

# TITLE:

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## CITATION:

Ueda, Katsumi. Structure of Plant Cells with Special Reference to Lower Plants (II): Feulgen's Nucleal Staining in Some Algae. Memoirs of the College of Science, University of Kyoto. Series B 1956, 23(1): 87-91

**ISSUE DATE:** 

1956-12-30

URL:

http://hdl.handle.net/2433/258473

**RIGHT:** 



Structure of Plant Cells with Special Reference to Lower Plants. II. Feulgen's Nucleal Staining in Some Algae\*

By

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(Received 20 Oct. 1956)

While many investigators have reported on Feulgen's nucleal staining in algae, some of the results obtained by them do not always agree one another, especially in case of cyanophyta and conjugatae (cf. Kiesel and Doinikowa, 1937, Yamaha and Suematsu, 1938, Hil'ary, 1939).

It is the purpose of the present investigation to employ the Feulgen staining in various divisions of algae with a special attention to cyanophyta and conjugatae.

#### Material and Methods

Nuclei of algae which belong to seven divisions were used for the material. Materials were fixed with ethanol, corrosive sublimate-ethanol mixture, or corrosive sublimate-acetic acid mixture, followed by washing with water. After washing, they were hydrolyzed with N-HCl in toto, or, if necessary, in microtome sections, for 3, 5, 10, 15, and 20 minutes. After the hydrolyzed materials were stained with Schiff's reagent \*\* from 2 to 4 hours at room temperature (18–25°C), they were washed well with SO<sub>2</sub>-water, then were observed with a microscope. In many species, however, the materials were sealed with canada balsam after dehydration and clearing.

Besides the nucleal staining, pyronin-methylgreen double staining was carried out to see the relation between the nucleal staining and the double staining.

#### Observations

Results of the observation are presented in the following table.

<sup>\*</sup> This investigation was performed with the Grant for Fundamental Scientific Research.

<sup>\*\*</sup> Schiff's reagent was prepared by reducing "Merck" basic fuchsin with natrium bisulfite,

	Species	Fixatives	Time of optimum hydrolysis	Feulgen staining	Preparations	Pyronin methlgreen staining
Cyanophyta	Nostoc commune	E,S-A,S-E		-	in toto	Cells are stained diffusely, but not differentially.
	N. carneum	"		_		
	N. piscinale	"		_		
	Oscillatoria Annae	"		_		
	O. tenuis	"		_		
	O. limosa	"			in toto &	
	O. princeps	"	4 .	_	microtome sections*	
	O. putrida	"		_	in toto	
	Cylindrospermum Michailovskoënse	"		_		
	Brachytrichia balami	"		_		
	Gelidium elegans	S-A	5	+	tetraspore mother cells microtome	Chromonemata and nucleoli are stained green and red respectively, but the color intensity is very faint.
Rh	Nemastoma Nakamurae	"	5	+		
Rhodophyta	Grateloupia affinis	"	5	4-	sect ons *	
	$Gloiosiphonia \ calmichael$	″	5	+ .	vegetative cells in toto	
	Corallina sp.	″	10	+		
Ch.	Synedra ulna	S-A	5	-+-	in toto	Chromonemata and nucleoli are well differentially
	Gyrosigmasp.	E	5	+		
	Cymbella tumida	S-A, E	5	+		
	C. affinis	E	5	+		
Chrysophyte	Surirella elegans	S-A,S-E	5	+		
hqc	S. lanceolata	S-A	5	+		stained green and red respectively.
yta	Dityrum Brightwelli	E	5			
	$Planktoniella\ sol$	"	5	+		
	Biddulphia sinensis	"	5	+		
	Coscinodiscus gigas	"	10	+		
Pyrrophyta	Dinophisis candina	E-A, E	10	+	in toto	Chromonemata and nucleoli are differentially stained green and red respectively.
	Ceratium hirundinella	S-A	15	+		
	C. trichoceros	S-E, E	15	+		
	C. furca	"	15	+		
	C. fusus	• #	15	-+-		
	Sargassum Horneri					
	egg epid.	S-A	5	± +	microtome sections	

Phaeophyta	S. piluliferum					
	egg epid.	"	5	± +		
	S. Thunbergii					
	egg epid.	"	5	± +		Egg nuclei are
	S. tortile		acaman acaman			diffusely stained
	egg epid.	"	5	± +		but chromonemate and nucleoli in a cell of conceptacle epidermis are differentially stained.
	S. hemiphyllum					
	egg epid.	"	5	± +		
	Cystophyllum sisymbrioides egg	ır		==		
	epid.	"	5	+		
	Hijikia fusiforme		mi constanta			
	egg	"	_	±		,
	epid.		5	+		
	Padina arborescens	"	10	+	in toto	
	Colpomenia sinosa	"	5	+		
Euglenophyta	Euglena gracilis	E, S-A, S-E	5	+	in toto & microtome sections*	Chromonemata and a nucleolus are well differentially stained green and red respectively.
	E. spirogyra	S-A	5	+	in toto	
	Facus sp.		5	+		
Chlorophyta	Closterium sp.	S-A			in toto & microtome	
	Spirogyra setiformis	S-A, E		_	sections *	
	S. sp.	S-A, E				· .
	Zygnema sp.	S-A			in toto & microtome	
	Micrasterias sp.	"		_	sections	
	Nitella sp.	"	10	+		:

#### Abbreviations:

S-A; corrosive sublimate-acetic acid mixture. S-E; corrosive sublimate-ethanol mixture.

It is seen in this table that the nuclei of whole species in cyanophyta and conjugatae were negative with the nucleal staining.

Among Feulgen positive algae, it was observed in general that related species took nearly equal time of optimum hydrolysis and were stained with similar tone and intensity, and that chromonemata and nucleoli were stained differentially green and red respectively when they were stained by methylgreen and pyronin.

E; ethanol. epid.; conceptacle epidermis.

<sup>\*</sup> Microtome sections were sealed by canada balsam after dehydration and clearing.

Results of the Feulgen staining in permanent preparations were not different from those obtained by the observation immediately after washing with SO<sub>2</sub>-water.

The nuclei of the algae belonging to pyrrophyta were colored most intensely with the Feulgen staining, while those of rhodophyta and phaeophyta were stained very faintly, so far as the material used in the present study concerns.

Generally speaking, marine chrysophyta seem to be stained more weakly than the species in freshwater.

#### Discussion and Conclusion

It has been reported by Poljansky and Petruschewsky (1929), Yamaha and Suematsu (1938), Hillary (1939), and Drawert\* (1949), that cells of Oscillatoria are Feulgen positive. Other genera among cyanophyta have also been reported that they show positive nucleal staining by Poljansky and Petruschewsky (1929) in Spirulina, Tolypothrix, Gloeotrichia, and by Bringmann (1950) in Lyngbya. On the other hand, it has been stated by Kiesel and Doinikowa (1937) in Oscillatoria and Tolypothrix, by Herbst (1953, 1954) in Oscillatoria, Aphanotheca, Pseudoanabaena, and by Shinke (unpublished) and Oura (unpublished) in Oscillatoria and other blue green algae that the cells are Feulgen negative. In the present investigation, it was observed that many species of cyanophyta showed negative nucleal staining in sectioned preparations as well as in toto preparations. Bringmann (1950) has reported that cells of Oscillatoria are Feulgen positive when the cells are treated with lanthanum acetate before hydrolysis. In the present investigation, however, the cells of Oscillatoria and Nostoc did not show positive Feulgen staining though they were treated with lanthanum acetate.

Besides cyanophyta, conjugatae is also a problematic group. For instance, Geitler (1935), Yamaha and Suematsu (1938), and Hillary (1939) have reported that the nuclei of Spirogyra are Feulgen positive, while Shinke and Shigenaga (1933), Kiesel and Doinikowa (1937) have reported that Spirogyra is not stained by the Feulgen method. Besides Spirogyra, Yamaha and Suematsu (1938) in Zygnema and Hillary (1939) in Closterium have observed that these algae show positive nucleal staining. Four genera of conjugatae used in the present work, however, showed negative nucleal staining. It must be noted here that the nuclei of Spirogyra setiformis was Feulgen negative throughout whole mitotic cycle in permanent preparations (Ueda, in press).

Westbrook (1930, 1935), Kiesel and Doinikowa (1937), and Yamaha and Suematsu (1938) in a few species of rhodophyta, Kiesel and Doinikowa (1937) and Hillary (1939) in a few species of chrysophyta, Papenfuss (1934) and Kiesel and Doinikowa (1937) in a few species of phaeophyta have reported that these algae show positive nucleal staining. These results seem to be not disaccord with those of the present investigation.

In egg nuclei of Sargassum, the coloration was so faint that the present

<sup>\*</sup> cited from Bringmann (1950)

author was not able to determine whether the Feulgen staining was positive or negative.

## Summary

Feulgen's nucleal staining was carried out in forty eight species among seven divisions of algae.

Whole species of cyanophyta and conjugatae were Feulgen negative, and other species were positive so far as the material used in the present investigation concerns.

Related species seem to take an equal time of optimum hydrolysis and to stain similar tone and intensity.

Nucleoli were Feulgen negative in all species.

The author wishes to express his sincere thanks to Professor N. Shinke for his advice and criticism throughout this work.

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