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## Draft Genome Sequence of the Putative Marine Pathogen *Thalassobius* sp. I31.1

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# Draft Genome Sequence of the Putative Marine Pathogen *Thalassobius* sp. I31.1

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**ABSTRACT** *Thalassobius* sp. I31.1 is a putative pathogen involved in epizootic shell disease in the American lobster (*Homarus americanus*). We report here the draft genome sequence for *Thalassobius* sp. I31.1 and provide insight into its metabolism and links to environmental pollutant degradation.

*Thalassobius* sp. I31.1 is a Gram-negative, aerobic, motile, rod-shaped bacterium. This genus is of the class *Alphaproteobacteria*, family *Rhodobacteraceae*, and order *Rhodobacterales*. *Thalassobius* sp. I31.1 was isolated from a lesion on the carapace of an American lobster (*Homarus americanus*) (1). Lesions are the characteristic symptom of epizootic shell disease (ESD), and *Thalassobius* has been indicated as a putative pathogen in lesion formation (1). Although *Thalassobius* strains may be unable to utilize various carbohydrates for growth (2), they can possess the ability to degrade the potential carcinogen phthalate, a chemical widely used in the manufacture of plastics, lubricants, and textiles (3). Such strains contain genes required for poly- $\beta$ -hydroxybutyrate (PHB) synthesis and genes relating to the degradation of phenylacetates, an abundant class of environmental pollutants (4).

*Thalassobius* sp. I31.1 was grown in artificial seawater (Instant Ocean) supplemented with yeast extract (1 g/liter) and peptone (5 g/liter) at 25°C on an elliptical shaker (New Brunswick) for 48 h. Genomic DNA was isolated with the Promega Wizard DNA purification kit, and DNA was resuspended in 2 mM Tris-HCl buffer (Bio Basic). DNA was quantified with a NanoDrop 1000 spectrophotometer (ND-1000) and checked for quality on a 1% agarose gel stained with ethidium bromide. DNA was sequenced on an Illumina MiSeq sequencer at the Genomics and Sequencing Center at the University of Rhode Island. Total genomic DNA was sheared with sonication (Covaris S220), and libraries were prepared with the SMARTer PrepX DNA Library kit on a SMARTer Apollo system (TaKaRa Bio, USA). Reads were trimmed with the CLC Genomic Workbench (version 9.5.3) for quality, ambiguous base pairs, adaptors, duplicates, and size with default parameters, resulting in 2,277,834 paired-end reads. The draft genome was assembled using the *de novo* assembly algorithm of SPAdes assembler (version 3.1.1) (5). Contigs with a coverage of >44 reads were processed with the CLC Microbial Genome Finishing module. The completed draft genome is composed of 65 contigs ( $N_{50}$  contig length, 182,141 bp), averaging 57,385 bp in size (total genome, 3,729,993 bp), with an average G+C content of 55.1%. The draft genome was annotated with the Rapid Annotations using Subsystem Technology (RAST) server and resulted in 3,749 open reading frames (6).

The annotated draft genome of *Thalassobius* sp. I31.1 encodes type I and II/IV secretion systems. RAST annotation identified a suite of *paa* genes involved in phenyl-

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acetate degradation, two alkane monooxygenase genes, *phbF*, a transcriptional regulator involved in PHB synthesis (7), and one toluenesulfonate dehydrogenase. The RAST-annotated draft genome was entered into Antibiotics and Secondary Metabolite Analysis Shell (antiSMASH) for secondary metabolite biosynthesis gene cluster analysis (8). Six clusters for secondary metabolism were identified, including those for homoserine lactone, bacteriocin, and lantipeptide-bacteriocin biosynthesis.

**Data availability.** This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number [PWAA00000000](https://www.ncbi.nlm.nih.gov/nuclseq/PWAA00000000). The version described in this paper is the first version, PWAA01000000. The SRA/DRA/ERA accession number is [PRJNA437474](https://www.ncbi.nlm.nih.gov/sra/PRJNA437474).

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## REFERENCES

- Chistoserdov AY, Smolowitz R, Mirasol F, Hsu A. 2005. Culture-dependent characterization of the microbial community associated with epizootic shell disease lesions in American lobster *Homarus americanus*. *J Shellfish Res* 24:741–747. [https://doi.org/10.2983/0730-8000\(2005\)24\[741:CCOTMC\]2.0.CO;2](https://doi.org/10.2983/0730-8000(2005)24[741:CCOTMC]2.0.CO;2).
- Rodrigo-Torres L, Pujalte MJ, Arahal DR. 2016. Draft genome sequence of *Thalassobius mediterraneus* CECT 5383<sup>T</sup>, a poly-beta-hydroxybutyrate producer. *Genom Data* 7:237–239. <https://doi.org/10.1016/j.gdata.2016.01.013>.
- Iwaki H, Nishimura A, Hasegawa Y. 2012. Isolation and characterization of marine bacteria capable of utilizing phthalate. *World J Microbiol Biotechnol* 28:1321–1325. <https://doi.org/10.1007/s11274-011-0925-x>.
- Teufel R, Mascaraque V, Ismail W, Voss M, Perera J, Eisenreich W, Haehnel W, Fuchs G. 2010. Bacterial phenylalanine and phenylacetate catabolic pathway revealed. *Proc Natl Acad Sci USA* 107:14390–14395. <https://doi.org/10.1073/pnas.1005399107>.
- Nurk S, Bankevich A, Antipov D, Gurevich AA, Korobeynikov A, Lapidus A, Prjibelski AD, Pyshkin A, Sirotkin A, Sirotkin Y, Stepanauskas R, Clingenpeel SR, Woyke T, McLean JS, Lasken R, Tesler G, Alekseyev MA, Pevzner PA. 2013. Assembling single-cell genomes and mini-metagenomes from chimeric MDA products. *J Comput Biol* 20:714–737. <https://doi.org/10.1089/cmb.2013.0084>.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.
- Kadowaki MAS, Müller-Santos M, Rego FGM, Souza EM, Yates MG, Monteiro RA, Pedrosa FO, Chubatsu LS, Steffens MBR. 2011. Identification and characterization of PhbF: a DNA binding protein with regulatory role in the PHB metabolism of *Herbaspirillum seropedicae* SmR1. *BMC Microbiol* 11:230. <https://doi.org/10.1186/1471-2180-11-230>.
- Weber T, Blin K, Duddela S, Krug D, Kim HU, Brucoleri R, Lee SY, Fischbach MA, Müller R, Wohlleben W, Breitling R, Takano E, Medema MH. 2015. antiSMASH 3.0—a comprehensive resource for the genome mining of biosynthetic gene clusters. *Nucleic Acids Res* 43:W237–W243. <https://doi.org/10.1093/nar/gkv437>.