

## REVIEW

## Open Access



# Restoring Wnt/ $\beta$ -catenin signaling is a promising therapeutic strategy for Alzheimer's disease

Lin Jia<sup>1,2</sup>, Juan Piña-Crespo<sup>3</sup> and Yonghe Li<sup>1\*</sup> 

## Abstract

Alzheimer's disease (AD) is an aging-related neurological disorder characterized by synaptic loss and dementia. Wnt/ $\beta$ -catenin signaling is an essential signal transduction pathway that regulates numerous cellular processes including cell survival. In brain, Wnt/ $\beta$ -catenin signaling is not only crucial for neuronal survival and neurogenesis, but it plays important roles in regulating synaptic plasticity and blood-brain barrier integrity and function. Moreover, activation of Wnt/ $\beta$ -catenin signaling inhibits amyloid- $\beta$  production and tau protein hyperphosphorylation in the brain. Critically, Wnt/ $\beta$ -catenin signaling is greatly suppressed in AD brain via multiple pathogenic mechanisms. As such, restoring Wnt/ $\beta$ -catenin signaling represents a unique opportunity for the rational design of novel AD therapies.

**Keywords:** Wnt, Alzheimer's disease, Neuronal survival, Neurogenesis, Synaptic plasticity, Drug target

## Introduction

Alzheimer's disease (AD) is the most common form of dementia accompanied by detrimental cognitive deficits and pathological accumulation of amyloid- $\beta$  (A $\beta$ ) plaques and tau-containing neurofibrillary tangles [1]. As one of the most important medical and social problems, there is an urgent need for effective therapies. The amyloid hypothesis is based on neuropathological evidence showing A $\beta$  aggregates (amyloid plaques) in AD brain and on the identification of over 200 mutations in the amyloid precursor protein (APP) and presenilin (PSEN) genes that cause familial AD (FAD) [1, 2]. The amyloid hypothesis has been the main driver of drug discovery efforts in the past 25 years; however, all clinical trials using anti-A $\beta$  drugs as a treatment for AD have ended in failure [3]. Therefore, current paradigms in AD drug discovery have shifted to the development of drugs that target the multiple disease processes that support the progression of AD pathology, and novel targeted therapies are urgently needed to prevent and treat AD [3–5].

The Wnt/ $\beta$ -catenin signaling pathway is a significant pathway regulating cell proliferation, migration and

differentiation, and Wnt proteins are key drivers of adult stem cells in mammals [6]. Studies have shown that dysregulated Wnt/ $\beta$ -catenin signaling plays an important role in the pathogenesis of AD [7]. In this review, we summarize our current understanding of regulation and function of the Wnt/ $\beta$ -catenin signaling pathway in AD brain and provide evidence indicating that the Wnt/ $\beta$ -catenin signaling pathway represents a new attractive therapeutic target for drug discovery in AD.

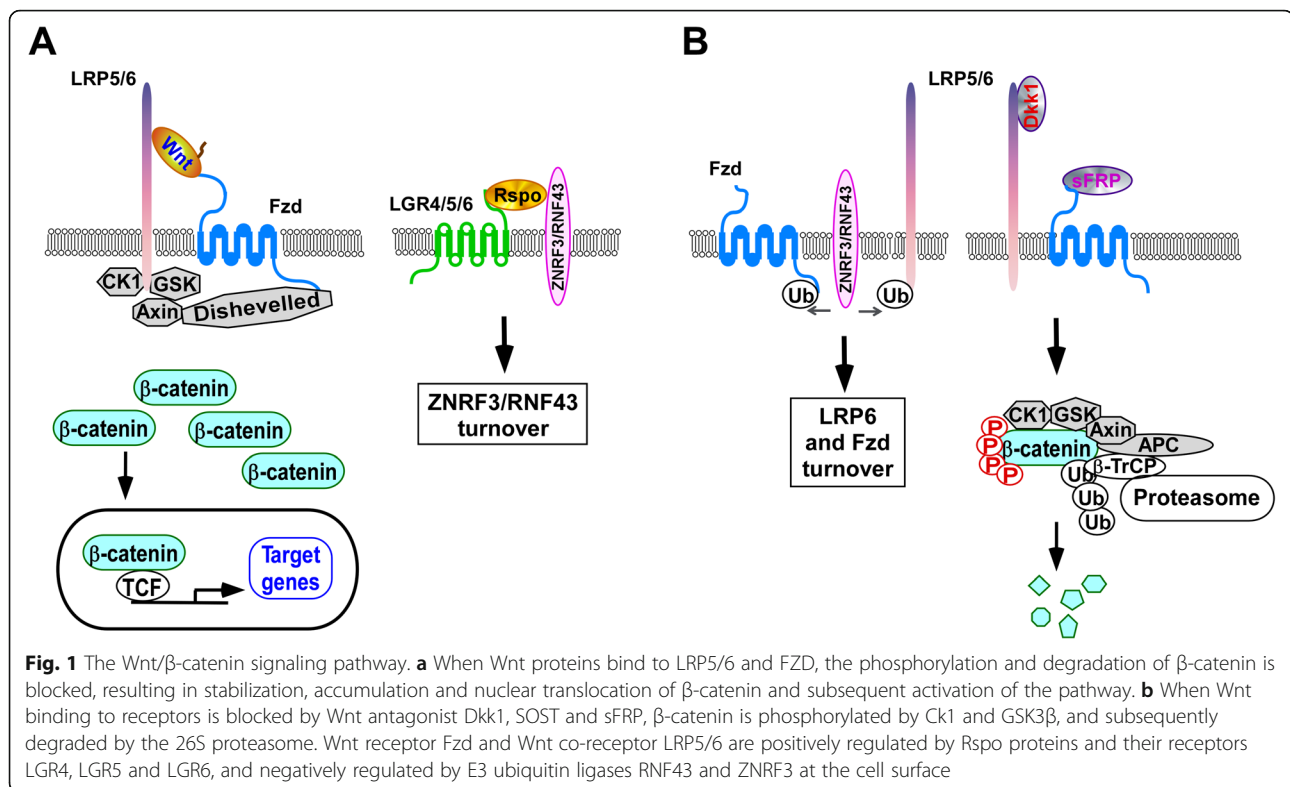
## Roles of Wnt/ $\beta$ -catenin signaling in physiological and pathophysiological processes in the brain

Wnt proteins are secreted glycoproteins that bind to the extracellular cysteine-rich domain of the Frizzled (Fzd) receptor family and Wnt co-receptor low density lipoprotein receptor-related protein 5 (LRP5) or LRP6 to activate the canonical Wnt/ $\beta$ -catenin signaling pathway. Binding of Wnt to the Fzd/LRP5/6 receptor complex results in inhibition of glycogen synthase kinase 3 $\beta$  (GSK3 $\beta$ ) and stabilization of cytosolic  $\beta$ -catenin. Stabilized  $\beta$ -catenin then translocates into the nucleus, interacts with T-cell factor/lymphoid enhancing factor (TCF/LEF), and induces the expression of specific target genes (Fig. 1) [6]. Wnt/ $\beta$ -catenin signaling is tightly regulated at the cell surface by various secreted proteins and

\* Correspondence: [Li.Yonghe@mayo.edu](mailto:Li.Yonghe@mayo.edu)

<sup>1</sup>Department of Neuroscience, Mayo Clinic, Jacksonville, FL 32224, USA  
Full list of author information is available at the end of the article





receptors. While Zinc and ring finger 3 (ZNRF3) and ring finger protein 43 (RNF43) promote LRP5/6 degradation [8–10], the extracellular molecule R-spondin (Rspo) together with its receptors leucine rich repeat containing G protein-coupled receptor 4/5/6 (LGR4/5/6) induces ZNRF3/RNF43 turnover, making LRP5/6 available on the cell surface for activation of the Wnt/β-catenin signaling pathway (Fig. 1) [11]. Moreover, Dickkopf (DKK) and soluble Frizzled-related protein (sFRP) bind to LRP5/6 and Fzd, respectively, and prevent LRP-Wnt-Fz complex formation in response to Wnts (Fig. 1 b) [6].

In the rest of this section, we summarize our current understanding of the roles of Wnt/β-catenin signaling on multiple physiological and pathophysiological processes in AD brain (Fig. 2).

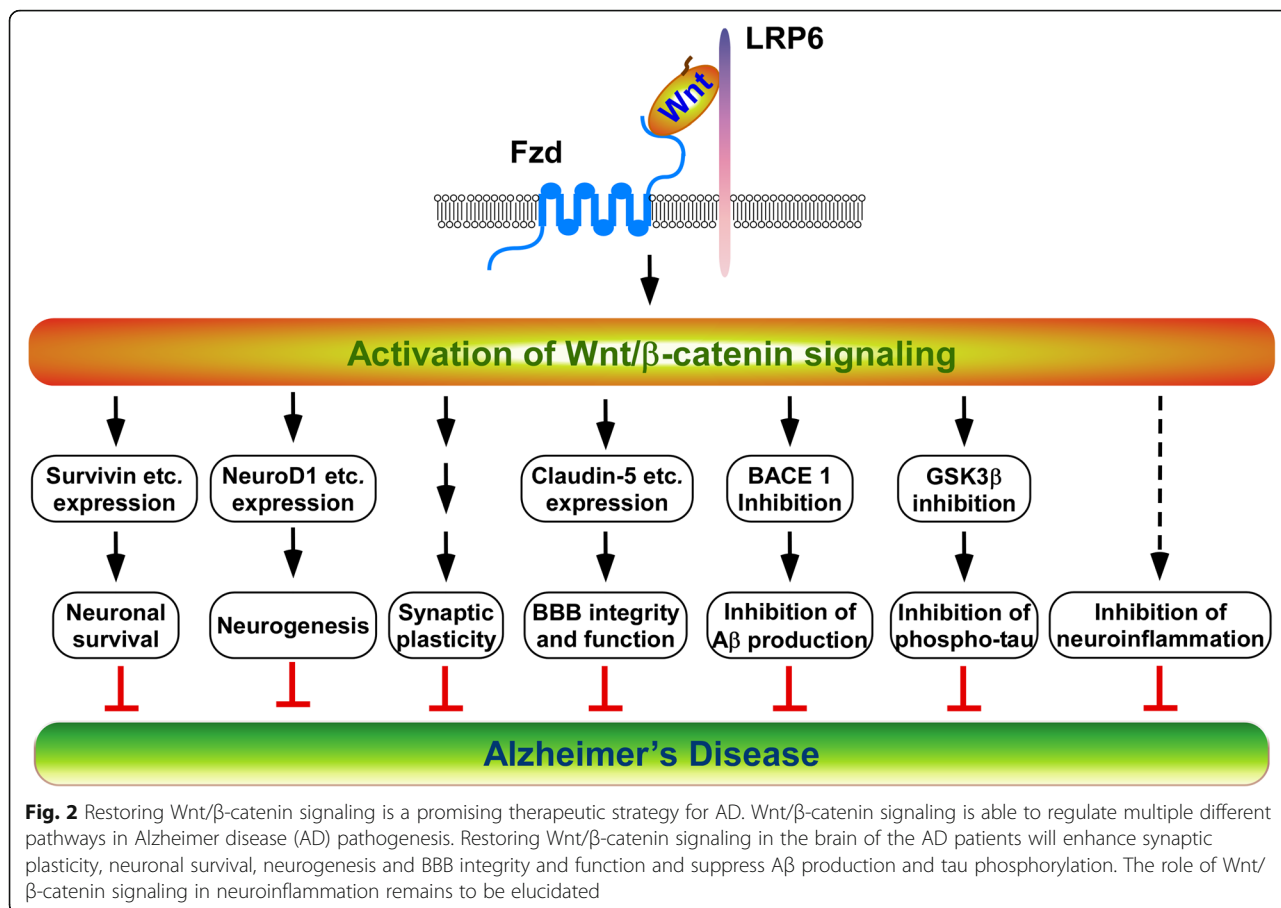
**Wnt/β-catenin signaling promotes neuronal survival and neurogenesis**

The neurodegenerative process in AD is initially characterized by synaptic damage followed by neuronal loss [12]. The Wnt/β-catenin signaling pathway is a key pathway controlling cell death and survival [6]. Indeed, loss of Wnt/β-catenin signaling renders neuron more susceptible to Aβ-induced apoptosis [13], and activation of Wnt/β-catenin signaling rescues Aβ-induced neuronal death and behavioral deficits [14–17].

While there is a debate of the presence of neurogenesis in human adult brain [18], emerging evidence suggests that human hippocampus neurogenesis persists in aged adult brain and declines dramatically in AD brain [19–23]. Importantly, numerous studies have demonstrated that Wnt/β-catenin signaling is a key regulator of adult hippocampal neurogenesis [24–34]. Wnt7a plays a critical role in multiple steps of neurogenesis by activating Wnt/β-catenin signaling and specific downstream target genes involved in cell cycle control and neuronal differentiation [32]. Moreover, astrocyte-secreted Wnt proteins are decreased in aged mice, leading to suppression of Wnt/β-catenin signaling, down-regulation of survivin levels in neural progenitor cells (NPCs) and impaired adult neurogenesis during aging [29, 33]. Interestingly, neurogenesis induced by anti-aggregant tau mutant is associated with the activation of Wnt/β-catenin signaling [34]. Mechanistically, transcriptional activation of the mitotic regulator survivin, the basic helix-loop-helix transcription factor NeuroD1 and prospero-related homeodomain transcription factor Prox1, which all are essential for the generation of granule cells in the hippocampus, is dependent on activation of the Wnt/β-catenin signaling pathway [25, 26, 33, 35].

**Wnt/β-catenin signaling enhances synaptic plasticity**

Synaptic plasticity is associated with higher brain functions such as learning and memory. Synapse loss,



which occurs prior to neuronal death at early stages in AD brain, is a major correlate of cognitive impairment in AD brain [36, 37]. Recent studies have found that Wnt/β-catenin signaling is essential for synaptic plasticity [38, 39]. Wnt proteins are not only required for synapse formation, but they can modulate neurotransmission by acting both pre- and post-synaptically [38]. Long-term potentiation (LTP) is considered a cellular correlate of learning [40], and studies have demonstrated that Wnt proteins can promote LTP [41–44]. Significantly, neuronal activity can induce the release of several Wnt proteins such as Wnt1, Wnt2, Wnt3A and Wnt7a/b [41, 44–46] and decrease the expression of Wnt antagonist sFRP3 [28]; while LTP is severely impaired by functional blockade of endogenous Wnt proteins with Wnt antagonists DKK1 and SFRPs [43, 44, 47].

LRP6 is an essential Wnt co-receptor for activation of Wnt/β-catenin signaling on the cell surface. LRP6 is selectively localized to excitatory synapses, and is required for excitatory synapse development in vitro and in vivo [48]. Moreover, neuronal deficiency of LRP6 results in synaptic and cognitive abnormalities in aged mice [49]. All together, these studies indicate

that neuronal LRP6-mediated Wnt/β-catenin signaling plays an important role in synaptic function and cognition.

DKK1 binds to LRP6 and blocks Wnt/β-catenin signaling on the cell surface. Mice with a dorsal hippocampal infusion of DKK1 exhibited impaired hippocampal-dependent novel object recognition memory with rapidly decreasing levels of key Wnt/β-catenin signaling proteins, including β-catenin, Cyclin D1, c-myc, Wnt7a, and PSD95 [50]. Induction of DKK1 expression in the hippocampus triggers synapse loss, synaptic dysfunction and memory impairment, all of which can be fully restored by reactivation of Wnt/β-catenin signaling after cessation of DKK1 expression in the hippocampus [43]. Collectively, these findings further demonstrate the critical role of LRP6-mediated Wnt/β-catenin signaling in synaptic plasticity.

**Wnt/β-catenin signaling is essential for the integrity and function of the blood-brain barrier (BBB)**

The BBB protects the brain from exposure to neurotoxic blood-derived debris, cells and microbial pathogens. Therefore, BBB disruption allows influx of harmful substances into the brain, induces inflammatory and immune

responses, and may subsequently initiate multiple pathways of neurodegeneration [51, 52]. BBB breakdown is an early biomarker of human cognitive impairment in AD [53]. It is observed before dementia, neurodegeneration and/or brain atrophy occur [54, 55]. BBB disruption is a key pathogenic feature of AD, which includes increased BBB permeability, microbleeding, diminished glucose transport, impaired Pgp-1 function (A $\beta$  clearance), perivascular accumulation of neurotoxic blood-derived products, and cellular infiltration and degeneration of pericytes and endothelial cells [51, 53, 56]. Therefore, developing novel approaches that target BBB repair is a promising strategy for AD therapy.

In the past decade, studies have established that the Wnt/ $\beta$ -catenin pathway is a key pathway required not only for BBB formation but also for BBB integrity and function [57, 58]. By binding to Wnt receptor Fzd4 and Wnt co-receptor LRP5/6, Wnt ligands Wnt7a and Wnt7b, which are mainly produced by neurons and astrocytes in brain [59], activate Wnt/ $\beta$ -catenin signaling in BBB endothelial cells (ECs) [60–62], and activation of Wnt/ $\beta$ -catenin signaling is a key driver of BBB formation and function [60–62]. In addition, Reck, a GPI-anchored membrane protein, and Gpr124, an orphan GPCR, are essential cofactors on the cell surface for Wnt7a/Wnt7b-specific signaling in mammalian CNS angiogenesis, BBB integrity and function [63–68].

Brain ECs are held together by tight junctions, in which claudins are the main constituent. In addition, glucose transporter 1 (GLUT1), which is specifically expressed in BBB ECs, is responsible for the transport of glucose from the blood into the brain; and p-glycoprotein (Pgp-1) is an active efflux transporter highly expressed on the luminal surface of BBB endothelial cells. Mechanically, claudin-1, -3 and -5, the three major claudins expressed in brain ECs [69], are the transcriptional targets of Wnt/ $\beta$ -catenin signaling in BBB ECs [60, 67, 70]. Moreover, Wnt/ $\beta$ -catenin signaling drives the expression of the BBB-specific glucose transporter GLUT1 [61] and efflux transporter Pgp-1 in BBB ECs [71].

#### **Wnt/ $\beta$ -catenin signaling inhibits BACE1 expression and suppresses A $\beta$ production/aggregation**

One of the two major hallmarks of AD is the accumulation of amyloid plaques between neurons in the brain [72, 73]. Recent studies have found that Wnt/ $\beta$ -catenin signaling is able to inhibit amyloidogenic processing of APP by suppressing the transcription of the  $\beta$ -site APP cleaving enzyme (BACE1) [74, 75]. While activation of Wnt/ $\beta$ -catenin signaling reduces A $\beta$ 42 production and aggregation, Wnt inhibition induces opposite effects on APP processing and A $\beta$ 42 production/aggregation in a cellular model [75]. Moreover, loss of Wnt/ $\beta$ -catenin

signaling induces AD-like neuropathological hallmarks in wild-type mice, and accelerates the development of AD-like pathology in an AD mouse model overexpressed human APP with two FAD mutations [76].

#### **Wnt/ $\beta$ -catenin signaling suppresses tau phosphorylation**

Another major hallmark of AD is the presence of intracellular neurofibrillary tangles (NFTs) composed of hyperphosphorylated forms of the microtubule-associated protein tau (MAPT) in neurons [72, 73, 77]. GSK3 $\beta$  is an important kinase associated with hyperphosphorylation of tau protein (p-tau) at AD-relevant phosphorylation sites [78]. Activation of Wnt/ $\beta$ -catenin signaling results in the inhibition of GSK3 $\beta$  activity and subsequent suppression of tau phosphorylation. Indeed, the Wnt antagonist DKK1 is able to inhibit Wnt/ $\beta$ -catenin signaling and induce both tau hyperphosphorylation and neuronal death [79, 80]. In contrast, activation of Wnt/ $\beta$ -catenin signaling can inhibit A $\beta$ -induced tau hyperphosphorylation and neuronal death [14, 17].

#### **Wnt/ $\beta$ -catenin signaling in microglia activity and neuroinflammation**

Glia-mediated neuroinflammation is another pathological hallmark of AD [81–83]. Genetic factors such as rare variants of TREM2 (triggering receptor expressed on myeloid cells-2) strongly increase the risk of developing AD, confirming a role of neuroinflammation as a driving force in AD [84–88]. Interestingly, TREM2, which is exclusively expressed by microglia in brain, can promote microglial survival by activating Wnt/ $\beta$ -catenin signaling through posttranslational regulation of  $\beta$ -catenin [89]. On the other hand, Wnt antagonist sFRP1 and sFRP2 act as negative modulators of the disintegrin and metalloproteinase domain 10 protein (ADAM10) [90], which is an  $\alpha$ -secretase responsible for shedding of the TREM2 ectodomain to produce soluble TREM2 (sTREM2) [86]; and recent studies indicate that sTREM2 displays a protective role in AD brain [91–93]. Moreover, activation of Wnt/ $\beta$ -catenin signaling with Wnt3a protein, LiCl, or TDZD-8 rescued microglia survival and microgliosis in *Trem2*<sup>-/-</sup> microglia and *Trem2*<sup>-/-</sup> mouse brain [89]. In addition, postnatal neuronal deletion of Wnt co-receptor LRP6 leads to microglial activation and neuroinflammation [49]. However, there are conflicting results regarding the roles of Wnt/ $\beta$ -catenin signaling on microglial activation and neuroinflammation [94]. Wnt/ $\beta$ -catenin signaling is active in microglia during neuroinflammation, raising the question as to whether enhanced Wnt/ $\beta$ -catenin signaling in microglia is harmful in AD brain [94], and further experimental work will be required to resolve this controversy.



### Wnt/ $\beta$ -catenin signaling is diminished in AD brain

While the Wnt/ $\beta$ -catenin signaling pathway is essential for brain function, this pathway is greatly suppressed via multiple pathogenic mechanisms in AD brain.

### Wnt/ $\beta$ -catenin signaling is down-regulated in the aging brain

It is well established that increasing age is the greatest risk factor for AD [95, 96]. Mounting evidence indicates a down-regulation of Wnt/ $\beta$ -catenin signaling in the aging brain, which may contribute to reduced neurogenesis and cognitive impairment [97]. In the aging brain, expression of Wnt proteins (such as Wnt 2, 3, 4, Wnt7b and Wnt10b) and disheveled (Dvl) proteins (such as Dvl2 and Dvl3) is down-regulated, while expression of Wnt antagonist DKK1 is up-regulated; leading to the suppression of Wnt/ $\beta$ -catenin signaling [29, 33, 98–100]. Importantly, the age-associated reduction in astrocytic levels of Wnt proteins impairs adult neurogenesis [29, 33], and rescue of secreted Wnt protein levels by exercise promotes adult neurogenesis [29].

### Dysregulation and malfunction of Wnt co-receptor LRP6 in AD brain

A growing body of evidence shows dysregulation and loss of function of Wnt co-receptor LRP6 contributes to down-regulation of Wnt/ $\beta$ -catenin signaling in AD. Firstly, two LRP6 SNPs and an alternatively splice variant that display impaired Wnt/ $\beta$ -catenin signaling activity, are associated with increased risk of developing AD [101, 102]. Secondly, expression of LRP6 is downregulated in AD brain [49], and deficiency in LRP6-mediated Wnt/ $\beta$ -catenin signaling contributes to synaptic dysfunction and amyloid pathology in AD [49]. Thirdly, apolipoprotein E4 (ApoE4), the most important risk factor for late-onset AD [103, 104], can inhibit Wnt/ $\beta$ -catenin signaling in neuronal LRP6-expressing PC-12 cells [105]. Finally, LRP6 physically interacts with APP and suppresses A $\beta$  production [49, 106], while the Swedish familial AD variant of APP (APP<sup>Swe</sup>) displays reduced activation of Wnt/ $\beta$ -catenin signaling [106].

### Up-regulation of DKK1 expression results in suppression of Wnt/ $\beta$ -catenin signaling in AD brain

A $\beta$  peptides can induce DKK1 expression and inhibit Wnt/ $\beta$ -catenin signaling in primary cortical neurons [80], and DKK1 expression in the adult hippocampus can induce synapse degeneration [43, 50]. Moreover, A $\beta$ -induced synaptic loss can be attenuated by DKK1-neutralizing antibodies in mouse brain slices [107]. DKK1 is upregulated in postmortem AD brain, where it colocalizes with neurofibrillary tangles and dystrophic neurites [80]. The upregulation DKK1 in AD brain and its colocalization with hyperphosphorylated tau have

been also demonstrated in transgenic AD-like mouse models [108]. Critically, there is a pathogenic-positive feedback loop with A $\beta$  stimulating DKK1 expression, thereby promoting synapse loss and driving further A $\beta$  production [106].

### Activation of GSK3 $\beta$ in AD brain

The binding of Wnt protein to Fzd/LRP results in inhibition of GSK3 $\beta$  and consequent activation of Wnt/ $\beta$ -catenin signaling [6, 109]. GSK3 $\beta$  is one of two major kinases responsible for  $\beta$ -catenin phosphorylation, and activation of GSK3 $\beta$  induces  $\beta$ -catenin phosphorylation and degradation [110]. The increased activity of GSK3 $\beta$  has been found in the brain of AD patients [111, 112], which could be resulted from the up-regulation of DKK1 and down-regulation of LRP6 in the AD brain. A recent study shows that a significant decrease in  $\beta$ -catenin protein levels is inversely associated with increased activation of GSK3 $\beta$  in the prefrontal cortical lobe structures of human AD brains [113], further strengthening the notion that GSK3 $\beta$  activity is associated with Wnt/ $\beta$ -catenin signaling in AD brain. Notably, GSK3 $\beta$  is a key kinase for tau phosphorylation, and over-activation of GSK3 $\beta$  is intimately linked to tau hyperphosphorylation, A $\beta$  deposition, plaque-associated microglial-mediated inflammatory responses and memory impairment [111, 112, 114].

### AD-associated APP mutants suppress Wnt/ $\beta$ -catenin signaling in AD brain

APP mutations can cause early-onset FAD [115, 116]. While studies using wild-type APP produced conflicting results regarding the activity of Wnt/ $\beta$ -catenin signaling, studies with FAD-associated APP mutants consistently revealed that Wnt/ $\beta$ -catenin signaling is inhibited by FAD-associated APP mutants [106, 117]. Studies in APP transgenic and knockout animal models and human AD brains demonstrated that APP and  $\beta$ -catenin co-localize and form a physical complex that is not present in healthy controls [118], and that  $\beta$ -catenin expression is greatly increased in hippocampal CA1 pyramidal cells from APP knockout mice [117]. Studies in primary neurons showed that overexpression of APP and its mutants promoted  $\beta$ -catenin degradation, while APP knockdown produced opposite effects [117].

### Regulation of Wnt/ $\beta$ -catenin signaling by PSEN1 and its AD-associated mutants in AD brain

Mutations in *PSEN1* are among the major causes of early-onset FAD [116, 119]. In the hippocampus, PSEN1 and PSEN2 play an important role in the regulation of synaptic plasticity, A $\beta$  production and intracellular Ca<sup>2+</sup> homeostasis [120, 121]. Many studies support the notion that PSEN1 and its mutants associated with FAD are

negative regulators of Wnt/ $\beta$ -catenin signaling [13, 122–128], although inconsistent results with respect to the effects of FAD-associated PSEN1 mutants on Wnt/ $\beta$ -catenin signaling have been reported [129]. In a genetic modifier screening, *Drosophila* PSEN was identified as a suppressor of wingless/Wnt signaling [125]. PSEN deficiency enhances Wnt/ $\beta$ -catenin signaling through relocalization of GSK3 to the late-endosomal compartment [130], and facilitates the stepwise phosphorylation of  $\beta$ -catenin independently of the Wnt-controlled Axin complex [126]. Moreover, the expression of  $\beta$ -catenin is reduced in AD patients carrying *PSEN1* mutations [13], and *PSEN1* mutations associated with AD cause a perturbation in the intracellular trafficking of  $\beta$ -catenin [122], decrease the stability and/or enhance the degradation of  $\beta$ -catenin [123, 124]. However, some FAD-associated PSEN1 mutants such as FAD-PSEN1<sup>L286V</sup> and -PSEN1<sup>M146L</sup> fail to induce  $\beta$ -catenin degradation [62, 131, 132]. Instead, FAD-PSEN1<sup>L286V</sup> can upregulate a subset of TCF/ $\beta$ -catenin transcription by enhancing the level of cAMP-response element-binding protein (CREB)-binding protein (CBP) [131].

### Targeting Wnt/ $\beta$ -catenin signaling in AD therapy

Giving that the Wnt/ $\beta$ -catenin pathway is greatly suppressed in the brain of AD patients, restoring Wnt/ $\beta$ -catenin signaling represents a unique opportunity for rational AD therapy (Fig. 2).

### The active lifestyle-induced cognitive improvement is associated with activation of Wnt/ $\beta$ -catenin signaling

A physically active lifestyle in adults and the elderly can improve brain health and reduce cognitive impairment associated with aging [133]. It has been reported that the enhancement of cognitive function by lifelong exercise is associated with induction of Wnt gene expression in the hippocampus [134]. Particularly, long-term moderate exercise and environmental enrichment can stimulate Wnt/ $\beta$ -catenin signaling by reducing DKK1 protein levels and increasing LRP6 and Wnt3a protein levels in hippocampus of adult animals [29, 135]. These findings suggest that activation of Wnt/ $\beta$ -catenin signaling is a potential mechanism underlying the cognitive improvement associated with an active lifestyle.

### Estrogen-induced neuroprotection is associated with inhibition of DKK1 expression

Estrogens can exert numerous protective actions in the adult brain, and reduced estrogen levels in adulthood are associated with increased risk of AD in women [136, 137]. In female rats, long-term estrogen deprivation leads to elevation of basal DKK1 expression and suppression of Wnt/ $\beta$ -catenin signaling in the CA1 hippocampal region [138]. Moreover, estrogen-induced neuroprotection and attenuation of tau

phosphorylation are associated with DKK1 inhibition and subsequent activation of Wnt/ $\beta$ -catenin signaling [139]. Together, these findings suggest that inhibition of DKK1 is a potential mechanism for estrogen-induced neuroprotection.

### GSK3 $\beta$ inhibitors

The activity of GSK3 $\beta$  is negatively regulated by Wnt/ $\beta$ -catenin signaling [6, 109]. Given the key role of GSK3 activity on the pathogenesis of AD, various GSK3 $\beta$  inhibitors have been shown to inhibit tau hyperphosphorylation and reduce A $\beta$  levels in both neuronal and nonneuronal cells, and rescue cognitive deficits in several murine models of AD [112, 140]. However, due to the wide range of GSK3 $\beta$  substrates and physiological actions, the use of GSK3 $\beta$  inhibitors in clinical studies in AD patients has been disappointing [112, 141]. Therefore, novel GSK3 $\beta$  inhibitors that selectively regulate the activity of this kinase in Wnt/ $\beta$ -catenin signaling in brain are highly desirable.

### DKK1 inhibitors

Suppression of Wnt/ $\beta$ -catenin signaling by A $\beta$ -induced up-regulation of DKK1 expression in AD brain suggests DKK1 inhibition is a potential therapeutic strategy for restoring Wnt/ $\beta$ -catenin signaling in AD [142]. Indeed, it has been found that DKK1 anti-sense oligonucleotides (ASO) attenuate neuronal apoptosis and prevent tau hyperphosphorylation in A $\beta$ -treated neurons [80], and that DKK1-neutralizing antibodies attenuate synapse loss induced by A $\beta$  in mouse brain slices [107].

A virtual screen of the National Cancer Institute database for chemical compounds identified a small molecule, IIC3 (NCI8642, galloycyanine), as a DKK1 inhibitor [143]. IIC3 can inhibit DKK1 binding to LRP6 with an IC<sub>50</sub> of 3  $\mu$ M [143], and revert DKK1-mediated inhibition of Wnt/ $\beta$ -catenin signalling [143, 144]. Moreover, IIC3 can reduce basal blood-glucose concentrations and improve glucose tolerance in mice [143]. Interestingly, IIC3 and its derivatives can decrease DKK1-induced Tau phosphorylation [145, 146]. However, it is unclear whether these galloycyanine inhibitors of DKK1 can cross the BBB.

### Other activators of Wnt/ $\beta$ -catenin signaling

WASP-1 is a small molecule Wnt activator with an EC<sub>50</sub> of about 250 nM in the Wnt reporter assays [147]. Although the exact mechanism of action of this compound is unclear, activation of Wnt/ $\beta$ -catenin signaling by bilateral intra-hippocampal infusion of WASP-1 rescues memory loss and improves synaptic dysfunction in murine models of AD [148, 149].

Curcumin, a natural compound found in the plant turmeric (*Curcuma longa*), displays protective effects in various animal models of AD [150, 151]. Studies have

shown that curcumin can potentially promote Wnt/ $\beta$ -catenin signaling by increasing the expression of Wnt proteins and Wnt co-receptor LRP5/6 and suppressing the expression of Wnt antagonist DKK1 [152, 153]. However, because of its poor brain bioavailability, curcumin is of limited use in human AD patients, and there is currently lack of clinical evidence to support its therapeutic use in AD patients [150, 151]. Recently, it has been reported that curcumin nanoparticles, which exhibit increased brain bioavailability, potently stimulate adult neurogenesis and mitigate cognitive impairment in the AD model via activation of the Wnt/ $\beta$ -catenin pathway [153].

Statins are a class of drugs typically used to lower blood levels of cholesterol by reducing the production of cholesterol by the liver, and many studies suggest that statin use might protect against AD pathology [154–158]. Several studies have shown that statins are activators of Wnt/ $\beta$ -catenin signaling [159–164]. Mechanistically, statins enhance Wnt/ $\beta$ -catenin signaling through regulation of isoprenoid synthesis, which is not associated with cholesterol levels [163]. Interestingly, it has been demonstrated that lovastatin protects neuronal cells from A $\beta$ -induced apoptosis via activation of Wnt/ $\beta$ -catenin signaling [159], and that simvastatin suppresses neural cell apoptosis and enhances locomotor recovery by stimulating Wnt/ $\beta$ -catenin signaling after spinal cord injury [164]. Moreover, simvastatin can promote Wnt/ $\beta$ -catenin signaling in the hippocampus of adult mice, and enhance neurogenesis both in cultured adult neural stem cells and the mouse hippocampus [163]. All together, these findings suggest that activation of Wnt/ $\beta$ -catenin signaling is one of the mechanisms by which statins are beneficial in AD and other neurological disorders.

## Conclusion and perspectives

Compared to a large number of Wnt inhibitors as potential agents for cancer prevention and treatment, there are only a few Wnt activators reported in the literature [6, 165]. Particularly, there are no specific BBB-permeant Wnt activators that can be used as potential candidates for the treatment of AD or other neurological disorders. While Wnt/ $\beta$ -catenin signaling is critical for synaptic plasticity, neuronal survival, neurogenesis and many other brain functions, it is greatly diminished in the brain of AD patients. Therefore, small molecule Wnt activators that restore Wnt/ $\beta$ -catenin signaling in brain, particularly those targeting Wnt antagonist DKK1, Wnt receptor LRP6 and tau regulator GSK3 $\beta$ , could represent novel therapeutic tools for the treatment for AD. In addition, emerging evidence indicates that Wnt/ $\beta$ -catenin signaling is also disrupted in other neurodegenerative disorders such as Parkinson's disease [166–171].

Thus, Wnt activators hold a great therapeutic potential for other neurological disorders.

It is well established that Wnt/ $\beta$ -catenin signaling plays a key role in the regulation of bone mineral density, and that the Wnt/ $\beta$ -catenin signaling pathway is an attractive target for therapeutic intervention to restore bone strength in patients with osteoporosis disorders [172, 173]. Interestingly, AD patients have a much greater risk of suffering osteoporosis [174–177]. In addition, low bone mineral density phenotypes are manifested in AD mouse models [178–181]. Particularly, a recent study demonstrated that Wnt/ $\beta$ -catenin signaling is disrupted both in brain and bone of the htau mouse model of tauopathy, which has an early low bone mineral density phenotype [179]. Therefore, osteoporosis and AD could share a key mechanism of pathogenesis [182], and Wnt activators might not only reduce cognitive impairment but also prevent bone loss in the AD patients.

There is always a concern that overstimulation of Wnt/ $\beta$ -catenin signaling can promote cancer because aberrant activation of Wnt/ $\beta$ -catenin signaling can lead to tumor formation [6, 109]. However, there are no reports of increased incidence of cancer in families carrying *LRP5* gain-of-function mutations, and *Sost*- or *Dkk1*-deficient animals do not have an increased risk of tumor developments [183]. Nevertheless, the therapeutic application of Wnt activators should be given precisely to restore, but not overactivate, the Wnt/ $\beta$ -catenin signaling pathway in AD patients.

## Abbreviations

AD: Alzheimer's disease; ADAM10: The disintegrin and metalloproteinase domain 10 protein; ApoE4: Apolipoprotein E4; APP: Amyloid precursor protein; ASO: Anti-sense oligonucleotides; BACE1: The  $\beta$ -site APP cleaving enzyme; BBB: Blood-brain barrier; CBP: CAMP-response element-binding protein (CREB)-binding protein; DKK: Dickkopf; Dvl: Disheveled; ECs: Endothelial cells; FAD: Familial Alzheimer's disease; Fzd: Frizzled; GLUT1: Glucose transporter 1; GSK3 $\beta$ : Glycogen synthase kinase 3 $\beta$ ; LGR: G protein-coupled receptor; LRP: Low density lipoprotein receptor-related protein; LTP: Long-term potentiation; MAPT: The microtubule-associated protein tau; NFTs: Intracellular neurofibrillary tangles; NPCs: Neural progenitor cells; Pgp-1: P-glycoprotein; PSEN: Presenilin; Rspo: R-spondin; RNF43: Ring finger protein 43; sFRP: Soluble Frizzled-related protein; sTREM2: Soluble TREM2; TCF/LEF: T-cell factor/lymphoid enhancing factor; TREM2: Triggering receptor expressed on myeloid cells-2; ZNRF3: Zinc and ring finger 3

## Acknowledgements

We thank Guojun Bu and Hongmei Li for their critical reading of the manuscript.

## Authors' contributions

YL contributed to the conception of the review, and LJ, JPC and YL contributed to the writing of the manuscript. All authors read and approved the final manuscript.

## Funding

Support for work conducted in the authors' laboratory was provided by the Ed and Ethel Moore Alzheimer's Disease Research Program Pilot Grant from the Florida Department of Health (9AZ09 to Y.LJ).

**Availability of data and materials**

Not applicable.

**Ethics approval and consent to participate**

Not applicable.

**Consent for publication**

Not applicable.

**Competing interests**

The authors declare that they have no competing interests.

**Author details**

<sup>1</sup>Department of Neuroscience, Mayo Clinic, Jacksonville, FL 32224, USA. <sup>2</sup>Fujian Provincial Key Laboratory of Neurodegenerative Disease and Aging Research, Institute of Neuroscience, Medical College, Xiamen University, Xiamen 361102, China. <sup>3</sup>Neuroscience Initiative, Sanford Burnham Prebys Medical Discovery Institute, La Jolla, CA 92037, USA.

Received: 30 October 2019 Accepted: 26 November 2019

Published online: 04 December 2019

**References**

- Selkoe DJ, Hardy J. The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO Mol Med*. 2016;8:595–608.
- Ryman DC, Acosta-Baena N, Aisen PS, Bird T, Danek A, Fox NC, et al. Symptom onset in autosomal dominant Alzheimer disease: a systematic review and meta-analysis. *Neurol*. 2014;83:253–60.
- Long JM, Holtzman DM. Alzheimer disease: an update on pathobiology and treatment strategies. *Cell*. 2019;179:312–39.
- Cao J, Hou J, Ping J, Cai D. Advances in developing novel therapeutic strategies for Alzheimer's disease. *Mol Neurodegener*. 2018;13:64.
- Futch HS, Croft CL, Truong VQ, Krause EG, Golde TE. Targeting psychological stress signaling pathways in Alzheimer's disease. *Mol Neurodegener*. 2017;12:49.
- Nusse R, Clevers H. Wnt/beta-catenin signaling, disease, and emerging therapeutic modalities. *Cell*. 2017;169:985–99.
- Inestrosa NC, Varela-Nallar L. Wnt signaling in the nervous system and in Alzheimer's disease. *J Mol Cell Biol*. 2014;6:64–74.
- Hao HX, Xie Y, Zhang Y, Charlat O, Oster E, Avello M, et al. ZNRF3 promotes Wnt receptor turnover in an R-spondin-sensitive manner. *Nat*. 2012;485:195–U76.
- Koo BK, Spit M, Jordens I, Low TY, Stange DE, van de Wetering M, et al. Tumour suppressor RNF43 is a stem-cell E3 ligase that induces endocytosis of Wnt receptors. *Nat*. 2012;488–665.
- Jiang X, Charlat O, Zamponi R, Yang Y, Cong F. Dishevelled promotes Wnt receptor degradation through recruitment of ZNRF3/RNF43 E3 ubiquitin ligases. *Mol Cell*. 2015;58:522–33.
- de Lau W, Peng WC, Gros P, Clevers H. The R-spondin/Lgr5/Rnf43 module: regulator of Wnt signal strength. *Genes Dev*. 2014;28:305–16.
- Serrano-Pozo A, Frosch MP, Masliah E, Hyman BT. Neuropathological alterations in Alzheimer disease. *Cold Spring Harb Perspect Med*. 2011;1:a006189.
- Zhang Z, Hartmann H, Do VM, Abramowski D, Sturchler-Pierrat C, Staufenbiel M, et al. Destabilization of beta-catenin by mutations in presenilin-1 potentiates neuronal apoptosis. *Nat*. 1998;395:698–702.
- Esposito G, De Filippis D, Carnuccio R, Izzo AA, Iuvone T. The marijuana component cannabidiol inhibits beta-amyloid-induced tau protein hyperphosphorylation through Wnt/beta-catenin pathway rescue in PC12 cells. *J Mol Med (Berl)*. 2006;84:253–8.
- Quintanilla RA, Munoz FJ, Metcalfe MJ, Hirschfeld M, Olivares G, Godoy JA, et al. Trolox and 17beta-estradiol protect against amyloid beta-peptide neurotoxicity by a mechanism that involves modulation of the Wnt signaling pathway. *J Biol Chem*. 2005;280:11615–25.
- De Ferranti GV, Chacon MA, Barria M, Garrido JL, Godoy JA, Olivares G, et al. Activation of Wnt signaling rescues neurodegeneration and behavioral impairments induced by beta-amyloid fibrils. *Mol Psychiatry*. 2003;8:195–208.
- Alvarez AR, Godoy JA, Mullendorff K, Olivares GH, Bronfman M, Inestrosa NC. Wnt-3a overcomes beta-amyloid toxicity in rat hippocampal neurons. *Exp Cell Res*. 2004;297:186–96.
- Sorrells SF, Paredes MF, Cebrian-Silla A, Sandoval K, Qi D, Kelley KW, et al. Human hippocampal neurogenesis drops sharply in children to undetectable levels in adults. *Nature*. 2018;555:377–81.
- Moreno-Jimenez EP, Flor-Garcia M, Terreros-Roncal J, Rabano A, Cafini F, Pallas-Bazarra N, et al. Adult hippocampal neurogenesis is abundant in neurologically healthy subjects and drops sharply in patients with Alzheimer's disease. *Nat Med*. 2019;25:554–60.
- Tobin MK, Musaraca K, Disouky A, Shetti A, Bheri A, Honer WG, et al. Human hippocampal neurogenesis persists in aged adults and Alzheimer's disease patients. *Cell Stem Cell*. 2019;24:974–82 e3.
- Meyer K, Feldman HM, Lu T, Drake D, Lim ET, Ling KH, et al. REST and neural gene network Dysregulation in iPSC models of Alzheimer's disease. *Cell Rep*. 2019;26:1112–27 e9.
- Hollands C, Tobin MK, Hsu M, Musaraca K, Yu TS, Mishra R, et al. Depletion of adult neurogenesis exacerbates cognitive deficits in Alzheimer's disease by compromising hippocampal inhibition. *Mol Neurodegener*. 2017;12:64.
- Boldrini M, Fulmore CA, Tartt AN, Simeon LR, Pavlova I, Poposka V, et al. Human hippocampal neurogenesis persists throughout aging. *Cell Stem Cell*. 2018;22:589–99 e5.
- Lie DC, Colamarino SA, Song HJ, Desire L, Mira H, Consiglio A, et al. Wnt signalling regulates adult hippocampal neurogenesis. *Nat*. 2005;437:1370–5.
- Kuwabara T, Hsieh J, Muotri A, Yeo G, Warashina M, Lie DC, et al. Wnt-mediated activation of NeuroD1 and retro-elements during adult neurogenesis. *Nat Neurosci*. 2009;12:1097–105.
- Karalay O, Doberauer K, Vadodaria KC, Knobloch M, Berti L, Miquelajaurregui A, et al. Prospero-related homeobox 1 gene (Prox1) is regulated by canonical Wnt signaling and has a stage-specific role in adult hippocampal neurogenesis. *Proc Natl Acad Sci U S A*. 2011;108:5807–12.
- Mardones MD, Andaur GA, Varas-Godoy M, Henriquez JF, Salech F, Behrens MI, et al. Frizzled-1 receptor regulates adult hippocampal neurogenesis. *Mol Brain*. 2016;9:29.
- Jang MH, Bonaguidi MA, Kitabatake Y, Sun J, Song J, Kang E, et al. Secreted frizzled-related protein 3 regulates activity-dependent adult hippocampal neurogenesis. *Cell Stem Cell*. 2013;12:215–23.
- Okamoto M, Inoue K, Iwamura H, Terashima K, Soya H, Asashima M, et al. Reduction in paracrine Wnt3 factors during aging causes impaired adult neurogenesis. *FASEB J*. 2011;25:3570–82.
- Zhu Y, Demidov ON, Goh AM, Virshup DM, Lane DP, Bulavin DV. Phosphatase WIP1 regulates adult neurogenesis and WNT signaling during aging. *J Clin Invest*. 2014;124:3263–73.
- Qiu CW, Liu ZY, Hou K, Liu SY, Hu YX, Zhang L, et al. Wip1 knockout inhibits neurogenesis by affecting the Wnt/beta-catenin signaling pathway in focal cerebral ischemia in mice. *Exp Neurol*. 2018;309:44–53.
- Qu Q, Sun G, Murai K, Ye P, Li W, Asuelime G, et al. Wnt7a regulates multiple steps of neurogenesis. *Mol Cell Biol*. 2013;33:2551–9.
- Miranda CJ, Braun L, Jiang Y, Hester ME, Zhang L, Riolo M, et al. Aging brain microenvironment decreases hippocampal neurogenesis through Wnt-mediated survivin signaling. *Aging Cell*. 2012;11:542–52.
- Joseph M, Anglada-Huguet M, Paesler K, Mandelkow E, Mandelkow EM. Anti-aggregant tau mutant promotes neurogenesis. *Mol Neurodegener*. 2017;12:88.
- Schneider R, Koop B, Schroter F, Cline J, Ingwersen J, Berndt C, et al. Activation of Wnt signaling promotes hippocampal neurogenesis in experimental autoimmune encephalomyelitis. *Mol Neurodegener*. 2016;11:53.
- DeKosky ST, Scheff SW. Synapse loss in frontal cortex biopsies in Alzheimer's disease: correlation with cognitive severity. *Ann Neurol*. 1990;27:457–64.
- Terry RD, Masliah E, Salmon DP, Butters J, DeTeresa R, Hill R, et al. Physical basis of cognitive alterations in Alzheimer's disease: synapse loss is the major correlate of cognitive impairment. *Ann Neurol*. 1991;30:572–80.
- McLeod F, Salinas PC. Wnt proteins as modulators of synaptic plasticity. *Curr Opin Neurobiol*. 2018;53:90–5.
- Buechler J, Salinas PC. Deficient Wnt signaling and synaptic vulnerability in Alzheimer's disease: emerging roles for the LRP6 receptor. *Front Synaptic Neurosci*. 2018;10:38.
- Titley HK, Brunel N, Hansel C. Toward a Neurocentric view of learning. *Neuron*. 2017;95:19–32.
- Chen J, Park CS, Tang SJ. Activity-dependent synaptic Wnt release regulates hippocampal long term potentiation. *J Biol Chem*. 2006;281:11910–6.
- Cerpa W, Gambrelli A, Inestrosa NC, Barria A. Regulation of NMDA-receptor synaptic transmission by Wnt signaling. *J Neurosci*. 2011;31:9466–71.



43. Marzo A, Galli S, Lopes D, McLeod F, Podpolny M, Segovia-Roldan M, et al. Reversal of synapse degeneration by restoring Wnt signaling in the adult Hippocampus. *Curr Biol*. 2016;26:2551–61.
44. McLeod F, Bossio A, Marzo A, Ciani L, Sibilla S, Hannan S, et al. Wnt signaling mediates LTP-dependent spine plasticity and AMPAR localization through Frizzled-7 receptors. *Cell Rep*. 2018;23:1060–71.
45. Ataman B, Ashley J, Gorczyca M, Ramachandran P, Fouquet W, Sigris SJ, et al. Rapid activity-dependent modifications in synaptic structure and function require bidirectional Wnt signaling. *Neuron*. 2008;57:705–18.
46. Wayman GA, Impey S, Marks D, Saneyoshi T, Grant WF, Derkach V, et al. Activity-dependent dendritic arborization mediated by CaM-kinase I activation and enhanced CREB-dependent transcription of Wnt-2. *Neuron*. 2006;50:897–909.
47. Ciani L, Marzo A, Boyle K, Stamatakou E, Lopes DM, Anane D, et al. Wnt signalling tunes neurotransmitter release by directly targeting Synaptotagmin-1. *Nat Commun*. 2015;6:8302.
48. Sharma K, Choi SY, Zhang Y, Nieland TJ, Long S, Li M, et al. High-throughput genetic screen for synaptogenic factors: identification of LRP6 as critical for excitatory synapse development. *Cell Rep*. 2013;5:1330–41.
49. Liu CC, Tsai CW, Deak F, Rogers J, Penuliar M, Sung YM, et al. Deficiency in LRP6-mediated Wnt signaling contributes to synaptic abnormalities and amyloid pathology in Alzheimer's disease. *Neuron*. 2014;84:63–77.
50. Fortress AM, Schram SL, Tuscher JJ, Frick KM. Canonical Wnt signaling is necessary for object recognition memory consolidation. *J Neurosci*. 2013;33:12619–26.
51. Sweeney MD, Sagare AP, Zlokovic BV. Blood-brain barrier breakdown in Alzheimer disease and other neurodegenerative disorders. *Nat Rev Neurol*. 2018;14:133–50.
52. Sweeney MD, Kisler K, Montagne A, Toga AW, Zlokovic BV. The role of brain vasculature in neurodegenerative disorders. *Nat Neurosci*. 2018;21:1318–31.
53. Ma Q, Zhao Z, Sagare AP, Wu Y, Wang M, Owens NC, et al. Blood-brain barrier-associated pericytes internalize and clear aggregated amyloid-beta42 by LRP1-dependent apolipoprotein E isoform-specific mechanism. *Mol Neurodegener*. 2018;13:57.
54. Montagne A, Zhao Z, Zlokovic BV. Alzheimer's disease: a matter of blood-brain barrier dysfunction? *J Exp Med*. 2017;214:3151–69.
55. Nation DA, Sweeney MD, Montagne A, Sagare AP, LM D'O, Pachicano M, et al. Blood-brain barrier breakdown is an early biomarker of human cognitive dysfunction. *Nat Med*. 2019.
56. Marques F, Sousa JC, Sousa N, Palha JA. Blood-brain-barriers in aging and in Alzheimer's disease. *Mol Neurodegener*. 2013;8:38.
57. Engelhardt B, Liebner S. Novel insights into the development and maintenance of the blood-brain barrier. *Cell Tissue Res*. 2014;355:687–99.
58. Liebner S, Dijkhuizen RM, Reiss Y, Plate KH, Agalliu D, Constantin G. Functional morphology of the blood-brain barrier in health and disease. *Acta Neuropathol*. 2018;135:311–36.
59. Zhang Y, Chen K, Sloan SA, Bennett ML, Scholze AR, O'Keefe S, et al. An RNA-sequencing transcriptome and splicing database of glia, neurons, and vascular cells of the cerebral cortex. *J Neurosci*. 2014;34:11929–47.
60. Liebner S, Corada M, Bangsow T, Babbage J, Taddei A, Czupalla CJ, et al. Wnt/beta-catenin signaling controls development of the blood-brain barrier. *J Cell Biol*. 2008;183:409–17.
61. Daneman R, Agalliu D, Zhou L, Kuhnert F, Kuo CJ, Barres BA. Wnt/beta-catenin signaling is required for CNS, but not non-CNS, angiogenesis. *Proc Natl Acad Sci U S A*. 2009;106:641–6.
62. Stenman JM, Rajagopal J, Carroll TJ, Ishibashi M, McMahon J, McMahon AP. Canonical Wnt signaling regulates organ-specific assembly and differentiation of CNS vasculature. *Sci*. 2008;322:1247–50.
63. Zhou Y, Nathans J. Gpr124 controls CNS angiogenesis and blood-brain barrier integrity by promoting ligand-specific canonical wnt signaling. *Dev Cell*. 2014;31:248–56.
64. Cho C, Smallwood PM, Nathans J. Reck and Gpr124 are essential receptor cofactors for Wnt7a/Wnt7b-specific signaling in mammalian CNS angiogenesis and blood-brain barrier regulation. *Neuron*. 2017;95:1221–5.
65. Chang J, Mancuso MR, Maier C, Liang X, Yuki K, Yang L, et al. Gpr124 is essential for blood-brain barrier integrity in central nervous system disease. *Nat Med*. 2017;23:450–60.
66. Vallon M, Yuki K, Nguyen TD, Chang J, Yuan J, Siepe D, et al. A RECK-WNT7 receptor-ligand interaction enables isoform-specific regulation of Wnt bioavailability. *Cell Rep*. 2018;25:339–49 e9.
67. Tran KA, Zhang X, Predescu D, Huang X, Machado RF, Gothert JR, et al. Endothelial beta-catenin signaling is required for maintaining adult blood-brain barrier integrity and central nervous system homeostasis. *Circ*. 2016;133:177–86.
68. Vanhollebeke B, Stone OA, Bostaille N, Cho C, Zhou Y, Maquet E, et al. Tip cell-specific requirement for an atypical Gpr124- and Reck-dependent Wnt/beta-catenin pathway during brain angiogenesis. *Elife*. 2015;4.
69. Main BS, Villapol S, Sloley SS, Barton DJ, Parsadanian M, Agbaeugbu C, et al. Apolipoprotein E4 impairs spontaneous blood brain barrier repair following traumatic brain injury. *Mol Neurodegener*. 2018;13:17.
70. Zhou Y, Wang Y, Tischfield M, Williams J, Smallwood PM, Rattner A, et al. Canonical WNT signaling components in vascular development and barrier formation. *J Clin Invest*. 2014;124:3825–46.
71. Lim JC, Kania KD, Wijesuriya H, Chawla S, Sethi JK, Pulaski L, et al. Activation of beta-catenin signalling by GSK-3 inhibition increases p-glycoprotein expression in brain endothelial cells. *J Neurochem*. 2008;106:1855–65.
72. Bloom GS. Amyloid-beta and tau: the trigger and bullet in Alzheimer disease pathogenesis. *JAMA Neurol*. 2014;71:505–8.
73. Li H, Liu CC, Zheng H, Huang TY. Amyloid, tau, pathogen infection and antimicrobial protection in Alzheimer's disease -conformist, nonconformist, and realistic prospects for AD pathogenesis. *Transl Neurodegener*. 2018;7:34.
74. Parr C, Mirzaei N, Christian M, Sastre M. Activation of the Wnt/beta-catenin pathway represses the transcription of the beta-amyloid precursor protein cleaving enzyme (BACE1) via binding of T-cell factor-4 to BACE1 promoter. *FASEB J*. 2015;29:623–35.
75. Tapia-Rojas C, Burgos PV, Inestrosa NC. Inhibition of Wnt signaling induces amyloidogenic processing of amyloid precursor protein and the production and aggregation of amyloid-beta (Aβ)42 peptides. *J Neurochem*. 2016;139:1175–91.
76. Tapia-Rojas C, Inestrosa NC. Wnt signaling loss accelerates the appearance of neuropathological hallmarks of Alzheimer's disease in J20-APP transgenic and wild-type mice. *J Neurochem*. 2018;144:443–65.
77. Wu XL, Pina-Crespo J, Zhang YW, Chen XC, Xu HX. Tau-mediated Neurodegeneration and Potential Implications in Diagnosis and Treatment of Alzheimer's Disease. *Chin Med J (Engl)*. 2017;130:2978–90.
78. Hernandez F, Lucas JJ, Avila J. GSK3 and tau: two convergence points in Alzheimer's disease. *J Alzheimers Dis*. 2013;33(Suppl 1):S141–4.
79. Scali C, Caraci F, Gianfriddo M, Diodato E, Roncarati R, Pollio G, et al. Inhibition of Wnt signaling, modulation of tau phosphorylation and induction of neuronal cell death by DKK1. *Neurobiol Dis*. 2006;24:254–65.
80. Caricasole A, Copani A, Caraci F, Aronica E, Rozemuller AJ, Caruso A, et al. Induction of Dickkopf-1, a negative modulator of the Wnt pathway, is associated with neuronal degeneration in Alzheimer's brain. *J Neurosci*. 2004;24:6021–7.
81. Heneka MT, Carson MJ, El Khoury J, Landreth GE, Brosseron F, Feinstein DL, et al. Neuroinflammation in Alzheimer's disease. *Lancet Neurol*. 2015;14:388–405.
82. Leyns CEG, Holtzman DM. Glial contributions to neurodegeneration in tauopathies. *Mol Neurodegener*. 2017;12:50.
83. Rangaraju S, Dammer EB, Raza SA, Rathakrishnan P, Xiao H, Gao T, et al. Identification and therapeutic modulation of a pro-inflammatory subset of disease-associated-microglia in Alzheimer's disease. *Mol Neurodegener*. 2018;13:24.
84. Efthymiou AG, Goate AM. Late onset Alzheimer's disease genetics implicates microglial pathways in disease risk. *Mol Neurodegener*. 2017;12:43.
85. Gratzue M, Leyns CEG, Holtzman DM. New insights into the role of TREM2 in Alzheimer's disease. *Mol Neurodegener*. 2018;13:66.
86. Jay TR, von Saucken VE, Landreth GE. TREM2 in neurodegenerative diseases. *Mol Neurodegener*. 2017;12:56.
87. Bemiller SM, McCray TJ, Allan K, Formica SV, Xu G, Wilson G, et al. TREM2 deficiency exacerbates tau pathology through dysregulated kinase signaling in a mouse model of tauopathy. *Mol Neurodegener*. 2017;12:74.
88. Zhao Y, Wu X, Li X, Jiang LL, Gui X, Liu Y, et al. TREM2 is a receptor for beta-amyloid that mediates microglial function. *Neuron*. 2018;97:1023–31 e7.
89. Zheng H, Jia L, Liu CC, Rong Z, Zhong L, Yang L, et al. TREM2 promotes microglial survival by activating Wnt/beta-catenin pathway. *J Neurosci*. 2017;37:1772–84.
90. Esteve P, Sandonis A, Cardozo M, Malapeira J, Ibanez C, Crespo I, et al. SFRPs act as negative modulators of ADAM10 to regulate retinal neurogenesis. *Nat Neurosci*. 2011;14:562–U43.
91. Zhong L, Xu Y, Zhuo R, Wang T, Wang K, Huang R, et al. Soluble TREM2 ameliorates pathological phenotypes by modulating microglial functions in an Alzheimer's disease model. *Nat Commun*. 2019;10:1365.

92. Ewers M, Franzmeier N, Suarez-Calvet M, Morenas-Rodriguez E, Caballero MAA, Kleinberger G, et al. Increased soluble TREM2 in cerebrospinal fluid is associated with reduced cognitive and clinical decline in Alzheimer's disease. *Sci Transl Med*. 2019;11.
93. Deming Y, Filippello F, Cignarella F, Cantoni C, Hsu S, Mikesell R, et al. The MS4A gene cluster is a key modulator of soluble TREM2 and Alzheimer's disease risk. *Sci Transl Med*. 2019;11.
94. Halleskog C, Mulder J, Dahlstrom J, Mackie K, Hortobagyi T, Tanila H, et al. WNT signaling in activated microglia is proinflammatory. *Glia*. 2011;59:119–31.
95. Herrup K. Reimagining Alzheimer's disease—an age-based hypothesis. *J Neurosci*. 2010;30:16755–62.
96. Guerreiro R, Bras J. The age factor in Alzheimer's disease. *Genome Med*. 2015;7:106.
97. Garcia-Velazquez L, Arias C. The emerging role of Wnt signaling dysregulation in the understanding and modification of age-associated diseases. *Ageing Res Rev*. 2018;17:135–45.
98. Hofmann JW, McBryan T, Adams PD, Sedivy JM. The effects of aging on the expression of Wnt pathway genes in mouse tissues. *Age (Dordr)*. 2014;36–9618.
99. Orellana AM, Vasconcelos AR, Leite JA, de Sa LL, Andreotti DZ, Munhoz CD, et al. Age-related neuroinflammation and changes in AKT-GSK-3beta and WNT/ beta-CATENIN signaling in rat hippocampus. *Ageing (Albany NY)*. 2015;7:1094–111.
100. Bayod S, Felice P, Andres P, Rosa P, Camins A, Pallas M, et al. Downregulation of canonical Wnt signaling in hippocampus of SAMP8 mice. *Neurobiol Aging*. 2015;36:720–9.
101. De Ferrari GV, Papassotiropoulos A, Biechele T, Wavrant De-Vrieze F, Avila ME, Major MB, et al. Common genetic variation within the low-density lipoprotein receptor-related protein 6 and late-onset Alzheimer's disease. *Proc Natl Acad Sci U S A*. 2007;104:9434–9.
102. Alarcon MA, Medina MA, Hu Q, Avila ME, Bustos BI, Perez-Palma E, et al. A novel functional low-density lipoprotein receptor-related protein 6 gene alternative splice variant is associated with Alzheimer's disease. *Neurobiol Aging*. 2013;34:e9–18.
103. Najm R, Jones EA, Huang Y. Apolipoprotein E4, inhibitory network dysfunction, and Alzheimer's disease. *Mol Neurodegener*. 2019;14:24.
104. Yamazaki Y, Zhao N, Caulfield TR, Liu CC, Bu G. Apolipoprotein E and Alzheimer disease: pathobiology and targeting strategies. *Nat Rev Neurol*. 2019.
105. Caruso A, Motolese M, Iacovelli L, Caraci F, Copani A, Nicoletti F, et al. Inhibition of the canonical Wnt signaling pathway by apolipoprotein E4 in PC12 cells. *J Neurochem*. 2006;98:364–71.
106. Elliott C, Rojo AI, Ribe E, Broadstock M, Xia W, Morin P, et al. A role for APP in Wnt signalling links synapse loss with beta-amyloid production. *Transl Psychiatry*. 2018;8:179.
107. Purro SA, Dickens EM, Salinas PC. The secreted Wnt antagonist Dickkopf-1 is required for amyloid beta-mediated synaptic loss. *J Neurosci*. 2012;32:3492–8.
108. Rosi MC, Luccarini I, Grossi C, Fiorentini A, Spillantini MG, Prisco A, et al. Increased Dickkopf-1 expression in transgenic mouse models of neurodegenerative disease. *J Neurochem*. 2010;112:1539–51.
109. Niehrs C. The complex world of WNT receptor signalling. *Nat Rev Mol Cell Biol*. 2012;13:767–79.
110. Liu C, Li Y, Semenov M, Han C, Baeg GH, Tan Y, et al. Control of beta-catenin phosphorylation/degradation by a dual-kinase mechanism. *Cell*. 2002;108:837–47.
111. Hooper C, Killick R, Lovestone S. The GSK3 hypothesis of Alzheimer's disease. *J Neurochem*. 2008;104:1433–9.
112. Llorens-Martin M, Jurado J, Hernandez F, Avila J. GSK-3beta, a pivotal kinase in Alzheimer disease. *Front Mol Neurosci*. 2014;7:46.
113. Folke J, Pakkenberg B, Brudek T. Impaired Wnt signaling in the prefrontal cortex of Alzheimer's disease. *Mol Neurobiol*. 2019;56:873–91.
114. Zhu H, Zhang W, Zhao Y, Shu X, Wang W, Wang D, et al. GSK3beta-mediated tau hyperphosphorylation triggers diabetic retinal neurodegeneration by disrupting synaptic and mitochondrial functions. *Mol Neurodegener*. 2018;13:62.
115. Goate A, Chartier-Harlin MC, Mullan M, Brown J, Crawford F, Fidani L, et al. Segregation of a missense mutation in the amyloid precursor protein gene with familial Alzheimer's disease. *Nat*. 1991;349:704–6.
116. Scheuner D, Eckman C, Jensen M, Song X, Citron M, Suzuki N, et al. Secreted amyloid beta-protein similar to that in the senile plaques of Alzheimer's disease is increased in vivo by the presenilin 1 and 2 and APP mutations linked to familial Alzheimer's disease. *Nat Med*. 1996;2:864–70.
117. Chen Y, Bodles AM. Amyloid precursor protein modulates beta-catenin degradation. *J Neuroinflammation*. 2007;4:29.
118. Zhang N, Parr CJ, Birch AM, Goldfinger MH, Sastre M. The amyloid precursor protein binds to beta-catenin and modulates its cellular distribution. *Neurosci Lett*. 2018;685:190–5.
119. Sherrington R, Rogaeve EI, Liang Y, Rogaeve EA, Levesque G, Ikeda M, et al. Cloning of a gene bearing missense mutations in early-onset familial Alzheimer's disease. *Nat*. 1995;375:754–60.
120. Lee SH, Lutz D, Mossalam M, Bolshakov VY, Frotscher M, Shen J. Presenilins regulate synaptic plasticity and mitochondrial calcium homeostasis in the hippocampal mossy fiber pathway. *Mol Neurodegener*. 2017;12:48.
121. Zoltowska KM, Maesako M, Lushnikova I, Takeda S, Keller LJ, Skibo G, et al. Dynamic presenilin 1 and synaptotagmin 1 interaction modulates exocytosis and amyloid beta production. *Mol Neurodegener*. 2017;12:15.
122. Nishimura M, Yu G, Levesque G, Zhang DM, Ruel L, Chen F, et al. Presenilin mutations associated with Alzheimer disease cause defective intracellular trafficking of beta-catenin, a component of the presenilin protein complex. *Nat Med*. 1999;5:164–9.
123. Weihl CC, Miller RJ, Roos RP. The role of beta-catenin stability in mutant PS1-associated apoptosis. *Neuroreport*. 1999;10:2527–32.
124. Weihl CC, Ghadge GD, Kennedy SG, Hay N, Miller RJ, Roos RP. Mutant presenilin-1 induces apoptosis and downregulates Akt/PKB. *J Neurosci*. 1999;19:5360–9.
125. Cox RT, McEwen DG, Myster DL, Duronio RJ, Loureiro J, Peifer M. A screen for mutations that suppress the phenotype of *Drosophila* armadillo, the beta-catenin homolog. *Genet*. 2000;155:1725–40.
126. Kang DE, Soriano S, Xia X, Eberhart CG, De Strooper B, Zheng H, et al. Presenilin couples the paired phosphorylation of beta-catenin independent of axin: implications for beta-catenin activation in tumorigenesis. *Cell*. 2002;110:751–62.
127. Noll E, Medina M, Hartley D, Zhou J, Perrimon N, Kosik KS. Presenilin affects arm/beta-catenin localization and function in *Drosophila*. *Dev Biol*. 2000;227:450–64.
128. Soriano S, Kang DE, Fu M, Pestell R, Chevallier N, Zheng H, et al. Presenilin 1 negatively regulates beta-catenin/T cell factor/lymphoid enhancer factor-1 signaling independently of beta-amyloid precursor protein and notch processing. *J Cell Biol*. 2001;152:785–94.
129. Boonen RA, van Tijn P, Zivkovic D. Wnt signaling in Alzheimer's disease: up or down, that is the question. *Ageing Res Rev*. 2009;8:71–82.
130. Dobrowolski R, Vick P, Ploper D, Gumper I, Snitkin H, Sabatini DD, et al. Presenilin deficiency or lysosomal inhibition enhances Wnt signaling through relocalization of GSK3 to the late-endosomal compartment. *Cell Rep*. 2012;2:1316–28.
131. Teo JL, Ma H, Nguyen C, Lam C, Kahn M. Specific inhibition of CBP/beta-catenin interaction rescues defects in neuronal differentiation caused by a presenilin-1 mutation. *Proc Natl Acad Sci U S A*. 2005;102:12171–6.
132. Kang DE, Soriano S, Frosch MP, Collins T, Naruse S, Sisodia SS, et al. Presenilin 1 facilitates the constitutive turnover of beta-catenin: differential activity of Alzheimer's disease-linked PS1 mutants in the beta-catenin-signaling pathway. *J Neurosci*. 1999;19:4229–37.
133. Kirk-Sanchez NJ, McGough EL. Physical exercise and cognitive performance in the elderly: current perspectives. *Clin Interv Aging*. 2014;9:51–62.
134. Stranahan AM, Lee K, Becker KG, Zhang Y, Maudsley S, Martin B, et al. Hippocampal gene expression patterns underlying the enhancement of memory by running in aged mice. *Neurobiol Aging*. 2010;31:1937–49.
135. Bayod S, Mennella I, Sanchez-Roige S, Lalanza JF, Escorihuela RM, Camins A, et al. Wnt pathway regulation by long-term moderate exercise in rat hippocampus. *Brain Res*. 2014;1543:38–48.
136. Pike CJ. Sex and the development of Alzheimer's disease. *J Neurosci Res*. 2017;95:671–80.
137. Merlo S, Spampinato SF, Sortino MA. Estrogen and Alzheimer's disease: still an attractive topic despite disappointment from early clinical results. *Eur J Pharmacol*. 2017;817:51–8.
138. Scott EL, Zhang QG, Han D, Desai BN, Brann DW. Long-term estrogen deprivation leads to elevation of Dickkopf-1 and dysregulation of Wnt/beta-catenin signaling in hippocampal CA1 neurons. *Steroids*. 2013;78:624–32.
139. Zhang QG, Wang R, Khan M, Mahesh V, Brann DW. Role of Dickkopf-1, an antagonist of the Wnt/beta-catenin signaling pathway, in estrogen-induced neuroprotection and attenuation of tau phosphorylation. *J Neurosci*. 2008;28:8430–41.

140. Maqbool M, Mobashir M, Hoda N. Pivotal role of glycogen synthase kinase-3: a therapeutic target for Alzheimer's disease. *Eur J Med Chem*. 2016;107:63–81.
141. Godyn J, Jonczyk J, Panek D, Malawska B. Therapeutic strategies for Alzheimer's disease in clinical trials. *Pharmacol Rep*. 2016;68:127–38.
142. Ren C, Gu X, Li H, Lei S, Wang Z, Wang J, et al. The role of DKK1 in Alzheimer's disease: a potential intervention point of brain damage prevention? *Pharmacol Res*. 2019;144:331–5.
143. Li X, Shan J, Chang W, Kim I, Bao J, Lee HJ, et al. Chemical and genetic evidence for the involvement of Wnt antagonist Dickkopf2 in regulation of glucose metabolism. *Proc Natl Acad Sci U S A*. 2012;109:11402–7.
144. Iozzi S, Remelli R, Lelli B, Diamanti D, Pileri S, Bracci L, et al. Functional characterization of a small-molecule inhibitor of the DKK1-LRP6 interaction. *ISRN Mol Biol*. 2012;2012:823875.
145. Mpousis S, Thysiadis S, Avramidis N, Katsamakos S, Efthimiopoulos S, Sarli V. Synthesis and evaluation of galloyl cyanine dyes as potential agents for the treatment of Alzheimer's disease and related neurodegenerative tauopathies. *Eur J Med Chem*. 2016;108:28–38.
146. Thysiadis S, Katsamakos S, Mpousis S, Avramidis N, Efthimiopoulos S, Sarli V. Design and synthesis of galloyl cyanine inhibitors of DKK1/LRP6 interactions for treatment of Alzheimer's disease. *Bioorg Chem*. 2018;80:230–44.
147. Beaumont V, Thompson SA, Choudhry F, Nuthall H, Glantschnig H, Lipfert L, et al. Evidence for an enhancement of excitatory transmission in adult CNS by Wnt signaling pathway modulation. *Mol Cell Neurosci*. 2007;35:513–24.
148. Vargas JY, Fuenzalida M, Inestrosa NC. In vivo activation of Wnt signaling pathway enhances cognitive function of adult mice and reverses cognitive deficits in an Alzheimer's disease model. *J Neurosci*. 2014;34:2191–202.
149. Vargas JY, Ahumada J, Arrazola MS, Fuenzalida M, Inestrosa NC. WSP-1, a canonical Wnt signaling potentiator, rescues hippocampal synaptic impairments induced by Abeta oligomers. *Exp Neurol*. 2015;264:14–25.
150. Farkhondeh T, Samarghandian S, Pourbagher-Shahri AM, Sedaghat M. The impact of curcumin and its modified formulations on Alzheimer's disease. *J Cell Physiol*. 2019;234:16953–65.
151. Sanei M, Saberi-Demneh A. Effect of curcumin on memory impairment: a systematic review. *Phytomedicine*. 2019;52:98–106.
152. Zhang X, Yin WK, Shi XD, Li Y. Curcumin activates Wnt/beta-catenin signaling pathway through inhibiting the activity of GSK-3beta in APPsw transfected SY5Y cells. *Eur J Pharm Sci*. 2011;42:540–6.
153. Tiwari SK, Agarwal S, Seth B, Yadav A, Nair S, Bhatnagar P, et al. Curcumin-loaded nanoparticles potently induce adult neurogenesis and reverse cognitive deficits in Alzheimer's disease model via canonical Wnt/beta-catenin pathway. *ACS Nano*. 2014;8:76–103.
154. Cramer C, Haan MN, Galea S, Langa KM, Kalbfleisch JD. Use of statins and incidence of dementia and cognitive impairment without dementia in a cohort study. *Neurol*. 2008;71:344–50.
155. Bettermann K, Arnold AM, Williamson J, Rapp S, Sink K, Toole JF, et al. Statins, risk of dementia, and cognitive function: secondary analysis of the ginkgo evaluation of memory study. *J Stroke Cerebrovasc Dis*. 2012;21:436–44.
156. Smith KB, Kang P, Sabbagh MN. Alzheimer's Disease Neuroimaging I. The Effect of Statins on Rate of Cognitive Decline in Mild Cognitive Impairment. *Alzheimers Dement (N Y)*. 2017;3:149–56.
157. Steenland K, Zhao L, Goldstein FC, Levey AI. Statins and cognitive decline in older adults with normal cognition or mild cognitive impairment. *J Am Geriatr Soc*. 2013;61:1449–55.
158. Heymann AD, Ravona-Springer R, Moshier EL, Godbold J, Beerli MS. Statin use is associated with better cognitive function in elderly with type 2 diabetes. *J Alzheimers Dis*. 2015;47:55–9.
159. Salins P, Shawesh S, He Y, Dibrov A, Kashour T, Arthur G, et al. Lovastatin protects human neurons against Abeta-induced toxicity and causes activation of beta-catenin-TCF/LEF signaling. *Neurosci Lett*. 2007;412:211–6.
160. Lin CL, Cheng H, Tung CW, Huang WJ, Chang PJ, Yang JT, et al. Simvastatin reverses high glucose-induced apoptosis of mesangial cells via modulation of Wnt signaling pathway. *Am J Nephrol*. 2008;28:290–7.
161. Biechele TL, Camp ND, Fass DM, Kulikauskas RM, Robin NC, White BD, et al. Chemical-genetic screen identifies riluzole as an enhancer of Wnt/beta-catenin signaling in melanoma. *Chem Biol*. 2010;17:1177–82.
162. Qiao LJ, Kang KL, Heo JS. Simvastatin promotes osteogenic differentiation of mouse embryonic stem cells via canonical Wnt/beta-catenin signaling. *Mol Cells*. 2011;32:437–44.
163. Robin NC, Agoston Z, Biechele TL, James RG, Berndt JD, Moon RT. Simvastatin promotes adult hippocampal neurogenesis by enhancing Wnt/beta-catenin signaling. *Stem Cell Rep*. 2014;2:9–17.
164. Gao K, Shen Z, Yuan Y, Han D, Song C, Guo Y, et al. Simvastatin inhibits neural cell apoptosis and promotes locomotor recovery via activation of Wnt/beta-catenin signaling pathway after spinal cord injury. *J Neurochem*. 2016;138:139–49.
165. Kahn M. Can we safely target the WNT pathway? *Nat Rev Drug Discov*. 2014;13:513–32.
166. Inestrosa NC, Arenas E. Emerging roles of Wnts in the adult nervous system. *Nat Rev Neurosci*. 2010;11:77–86.
167. Arenas E. Wnt signaling in midbrain dopaminergic neuron development and regenerative medicine for Parkinson's disease. *J Mol Cell Biol*. 2014;6:42–53.
168. Cantuti-Castelvetri I, Keller-McGandy C, Bouzou B, Asteris G, Clark TW, Froesch MP, et al. Effects of gender on nigral gene expression and parkinson disease. *Neurobiol Dis*. 2007;26:606–14.
169. L'Episcopo F, Tirolo C, Testa N, Caniglia S, Morale MC, Cossetti C, et al. Reactive astrocytes and Wnt/beta-catenin signaling link nigrostriatal injury to repair in 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine model of Parkinson's disease. *Neurobiol Dis*. 2011;41:508–27.
170. Berwick DC, Javaheri B, Wetzel A, Hopkinson M, Nixon-Abell J, Granno S, et al. Pathogenic LRRK2 variants are gain-of-function mutations that enhance LRRK2-mediated repression of beta-catenin signaling. *Mol Neurodegener*. 2017;12:9.
171. Salasova A, Yokota C, Potesil D, Zdrahal Z, Bryja V, Arenas E. A proteomic analysis of LRRK2 binding partners reveals interactions with multiple signaling components of the WNT/PCP pathway. *Mol Neurodegener*. 2017;12:54.
172. Baron R, Kneissel M. WNT signaling in bone homeostasis and disease: from human mutations to treatments. *Nat Med*. 2013;19:179–92.
173. Amjadi-Moheb F, Akhavan-Niaki H. Wnt signaling pathway in osteoporosis: epigenetic regulation, interaction with other signaling pathways, and therapeutic promises. *J Cell Physiol*. 2019.
174. Melton LJ 3rd, Beard CM, Kokmen E, Atkinson EJ, O'Fallon WM. Fracture risk in patients with Alzheimer's disease. *J Am Geriatr Soc*. 1994;42:614–9.
175. Johansson C, Skoog I. A population-based study on the association between dementia and hip fractures in 85-year olds. *Aging (Milano)*. 1996;8:189–96.
176. Looker AC, Borrud LG, Dawson-Hughes B, Shepherd JA, Wright NC. Osteoporosis or low bone mass at the femur neck or lumbar spine in older adults: United States, 2005–2008. *NCHS Data Brief*. 2012;1–8.
177. Amouzougan A, Lafaie L, Marotte H, Denarie D, Collet P, Pallot-Prades B, et al. High prevalence of dementia in women with osteoporosis. *Joint Bone Spine*. 2017;84:611–4.
178. Dengler-Criss CM, Smith MA, Wilson GN. Early evidence of Low bone density and decreased serotonergic synthesis in the dorsal raphe of a Tauopathy model of Alzheimer's disease. *J Alzheimers Dis*. 2017;55:1605–19.
179. Dengler-Criss CM, Ball HC, Lin L, Novak KM, Cooper LN. Evidence of Wnt/beta-catenin alterations in brain and bone of a tauopathy mouse model of Alzheimer's disease. *Neurobiol Aging*. 2018;67:148–58.
180. Yang MW, Wang TH, Yan PP, Chu LW, Yu J, Gao ZD, et al. Curcumin improves bone microarchitecture and enhances mineral density in APP/PS1 transgenic mice. *Phytomedicine*. 2011;18:205–13.
181. Cui S, Xiong F, Hong Y, Jung JU, Li XS, Liu JZ, et al. APPsw/Abeta regulation of osteoclast activation and RAGE expression in an age-dependent manner. *J Bone Miner Res*. 2011;26:1084–98.
182. Dengler-Criss CM, Eleftheriou F. Shared mechanisms: osteoporosis and Alzheimer's disease? *Aging-Us*. 2019;11:1317–8.
183. Monroe DG, McGee-Lawrence ME, Oursler MJ, Westendorf JJ. Update on Wnt signaling in bone cell biology and bone disease. *Gene*. 2012;492:1–18.

## Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.