## THE UNIVERSITY OF RHODE ISLAND

University of Rhode Island DigitalCommons@URI

**Biological Sciences Faculty Publications** 

**Biological Sciences** 

2020

# *Tomato Yellow Leaf Curl Virus* Infection Alters *Bemisia tabaci* MED (Hemiptera: Aleyrodidae) Vulnerability to Flupyradifurone

**Baiming Liu** 

Evan L. Preisser University of Rhode Island, preisser@uri.edu

Xiaoguo Jiao

Youjun Zhang

Follow this and additional works at: https://digitalcommons.uri.edu/bio\_facpubs

The University of Rhode Island Faculty have made this article openly available. Please let us know how Open Access to this research benefits you.

This is a pre-publication author manuscript of the final, published article.

Terms of Use

This article is made available under the terms and conditions applicable towards Open Access Policy Articles, as set forth in our Terms of Use.

### **Citation/Publisher Attribution**

Liu, B., Preisser, E. L., Jiao, X., & Zhang, Y. (2020). *Tomato Yellow Leaf Curl Virus* Infection Alters *Bemisia tabaci* MED (Hemiptera: Aleyrodidae) Vulnerability to Flupyradifurone. *Journal of Economic Entomology, 113*(4), 1922-1926. https://doi.org/10.1093/jee/toaa118 Available at: https://doi.org/10.1093/jee/toaa118

This Article is brought to you for free and open access by the Biological Sciences at DigitalCommons@URI. It has been accepted for inclusion in Biological Sciences Faculty Publications by an authorized administrator of DigitalCommons@URI. For more information, please contact digitalcommons@etal.uri.edu.

1	Youjun Zhang
2	Department of Entomology
3	Institute of Vegetables and Flowers
4	Chinese Academy of Agricultural Sciences
5	No. 12 Zhongguancun Nandajie
6	Haidian District, Beijing 100081, China
7	zhangyoujun@caas.cn
8	
9	TYLCV Infection Alters Bemisia tabaci MED (Hemiptera: Aleyrodidae) Vulnerability to
10	Flupyradifurone
11	
12	Baiming Liu <sup>1†</sup> , Evan L. Preisser <sup>2†</sup> , Xiaoguo Jiao <sup>3</sup> , and Youjun Zhang <sup>4</sup>
13	
14 15	<sup>1</sup> Institute of Plant Protection, Tianjin Academy of Agricultural Sciences, Tianjin 300381, China
16	<sup>2</sup> Department of Biological Sciences, University of Rhode Island, Kingston, RI 02881
17	USA
18	<sup>3</sup> State Key Laboratory of Biocatalysis and Enzyme Engineering, Center for Behavioral
19	Ecology & Evolution, School of Life Sciences, Hubei University, Wuhan, 430062, China
20	<sup>4</sup> Department of Entomology, Institute of Vegetables and Flowers, Chinese Academy of
21	Agricultural Sciences, Beijing 100081, China
22	†These authors contributed equally to this work.

#### 23 Abstract

The whitefly Bemisia tabaci (Hemiptera: Aleyrodidae) is a major phloem-feeding pest of 24 25 agricultural crops that is also an important vector of many plant diseases. The B. tabaci Mediterranean ('MED') biotype is a particularly effective vector of Tomato yellow leaf curl virus 26 (TYLCV), a devastating plant pathogen. While insecticides play an important role in the control 27 28 of MED and TYLCV, little is known about how TYLCV infection affects MED susceptibility to 29 insecticides. We conducted research addressing how MED susceptibility to flupyradifurone, the 30 first commercially available systemic control agent derived from the butenolide class of 31 insecticides, was affected by TYLCV infection. We first conducted bioassays determining the LC<sub>15</sub> and LC<sub>50</sub> for control and viruliferous MED feeding on either water- or insecticide-treated 32 plants. We next measured several demographic parameters of control and viruliferous MED 33 exposed to either insecticide- or water-treated plants. TYLCV infection increased MED tolerance 34 35 of flupyradifurone: the  $LC_{15}$  and  $LC_{50}$  of viruliferous MED were double that of uninfected MED. 36 Viral infection also altered MED demographic responses to flupyradifurone, but in an inconsistent manner. While the ability of TYLCV and other persistently-transmitted viruses to 37 benefit Bemisia via manipulation of host plant defense is well-known, this appears to be the first 38

39 example of virally-mediated changes in vector susceptibility to an insecticide.

40 Key Words

41 Insecticide, Sivanto, tolerance, *Bemisia*, TYLCV

#### 42 Introduction

The whitefly Bemisia tabaci Gennadius (Hemiptera: Aleyrodidae) is a major phloem-feeding 43 pest of both field and greenhouse crops worldwide (Stansly and Naranjo 2010). Its management 44 is complicated by the fact that *B. tabaci* contains over 30 phenotypically identical but genetically 45 distinct cryptic species (Liu et al. 2012, Hadjistylli et al. 2016) that vary widely in traits such as 46 47 insecticide resistance (Chen et al. 2016, Xie et al. 2017). Bemisia tabaci Mediterranean (MED) poses a particular threat to agriculture due to its invasiveness. Since its arrival in China in 2003 48 49 (Chu et al. 2006), it has displaced both native and invasive *B. tabaci* throughout the country (Teng et al. 2010). 50

Although *Bemisia* feeding can itself reduce plant growth, its primary threat to agriculture 51 occurs via its ability to transmit a wide variety of plant viruses. MED is particularly effective at 52 transmitting such viruses, and its invasion is often associated with plant disease outbreaks (Ning 53 et al. 2015). The *Tomato yellow leaf curl virus* (TYLCV) is a particularly damaging pathogen 54 55 that has caused significant damage worldwide (Jones 2003). TYLCV relies on B. tabaci as a vector to spread among plants (Fereres and Moreno 2009). As a result, Bemisia- plant-TYLCV 56 interactions have been the subject of intense interest and researchers have confirmed the 57 58 mutualistic relationship between B. tabaci and the virus. It is now known, for instance, that TYLCV can increase *Bemisia* fitness via its suppression of plant defense (Zhang et al. 2012, 59 60 Luan et al. 2013) and that MED benefits from feeding on TYLCV-infected hosts (Pan et al. 61 2013a, Shi et al. 2019).

Insecticides play an important role in an integrated pest management approach to
 controlling *Bemisia* and viral outbreaks in agricultural systems. Because the whitefly can rapidly
 develop insecticide resistance, the continued development and deployment of novel compounds

is essential for effective pest control. One such compound is flupyradifurone, the first 65 commercially available systemic control agent derived from the butenolide class of insecticides 66 67 (Nauen et al. 2015). This compound, an agonist on insect nicotinic acetylcholine receptors, differs structurally from other chemicals that target these receptors. As a result, it is effective 68 against neonicotinoid- and pymetrozine-resistant *Bemisia* populations (Nauen et al. 2015). 69 70 A recent assessment of MED survival and TYLCV transmission found that while flupyradifurone rapidly killed MED and reduced TYLCV transmission by 85%, treatment with 71 72 the neonicotinoid thiomethoxam only reduced viral transmission by 25% (Roditakis et al. 2017). 73 Other work confirming the general efficacy of flupyradifurone against B. tabaci Middle East-Asia Minor 1 (MEAM1) nonetheless found a few field populations with high levels of 74 flupyradifurone tolerance (Smith et al. 2016). While increasing insecticide tolerance in its vector 75 would clearly benefit TYLCV and similar viruses, there is no published research assessing 76 77 whether viruses can provide such benefits. Alternately, TYLCV could affect MED in a manner 78 similar to Rickettsia, which is correlated with increased *Bemisia* sensitivity to a range of different insecticides (Kontsedalov et al. 2008, but see Pan et al. 2013b). Understanding how 79 TYLCV affects the flupyradifurone tolerance of its vector is important to maximize the effective 80 81 use of this important insecticide.

We report the results of two experiments exploring how TYLCV infection affected MED susceptibility to flupyradifurone (trade name Sivanto). We first determined the  $LC_{15}$  and  $LC_{50}$  for control and viruliferous MED feeding on plants treated with either Sivanto or distilled water. We calculated both  $LC_{15}$  and  $LC_{50}$  because chemical degradation and dilution gradually reduce insecticide concentrations following application (e.g., Roditakis et al. 2017),. We next measured several demographic parameters of control and viruliferous MED exposed to either insecticideor water-treated plants. Although TYLCV infection has little direct effect on MED fitness (Pan et
al. 2013a, Su et al. 2015), recent work found a net downregulation of detoxification enzymes in
TYLCV-infected MED (Ding et al. 2019); we hypothesized that viruliferous MED would be
more sensitive to Sivanto than uninfected individuals.

92 Materials and Methods

Plants: Tomato plants (*Solanum lycopersicum* L, cv. Zhongza 9) were grown individually in twoliter pots in a greenhouse with natural lighting and controlled temperature (26±2°C). All plants
were grown in a 10:5:1 (by volume) mixture of peat moss, vermiculite, and organic 8-8-8
fertilizer. TYLCV-infected plants were produced with injection of *Agrobacterium tumefaciens*mediated TYLCV clones (Shanghai isolate) at the 3-4 true leaf stage (Zhang et al. 2009). The
plants were grown for four weeks post-injection to give them time to display infection-associated
pathological symptoms.

Insects: The whitefly *Bemisia tabaci* MED (Q) was first collected in 2009 from 100 101 poinsettia, Euphorbia pulcherrima Wild. (ex Klotz.), in Beijing, China. It was reared on poinsettia. In 2015, a portion of the population (~300 adults) was transferred to the Tianjin 102 Institute of Plant Protection and reared on cotton plants (Gossypium herbaceum L., cv DP99B) in 103 104 80 mesh nylon insect cages (45×45×60 cm) under 26±2°C, 60±10% RH, 14L:10D photoperiod. 105 A viruliferous MED population was produced by transferring ~300 whiteflies into a cage with four TYLCV-infected tomato plants; a parallel uninfected MED population was produced by 106 transferring >300 whiteflies into a cage with four healthy tomato plants. Both viruliferous and 107 uninfected MED were reared for two generations on their respective plants before being used for 108 109 experiments. Colony purity was monitored every 2-3 generations using a DNA marker (Khasdan 110 et al. 2005), and TYLCV infection was confirmed via PCR validation (Ghanim et al. 2007).

111	Flupyradifurone bioassay of viruliferous and uninfected MED: Sivanto 200SL (17.09%
112	flupyradifurone) was provided by Bayer Crop Science (China) Company Ltd. and diluted with
113	distilled water to five different concentrations: 200 mg[AI]kg <sup>-1</sup> , 100 mg[AI]kg <sup>-1</sup> , 50 mg[AI]kg <sup>-1</sup> ,
114	25 mg[AI]kg <sup>-1</sup> , and 12.5 mg[AI]kg <sup>-1</sup> . For each of the five concentrations and an additional
115	distilled water control (a total of six treatments), 200 mL was added to a 500 mL plastic spray
116	bottle. For each concentration, one spray bottle was used to spray four tomato plants that were
117	each at the 6-7 true leaf stage; plants were sprayed until drip-off. One day after spraying, 100
118	newly-emerged (within 24 hours) adult MED per plant were placed in clip cages attached to the
119	abaxial side of both the third and fourth leaves of each sprayed plant. Clip cages were kept on for
120	two days; the number of living and dead MED were then counted. This work was conducted in a
121	climate-controlled chamber at $26\pm1$ °C and $60\pm10\%$ RH with 14L:10D photoperiod.
122	Demographic responses of viruliferous and uninfected MED to flupyradifurone (LC <sub>15</sub> ):
123	Data from the above-mentioned experiment was used to calculate the $LC_{15}$ for viruliferous MED.
124	a solution of this concentration was sprayed on healthy tomato plants at the 6-7 true leaf stage
125	until drip-off. Another group of healthy tomato plants was sprayed to drip-off with distilled
126	water. After 24 hours, approximately 100 newly emerged (within one day) viruliferous or
127	uninfected MED were attached in separate clip cages to the abaxial side of the third and fourth
128	leaves of either a Sivanto-treated or control plant. This produced four treatments: TYLCV
129	(uninfected, viruliferous) crossed with insecticide (dH <sub>2</sub> O, Sivanto). This work were conducted in
130	a climate-controlled chamber at 26±1 $^\circ\!C$ and 60±10% RH with 14L:10D photoperiod. Clip cages
131	were removed after two days and the living and dead adult MED collected and counted.
132	Female adult longevity and first-week fecundity: Thirty female MED from each of the
133	four treatments (=120 total) were placed individually in clip cages. Each cage was then clipped

on the abaxial side of a middle leaf of an unsprayed healthy tomato plant (6-7 true leaf stage). A
total of two MED were clipped onto each plant, one per leaf, and both MED on a given plant had
the same infection status (i.e., they were both either uninfected or viruliferous). The clip cages
were checked each day for MED mortality; after one week, all surviving adults were individually
transferred to new unsprayed healthy tomato plants and the number of eggs laid during the first
week counted.

Egg-to-adult survival and developmental time: Five pairs of newly-emerged MED 140 (within one day; 1:1 sex ratio) from a given treatment were placed into a single clip cage and 141 clipped onto the abaxial side of a middle leaf of an unsprayed healthy tomato plant (6-7 true leaf 142 stage). Only one clip cage was attached to each plant. This was replicated 10 times in each of the 143 four treatments, for a total of 40 replicates. After one day, each clip cage was opened and the 144 adults were removed, leaving only the eggs and nymphs. Each clip cage was then inspected daily 145 146 and the number of nymphs and adults recorded. Daily inspections continued until the last nymph 147 had either entered adulthood or died.

Statistical analysis: Probit parameter estimation of the concentration-mortality response 148 for viruliferous and uninfected MED in the six concentrations were calculated using POLO-PC 149 150 (Russell et al. 1977, LeOra 1987). These parameters included LC<sub>15</sub> and LC<sub>50</sub> values expressed in mg[AI]kg-1 and their corresponding 95% confidence limit (CL) along with the slopes of the 151 152 probit regressions. Between-treatment differences in the mortality of viruliferous and uninfected 153 MED were calculated using 95% CLs; LC<sub>15</sub> or LC<sub>50</sub> values for viruliferous and uninfected MED were considered significantly different if their corresponding 95% CLs did not overlap. 154 Data on each of the demographic responses was analyzed using two-way ANOVA to 155 156 assess the main effects of TYLCV (uninfected, viruliferous) and insecticide (dH2O, Sivanto) as

well as their interaction. When one or more main effects or their interaction was significant at p 157 = 0.05, Tukeys' HSD was used for means separation tests. Data on adult longevity and survival 158 159 was sqrt transformed before analysis. All analyses were conducted using JMP 9.0.0 (SAS 2010). **Results** 160 Viruliferous MED were more tolerant of Sivanto than uninfected MED (Table 1). The 161 162  $LC_{15}$  of viruliferous MED was more than twice that of uninfected MED (11.8 versus 5.8, respectively), and the  $LC_{50}$  of viruliferous MED was almost twice as high (31.3 versus 17.3). 163 164 The 95% CLs of viruliferous and uninfected MED did not overlap, meaning that the two groups differed significantly in both their  $LC_{15}$  and  $LC_{50}$  values (Table 1). 165 Exposure to Sivanto (at LC<sub>15</sub> concentration determined for viruliferous MED) marginally 166 increased adult female longevity (Fig. 1A), increased first-week fecundity (Fig. 1B) and 167 decreased egg-adult development time (Fig. 1C) in both MED groups (Table 2). In contrast, the 168 only significant main effect of TYLCV was a 28% decrease in first-week fecundity (Fig. 1B). 169 170 The TYLCV\*Sivanto interaction was marginally significant (P = 0.067 - 0.085) for three of the four variables: Sivanto had a greater impact on the first-week fecundity and egg-adult 171 development time of uninfected MED than viruliferous MED (Fig. 1B, 1C), but increased the 172 173 adult female lifespan of viruliferous MED more than for uninfected MED (Fig. 1A). Survival from egg to adult (Fig. 1D) was not affected by either main effect or their interaction (Table 2). 174 Discussion 175 176 Contrary to expectations, we found that TYLCV did not increase MED vulnerability to flupyradifurone. Instead, both the LC<sub>15</sub> and LC<sub>50</sub> values for viruliferous MED were significantly 177 178 higher than those of uninfected MED (Table 1). In three of the four demographic variables, there 179 was also a marginally significant interaction between Sivanto and TYLCV: Sivanto tended to

increase adult longevity only in viruliferous MED and first-week fecundity only in uninfected
MED, and tended to decrease egg-adult development time only in uninfected MED (Fig.
1A,B,C). While the ability of TYLCV and other persistently-transmitted viruses to benefit *Bemisia* via manipulation of host plant defense is well-known, this appears to be the first
example of virally-mediated changes in vector susceptibility to an insecticide.

185 While our results were surprising, there have been other reports of microorganismmediated changes in insecticide susceptibility (Pietri and Liang 2018). Gut symbionts in both the 186 187 cigarette beetle Lasioderna serricorne (Shen and Dowd 1991) and the apple fly Rhagoletis pomonella (Lauzon et al. 2003) are involved with the detoxification of natural and synthetic 188 toxins. In contrast, the symbiotic microorganism Rickettsia increased Bemisia sensitivity to a 189 range of different insecticides (Kontsedalov et al. 2008, but see Pan et al. 2013b); later research 190 linked increases in *Bemisia* symbiont diversity and density to greater insecticide susceptibility 191 192 (Ghanim and Kontsedalov 2009). Similar results have been reported in the psyllid Diaphorina 193 *citri*, where infection with *Candidatus* Liberibacter asiaticus increased its vulnerability to several insecticides (Tiwari et al. 2011). A recent review (Pietri and Liang 2018) suggested these 194 variable results may partially reflect symbiont-specific effects on both host detoxification 195 196 enzymes and their immune/stress response. A transcriptomic analysis of gene regulation in TYLCV-infected MED found that while TYLCV generally downregulated detoxification 197 198 enzymes, genes involved in both stress and immune responses were upregulated (Ding et al. 199 2019). It seems likely that some of these upregulated genes alter MED susceptibility to 200 flupyradifurone.

The negative impact of flupyradifurone revealed in the LC<sub>15</sub> and LC<sub>50</sub> bioassays appears
at odds with its equivocal effect on various aspects of MED demography. MED that survived one

day of flupyradifurone exposure had slightly higher female longevity, higher first-week 203 fecundity, and a shorter egg-adult development time than MED in the control treatment. These 204 'benefits' of flupyradifurone are almost certainly an experiment artifact: a day of insecticide 205 exposure removed the weakest and/or most susceptible MED from the population that was 206 subsequently used for our demographic work. They may also reflect hormesis, a phenomenon in 207 208 which sublethal dosages of insecticide improve fecundity or provide other benefits to the targeted insects (Cutler 2012). It is also worth noting that both uninfected and viruliferous MED 209 210 were exposed to flupyradifurone at the  $LC_{15}$  concentration determined for viruliferous MED. 211 Because the LC<sub>15</sub> value for uninfected MED was lower than for viruliferous MED, this flupyradifurone concentration was more lethal to the uninfected population than to the 212 viruliferous one. Higher rates of exposure-related mortality in our uninfected group may have 213 had the unintended effect of minimizing differences between the uninfected and viruliferous 214 groups. It should also be noted that the recommended label rate of flupyradifurone, 150 mg/l, 215 216 was substantially higher than the concentrations we used; we chose to work with lower concentrations in order to assess MED that survive initial exposure. The effect of TYLCV on 217 MED insecticide tolerance may be reduced or eliminated at these higher concentrations. 218 219 Pesticides can indirectly control insect-vectored plant diseases via their impact on vector density. This control may be lessened, however, if vectors feeding on pesticide-sprayed plants 220 221 survive long enough to transmit TYLCV and other viruses. Viruliferous Bemisia efficiently 222 transmit TYLCV to uninfected plants. Less than two minutes of *Bemisia* salivation is necessary 223 to infect a healthy tomato plant (Jiang et al. 2000). As a result, thiamethoxam and other 224 insecticides that do not quickly kill Bemisia may prove inefficient at decreasing TYLCV

transmission (Roditakis et al. 2017). Flupyradifurone has a higher knockdown rate than 225 thiamethoxam and is more effective at reducing TYLCV transmission (Roditakis et al. 2017). 226 Research assessing the impact of flupyradifurone on *Bemisia* feeding behavior is 227 necessary to understand the mechanism(s) underlying its effect on viral transmission. Aphids 228 feeding on thiamethoxam-treated plants, for example, spend less time in the sieve element phase 229 230 required for viral transmission to an uninfected plant (Cho et al. 2011, Stamm et al. 2013). Although TYLCV increased MED tolerance to flupyradifurone (Table 1), it might still change 231 232 MED feeding behavior in ways that make this pesticide effective at reducing or eliminating viral 233 transmission. Alternately, TYLCV-linked increases in flupyradifurone tolerance may provide viruliferous MED an advantage over uninfected individuals in pesticide-treated fields. If so, 234 insecticide application could, under some conditions, favor viral outbreaks in agricultural 235 systems (Pan et al. 2015). 236

In summary, our work found that infection with TYLCV altered the susceptibility of *Bemisia tabaci* MED to flupyradifurone. While the mechanism underlying our results is unknown, our findings suggest that viral infection may be capable of changing population-level responses to current management practices. Even for novel insecticides, such interactions highlight how work exploring pesticide impacts on each part of the vector-virus-plant interaction can contribute to the development of effective strategies to control MED and TYLCV.

## 243 Acknowledgements

- 244 This work was funded by the National Natural Science Foundation of China (31772171,
- 245 31401785), Tianjin Natural Science Foundation (17JCZDJC33700), Innovative research and
- 246 experimental projects for young researchers of Tianjin Academy of Agricultural Science
- 247 (201903). The authors declare that no conflict of interest exists.

#### 248 **References Cited**

- 249 Chen, W., D. K. Hasegawa, N. Kaur, A. Kliot, P. V. Pinheiro, J. Luan, M. C. Stensmyr, Y.
- 250 Zheng, W. Liu, H. Sun, Y. Xu, Y. Luo, A. Kruse, X. Yang, S. Kontsedalov, G. Lebedev, T.
- 251 W. Fisher, D. R. Nelson, W. B. Hunter, J. K. Brown, G. Jander, M. Cilia, A. E. Douglas, M.
- 252 Ghanim, A. M. Simmons, W. M. Wintermantel, K.-S. Ling, and Z. Fei. 2016. The draft
- 253 genome of whitefly *Bemisia tabaci* MEAM1, a global crop pest, provides novel insights into
- virus transmission, host adaptation, and insecticide resistance. BMC Biol. 14: 110.
- 255 Chu, D., Y. J. Zhang, J. K. Brown, B. Cong, B. Y. Xu, Q. J. Wu, and G. R. Zhu. 2006. The
- introduction of the exotic Q biotype of *Bemisia tabaci* (Gennadius) from the Mediterranean
- region into China on ornamental crops. Fla Entomol 89: 168-174.
- 258 Cutler, G. C. 2012. Insects, insecticides and hormesis: evidence and considerations for study.
- 259 Dose-Response 11: 154-177.
- 260 Ding, T.-B., J. Li, E.-H. Chen, J.-Z. Niu, and D. Chu. 2019. Transcriptome profiling of the
- 261 whitefly *Bemisia tabaci* MED in response to single infection of tomato yellow leaf curl virus,
- tomato chlorosis virus, and their co-infection. Front. Physiol. 10: 302.
- **Fereres, A., and A. Moreno. 2009.** Behavioural aspects influencing plant virus transmission by
- homopteran insects. Virus Res. 141: 158-168.
- **Ghanim, M., and S. Kontsedalov. 2009.** Susceptibility to insecticides in the Q biotype of
- 266 *Bemisia tabaci* is correlated with bacterial symbiont densities. Pest Manag Sci 65: 939-942.
- **Ghanim, M., I. Sobol, M. Ghanim, and H. Czosnek. 2007.** Horizontal transmission of
- begomoviruses between *Bemisia tabaci* biotypes. Arthropod-Plant Interactions 1: 195-204.
- 269 Hadjistylli, M., G. K. Roderick, and J. K. Brown. 2016. Global population structure of a
- worldwide pest and virus vector: genetic diversity and population history of the *Bemisia tabaci*sibling species group. PLoS ONE 11: e0165105.
- Jiang, Y., C. de Blas, L. Barrios, and A. Fereres. 2000. Correlation between whitefly
- 273 (Homoptera: Aleyrodidae) feeding behavior and transmission of *tomato yellow leaf curl virus*.
- 274 Ann. Entomol. Soc. Am. 93: 573-579.
- Jones, D. 2003. Plant viruses transmitted by whiteflies. Eur. J. Plant Pathol. 109: 195-219.
- 276 Khasdan, V., I. Levin, A. Rosner, S. Morin, S. Kontsedalov, L. Maslenin, and A. R.
- 277 Horowitz. 2005. DNA markers for identifying biotypes B and Q of *Bemisia tabaci* (Hemiptera:
- Aleyrodidae) and studying population dynamics. Bull. Entomol. Res. 95: 605-613.
- 279 Kontsedalov, S., E. Zchori-Fein, E. Chiel, Y. Gottlieb, M. Inbar, and M. Ghanim. 2008. The
- 280 presence of *Rickettsia* is associated with increased susceptibility of *Bemisia tabaci* (Homoptera:
- Aleyrodidae) to insecticides. Pest Manag Sci 64: 789-792.
- Lauzon, C. R., S. E. Potter, and R. J. Prokopy. 2003. Degradation and detoxification of the
- dihydrochalcone phloridzin by *Enterobacter agglomerans*, a bacterium associated with the apple
- pest, *Rhagoletis pomonella* (Walsh) (Diptera: Tephritidae). Environ Entomol 32: 953-962.
- 285 LeOra (ed.) 1987. POLO-PC. A User's Guide to Probit or Logit Analysis. LeOra Software,
- 286 Berkeley, CA.
- 287 Liu, S.-S., J. Colvin, and P. J. De Barro. 2012. Species concepts as applied to the whitefly
- 288 Bemisia tabaci systematics: how many species are there? J. Integr. Agric. 11: 176-186.
- Luan, J. B., D. M. Yao, T. Zhang, L. L. Walling, M. Yang, Y. J. Wang, and S. S. Liu. 2013.
- Suppression of terpenoid synthesis in plants by a virus promotes its mutualism with vectors. EcolLett 16: 390-398.

- 292 Nauen, R., P. Jeschke, R. Velten, M. E. Beck, U. Ebbinghaus-Kintscher, W. Thielert, K.
- Wölfel, M. Haas, K. Kunz, and G. Raupach. 2015. Flupyradifurone: a brief profile of a new
   butenolide insecticide. Pest Manag Sci 71: 850-862.
- Ning, W., X. Shi, B. Liu, H. Pan, W. Wei, Y. Zeng, X. Sun, W. Xie, S. Wang, Q. Wu, J.
- 296 Cheng, Z. Peng, and Y. Zhang. 2015. Transmission of *Tomato yellow leaf curl virus* by
- *Bemisia tabaci* as affected by whitefly sex and biotype. Sci. Rep. 5: 10744.
- 298 Pan, H., E. L. Preisser, D. Chu, S. Wang, Q. Wu, Y. Carrière, X. Zhou, and Y. Zhang.
- 2015. Insecticides promote viral outbreaks by altering herbivore competition. Ecol. Appl. 25:
  1585-1595.
- 301 Pan, H. P., D. Chu, B. M. Liu, X. B. Shi, W. Xie, Y. Carriere, X. C. Li, and Y. J. Zhang.
- **2013a.** Differential effects of virus on its two closely-related vectors, *Bemisia tabaci* B and Q.
  Sci. Rep. 3: 2230.
- 304 Pan, H. P., D. Chu, B. M. Liu, W. Xie, S. L. Wang, Q. J. Wu, B. Y. Xu, and Y. J. Zhang.
- 2013b. Relative amount of symbionts in insect hosts changes with host-plant adaptation and
   insecticide resistance. Environ Entomol 42: 74-78.
- Pietri, J. E., and D. Liang. 2018. The links between insect symbionts and insecticide resistance:
   causal relationships and physiological tradeoffs. Ann. Entomol. Soc. Am. 111: 92-97.
- 309 Roditakis, E., M. Stavrakaki, M. Grispou, A. Achimastou, X. Van Waetermeulen, R.
- 310 Nauen, and A. Tsagkarakou. 2017. Flupyradifurone effectively manages whitefly *Bemisia*
- *tabaci* MED (Hemiptera: Aleyrodidae) and tomato yellow leaf curl virus in tomato. Pest Manag
   Sci 73: 1574-1584.
- **Russell, R. M., J. L. Robertson, and N. E. Savin. 1977.** POLO: A New Computer Program for
- Probit Analysis. Bull Entomol Soc Am 23: 209-213.
- **SAS 2010.** JMP user's guide, version 9.0 computer program, version By SAS, Cary NC.
- Shen, S. K., and P. F. Dowd. 1991. Detoxification spectrum of the cigarette beetle symbiont
   *Symbiotaphrina kochii* in culture. Entomol Exp Appl 60: 51-59.
- Shi, X., E. L. Preisser, B. Liu, H. Pan, M. Xiang, W. Xie, S. Wang, Q. Wu, C. Li, Y. Liu, X.
- Shi, A., E. L. Heisser, B. Liu, H. Han, W. Alang, W. Ale, S. Wang, Q. Wu, C. Li, T. Liu, A.
   Zhou, and Y. Zhang. 2019. Variation in both host defense and prior herbivory can alter plant-
- vector-virus interactions. BMC Plant Biol. 19: 556.
- 321 Smith, H. A., C. A. Nagle, C. A. MacVean, and C. L. McKenzie. 2016. Susceptibility of
- 322 *Bemisia tabaci* MEAM1 (Hemiptera: Aleyrodidae) to Imidacloprid, Thiamethoxam, Dinotefuran
- and Flupyradifurone in South Florida. Insects 7: 57.
- 324 Stansly, P. A., and S. E. Naranjo (eds.). 2010. *Bemisia*: Bionomics and Management of a
- 325 Global Pest. Springer, New York NY.
- Su, Q., E. L. Preisser, X. M. Zhou, W. Xie, B. M. Liu, S. L. Wang, Q. J. Wu, and Y. J.
- **Zhang. 2015.** Manipulation of host quality and defense by a plant virus improves performance of
- whitefly vectors. J. Econ. Entomol. 108: 11-19.
- 329 Teng, X., F. H. Wan, and D. Chu. 2010. *Bemisia tabaci* biotype Q dominates other biotypes
- across China. Fla Entomol 93: 363-368.
- 331 Tiwari, S., K. Pelz-Stelinski, and L. L. Stelinski. 2011. Effect of Candidatus Liberibacter
- asiaticus infection on susceptibility of Asian citrus psyllid, *Diaphorina citri*, to selected
- insecticides. Pest Manag Sci 67: 94-99.
- Xie, W., C. Chen, Z. Yang, L. Guo, X. Yang, D. Wang, M. Chen, J. Huang, Y. Wen, Y.
- Zeng, Y. Liu, J. Xia, L. Tian, H. Cui, Q. Wu, S. Wang, B. Xu, X. Li, X. Tan, M. Ghanim, B.
- 336 Qiu, H. Pan, D. Chu, H. Delatte, M. N. Maruthi, F. Ge, X. Zhou, X. Wang, F. Wan, Y. Du,
- 337 C. Luo, F. Yan, E. L. Preisser, X. Jiao, B. S. Coates, J. Zhao, Q. Gao, J. Xia, Y. Yin, Y. Liu,

- 338 J. K. Brown, X. J. Zhou, and Y. Zhang. 2017. Genome sequencing of the sweetpotato whitefly
- 339 *Bemisia tabaci* MED/Q. GigaScience 6: 1-7.
- **Zhang, H., H. Gong, and X. Zhou. 2009.** Molecular characterization and pathogenicity of
- tomato yellow leaf curl virus in China. Virus Genes 39: 249-255.
- 342 Zhang, T., J. B. Luan, J. F. Qi, C. J. Huang, M. Li, X. P. Zhou, and S. S. Liu. 2012.
- 343 Begomovirus-whitefly mutualism is achieved through repression of plant defences by a virus
- pathogenicity factor. Mol Ecol 21: 1294-1304.

345

**Table 1:** Median lethal concentration (LC<sub>15</sub> and LC<sub>50</sub>) of flupyradifurone (Sivanto) to uninfected

and viruliferous MED.  $LC_{15}$  and  $LC_{50}$  followed by different upper-case letters indicate that

uninfected and viruliferous MED are significantly different based on overlap of 95% CLs.

Treatment	atment N		LC <sub>15</sub> (mg[AI]kg <sup>-1</sup> ) (95% CL)	LC <sub>50</sub> (mg[AI]kg <sup>-1</sup> ) (95% CL)	$X^2$ (df)	P value
		3.64 ±		17.33 (14.08-20.48)	1.61	
Uninfected	478	0.39	5.78 (3.72-7.82) A	Α	(3)	0.66
		$4.07 \pm$	11.75 (8.91-14.44)	31.33 (27.22-35.82)		
Viruliferous	476	0.36	В	В	2.4 (3)	0.49

349

350

351 Table 2: Results of ANOVA assessing the impact of TYLCV infection, Sivanto exposure, and

their interaction on MED demographic variables.

				Egg-adult								
	Female longevity			First week fecundity			developmental time			Egg-adult survival		
	(d)			(# eggs)			(d)			(%)		
Treatment	F	$d\!f$	Р	F	$d\!f$	Р	F	df	Р	F	df	Р
TYLCV <sup>†</sup>	0.43	1,101	0.513	15.71	1,101	< 0.001	0.86	1,18	0.364	1.03	1,18	0.324
Sivanto	3.55	1,101	0.063	10.24	1,101	0.002	7.26	1,18	0.015	0.31	1,18	0.586
TYLCV*Sivanto	3.43	1,101	0.067	3.18	1,101	0.078	3.34	1,18	0.085	0.97	1,18	0.338
<sup>†</sup> Tomato yellow leaf curl virus												

353

#### 354 Figure Legends

- Figure 1. *Bemisia tabaci* MED feeding on *Lycopersicon esculentum*. Mean ± SE values for the
- demographic variables A) Female longevity (days); B) Eggs per female over one week; C) Egg-
- adult development time (days); and D) Egg-adult survival (%). Light gray bars: uninfected MED;
- dark gray bars: viruliferous MED. Unstriped bars (S-): plants sprayed with distilled water;
- striped bars (S+): plants sprayed with 11.75 mg[AI]kg<sup>-1</sup> Sivanto (LC<sub>15</sub> for viruliferous MED).
- 360 Different upper-case letters above bars indicate significant differences (Tukeys' HSD with  $\alpha =$
- 361 0.05); in figure 1D, there were no significant between-treatment differences.

