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Coláiste na hOllscoile Corcaigh

| 3            | Moulting season corticosterone correlates with winter season body-weight in                                    |
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| 4            | an Arctic migrant bird   |
| 5            |  |
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In vertebrates, the endocrine system translates environmental changes into 19 physiological responses on which natural selection can act to regulate individual 20 21 fitness and, ultimately, population dynamics. Corticosterone (CORT) and 22 dehydroepiandrosterone (DHEA) are important regulators of the avian endocrine system but relatively few studies have investigated their downstream effects on key 23 morphological fitness-related traits in free-living populations. This study quantified 24 25 endocrine-morphology relationships in free-living Greenland Barnacle Geese Branta *leucopsis* that breed in the high Arctic. CORT and DHEA were extracted from feather 26 27 and blood samples and tested for relationships with three morphological traits associated with survival and reproduction: body weight, body size and facial 28 plumage coloration. We expected CORT concentration to be higher in birds with less 29 favourable morphological traits (i.e. lighter, smaller and less attractive) and DHEA to 30 be higher in birds with more favourable traits (i.e. heavier, bigger and more 31 attractive). As expected, individuals with higher CORT during the post-breeding 32 moult (July/August) had significantly lower body weight during the following winter 33 (November - April). In contrast, we found no robust DHEA-morphology relationships 34 and no statistically significant relationship between CORT and body size or facial 35 36 plumage. Overall, this study provides evidence of a negative relationship between 37 CORT and body-weight extending across different seasons of the annual cycle in a long-distance migrant. This is of particular interest because body-weight fluctuates 38 rapidly in response to environmental resources and is closely linked to both survival 39 and reproductive success in this species. Understanding the relationship between 40 CORT and key morphological traits is important because endocrine-disrupting 41 contaminants in the Arctic increasingly interfere with CORT function in birds, 42

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including Barnacle Geese, and based on the results of this study may haveconsequences for body-weight regulation.

The endocrine system is an important modulator of fitness in vertebrates. 48 Environmental conditions affect fitness - an individual's ability to survive and 49 50 reproduce - which in turn regulates population dynamics (Orr 2009). Hormones play 51 a key role in this sequence by translating environmental information into individual physiological responses that have positive or negative fitness consequences (Norris 52 2018, Wingfield 2018). In birds, hormones modulate morphology such as body-53 weight (Awerman & Romero 2010), body size (Müller et al. 2009) and plumage 54 coloration (Jawor & Breitwisch 2003, Grunst et al. 2015) - morphological traits that 55 56 are strongly connected with survival and reproductive success (e.g. Poisbleau et al. 2006, Tombre et al. 2012, Badás et al. 2018). Importantly, natural selection acts on 57 the downstream physiological effects of hormones (i.e. weight, size and plumage) 58 rather than the circulating hormones themselves. As such, identifying endocrine-59 60 morphology relationships can provide insight into the sequence between environment, individuals and population. Although the major functions of the 61 endocrine system are highly conserved among vertebrates, endocrine-morphology 62 relationships are not always consistent among taxa or even within a species' life-63 history (Bonier et al. 2009), therefore studies on novel species or relationships can 64 provide new insights. 65

66

The glucocorticoid hormone corticosterone (CORT) and the androgen hormone
dehydroepiandrosterone (DHEA) are principal modulators of the avian endocrine
system (Hau *et al.* 2010, Wingfield *et al.* 2018). CORT principally modulates
homeostasis, energy balance and stress response via release by the hypothalamicpituitary-adrenal axis in response to changing conditions (Sapolsky *et al.* 2000, Creel *et al.* 2013). Mildly elevated CORT in the short term can be an adaptive mechanism

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for coping with environmental or physiological challenges, such as parasite infection 73 or increased energetic demand (Kouwenberg et al. 2013, Rivers et al. 2017, Grace & 74 75 Anderson 2018, Schoenle et al. 2018, Vágási et al. 2018). In contrast, chronically elevated CORT can negatively affect overall body condition (Harms et al. 2015, Latta 76 et al. 2016). Common Kestrel Falco tinnunculus nestlings experimentally implanted 77 with CORT (to represent an environmental stressor) were smaller, with shorter tarsi 78 79 and flight feathers before fledging compared to control nestlings (Müller et al. 2009). American Yellow Warblers Setophaga petechia with higher CORT had lower 80 81 carotenoid hues in plumage associated with sexual signalling (Grunst et al. 2015). Thus, chronic elevated CORT relative to an individual's conspecifics may be a 82 fitness disadvantage (Bonier et al. 2009). 83

84

DHEA principally modulates aggression and dominance behaviour (Soma et al. 85 2008, 2015). It is produced by the adrenal glands and other organs, then taken up by 86 the brain and converted into sex steroids that generate physiological changes (Soma 87 et al. 2015, Wingfield et al. 2018). Hau et al. (2004) found that aggressive 88 vocalisations by Spotted Antbirds Hylophylax naevioides naevioides during 89 simulated territorial intrusions were positively correlated with DHEA levels. Similarly, 90 91 male Song Sparrows *Melospiza melodia morphna* experimentally implanted with DHEA showed greater aggression in simulated territory intrusions than control 92 sparrows (Wacker et al. 2016). In contrast, Poisbleau et al. (2009) found a negative 93 correlation between DHEA and dominance score (the percentage of competitive 94 95 interactions "won" by an individual) in wintering Brent Geese Branta bernicla bernicla, indicating uncertainty in the role of DHEA in aggression among taxa. 96 However, DHEA also modulates potential negative impacts of glucocorticoids and 97

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has positive neuroprotective properties (Newman *et al.* 2010, 2013). It has also been
linked to alternative plumage colour morphs (Spinney *et al.* 2006). Based on the
literature, one could hypothesise that elevated DHEA relative to an individual's
conspecifics may offer a fitness advantage, both through its protective properties and
as dominant individuals would be more likely to access resources with positive
outcomes for body condition (Poisbleau *et al.* 2006).

104

Although the relationship between CORT and fitness is well studied in wild bird 105 106 populations, fewer studies concern its relationship with morphology. Additionally, the relationship between DHEA and fitness or morphology in wild bird populations is not 107 well studied. Quantifying endocrine-morphology relationships in free-living 108 populations can reveal patterns that may not be apparent in laboratory investigations 109 (Schoech et al. 2011, Wingfield 2018). Thus, this study aimed to quantify endocrine-110 morphology links in a free-living population of Greenland Barnacle Goose Branta 111 112 *leucopsis.* The majority of this population breeds in high-Arctic north-east Greenland 113  $(70-78 \,^{\circ}\text{N})$ , with a small emerging breeding population in south-east Iceland (64  $^{\circ}\text{N})$ . Barnacle Geese here undergo a post-breeding synchronous flight feather moult in 114 July/August before migrating to Ireland and Scotland for winter (October – April) 115 116 (Cabot et al. 1984). Increased body weight is a fitness advantage in geese, as more 117 reserves are available for migration, egg formation and nesting (Poisbleau et al. 2006, Hahn et al. 2011, Tombre et al. 2012). Increased body size is also a fitness 118 119 advantage as it improves social status and access to resources (Choudhury et al. 120 1996, Poisbleau et al. 2006, Kurvers et al. 2010). Barnacle Geese also have prominent white patches extending across the face contrasting with black plumage 121 around the eye and bill. These patches are an important social signal (including 122

during aggressive encounters) and, although the role is not fully understood, it
appears that, at least among females, individuals with darker faces are more
attractive as mates (Black *et al.* 2014). Hence, we expect hormone concentrations
associated with increased body weight, body size and smaller cheek patches to
improve overall fitness in Barnacle Geese.

128

129 We analysed the strength of relationships between these three morphological traits and CORT and DHEA. The analysis determines whether meaningful endocrine-130 131 morphology relationships are present but cannot confirm causality. We hypothesised that there would be a negative correlation between body weight and CORT and 132 between body size and CORT, but a positive correlation between cheek patch size 133 and CORT (i.e. individuals with higher CORT are lighter, smaller and less attractive). 134 We hypothesised that DHEA would correlate in the opposite way (i.e. individuals with 135 higher DHEA are heavier, bigger and more attractive). In addition, we used long-term 136 data on body-weight, body size and cheek patch size to investigate if these traits 137 have changed over time, which could signal changing selective pressures. CORT 138 and DHEA play a critical regulatory role in the avian endocrine system, so 139 determining consistent relationships with morphological traits associated with 140 141 survival and reproduction will improve our understanding of these systems and how birds are affected by and adapt to their changing environment. 142

143

## 144 METHODS

## 145 Study outline

During winter 2017/2018, 55 adult Barnacle Geese were captured on their wintering grounds in Ireland using cannon-netting. Four flocks were captured at four different

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locations in November 2017 (*n* = 5 at 54.12, -10.21), January 2018 (*n* = 33 at 55.29, 148 -7.28) and April 2018 (n = 8 at 55.28, -7.31 and n = 9 at 55.38, -7.37). Birds were 149 sexed by cloacal examination and morphological measurements were collected. 150 151 Feather and blood samples were then collected to extract CORT and DHEA, respectively. Morphological measurements from an additional 53 Barnacle Geese 152 were collected by one author (D.C.) using the same methods from 1970-2018 (see 153 154 Table S1 for annual breakdown). Capture and sampling of birds was carried out under licences by the National Parks and Wildlife Service of the Government of 155 156 Ireland and the British Trust of Ornithology. All feather and blood sampling procedures were approved by the University College Dublin Animal Research Ethics 157 Committee and the Health Products Regulatory Authority of Ireland (AE18982/P115). 158

159

#### 160 **Feather CORT**

CORT is deposited into segments of the feather vane and rachis containing the 161 blood guill during growth and can be measured from feather trimmings subsequently 162 (Jenni-Eiermann et al. 2015). Because CORT concentration in the feather is fixed 163 after passing the blood guill, assuming a constant growth rate, measurements from 164 feathers provide a smoothed average baseline of an individual Barnacle Goose's 165 166 hypothalamo-pituitary-adrenal activity during the synchronous growth period in 167 July/August (Jenni-Eiermann et al. 2015, Taff et al. 2018). Studies of Baltic Barnacle Geese found that primary feather growth rate was  $8 \pm 1.58$  mm/day (n = 52) in 168 females and 7.3  $\pm$  1.33 mm/day (n = 50) in males, indicating relatively consistent 169 growth rates (Larsson 1996). As such, CORT recovery from feathers is preferable to 170 recovery from blood when cannon-netting as CORT will not be affected by the long 171 and variable handling times associated with this capture procedure. 172

Approximately 25 mm was trimmed from the distal tip of the fifth secondary flight 174 feather using a scissors and stored in paper envelopes until CORT extraction. CORT 175 was extracted from feathers using a methanol-based extraction technique based on 176 Bortolotti et al. (2008). The feather sample was trimmed to a weight of ~15.5 g and 177 minced into <5 mm<sup>2</sup> pieces using a blade, then shaken for 2 minutes with 1.5 ml 178 HPLC grade 99.9% methanol in a Qiagen Tissue Lyser. This sample weight was a 179 compromise between maximising and standardising the number of samples while 180 181 removing as little feather from the bird as possible (whole feathers were not plucked for ethical reasons). The sample was transferred to a flask with an additional 8.5ml 182 methanol and incubated overnight in a shaking water bath at 50 °C to extract CORT. 183 The sample was run through a 0.45  $\mu$ m syringe filter to remove the feather particles 184 and then heated in a sand block at 50  $\,^{\circ}\mathrm{C}$  until the liquid fraction evaporated 185 completely. The dried extract was reconstituted in 500 µl phosphate-buffered saline 186 liquid and frozen at -20 °C until CORT concentration analysis. From the 55 birds 187 captured, we extracted CORT from 53 samples (2 samples excluded as they were 188 below the threshold weight of 15.5 g). As CORT was extracted in two batches, inter-189 extraction variation was measured as 0.06% using two pool samples from a single 190 191 homogenous pool created by mixing a number of feathers.

192

Analysis of CORT concentration was conducted in a single batch by
radioimmunoassay using the ImmuChem Corticosterone Double Antibody kit (MP
Biomedicals, Orangeburg, NY). The frozen samples, including pool samples, were
brought to room temperature and vortexed to mix before aliquoting 100 µl of each for
analysis. A standard calibration curve was created by serial dilution of a 1000 ng/ml

CORT calibrator and 100 µl of high and low CORT control reconstituted in de-ionised 198 water were aliquoted to provide reference value samples. We added 200 µl of 199 <sup>125</sup>Iodine labelled CORT tracer to all samples followed by 200 µl of rabbit anti-CORT. 200 201 The tracer was allowed to compete with CORT for a fixed amount of antibody sites for 2 h. The antibody-tracer complex was then separated by adding 500 μl 202 precipitation solution and centrifuging at 2 000 rpm at 4 °C for 20 mins. Unbound 203 204 tracer was removed by aspiration with a vacuum pump. A 60 s gamma count was conducted using a Wallac 1470 gamma counter, with the quantity of radiation 205 206 measured inversely proportionate to the quantity of CORT. CORT concentrations in each sample were determined by interpolation from the standard calibration curve. 207 All samples were analysed in duplicate and the mean final hormone concentration of 208 feather extracted samples was expressed as a function of feather length (ng/mm) 209 following Bortolotti et al. (2008). The intra-assay coefficient of variation (CV) was 210 4.8%, within the range of other similar studies (e.g. Bortolotti et al. 2008, Legagneux 211 212 et al. 2013, Johns et al. 2018).

213

## 214 Blood DHEA

DHEA concentration can be measured accurately from avian blood and, unlike other
hormone modulators of aggression such as testosterone, does not appear to be
affected by capture or variable handling time in geese (Poisbleau *et al.* 2009) making
it a suitable measurement in a cannon-netting procedure with long and variable
handling times. No literature was available on DHEA levels in geese during the
breeding season (when feathers are grown) therefore we chose to measure DHEA
from blood. However, because Newman *et al.* (2013) found that 30 min restraint of

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Song Sparrows altered DHEA concentration, we took the opportunity to include ananalysis of the effect of handling time on DHEA levels in our study.

224

225 A 1.5 ml blood sample was collected from the metatarsial vein using a syringe (23) G). Blood was centrifuged at 3000 rcf for 15 min to separate the extracellular fluid 226 from red blood cells and 50 µl serum was drawn off using a micropipette and sealed 227 228 in a culture tube. Tubes were frozen at -18 °C as soon as possible and transferred to -20 ℃ within 48 h until DHEA concentration analysis. From the 55 birds captured, 229 230 we recovered DHEA from 35 samples due to time constraints on blood sampling, as all birds in the captured flock must be released together within the maximum 231 permitted time following initial capture according to ethical guidelines. 232

233

DHEA concentration was analysed at NationWide Laboratories (Cambridge, UK) by 234 enzyme immunoassay using the DHEA ELIZA kit (IBL International GmbH, 235 Hamburg, Germany). All samples were checked for gross haemolysis prior to 236 237 analysis. The frozen samples were brought to room temperature and 20 µl was aliguoted into wells on a microtiter plate coated with polyclonal rabbit anti-DHEA. A 238  $20 \mu$ l high and low control sample were also aliquoted along with serial dilution of a 239 240 30 ng/ml DHEA calibrator to produce a standard calibration curve. Next, 100 µl of DHEA-horseradish peroxidase conjugate was added to each well. The conjugate 241 242 was allowed to compete with DHEA for a fixed amount of antibody sites for 1 hr. 243 Unbound conjugate was then washed off with phosphate buffer. Next, 100 µl TMB 244 Substrate Solution was added to each well and incubated for 30 min at room temperature before stopping the reaction with 100 μl TMB Stop Solution. The optical 245 density of the wells was measured using a microtiter plate reader at 450 nm, with the 246

intensity of colour developed during the TMB reaction inversely proportional to DHEA
concentration. DHEA concentrations in each sample were determined by
interpolation from the standard calibration curve. All samples were analysed in
duplicate, with the final hormone concentration expressed as a function of blood
volume (nmol/L).

252

#### 253 Morphology

Measurements of body-weight, body size and cheek patch size were collected from the 55 birds in 2017/18 (18 males and 37 females) and from an additional 53 birds in 1970-2018 (20 males and 33 females). Each individual was weighed to the nearest 10.0 g using a spring scale. The length of the tarsus was measured to the nearest 0.1 mm with a callipers and used as a proxy for body size because it remains fixed in adults (Choudhury *et al.* 1996). One observer (D.C.) carried out all measurements to minimise observer bias.

261

262 To measure cheek patch size, we took a photograph of the head at a 90° lateral angle. The area of brilliant white pigmentation on the forehead and lower mandible 263 was measured in pixels using the polygon tool in the ImageJ program 264 265 (https://imagej.net/Welcome; Abràmoff et al. 2004). One observer (Y.L.) carried out all measurements to minimise observer bias. Because the data included 1970-2018 266 photographs, images could not be standardised at the time of collection. To address 267 this, the eye, which differs little among individuals, was used as a standard reference 268 to calibrate photographs. A correction factor was calculated as the ratio of the eye 269 area in each photograph to the mean eye area of all photographs. This correction 270

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factor was used to scale white patch area, providing a relative patch size for eachbird.

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## 274 Statistical analysis

Preliminary analysis of the data indicated that body-weight, tarsus length and cheek 275 patch size were continuous data of a normal distribution. CORT was a continuous 276 277 measurement with a strong positive skew, as there were relatively few birds with high CORT concentrations. DHEA was a continuous measurement limited to a lower 278 279 boundary of 0.1 because the enzyme immunoassay detected concentrations to a lower limit of 0.1 nmol/L. To account for physiological fluctuations over winter, we 280 included the day of capture as a parameter in the data by counting the number of 281 days elapsed between the beginning of winter (November 1; marks the end of the 282 autumn migration period for this species) and the day of capture. We also included 283 sex as a parameter in the data because many physiological traits are known to differ 284 between males and females. Finally, we included handling time on the day of 285 capture, defined as minutes elapsed between initial capture and blood sampling, to 286 account for potential effects of handling on DHEA concentration. 287

288

We performed six models to test endocrine-morphology relationships using the R statistical language and environment 3.5.1 (R Core Team 2018). Linear regression was used to test body weight as a function of (1) CORT and (2) DHEA. These models included the additional parameters sex, tarsus (to account for bigger birds being inherently heavier) and the interaction between hormone and linear and quadratic effects of day of capture. Linear regression was also used to test patch size as a function of (3) CORT and (4) DHEA. These models included the additional

parameters sex and the interaction between hormone and linear and quadratic 296 effects of day of capture. A generalized linear model with gamma errors and a log 297 298 link was used to test (5) CORT as a function of tarsus length. This model included 299 the additional parameter sex. Finally, a Tobit regression with a lower boundary of 0.1 (VGAM package 1.1-1; Yee 2015) was used to test (6) DHEA as a function of tarsus 300 length. This model included the additional parameters sex, linear and quadratic 301 302 effects of day of capture and the effect of handling time. Table 1 provides a summary of the six initial models. 303

304

In all models, numerical explanatory variables were standardised by subtracting the
mean and dividing by the standard deviation to allow for meaningful comparisons
between variables on different scales. Likelihood ratio tests (LRT) were used to
determine whether each parameter significantly improved the model fit. Parameters
were only retained in the final model if the LRT value was significant (<0.05) or</li>
borderline significant (<0.099). Table S2 provides all LRT results.</li>

311

In addition to the above analyses, we combined 2017/18 and 1970-2018 312 morphological data to test for long-term changes in the three Barnacle Goose 313 314 morphological traits over time. Firstly, we performed two linear mixed models (Ime4 315 package 1.1-21; Bates et al. 2015) to test body-weight and patch size as a function of year. A random intercept for the month of measurement was included in these 316 models to account for seasonal variation in physiology. Secondly, we performed a 317 linear regression to test tarsus length as a function of year. All three models included 318 sex as an additional parameter. Table 1 provides a summary of the three models. 319 320

#### 321 **RESULTS**

Mean CORT concentration was 3.51 ng/mm (range 0.54 - 13.95 ng/mm, n = 52; we 322 excluded one sample with an individual CV >20% from the analysis, as 323 324 recommended by Reed et al. (2002)). It should be noted that some bird species suppress CORT during moult (Romero 2002), therefore CORT concentration in 325 these samples could be suppressed relative to concentration during other life history 326 327 stages. Interestingly, above-average values of CORT were only recorded among females, although LRT indicated that there was not statistically significant effect of 328 329 sex. Broader variation in CORT was also recorded in early winter (January and November) compared to late winter (April). 330

331

The final body-weight model included additive effects of CORT, tarsus (i.e. body 332 size), sex and day of capture (Table 2). There was a significant negative relationship 333 between body weight and CORT (Fig. 1(a)). After accounting for sex, body size and 334 day of capture, a 1 sd (standard deviation) increase in CORT was associated with a 335 ~60 g decrease in body-weight (almost 5% of the body-weight of an average bird). 336 Bigger birds were heavier; a 1 sd increase in tarsus length was associated with a 337  $\sim$ 40 g increase in weight. Males were, on average,  $\sim$ 120 g heavier than females. 338 339 Finally, body weight increased linearly between November and April. The final patch size model included only the variable day of capture, indicating no relationship 340 between patch size and CORT (Table 2). Similarly, the final tarsus length model was 341 the null model, indicating no relationship between tarsus length and CORT. 342 343

Mean blood DHEA concentration was 0.75 nmol/L (range 0.10 nmol/L - 4.65 nmol/L, n = 35). The final body-weight model included additive effects of DHEA, sex, tarsus

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(i.e. body size) and day of capture (Table 2). After accounting for sex, body size and 346 day of capture, a 1 sd increase in DHEA was associated with a ~50 g decrease in 347 body weight, but this relationship was not statistically significant (Fig. 1(b)). As 348 349 before, bigger birds were heavier (a 1 sd increase in tarsus length was associated 350 with a ~55 g increase in body weight), males were, on average, ~115 g heavier than females and body weight increased linearly between November and April. The final 351 352 patch size model included the variables DHEA and day of capture. A 1 sd increase in DHEA was associated with a ~3.5 decrease in patch size (pixels), but this 353 354 relationship was not significant (Fig. 1(c)). Patch size decreased linearly between November and April. As before, the final tarsus length model was the null model, 355 indicating no relationship between tarsus length and DHEA. Note that the LRT for 356 this model also indicates no significant effect of handling time on blood DHEA 357 concentrations. 358

359

The models combining 1970-2018 and 2017/18 morphological data (n = 108) detected no change in mean Barnacle Goose body weight, patch size or tarsus length since 1970. After accounting for variation between the sexes and seasonal fluctuations, none of the three models detected a significant relationship between morphological trait and year (Table 2).

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366

## 367 **DISCUSSION**

This is the first study to quantify endocrine-morphology relationships in Barnacle Geese, to our knowledge. Individuals with higher CORT during body moult had a lower body weight early the following winter. It is one of few studies to demonstrate this effect in a free-living population outside controlled laboratory conditions. In
contrast, we did not find robust evidence of DHEA-morphology relationships or
CORT-tarsus and CORT-patch relationships. We also show that mean Barnacle
Goose body weight, tarsus length and patch size have not varied significantly since
1970.

376

377 Our CORT-weight results are consistent with experimental observations from other bird species and vertebrate taxa. A negative CORT-weight relationship has been 378 379 observed in aviary enclosure tests of captive House Sparrow Passer domesticus (Vágási et al. 2018), Gambel's White-crowned Sparrow Zonotrichia leucophrys 380 gambelii (Busch et al. 2008) and Starling Sturnus vulgaris (Awerman & Romero 381 382 2010). Farm-bred Japanese Quail Coturnix coturnix japonica treated with CORT had lower body weights than controls (Hull et al. 2007). Among mammals, stressed 383 laboratory Striped Field Mice Apodemus agrarius had higher cortisol (the mammalian 384 equivalent of CORT) and lower body weight than their unstressed counterparts 385 (Wang et al. 2011). Simulated chronic stress tests on laboratory rats showed that a 386 surge in CORT caused a sustained decrease in body weight (Scherer et al. 2011). A 387 negative CORT-weight relationship in free-living Common Eider Somateria 388 389 mollissima borealis has been observed (Harms et al. 2015), but further examples 390 using free-living populations are limited.

391

Because this study is cross-sectional rather than longitudinal, it is not possible to
confirm whether body weight affects CORT or visa-versa. Heavier birds could
experience fewer homeostatic challenges or could be better able to cope with such
challenges and maintain lower CORT (López-Jiménez *et al.* 2017). Nutritional stress

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could both reduce body weight and increase allostatic load and increased allostatic 396 load has been shown to upregulate CORT (Johns et al. 2018). Alternatively, the 397 398 results could indicate carry-over effects of stressful environmental conditions during 399 moult (Legagneux et al. 2013, Latta et al. 2016). For instance, unusually arid conditions were associated with increased CORT in House Sparrows (Treen et al. 400 2015). Awerman and Romero (2010) and Busch et al. (2008) show that elevated 401 402 CORT increases protein metabolism and muscle-wasting, providing a physiological mechanism for weight loss. Research on other Arctic-breeding waterfowl suggests 403 404 that external environmental factors have a stronger influence on CORT than intrinsic factors (Legagneux et al. 2013). If the same is true for Barnacle Geese, CORT could 405 provide a medium for carry-over effects across the annual cycle. By the same 406 process, the results could indicate a cost of reproduction: at the end of the breeding 407 408 season, successful breeders may have higher CORT following the stress of raising offspring compared to failed or sabbatical breeders (Crossin et al. 2017, Ibañez et al. 409 410 2018, Ramos et al. 2018). Although Barnacle goslings are precocial, offspring typically remain with the parents for their first year, requiring some form of care 411 412 (Black et al. 2014). Ramos et al. (2018) show that the cost of reproduction can be physiologically mediated by CORT from breeding to winter season in Cory's 413 414 Shearwater *Calonectris borealis*. However, due to the remote breeding sites of Barnacle Geese, it was not possible to know if individuals in this study successfully 415 reared offspring that were lost prior to sampling (i.e. prior to arrival on the winter 416 grounds), therefore the relationship between breeding success and CORT could not 417 be tested. 418

Understanding endocrine-morphology relationships is useful in the current period of 420 rapid global change. Endocrine-disrupting contaminants released by agricultural and 421 422 industrial activity are increasingly known to interfere with natural endocrinological 423 function and are exacerbated by weather extremes associated with climate change (Jenssen 2006, Norris 2018). Climate change and accumulation of endocrine-424 disrupting contaminants are amplified in the Arctic compared to lower latitudes as 425 426 heat and contaminants are transported northward on natural currents (Bekryaev et al. 2010, AMAP 2015, Praetorius et al. 2018). Glucocorticoids (including CORT) that 427 428 regulate adaptation to stress are among the most susceptible hormones to contaminant disruption (Jenssen 2006). Indeed, there is already evidence that 429 exposure to endocrine-disrupting contaminants interferes with the normal 430 behavioural and physiological responses to acute stress in Svalbard Barnacle 431 Geese; individuals exposed to trace metals at coal-mining sites were found to 432 erratically increase CORT secretion during subsequent stress tests (Scheiber et al. 433 2018). The effects of environmental change are evident in other systems: Carolina 434 435 Chickadees *Poecile carolinensis* in forest disturbed by logging had elevated CORT and lower body mass compared to chickadees from undisturbed forest (Lucas et al. 436 2006). Adélie Penguins Pygoscelis adeliae confronted with unusual sea ice 437 438 conditions in the Antarctic had elevated CORT and lower body-weight (Cockrem et 439 al. 2006). Barn Owl Tyto alba nestlings growing up in intensive agricultural areas in Switzerland also had elevated baseline CORT and lower body-weight (Almasi et al. 440 2015). Upregulation of CORT could be an adaptive response to maintain condition in 441 442 the face of stressors (e.g. Rivers et al. 2017, Vágási et al. 2018) but could also negatively impact, for example, future reproductive investment (Kouwenberg et al. 443 2013, Crossin et al. 2017) and survival (Koren et al. 2012). 444

This study is one of few to investigate DHEA-morphology relationships in birds. 446 Although a small number of studies have established DHEA-morphology links in 447 448 birds and mammals (e.g. Spinney et al. 2006, de Heredia et al. 2007) and we hypothesised that DHEA-morphology relationships might manifest via dominant 449 behaviour and improved access to resources, our analysis did not provide robust 450 451 evidence to relate winter DHEA to winter body-weight, size or facial plumage in Barnacle Geese. Poisbleau et al. (2009) also found no correlation between DHEA 452 453 and body condition in Brent Geese, adding to evidence that DHEA is not strongly associated with these morphological traits. DHEA may have a stronger relationship 454 with behavioural traits or other morphological traits not tested in this study. DHEA 455 may also play different roles depending on the territoriality or gregariousness of a 456 species (compare Hau et al. 2004 to Poisbleau et al. 2009). Future studies on a 457 greater taxonomic range of birds are warranted to shed light on this. The analysis 458 459 also detected no relationship between CORT and tarsus length or cheek patch size, providing no support for a link between CORT and body size or a role of CORT in 460 social signalling in Barnacle Geese, despite strong evidence of such links in other 461 bird species (e.g. Grunst et al. 2015, López-Jiménez et al. 2017). 462

463

Understanding the mechanistic role hormones such as CORT and DHEA play in
free-living birds provides insight into the links between environment, individual and
population. These hormones are principal regulators of the avian endocrine system.
However, the downstream effects of CORT on morphological traits on which natural
selection can act are less well studied in free-living populations and the downstream
effects of DHEA on both morphology and fitness are not well studied. This cross-

sectional study provides evidence of a correlation between moulting season feather
CORT and subsequent winter body weight in free-living Barnacle Geese. It is widely
accepted that chronically elevated CORT has negative effects on fitness in birds, but
our results suggest that, in Barnacle Geese, this effect could be associated with
variation in body-weight. It may also broaden our understanding of some of the ways
in which bird physiology may be affected by or adapt to environmental change.

476

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#### 489 **Conflict of interest**

490 The authors declare they have no conflicts of interest.

491

#### 492 Author contributions

493 SD, DC and BM conceived ideas and designed methodology. SD, DC, BM, AW and

- 494 KC captured birds and collected samples, with DC responsible for all historic data.
- 495 SD and JF conducted corticosterone measurements and SD and YL conducted
- 496 plumage measurements. SD analysed the data, with assistance from BM, and led

writing of the manuscript. All authors contributed critically to drafts and gave finalapproval for publication.

## **Data availability statement**

The data that support the findings of this study are available from the correspondingauthor upon reasonable request.

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- Table 1. Table 1. Summary of the six endocrine-morphology models and three longterm change models. Models include additive (+), synergistic (\*), quadratic (^2) or
- random (1) effects.

| Model                              | Response variable   | Explanatory variables                        |  |
|------------------------------------|---------------------|--|--|
| endocrine-morphology relationships |                     |  |  |
| 1                                  | weight              | ~ sex + tarsus + CORT * days + CORT * days^2 |  |
| 2                                  | patch               | ~ sex + CORT * days + CORT * days^2          |  |
| 3                                  | CORT                | ~sex + tarsus                                |  |
| 4                                  | weight              | ~ sex + tarsus + DHEA * days + DHEA * days^2 |  |
| 5                                  | patch               | ~sex + DHEA * days + DHEA * days^2           |  |
| 6                                  | DHEA                | ~sex + handling + days + days^2 + tarsus     |  |
| long-term                          | n changes in morpho | logical traits                               |  |
|                                    | weight              | ~sex + years + (1   month)                   |  |
|                                    | patch               | ~sex + years + (1   month)                   |  |
| 1                                  | tarsus              | ~sex + years                                 |  |
|                                    |                     |  |  |

**Table 2.** The formula of each final model is provided, along with the percentage variance/deviance explained (%) and the parameter estimates ( $\beta$  ± standard error) for each explanatory variable.

| Model | Final model                         | %        | Variable | Estimate         |
|-------|-------------------------------------|----------|----------|------------------|
|       | weight ~ sex + tarsus + CORT + days | 36 —<br> | sex      | 118.69 ± 47.19g  |
|       |                                     |          | tarsus   | 40.86 ± 23.42g   |
| 5     |                                     |          | CORT     | -59.81 ± 22.79g  |
|       |                                     |          | days     | 53.38 ± 23.65g   |
| 2     | patch ~ days                        | 14       | days     | -5.71 ± 1.96     |
|       |                                     |          | sex      | 115.93 ± 67.87g  |
| 1     | weight ~ sex + tarsus + DHEA + days | 26       | tarsus   | 55.46 ± 32.96g   |
|       |                                     |          | DHEA     | -52.09 ± 28.56g  |
|       |                                     |          | days     | 79.02 ± 30.97g   |
| 5     | patch ~ DHEA $\pm$ days             | 17       | DHEA     | -3.56 ± 2.13     |
| J     | pater brief + days                  |          | days     | -4.97 ± 2.15     |
| 5     | weight ~ sex + years + (1   month)  | 05       | sex      | 159.94 ± 33.72g  |
|       |                                     | 30       | years    | 0.70 ± 1.08g     |
|       | patch ~ sex + years + (1   month)   | 10       | sex      | 6.55 ± 2.95      |
|       |                                     | 15       | years    | $-0.05 \pm 0.09$ |
|       |                                     | 11       | sex      | 3.16 ± 0.85mm    |
|       | a = b = b = b = b                   | 11       | years    | -0.03 ± 0.03mm   |



**Figure 1.** Endocrine-morphology relationships in Barnacle Geese. (a) There is a significant negative relationship between feather CORT (ng/mm) and body weight (g). There are negative relationships between blood DHEA (nmol/L) and (b) body weight and (c) cheek patch size (measured in pixels), but these relationships are not statistically significant. The solid line represents least-squares fit with the shaded area representing standard error around the mean.

| 709 | SUPPORTING INFORMATION   |
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| 710 |  |
| 711 | Table S1. Annual breakdown of the number of Barnacle Geese captured from 1970- |
| 712 | 2018.  |
| 713 |  |
| 714 | Table S2. Results of likelihood ratio tests                                    |
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