in simulations where other
353 traits are constrained alongside virus shedding, despite this differing substantially from the effects of
354 social aggregation and mortality rate when constrained alone (Figures 4 & S3). A similar, overruling
355 effect was seen when social aggregation and mortality rate were constrained simultaneously, and virus
356 shedding varied freely; outbreak dynamics were similar to the cases where only aggregation is
357 constrained (Figures 4 & S3).

358

359

360 Discussion

361 Here, we investigated how host genetic background and sex may contribute to the variance in social
362 aggregation, infectiousness and mortality and how this variation may scale up to population level
363 disease dynamics. We found substantial between-individual differences in pathogen transmission,
364 constituting genetic and sex-specific variation in transmission potential. Crucially, in relatively
365 homogenous populations comprised of single sex and genotype combinations, heterogeneity in the
366 index case produced major differences in population-level outbreak dynamics, including making
367 outbreaks more likely, broader reaching, and longer lasting. However, variation in the index case's
368 transmission potential exerted little influence over population-level outbreak dynamics in diverse host
369 populations. We also found that population-level variation in social aggregation, virus shedding, and
370 disease-related mortality affected outbreak dynamics in starkly contrasting ways. This effect appeared
371 to be linked to the population-level distribution of each respective host trait, with factors such as
372 skewness and zero-inflation influencing how variation in each trait affected outbreak dynamics.

373

374 In simulation experiment #1, males from the RAL-818 genetic background were not only more likely
375 to start an outbreak of infectious disease, but these outbreaks were also more severe than in other
376 populations. This suggests these males represent a class of individuals with a high transmission risk.
377 Interestingly, high-risk males are seen in a number of host-pathogen systems [35,36]. While high-risk
378 male classes can be produced by a range of traits pertaining to sex-specific ecology or physiology,
379 their occurrence across systems is likely driven by sexual selection shaping male traits affecting
380 transmission [37]. For example, in the yellow-necked mouse, *Apodemus flavicollis*, males are thought
381 to be a high-risk class due to a range of sex differences in their immune response, home range and
382 contact rates [35]. Moreover, as male *Drosophila* exhibit a number of other traits with the potential to
383 alter their transmission potential, such as male-male fighting [38], the transmission risk of RAL-818
384 males could increase further. Focussing on classes of high-risk individuals is a more pragmatic
385 approach to reducing the effect of heterogeneity in transmission potential, requiring less intensive
386 monitoring protocols [4]. Additionally, as classes of individuals are identified using ranges of
387 physiological or behavioural traits, classes are potentially more generalisable to other host-pathogen

388 systems (e.g. sex, social dominance). Many studies of transmission heterogeneity in natural systems
389 focus on using either behavioural or physiological traits to infer transmission dynamics and identify
390 high-risk individuals [2,4]. Our results highlight the importance of disentangling the relative
391 contributions made by behavioural and physiological traits together in order to infer variation in
392 transmission potential.

393

394 High-risk individuals, such as superspreaders, present a challenge to current methods of disease
395 control because they are capable of starting outbreaks of infectious disease that are difficult to predict
396 and amplifying them once transmission begins [39,40]. Pre-emptively identifying high-risk
397 individuals is therefore a major aim of epidemiology and disease ecology. However, in the second
398 theoretical experiment we conducted, we found that starting outbreaks with individuals that differed
399 in transmission potential did not affect outbreak dynamics when susceptible populations are
400 genetically diverse. Our results therefore suggest outbreaks are not solely driven by the traits of rare,
401 high-risk individuals, but are also affected by the traits of the susceptible population. High-risk
402 individuals were unable to cause explosive outbreaks of infectious disease when surrounded by low-
403 risk individuals as presumably, once infected, low-risk individuals failed to transmit disease to the rest
404 of the population. Similar transmission dynamics have also been observed in laboratory populations
405 of the social spider, *Stegodyphus dumicola*, where transmission of a bacterial pathogen was affected
406 by the boldness of the index case and the individuals it interacted with [14], but ultimately traits of the
407 index case did not alter transmission dynamics compared to the collective traits of the susceptible
408 population. Together with our results, these findings do not suggest diversity in the susceptible
409 population is a universal buffer to the effects of between-individual heterogeneity in disease
410 transmission. Instead, this work highlights the necessity to characterise population diversity in the
411 context of social interactions and networks as these may determine the relevance of this diversity.
412 Population-level diversity is particularly important in host-pathogen systems where behavioural
413 changes occur following infection. In populations of the guppy, *Poecilia reticulata*, for example,
414 male, but not female, sociality has been shown to increase following infection. As a result, females
415 social males are more likely to interact with, and infect, females [12]. There are many traits across

416 species that bias social interactions, such as sexual receptivity or personality type [41]. Should these
417 traits bias contact between transmission classes, this may explain why social and transmission
418 networks rarely match.

419

420 Extreme phenotypes often play a key role in between-individual heterogeneity in disease
421 transmission. However, being a relative term, ‘extreme’ phenotypes are defined by population-level
422 variation. Constraining population-level variation in the amount of virus shed following infection to
423 the population mean increased outbreak likelihood and severity. This was likely a result of the huge
424 zero-inflation of the distribution of virus shedding, where many infected individuals did not shed
425 virus. These individuals, previously termed ‘supersponges’ [42], represent the left-most extreme of
426 the population distribution, and bore no transmission risk. While some of the individuals that do not
427 transmit infection may simply not get any transmission opportunity, others may be supersponges and
428 therefore incapable of transmitting disease. The presence of supersponges also demonstrates the
429 importance of measuring variation in both behavioural and physiological traits when seeking to
430 understand heterogeneity in disease transmission. Characterising extreme forms of population-level
431 variation, particularly in natural systems where experiments are less controlled, should certainly be
432 prioritised in order to understand individual heterogeneity in disease transmission.

433

434 An important caveat of our results is that because we did not measure social aggregation, virus
435 shedding and lifespan simultaneously we cannot account for how they might covary within
436 individuals. We therefore allow them to co-occur in hosts randomly, which may not reflect
437 associations produced in nature or potential combinations of traits that are not likely due to
438 physiological or evolutionary constraints. This is particularly true for how we estimated contact
439 behaviour from social aggregation arenas containing 10-12 flies and measuring 55mm wide. For our
440 simulations, we scaled-up these smaller populations to create theoretical populations of 1000
441 individuals. This approach was required by the experimental demands of measuring social
442 aggregation, although it is known that social aggregation changes may change with population size
443 and sex ratio [43,44].

444

445 Threshold radius was a singularly important parameter across our theoretical experiment.
446 Understanding how distance affects pathogen transmission or definitions of what constitutes a contact
447 remains poorly described in many host-pathogen systems [7]. Moreover, real networks may have
448 different structures not accounted for here, such as a modular structure which has been shown to
449 facilitate or prevent the spread of disease [44,45]. As our social aggregation data comes from Petri
450 dishes containing only males or females from a single genetic background, we cannot account for how
451 aggregation might change in more diverse and larger populations[43].

452

453 Our work bears a number of consequences for understanding how between-individual heterogeneity
454 in disease transmission is determined and how it could affect outbreak dynamics. We show that
455 variation in key individual traits can dramatically affect population-level transmission, surmounting to
456 genetic and sex-specific variation in transmission potential. Importantly, the influence of this variation
457 is dramatically affected by susceptible population diversity and the distribution of population-level
458 variation. These results support the observations of other systems that suggest the traits of susceptible
459 individuals can exert significant influence over transmission. This is particularly relevant to
460 populations with low genetic diversity, such as agricultural monocultures, as this lack of diversity
461 increases the risk of explosive outbreaks [46]. Our work posits the merits of integrating data collected
462 in highly controlled laboratory experiments with simulations capable of extrapolating this information
463 to larger populations.

464

465 **Funding**

466 J.A.S-J was funded by a NERC E3 DTP PhD studentship awarded to the University of Edinburgh.
467 L.A.W. was funded by the National Science Foundation (GRFP-00039202 and DEB-1701069), the
468 University of Minnesota Informatics Institute, and the National Socio-Environmental Synthesis
469 Center (SESYNC) under funding received from the National Science Foundation DBI-1639145.
470 J.A.S-J and L.A.W. received funding for a research exchange from the Infectious Disease Evolution
471 Across Scales RCN funded by NSF. M.E.C. was funded by National Science Foundation (DEB-

472 1413925, 1654609, and 2030509). The authors acknowledge the Minnesota Supercomputing Institute
473 (MSI) at the University of Minnesota for providing resources that contributed to the research results
474 reported within this paper. URL: <http://www.msi.umn.edu>. P.F.V was supported by a Branco Weiss
475 fellowship (<https://brancoweissfellowship.org/>) and a Chancellor's Fellowship (School of Biological
476 Sciences, University of Edinburgh).

477

478 **References**

- 479 1. Craft ME. 2015 Infectious disease transmission and contact networks in wildlife and
480 livestock. *Philos. Trans. R. Soc. Lond. B. Biol. Sci.* **370**, 20140107-
481 (doi:10.1098/rstb.2014.0107)
- 482 2. White LA, Forester JD, Craft ME. 2017 Using contact networks to explore mechanisms of
483 parasite transmission in wildlife. *Biol. Rev.* **92**, 389–409. (doi:10.1111/brv.12236)
- 484 3. Lloyd-Smith JO, Schreiber S, Kopp P, Getz W. 2005 Superspreading and the effect of
485 individual variation on disease emergence. *Nature* **438**, 355–359.
486 (doi:10.1038/nature04153)
- 487 4. VanderWaal KL, Ezenwa VO. 2016 Heterogeneity in pathogen transmission: mechanisms
488 and methodology. *Funct. Ecol.* **30**, 1606–1622. (doi:10.1111/1365-2435.12645)
- 489 5. McCallum H *et al.* 2017 Breaking beta: deconstructing the parasite transmission
490 function. *Philos. Trans. B* **372**, 20160084. (doi:10.1098/rstb.2016.0084)
- 491 6. Hawley DM, Altizer SM. 2011 Disease ecology meets ecological immunology:
492 understanding the links between organismal immunity and infection dynamics in natural
493 populations. *Funct. Ecol.* **25**, 48–60. (doi:10.1111/j.1365-2435.2010.01753.x)
- 494 7. White LA, Forester JD, Craft ME. 2018 Covariation between the physiological and
495 behavioral components of pathogen transmission: host heterogeneity determines
496 epidemic outcomes. *Oikos* **127**, 538–552. (doi:10.1111/oik.04527)
- 497 8. Cousineau SV, Alizon S. 2014 Parasite evolution in response to sex-based host
498 heterogeneity in resistance and tolerance. *J. Evol. Biol.* **27**, 2753–2766.
499 (doi:10.1111/jeb.12541)
- 500 9. Vale Pedro F., Choisy Marc, Little Tom J. 2013 Host nutrition alters the variance in
501 parasite transmission potential. *Biol. Lett.* **9**, 20121145. (doi:10.1098/rsbl.2012.1145)
- 502 10. Shocket MS, Strauss AT, Hite JL, Šljivar M, Civitello DJ, Duffy MA, Cáceres CE, Hall SR.
503 2018 Temperature Drives Epidemics in a Zooplankton-Fungus Disease System: A Trait-
504 Driven Approach Points to Transmission via Host Foraging. *Am. Nat.* **191**, 435–451.
505 (doi:10.1086/696096)

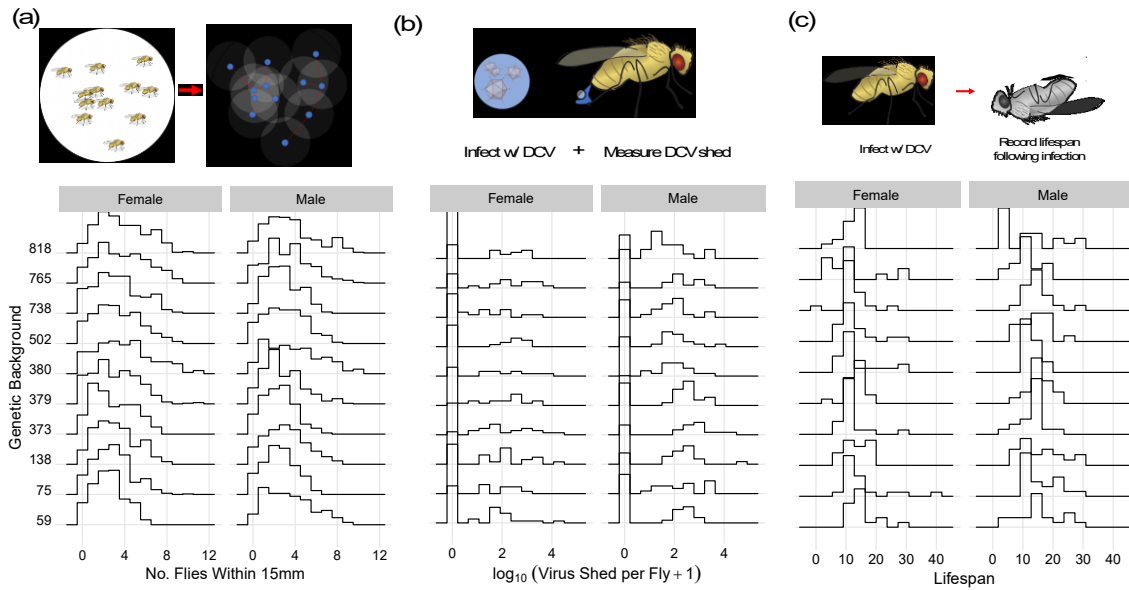
- 506 11. Susi H, Vale PF, Laine A-L. 2015 Host Genotype and Coinfection Modify the Relationship
507 of within and between Host Transmission. *Am. Nat.* **186**, 252–263.
508 (doi:10.1086/682069)
- 509 12. Stephenson JF. 2019 Parasite-induced plasticity in host social behaviour depends on sex
510 and susceptibility. *Biol. Lett.* **15**, 20190557. (doi:10.1098/rsbl.2019.0557)
- 511 13. Siva-Jothy JA, Vale PF. 2019 Viral infection causes sex-specific changes in fruit fly social
512 aggregation behaviour. *Biol. Lett.* , 630913. (doi:10.1101/630913)
- 513 14. Keiser CN, Pinter-Wollman N, Augustine DA, Ziemba MJ, Hao L, Lawrence JG, Pruitt JN.
514 2016 Individual differences in boldness influence patterns of social interactions and the
515 transmission of cuticular bacteria among group-mates. *Proc. R. Soc. B Biol. Sci.* **283**,
516 20160457–20160457. (doi:10.1098/rspb.2016.0457)
- 517 15. Susi H, Barrès B, Vale PF, Laine A-L. 2015 Co-infection alters population dynamics of
518 infectious disease. *Nat. Commun.* **6**. (doi:10.1038/ncomms6975)
- 519 16. Lloyd-Smith JO, George D, Pepin KM, Pitzer VE, Pulliam JRC, Dobson AP, Hudson PJ,
520 Grenfell BT. 2009 Epidemic dynamics at the human-animal interface. *Science* **326**, 1362–
521 1367. (doi:10.1126/science.1177345)
- 522 17. Siva-Jothy JA, Vale PF. 2019 Dissecting genetic and sex-specific host heterogeneity in
523 pathogen transmission potential. *bioRxiv* , 733915. (doi:10.1101/733915)
- 524 18. Kapun M, Nolte V, Flatt T, Schlötterer C. 2010 Host Range and Specificity of the
525 *Drosophila C Virus*. *PLoS ONE* **5**, e12421. (doi:10.1371/journal.pone.0012421)
- 526 19. Webster CL *et al.* 2015 The Discovery, Distribution, and Evolution of Viruses Associated
527 with *Drosophila melanogaster*. *PLOS Biol* **13**, e1002210.
528 (doi:10.1371/journal.pbio.1002210)
- 529 20. Arnold PA, Johnson KN, White CR. 2013 Physiological and metabolic consequences of
530 viral infection in *Drosophila melanogaster*. *J. Exp. Biol.* **216**, 3350–3357.
531 (doi:10.1242/jeb.088138)
- 532 21. Chtarbanova S *et al.* 2014 *Drosophila C virus* systemic infection leads to intestinal
533 obstruction. *J. Virol.* (doi:10.1128/JVI.02320-14)
- 534 22. Gupta V, Stewart CO, Rund SSC, Monteith K, Vale PF. 2017 Costs and benefits of
535 sublethal *Drosophila C virus* infection. *J. Evol. Biol.* **30**, 1325–1335.
536 (doi:10.1111/jeb.13096)
- 537 23. Siva-Jothy JA, Monteith KM, Vale PF. 2018 Navigating infection risk during oviposition
538 and cannibalistic foraging in a holometabolous insect. *Behav. Ecol. Off. J. Int. Soc. Behav.*
539 *Ecol.* **29**, 1426–1435. (doi:10.1093/beheco/ary106)
- 540 24. Anderson R, May R. 1991 *Infectious diseases of humans*. Oxford: Oxford University
541 Press.

- 542 25. Siettos CI, Russo L. 2013 Mathematical modeling of infectious disease dynamics.
543 *Virulence* **4**, 295–306. (doi:10.4161/viru.24041)
- 544 26. Mackay TFC *et al.* 2012 The *Drosophila melanogaster* Genetic Reference Panel. *Nature*
545 **482**, 173–8. (doi:10.1038/nature10811)
- 546 27. Huszar T, Imler J. 2008 *Drosophila* Viruses and the Study of Antiviral Host-Defense. In
547 *Advances in Virus Research*, pp. 227–265. Academic Press.
- 548 28. Webster CL *et al.* 2015 The Discovery, Distribution, and Evolution of Viruses Associated
549 with *Drosophila melanogaster*. *PLoS Biol* **13**, e1002210–e1002210.
550 (doi:10.1371/journal.pbio.1002210)
- 551 29. Kapun M, Nolte V, Flatt T, Schlötterer C. 2010 Host Range and Specificity of the
552 *Drosophila C* Virus. *PLoS ONE* **5**. (doi:10.1371/journal.pone.0012421)
- 553 30. Csardi G, Nepusz T. 2006 The igraph software package for complex network research.
554 *InterJournal Complex Systems*, 1695.
- 555 31. Lloyd-Smith JO, Cross PC, Briggs CJ, Daugherty M, Getz WM, Latto J, Sanchez MS, Smith
556 AB, Swei A. 2005 Should we expect population thresholds for wildlife disease? *Trends*
557 *Ecol. Evol.* **20**, 511–519. (doi:10.1016/j.tree.2005.07.004)
- 558 32. Cutler DR, Edwards TC, Beard KH, Cutler A, Hess KT, Gibson J, Lawler JJ. 2007 Random
559 Forests for Classification in Ecology. *Ecology* **88**, 2783–2792. (doi:10.1890/07-0539.1)
- 560 33. Breiman L. 2001 Random forests. *Mach. Learn.* **45**, 5–32.
561 (doi:10.1023/A:1010933404324)
- 562 34. Ostfeld RS, Keesing F. 2012 Effects of Host Diversity on Infectious Disease. *Annu. Rev.*
563 *Ecol. Evol. Syst.* **43**, 157–182. (doi:10.1146/annurev-ecolsys-102710-145022)
- 564 35. Ferrari N, Cattadori IM, Nespereira J, Rizzoli A, Hudson PJ. 2004 The role of host sex in
565 parasite dynamics: field experiments on the yellow-necked mouse *Apodemus flavicollis*.
566 *Ecol. Lett.* **7**, 88–94. (doi:10.1046/j.1461-0248.2003.00552.x)
- 567 36. Grear DA, Perkins SE, Hudson PJ. 2009 Does elevated testosterone result in increased
568 exposure and transmission of parasites? *Ecol. Lett.* **12**, 528–537. (doi:10.1111/j.1461-
569 0248.2009.01306.x)
- 570 37. Zuk M, McKean KA. 1996 Sex differences in parasite infections: Patterns and processes.
571 *Int. J. Parasitol.* **26**, 1009–1024. (doi:10.1016/S0020-7519(96)80001-4)
- 572 38. Baxter CM, Barnett R, Dukas R. 2015 Aggression, mate guarding and fitness in male fruit
573 flies. *Anim. Behav.* **109**, 235–241. (doi:10.1016/j.anbehav.2015.08.023)
- 574 39. Craft ME, Caillaud D. 2011 Network Models: An Underutilized Tool in Wildlife
575 Epidemiology? *Interdiscip. Perspect. Infect. Dis.* (doi:10.1155/2011/676949)

- 576 40. Keiser CN, Pinter-Wollman N, Ziemba MJ, Kothamasu KS, Pruitt JN. 2017 The index case
577 is not enough: Variation among individuals, groups and social networks modify bacterial
578 transmission dynamics. *J. Anim. Ecol.* , 1–10. (doi:10.1111/1365-2656.12729)
- 579 41. Keiser CN, Howell KA, Pinter-Wollman N, Pruitt JN. 2016 Personality composition alters
580 the transmission of cuticular bacteria in social groups. *Biol. Lett.* **12**, 20160297.
581 (doi:10.1098/rsbl.2016.0297)
- 582 42. Barron DG, Gervasi SS, Pruitt JN, Martin LB. 2015 Behavioral competence: How host
583 behaviors can interact to influence parasite transmission risk. *Curr. Opin. Behav. Sci.* **6**,
584 35–40. (doi:10.1016/j.cobeha.2015.08.002)
- 585 43. Keiser CN, Rudolf VHW, Sartain E, Every ER, Saltz JB. 2018 Social context alters host
586 behavior and infection risk. *Behav. Ecol.* **29**, 869–875. (doi:10.1093/beheco/ary060)
- 587 44. Nunn CL, Jordán F, McCabe CM, Verdolin JL, Fewell JH. 2015 Infectious disease and
588 group size: more than just a numbers game. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **370**.
- 589 45. Sah P, Leu ST, Cross PC, Hudson PJ, Bansal S. 2017 Unraveling the disease consequences
590 and mechanisms of modular structure in animal social networks. *Proc. Natl. Acad. Sci.*
591 **114**, 4165–4170. (doi:10.1073/pnas.1613616114)
- 592 46. Wallace R, Wallace RG. 2015 Blowback: new formal perspectives on agriculturally driven
593 pathogen evolution and spread. *Epidemiol. Infect.* **143**, 2068–2080.
594 (doi:10.1017/S0950268814000077)
- 595 47. Siva-Jothy JA, White LA, Craft ME, Vale PF. 2019 Population-Level Disease Dynamics
596 Reflect Individual Heterogeneities in Transmission. *bioRxiv* , 735480.
597 (doi:[10.1101/735480](https://doi.org/10.1101/735480))

598
599

600



601

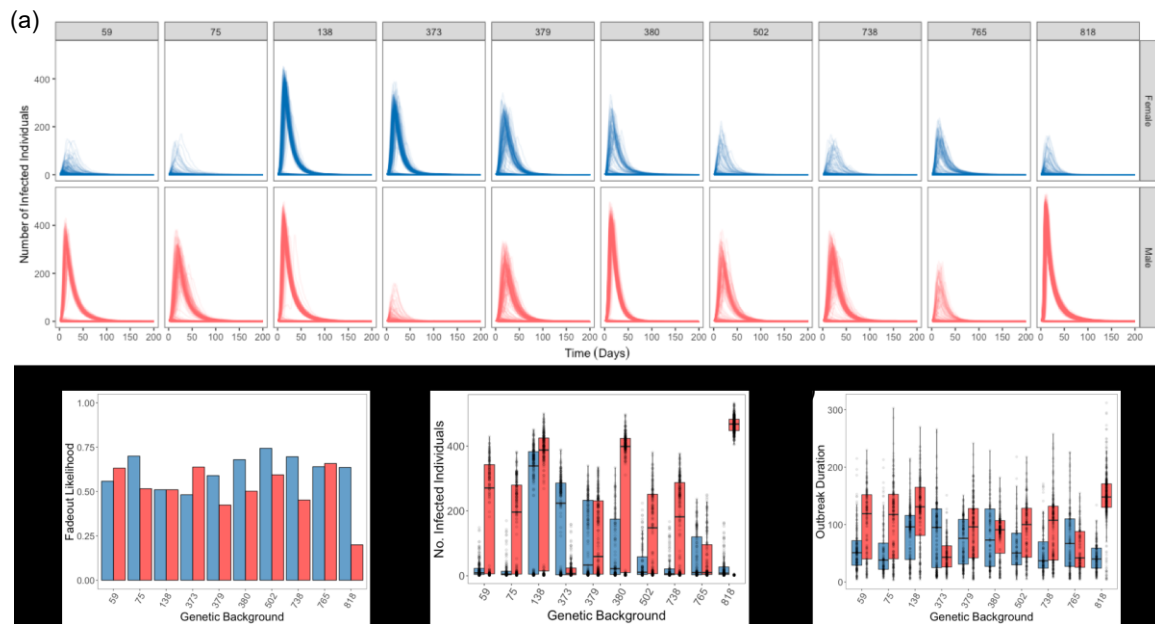
602 **Figure 1** – The epidemiological model was parameterised by sampling frequency distributions of
 603 experimental data collected from female and male ten *Drosophila* Genetic Reference Panel lines
 604 infected with DCV published previously [13,17]. Here, we provide a qualitative description of these
 605 data. a) social aggregation: the average number of neighbouring flies present within a 15mm radius
 606 of each focal fly; b) infectiousness: the number of viral copies shed per fly within the first 3 days
 607 following infected, as measured by DCV-specific qPCR; c) the day of death of each individual
 608 infected fly. Detailed analysis showing extensive line-by-sex interactive effects are reported in
 609 [13,17].

610

611

612

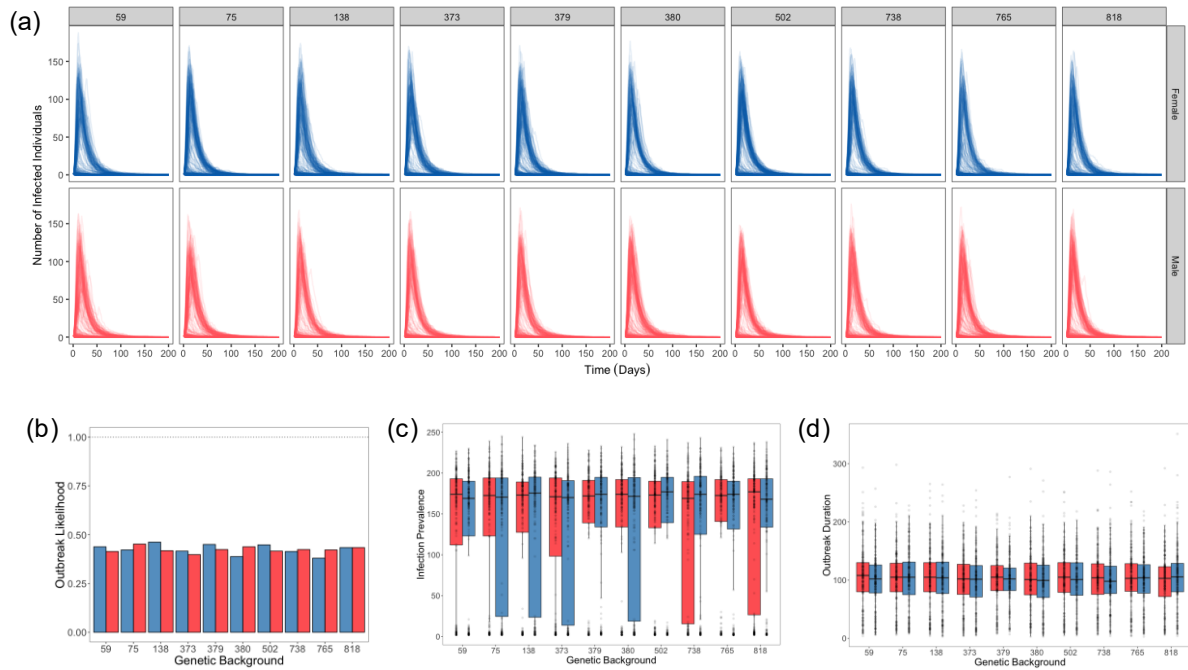
613



614

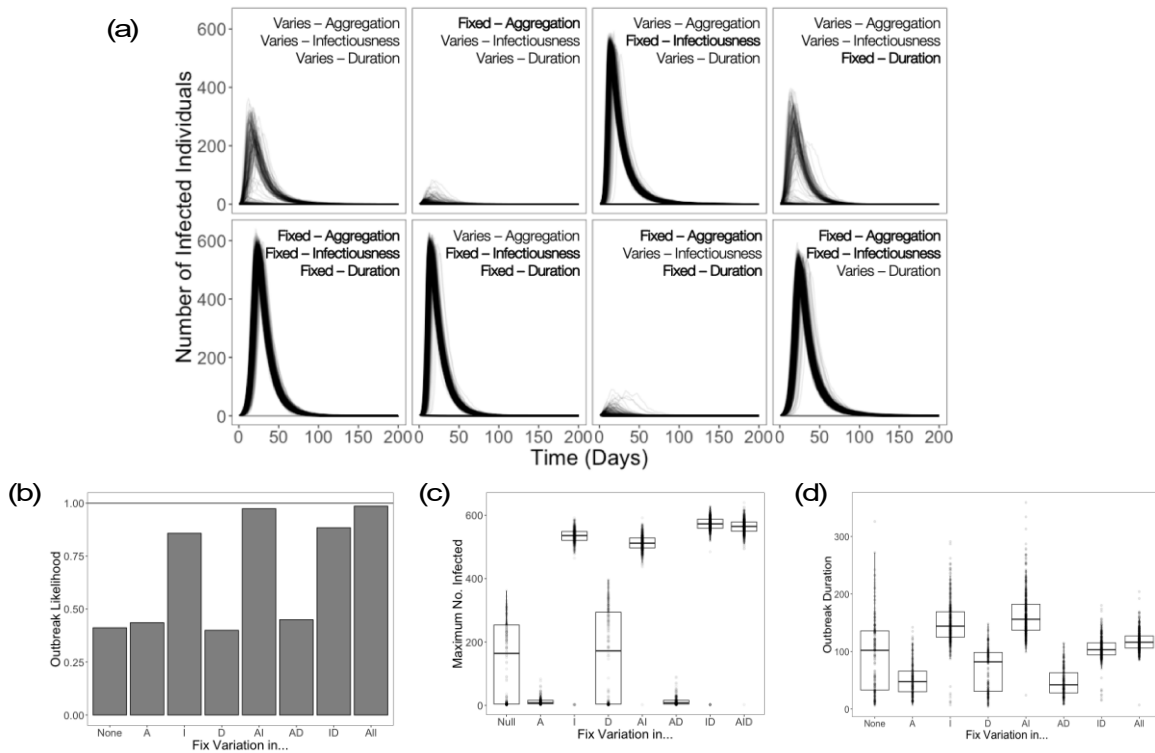
615 **Figure 2** – a) Simulation time courses of populations comprised of either male (red) or female (blue)
 616 individuals of the same sex and genetic background (columns) for simulation experiment #1. Across
 617 all of these simulations, parameters outside of host genetic background and sex are fixed; threshold
 618 radius (r)= 15mm, transmission efficiency (τ)=1 and scaled infectiousness (η)=2. (b-d) Summary
 619 statistics of simulations of populations comprised of male (red) or female (blue) individuals of the
 620 same genetic background (x-axis) for (b) the proportion of simulations that resulted in fadeout; and, in
 621 the subset of simulations where fadeout did not occur and disease spread from the index case; (c) the
 622 maximum number of infected individuals at any given time step; and (d) the number of time steps
 623 infected by the index case. Shown for threshold radius (r)= 15mm, transmission efficiency (τ)=1 and
 624 scaled infectiousness (η)=2. A random forest analysis was used to determine the relative importance
 625 of genetic background and sex to each summary statistic used to describe outbreak dynamics (Figure
 626 S1).

627
628
629



630
631
632
633
634
635
636
637
638
639

Figure 3 – Simulation time courses of populations comprised of all ten genetic backgrounds and males (red), and females (blue) in equal proportion, where the index case of an outbreak is an individual of a specific genetic background and sex (simulation experiment #2). Across all of these simulations, other parameters are fixed: threshold radius (r)= 15mm, transmission efficiency (τ)=1 and scaled infectiousness (η)=2. A random forest analysis was used to determine the relative importance of genetic background and sex to each summary statistic used to describe outbreak dynamics (Figure S2).



640

641 **Figure 4** – a) Simulation time courses of populations where aggregation, infectiousness and duration

642 variation are derived from the entire population's variation rather than for a single genetic line and sex

643 combination (simulation experiment #3). In each panel, the variation of a particular set of components

644 is confined to the population's mean. Across all of these simulations, parameters outside of host

645 genetic background and sex are fixed: threshold radius = 15mm, transmission efficiency =1 and

646 scaled infectiousness =2. **(b-d)** Summary statistics of time course simulations where individual

647 variation is determined by the variation seen across all genetic backgrounds and sexes (simulation

648 experiment #3). The x-axis of all panels sees variation in aggregation (A), infectiousness (I) and

649 mortality rate (D), and all their combinations fixed to the population mean. Outbreak metrics include:

650 b) the proportion of simulations that resulted in fadeout; c) the maximum number of individuals

651 infected during the simulation; and d) the time until maximum prevalence was reached. Shown for

652 threshold radius = 15mm, transmission efficiency =1, and scaled infectiousness =2. A random forest

653 analysis was used to determine the relative importance of genetic background and sex to each

654 summary statistic used to describe outbreak dynamics (Figure S3).

655

656

Parameter	Levels	Simulation 1	Simulation 2	Simulation 3
Population genetic background	RAL-59, RAL-75, RAL-138, RAL-373, RAL-379, RAL-380, RAL-502, RAL-738, RAL-765, RAL-818	X		
Population sex	Female, Male	X		
Index genetic background	RAL-59, RAL-75, RAL-138, RAL-373, RAL-379, RAL-380, RAL-502, RAL-738, RAL-765, RAL-818		X	
Index sex	Female, Male		X	
Threshold radius (r)	10mm, 15mm, 20mm	X	X	X
Pathogen transmission efficiency (τ)	0.1, 0.5, 1	X	X	X
Scaled infectiousness (η)	1, 2	X	X	X
Vary social aggregation	TRUE, FALSE			X
Vary infectiousness	TRUE, FALSE			X
Vary infection duration	TRUE, FALSE			X

657
658 **Table 1.** Parameters used to simulate outbreaks of infectious disease in simulations 1-3. Simulation 1
659 tested the effect of genetic and sex-specific variation in social aggregation, viral shedding and
660 susceptibility on population-level disease dynamics. Simulation 2 tested the effect of susceptible host
661 diversity on disease transmission potential. Simulation 3 tested the effect of variation in social
662 aggregation, infectiousness and infection duration on population-level disease transmission dynamics.
663 We conducted 500 replicates per parameter set with 1000 individuals in the network. Simulations
664 were allowed to run for 1000-time steps.