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1 2	Selection of new chemicals to be used in conditioned aversion for non-lethal predation control					
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16 Abstract

The use of conditioned food aversion (CFA) can reduce the predation conflict and 17 therefore the incidence of illegal poisoning, which is one of the most important 18 conservation threats for predators and scavengers around the world. CFA is a robust 19 learning paradigm that occurs when animals associate a food with a discomfort 20 21 induced by a chemical, thereby avoiding that food in subsequent encounters. We reviewed the potential of 167 chemical compounds to be used in CFA, considering 22 23 effects, margin of safety, accessibility, and detectability. After the review, 15 compounds fulfilled the required characteristics, but only five were finally selected to 24 be tested in CFA assays with dogs. Of the tested compounds, thiabendazole, thiram 25 26 and levamisole caused target food rejection by dogs and reduced the time spent eating 27 during post-conditioning. However, despite being microencapsulated, levamisole appeared to be detectable by dogs, whereas thiram and thiabendazole were not. 28 Fluconazole and fluralaner did not produce any CFA effect. Thiabendazole, thiram and 29 30 levamisole can therefore induce CFA, and thus are potential candidates as aversive compounds for wildlife management. Thiram is a new undetectable, safe and 31 32 accessible compound that can induce CFA in canids, and opens new possibilities to 33 develop methods of non-lethal predation control.

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Keywords: Conditioned taste aversion; learned aversion; predation conflict; non-lethal
 predator control; wildlife management

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38 1. Introduction

40 Predation conflict between humans and predators has been occurring since the prehistoric age. This conflict is especially pronounced almost everywhere where the 41 42 medium-large wild canids such as red foxes (Vulpes vulpes), coyotes (Canis latrans) or grey wolves (Canis lupus) coexist with livestock or game species (Macdonald and 43 Sillero-Zubiri, 2004). Humans try to avoid damages caused by wildlife to their crops, 44 45 livestock and game species, by using mainly lethal predator control (Reynolds & Tapper 1996; Bergstrom et al. 2014). Lethal predator control has negative effects on the 46 47 ecosystems (Gordon et al. 2017; Wallach et al. 2017) and endangered species (Margalida and Mateo 2019), and nowadays, its social acceptability is low and the 48 public demands non-lethal approaches for wildlife control (Cowan et al. 2000; 49 50 Bergstrom 2017).

51 Conditioned food aversion (CFA) is a non-lethal predation control method that has been rarely used but is considered a potential tool to reduce predation of wildlife 52 (Nicolaus et al. 1989a; Cowan et al. 2000). CFA is a natural mechanism in animals to 53 prevent poisoning and intoxications (Gustavson et al. 1974). Thus, a toxic food is 54 avoided after an illness induced by the ingestion of a non-lethal dose of that food 55 56 (Garcia et al. 1974). CFA can be induced experimentally by adding a chemical 57 substance in a specific food to which it is intended to create an aversion. The correct 58 selection of the aversive compound is very important to induce an effective CFA, which must comply with several characteristics: (1) to induce slight gastrointestinal adverse 59 effects as vomit or diarrhoea; (2) to have a wide (or great) acute margin of safety 60 61 (MOS), which means a high toxic dose together with a low effective dose, that is also 62 required to avoid intoxications in case that a single individual monopolizes and 63 consumes numerous doses; (3) to have a short period of latency (30 min-two hours) to

assure the correct learning of CFA (Garcia et al. 1974); and (4) to be odourless and
tasteless to avoid its detection by the predators (Gentle et al. 2004; Nielsen et al.
2015).

CFA has been experimentally studied in rodents (Gill et al. 2000; Massei and 67 Cowan 2002), wild birds (Nicolaus et al. 1989b; Avery et al. 1995), wild mammals 68 69 (Nicolaus et al. 1989c; Norbury et al. 2005) and reptiles (Price-Rees et al. 2013). Several chemical compounds have been shown effective to induce CFA, but most of them do 70 71 not accomplished with all characteristics required for CFA, and more effective CFA 72 compounds need to be found (Gill et al. 2000). Issues of safety and detectability severely limit the number of compounds that may be used for practical applications of 73 74 CFA in wildlife management. The application of the CFA with safety must be one of the 75 main characteristics of the potential candidates for its use in wildlife management. Up to now, the CFA experiments performed have not taken into consideration this 76 77 important issue, and some had to be discarded due to their high toxicity or detrimental 78 effects on animal health (Conover 1989; Dueser et al. 2018). The other major problem 79 is that the detectability of the compounds by the animals causes rapid discrimination 80 between foods with and without CFA compounds (Burns 1980; Nicolaus and Nellis 1987; Gentle et al. 2004; Nielsen et al. 2015). In this case, if the compound added to 81 82 the bait is detected, the animals associate illness with the substance and avoids only treated baits, acting as secondary repellent (Sayre and Clark 2001; Cagnacci et al. 83 84 2005). Microencapsulation is a potential way of masking the odour or taste, but it 85 requires experimental research to test their effectivity (Cotterill et al. 2006; Shukla et 86 al. 2011). Expensive chemical compounds that need to be fabricated and/or handled 87 under special conditions, and/or cannot be preserved in the field for a long time, are

not suitable for their use as aversive for wildlife management. Hence, the search for new, safe, accessible and undetectable compounds is paramount for the development of CFA for wildlife management. For these reasons, we reviewed the compounds previously used in CFA and assessed potential new candidates that accomplish the desirable characteristics as aversive compounds for canids. Finally, we evaluated the potential of five selected compounds in a pilot study to induce CFA in penned dogs, to test them as potential candidates to be applied as aversive for wildlife management.

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96 2. Material and methods

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98 2.1 Review and selection of aversive compounds

We conducted a literature review using Web of Science, Scopus, Toxnet 99 databases, Google Scholar and databases of chemical substances registered in Spain 100 101 (i.e. pesticides, biocides and pharmaceuticals) in order to identify: (1) substances that 102 have been used in CFA in canids or other wildlife species, (2) substances that have been described as potential CFA inducers in rats, (3) other new substances of the same 103 104 chemical family as the known aversive substances that could be candidates to be 105 tested in CFA, (4) LD₅₀ in rat and dog of the selected substances, (5) odour and taste of 106 the selected substances, (6) potential doses to produce CFA in canids, and (7) commercial availability of the product. Based on the LD₅₀ values in rat and the 107 108 potential doses to induce CFA we calculated the expected acute MOS of each 109 substance. The acute MOS has been calculated as the ratio between LD₅₀ in rat and the 110 potential dose to induce CFA. The purpose of this literature review was to identify 111 substances that have been successfully used to induce CFA and to identify new

112 candidates that could be used in further experimental CFA tests with canids. 113 Therefore, the selected substances should have an acute MOS above 10 (i.e. the lethal 114 dose should be at least 10-fold higher than the potential CFA dose). Moreover, the 115 selected compounds should be also odourless and tasteless, and available in the 116 market with an approved use as a veterinary drug or other use.

117 A total of 167 substances were included in the literature review (see full list in supplementary material, Table S1). Based on the data obtained from all these 118 119 substances, we first selected the 15 chemical compounds that have been or could be used to induce CFA (Table 1). These substances were chosen because among their 120 adverse effects they include gastrointestinal symptoms related to the CFA mechanism 121 122 (i.e. nausea, vomits). From these compounds we finally selected five compounds that 123 could be good candidates to induce CFA in wild canids with a low risk of lethality as expected by the MOS > 10. In a second phase of the study, the five selected substances 124 125 were tested in a pilot experiment to induce CFA in penned dogs. Two of these 126 compounds had been successfully used before in CFA with canids (i.e. thiabendazole and levamisole). Thiabendazole was taken because of the available literature on this 127 128 substance as CFA inducer. Levamisole has also been tested as CFA inducer, but 129 important practical limitations may exist with this compound because of its potential 130 detectability by taste and odour. The other three substances (thiram, fluconazole and 131 furalaner) fulfilled most of the requirements to be used as aversive but have not been 132 previously tested.

133

134 *2.2. Animals*

135 Five males and seven females of adult Beagle dogs (Canis lupus familiaris) were used in the experiment. Dog's body weight ranged from 9.3 to 15.7 kg. The experiment 136 137 was performed following the appropriate European regulations in the Laboratory Animals Section (Research Support Service, University of Murcia). The project has been 138 evaluated by the Ethics Committee of the University of Murcia and approved 139 140 subsequently by the Government of the Region of Murcia (Spain) with the permit № A13170703. Animals were fed every morning with dry (Gosbi[®] Premium Performance) 141 and/or wet (Gosbi® Fresko Chicken) dog food and they were fasted during 24 hours 142 143 before each test. The dogs were housed individually in separate indoor pens (size: 1.6 \times 4.3 \times 3 m), following the "Guidelines for accommodation and care of animals of the 144 145 European Convention for the protection of vertebrate animals used for experimental and other scientific purposes" (European Commission 2007), conforming to Directive 146 147 2010//63/EU: room temperature: 20–24 °C; relative humidity: 45–65%; air exchange: 148 10–15 times/h; 12 h light/darkness cycle. Tap water was provided ad libitum and the 149 dogs were released for exercise and physical contact with their roommates for 30 150 minutes every day.

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152 2.3. Tested aversive compounds

The selection of substances and doses were based on literature as described above (Table 1). Thiabendazole and levamisole have been tested in the past for their CFA effects. To our knowledge, thiram, fluconazole and fluralaner have never been used as CFA agents. Thiabendazole is a benzimidazole with anthelmintic properties that has been previously used to generate CFA in several mammalian species (Gustavson et al. 1983; Conover 1989; Ternent and Garshelis 1999). Levamisole

159 chlorhydrate (levamisole hereafter) is an imidazothiazole with anthelmintic properties 160 that has been used in CFA studies with laboratory rats (Massei and Cowan 2002), foxes 161 (Vulpes vulpes and Pseudalopex griseus) (Massei et al. 2003a; Nielsen et al. 2015) and Eurasian badgers (Meles meles) (Cagnacci et al. 2005). Thiram is a dithiocarbamate 162 fungicide that has been used as a repellent both in birds and mammals (Nolte and 163 164 Barnett, 2000; Werner et al. 2010). Fluralaner, one of the new and structurally-unique 165 isoxazolines, is an ectoparasiticide selected because it is considered a safe drug causing 166 vomiting, decreased appetite and diarrhoea as the most common adverse reactions at the recommended therapeutic doses (25–56 mg/kg) in dogs (EMA, 2014). Fluconazole 167 is a triazole used as antifungal with a wide acute MOS. Nausea, vomiting and anorexia 168 169 have been described at therapeutic doses of this in dogs (Mueller, 2007).

170 The effective doses of the aversive substances were obtained from previous toxicity studies based on their ability to cause digestive symptoms (vomiting, nausea 171 172 and/or diarrhoea), but without causing severe adverse effects (Table 1). These single 173 oral doses were 200 mg/kg for thiabendazole, 50 mg/kg for levamisole, 40 mg/kg for thiram, 200 mg/kg for fluralaner and 30 mg/kg for fluconazole (see references in Table 174 175 1). Because no vomits or food avoidance were found during conditioning with 176 fluconazole and fluralaner, the dose during the reinforcement phase (see section 2.3) 177 was increased to 70 mg/kg for fluconazole (seven times the maximum therapeutic 178 dose for dogs, Kukanich, 2008) and to 300 mg/kg for fluralaner (about six times the 179 maximum therapeutic dose for dogs; Walther et al., 2014), but in both cases well 180 below LD₅₀ values (Table 1).

181 In order to reduce the levamisole bitter taste described in previous studies with 182 canids (Massei et al. 2003a; Gentle et al. 2004), it was microencapsulated with

Precirol[®] Ato 5 (glyceryl palmitostearate) as the hydrophobic binder using a meltgranulation technique (Hamdani et al. 2003; Mašić et al. 2012). The other chemicals were used in the pure composition, except for fluralaner that was used in the commercial form. Levamisole was microencapsulated by the Drug Development Service, Faculty of Pharmacy, University of Barcelona. Fluconazole and thiram were purchased from Sigma-Aldrich[®], fluralaner (Bravecto[™]) from MSD Animal Health, and thiabendazole from Alfa Aesar[®].

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191 *2.4. Experimental design of the aversive compounds assay*

In order to evaluate the treatment effect of the compounds on the dogs 192 193 feeding behaviour, we performed a Before-After Control-Impact (BACI) design 194 (Underwood 1994). BACI analysis approaches include generalized linear mixed models 195 (McDonald et al. 2000), where a significant interaction between treatments (each substance and control group) × time (pre-conditioning and post-conditioning) indicates 196 197 that the experimental treatment had an effect on dogs feeding behaviour. We followed the typical phases in CFA experiments: pre-conditioning (only food, four 198 199 trials); conditioning (food + aversive agent, single trial); post-conditioning (only food, 200 four trials); reinforcement (food + aversive agent, single trial). This initial assay was 201 performed only with a pair of dogs per each chemical tested, in order to test the potential of these substances to induce CFA on dogs. This decision was made following 202 the current ethical and animal welfare standards to reduce the number of individuals 203 204 used in the animal experimentation. Those substances that yielded best CFA results in 205 this study will be tested with a larger number of animals in further studies (see Tobajas 206 et al. in press).

207 Dogs were enrolled in the experiment on day 1 (start of pre-conditioning). During pre-conditioning period, they were fed with two types of food, dry and wet, 208 and the amount of consumed food was calculated daily. Wet food was usually 209 210 preferred over dry food and then this wet food was chosen as the target food against which we wanted to induce aversion. Although it is known that prior exposure of a 211 212 food before conditioning reduces the aversion acquisition (Kalat and Rozin, 1973), we decided to perform the pre-conditioning phase with the target food to achieve a 213 214 conservative experiment. On day 15 (conditioning trial), the dogs were randomly 215 assigned to each substance or control group. A male and a female of Beagle dogs were 216 conditioned with each substance, except for thiram for which 2 females were used (as 217 no more males were available for the test). Prescribed amounts of aversive compounds were mixed homogenously with the wet food and offered for 30 minutes 218 219 to each dog. A control pair (a male and a female) received the same amount of wet 220 food, but without any substance added, and were handled in the same conditions. The 221 dogs were checked by a veterinarian every 2 h for signs of illness such as nausea, 222 vomiting and diarrhoea for 8 hours after exposure, and 24 h later for confirm no 223 consumption changes of their normal diet.

During the post-conditioning, two-choice tests were performed on days 19, 26, 33 and 45 to compare consumption of the previously conditioned food (wet food, but without the aversive substance) versus the non-conditioned dry food. Reinforcement of aversive conditioning with wet food containing the chemical was performed on day 228 22. In each trial, dry and wet food was weighed (± 1 g) with a precision balance (Mettler[®] PJ15, Mettler Instrumente[®], Greifensee-Zurich, Switzerland) in stainlesssteel dog bowls and was offered to each dog for 30 minutes. Bowls were then removed

231 from the dog pens and weighted to calculate the amount of food eaten and the proportion of food rejected (PFR). Dog behaviour was recorded with a video camera 232 233 (Spartan, HCO Outdoor Products, Norcross, GA, USA) to observe the signs of adverse 234 effects of conditioning and to estimate the latency time (LT; time from food offer to 235 start eating) and the time spent eating all food (TSE). LT and TSE (in min) were used as 236 CFA indicators (Massei et al. 2002; Webb et al. 2008). If at the end of the 30-min presentation dogs had not started or eaten all the food, LT and TSE, respectively, were 237 238 recorded as 30 min. Feeding behaviour was also video recorded during the pre-239 conditioning phase.

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241 2.5. Haematology and serum biochemistry analysis

Haematology and serum biochemistry were studied to evaluate the possibility of detrimental effects on health. Blood samples were obtained from all the dogs, including controls, one day before and one day after the conditioning and reinforcement with the aversive substances. Blood samples (4–5 mL) were obtained by puncturing the brachial vein, using a 5 mL syringe and a 21 G needle. All the analyses were made at the Interdisciplinary Laboratory of Clinical Pathology, Interlab-UMU, Campus of Excellence Mare Nostrum, University of Murcia, Spain.

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250 2.6. Data analysis

The effect of treatment (fixed factor) on PFR, LT and TSE were analysed by a generalized linear mixed model (GLMM) to investigate differences between treatments in the pre-conditioning, and in the post-conditioning phase. The relationships between pre-conditioning and post-conditioning phases of PFR, LT and

TSE were modelled by GLMM using the interaction "treatment × phase" as a fixed 255 effect. Individual dogs were fitted as a random effect in all models. Where differences 256 between treatments or significant effects of "treatment × phase" were found, pair-257 wise comparisons for each group were performed. Paired t-tests were used to 258 compare haematology and serum biochemical parameters before and after treatment 259 260 with each aversive substance. Normality of residuals was checked, and non-normal data were logit transformed for the PFR and log transformed for LT and TSE. 261 Significance of statistical tests was considered at $p \le 0.05$. All statistical analyses were 262 carried out with the SPSS statistical package 24.0 software (IBM Inc., Chicago, USA). 263

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265 3. Results
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267 3.1. Potential aversive compounds

268 From 167 substances evaluated during the review of the available literature 269 (Table S1), 15 were considered as potential candidates to be used in CFA studies with canids. These included anthelmintics, fungicides, insecticides and mollusquicides of 12 270 271 chemical groups: salicylanilides (i.e. niclosamide), pyrazinobenzazepines (i.e. 272 epsiprantel), pyrazinoquinolines (i.e. praziquantel), tetrahydropyrimidines (i.e. pyrantel 273 and oxantel), imidazoles (i.e. clotrimazole), benzimidazoles (i.e. fenbendazole and thiabendazole), imidazothiazoles (i.e. levamisole), triazoles (i.e. fluconazole) 274 275 dithiocarbamates (i.e. thiram), isoxazolines (i.e. fluralaner and afoxolaner), spinosyns 276 (i.e. spinosad) and aldehydes (i.e. metaldehyde). Five of these compounds were 277 odourless and tasteless, for other four this information was not available and the other 278 six have some odour and taste that may affect the conditioning process if the animals

associate the adverse effect with these physical properties of the substances (Table 1). Only three of these substances had been previously used in CFA assays (i.e. thiabendazole, levamisole and clotrimazole). The oral acute LD_{50} in rat was available for all the considered compounds, ranging from 480 mg/kg of levamisole to >10,000 mg/kg of fenbendazole. The oral acute LD_{50} in dog was only available for nine of the considered compounds (Table 1). The acute MOS was <10 in four substances, between 10 and 100 in nine substances and >100 in two substances (Fig. 1).

286 Only those substances with an acute MOS > 10 were considered to be included as CFA inducers. Thus afoxolaner, clotrimazole, metaldehyde, niclosamide and 287 excluded. According to the 288 praziguantel were firstly MOS calculated, 289 pyrazinobenzazepines and tetrahydropyrimidines would be the best candidates to be 290 tested as CFA inducers, but these anthelmintics also show important differences in LD₅₀ values between rat and dog (i.e. oxantel, Table 1). Hence these substances were 291 not finally selected following a precautionary principle. For the same reason, 292 293 fenbendazole was also excluded. We finally selected 5 substances to be tested experimentally with penned dogs: thiabendazole, microencapsulated levamisole, 294 295 thiram, fluconazole and fluralaner. Thiabendazole was included in the experimental 296 tests because it is a confirmed CFA inducer in canids. We also included levamisole 297 because it is a CFA inducer in several mammal species, but here we have employed a microencapsulation to reduce its detectability by canids. Finally, three other 298 299 substances with MOS > 10 that can produce gastrointestinal symptoms related to the 300 CFA mechanism (nausea, vomits) were selected for the experimental tests with dogs, 301 two odourless and tasteless (i.e. thiram, fluconazole) and another one with unknown 302 organoleptic characteristics (i.e. fluralaner) (Fig. 1).

304 3.2. Conditioned food aversion

305 During the first two-choice test (day 19) after conditioning, one dog from the 306 levamisole group, and another one from the thiram group rejected the target food 307 (wet food). Moreover, both dogs of thiabendazole, thiram and levamisole group 308 increased LT and TSE compared to control group and also respect to pre-conditioning phase. The dogs of these treatment groups lasted more time to start tasting the food 309 310 and ate the food by sticking small nibbles, stopping to eat and recede from food several times, often stopping to observe the food, which could be interpreted as a 311 misgiving behaviour. In contrast, the other dogs from the fluconazole, fluralaner and 312 313 control group ate the food without stopping in a shorter time. During the reinforcement on day 22, all the unconditioned dogs (one from thiram and 314 315 thiabendazole group and both from fluconazole and fluralaner group) ate all the wet food except the dog exposed to levamisole, which rejected the food. After 316 317 reinforcement, previously conditioned dogs with levamisole and thiram continued showing aversion in the two-choice tests performed at days 26 and 33, and one 318 319 additional dog from the thiabendazole group showed food aversion at day 26. As 320 during the first two-choice tests, the dogs from the thiabendazole, thiram and 321 levamisole showed a misgiving behaviour and they increased the LT and TSE. No signs of food aversion in the fluralaner and fluconazole were found during the experiment. 322

Comparing PFR between pre-conditioning and post-conditioning phases (Fig. 2), we found a significant effect of the "treatment x phase" interaction ($F_{11, 84} = 2.756$, p = 0.004). Differences between pre- and post-conditioning phases for PFR were significant for the levamisole (p = 0.002). The effect of the interaction "treatment x phase" was also significant for LT ($F_{11, 84} = 106.55$, p < 0.001; Fig. 3), and TSE ($F_{11, 84} = 3.903$, p < 0.001; Fig. 4). In the case of LT, the difference between the pre and post-conditioning phase in each treatment group was significant for levamisole (p < 0.001), thiram (p = 0.001) and thiabendazole (p < 0.001; Fig. 3). In the case of TSE (Fig. 4), differences were significant for levamisole (p = 0.035), thiram (p = 0.029) and thiabendazole (p = 0.002). No significant differences were found in PFR, LT and TSE for the fluconazole and fluralaner treatment groups.

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335 *3.3. Clinical signs related with aversive ingestion*

During conditioning, although all the dogs ingested the total dose of aversive 336 337 substances by eating 100% of the target food, vomiting and/or diarrhoea were only 338 found in dogs exposed to levamisole, thiram and thiabendazole (Supplementary material, Table S2). Dogs exposed to fluralaner and fluconazole did not show any 339 340 symptoms. First vomiting after exposure to each substance occurred at different times: 341 between 30-40 min after exposure to thiram; between 1.5-2.5 h after exposure to 342 thiabendazole; and 2 h 20 min–3 h after exposure to levamisole (Table S2). Diarrhoea 343 was found in one dog 5 h 15 min after exposure to thiram and in another dog 3 h after 344 exposure to levamisole (Table S2). For the reinforcement, as mentioned above, doses 345 of fluralaner and fluconazole were increased due to the lack of conditioning related 346 symptoms. Despite this, no conditioning related symptoms were found in dogs after 347 reinforcement with these substances. Levamisole exposed dogs in the reinforcement 348 barely ate the food (about 4% food ingested) as did a thiram-exposed dog (17% food 349 ingested). The other thiram exposed dog ingested all food and vomited 20 minutes 350 later.

352 *3. 4. Haematology and serum biochemistry*

Exposure to these substances did not seem to cause physiological adverse effects in the dogs, as both haematological and serum biochemical parameters did not significantly differ between before and after treatment (Supplementary material, Table S3). Only levamisole caused a significant increase of neutrophils in the dogs after conditioning (female: 5.3 to 10.55 x10³ cells/µL; male: 4.2 to 10.02 x10³ cells/µL, p =0.03). Despite this increase, the values were within the reference values established in the laboratory for dogs and decreased back after the reinforcement.

360

361 **4. Discussion**

362

The selection of substances with the literature review and the subsequent 363 experimental tests have yielded three substances than could be used as CFA agents to 364 365 reduce the conflicts with wild predators. Thiabendazole and levamisole were already 366 known as CFA inducers in canids, and thiram is a new candidate with a very high acute 367 MOS (aprox. 100), which should be confirmed as CFA inducer in further experimental 368 studies with more animals. Moreover, thiram was not detected by dogs during the 369 conditioning process, which is an important aspect to be considered in case thiram was used with wild canids to reduce the predatory conflict. 370

371

372 4.1 Review and selection of substances

The list of substances reviewed in the present study reveals many potential candidates to be CFA inducers in canids. However, the adverse effects produced by

375 some of the substances reviewed and used in CFA may have negative consequences in the health of the animals. Some of the aversive compounds used, such as 376 377 amphetamine, amitriptyline, bupropion (Miller and Miller 1983; Bryant et al. 1993), 378 and more recently, fluoxetine hydrochloride (Massei and Cowan 2002), affect the 379 central neural system. Although some of these compounds can induce CFA in rats, all 380 of them modify their natural behaviour exciting or depressing the central neural system, which could lead vulnerability and risk situations for the conditioned 381 individuals in the wild. Also, many repellents, such as anthraquinone, d-pulegone, 382 383 cinnamic aldehyde, cinnamamide and capsaicin, have been used (Avery et al. 1998; Gill et al. 2000; Conover and Lyons 2003), but these act differently to CFA agents because 384 385 they only prevent predation in the presence of the chemical. Oestrogens, like 17αethinyloestradiol, have also been used as aversive compounds with good results in rats 386 387 (Gill et al. 2000) and carnivore species (Nicolaus et al. 1989c; Semel and Nicolaus 1992; Dueser et al. 2018). However, their ability to induce abortion or even death in 388 389 pregnant individuals (Yasuda et al. 1981, Dueser et al. 2018) makes them inappropriate candidates for CFA. Finally, other compounds such as insecticides and fungicides have 390 391 been tested for CFA, mainly causing an agonist cholinergic effect and gastrointestinal 392 irritation (Dimmick and Nicolaus 1990; Massei and Cowan 2002; Cox et al. 2004; 393 Maguire et al. 2009). These groups of substances, especially those with an agonist cholinergic effect, (e.g. levamisole, thiabendazole, trimethacarb and carbachol) have 394 shown good results in CFA studies (Gustavson et al. 1983; Nicolaus and Nellis 1987; 395 396 Dimmick and Nicolaus 1990; Massei et al. 2003a). Due to the high toxicity of trimethacarb and carbachol, these are not recommended for being used in the field as 397 398 CFA inducers (Schafer 1972; Conover 1990).

399 Taste and odour of most compounds used in CFA studies modify the original food and taste of foods (Burns 1980; Nicolaus and Nellis 1987; Gentle et al. 2004; 400 401 Nielsen et al. 2015). Only 17 α -ethinyloestradiol and thiabendazole appear to be 402 undetectable (Gustavson et al. 1983; Nicolaus et al., 1989a; Gentle et al. 2006; Dueser 403 et al. 2018). To solve detection problems, a masking odour or taste has been used, 404 although with limited success on baits (Cotterill et al. 2006; Nielsen et al. 2015). Other possibility suggested with few positive results is the microencapsulation technique 405 406 (Burns 1983; Mašić et al. 2012), but it requires an increase of cost and specialized 407 machinery. However, the new formulations and manufacturing methods could enable its use in an effective and economical way. 408

409

410 *4.2 Assays with penned dogs*

411 The assay results with dogs showed that levamisole, thiabendazole and thiram produced CFA in dogs. In contrast, fluralaner and fluconazole at 6 and 7 times the 412 413 therapeutic dose, respectively, did not produce CFA. Contrary to our expectations, these two substances apparently did not cause any adverse effect in the dogs, neither 414 415 vomits nor diarrhoea. In the case of fluconazole, the lack of adverse signs may be due 416 to the low doses used here, but it seems to be undetectable by dogs and may have a 417 potential to produce CFA at higher doses. Fluralaner, in its commercial form, has a 418 strong smell so it is susceptible to modify importantly the organoleptic characteristics 419 of the food. The dose of fluralaner was increased until 300 mg/kg, this means a large 420 amount of commercial product that modified the characteristics of the amount of the 421 target food used. Therefore, the use of fluralaner as an aversive agent can be ruled 422 out.

423 Thiabendazole showed an unexpected low aversive effect on dogs, contrary to 424 previous studies on canids (Ziegler et al. 1982; Gustavson et al. 1983; Massei et al. 425 2003a) and other species (Massei and Cowan 2002; Norbury et al. 2005; Gentle et al. 2006; O'Donnell et al. 2010). The dose used was the same or higher than in other 426 studies with canids (Ziegler et al. 1982; Massei et al. 2003a), so the reduced effect may 427 428 be due to the individual variability and differences in behaviour between domestic and wild canids. Massei et al. (2003a) found individual variability in the response of 429 430 aversion to thiabendazole by red foxes, as other authors found with other species (Conover 1989; Ternent and Garshelis 1999). They suggested that the lack of effect in 431 some individuals could be due to the detection of thiabendazole and subsequent 432 433 aversion to this agent rather than to the test food. In our case, the dogs from the 434 thiabendazole group did not detect the chemical. However, our results correspond to a 435 pilot study with a small sample, so we should take them with caution.

436 Thiram is used in agriculture as a fungicide, but it also protects seeds sown 437 seeds from birds and mammals due to its repellent action (Nolte and Barnett, 2000; Lopez-Antia et al. 2014), but it has never been used as CFA agent. One dog acquired 438 439 CFA to the target wet food, which was rejected during the post-conditioning, while the other dog ate twice the treated food and had vomits in both cases without acquiring 440 441 CFA. Therefore, thiram was apparently undetectable by dogs. This fact makes thiram a 442 potential candidate as an aversive substance in predation control, since detectability is 443 one of the main handicaps in the CFA applicability (Burns 1980; Nicolaus and Nellis 444 1987; Gentle et al. 2004). Another advantage of thiram is its low toxicity (Table 1). 445 Accordingly, no negative effects on blood parameters were observed after two 446 ingestions (Table S3).

447 Levamisole has been used as CFA agent in several studies with controversial results. It has induced long-lasting CFA in rats (Massei and Cowan 2002) and grey foxes 448 449 (Pseudalopex griseus) (Nielsen et al. 2015), but it failed in ferrets (Mustela furo) (Massei et al. 2003b; Norbury et al. 2005) and badgers (Cagnacci et al. 2005), and 450 produced contrasting results in red foxes (Massei et al. 2003a; Gentle et al. 2004). The 451 452 failures in the generation of aversion happened because animals detected the levamisole and only avoided consuming the food when the levamisole was present. 453 454 The differences between the studies may be due to the ability of certain strong 455 flavoured foods to mask the taste and smell of levamisole. On the other hand, the ratio between the amount of levamisole and the food may mask the flavour of levamisole to 456 457 a greater or lesser extent (Nielsen et al. 2015). In our study, one of the dogs possibly 458 detected the levamisole despite it was microencapsulated, because it only rejected the target wet food when levamisole was present in the reinforcement. However, the 459 460 other dog exposed to levamisole developed CFA behaviour, increasing PFR, LT and TSE. 461 The different results between both dogs may be due to individual aversion behavioural 462 differences, and a new experiment with a larger sample size is necessary to confirm its 463 potential as aversive.

LT and TSE increased significantly after conditioning with thiabendazole, thiram and levamisole, indicating that the dogs had an internal conflict between the awareness of the consequences of eating and the food palatability (Forbes 1998). If these results should be applied to CFA generation in the wild, we can expect that, unlike penned dogs, wild animals could suffer a disruptive effect at early phases of predation, and this could favour prey escape.

470 The conclusions of the present study are limited by the reduced number of 471 dogs used for each compound, and by the conservative design of the experiment. In 472 this sense, the pre-exposure (pre-conditioning phase) to the target food reduces the strength of the aversion (Revusky and Bedarf, 1967; Mikulka and Klein, 1977). The dogs 473 were used to the offered target food, which is then assumed as "learned safe" food, 474 475 thus reducing the CFA (Kalat and Rozin, 1973). In the same way, Mikulka and Klein (1977) observed that leaving the food available for long periods of time in the aversion 476 477 tests can mask a weak aversion, based on similar studies carried out by Fenwick et al. (1975) with short test intervals. Also, Carroll et al. (1975) observed that neophobia can 478 be found with short test intervals but is not apparent in long test periods. Another 479 480 consideration to keep in mind is that our experimental subjects were domestic animals 481 fed by humans during all their life, thus they associate the food coming from humans as safe. In the case of wild animals, the processes of neophobia associated with illness 482 483 after consumption of foods would surely cause an increase in aversion in comparison 484 with domestic animals (Mitchell 1976).

In summary, the results provided here and in previous studies show that thiabendazole, thiram and levamisole can cause aversion in canids and that they are good candidates for use as aversive compounds in the wild. This pilot study identified thiram as a safe, accessible, cheap and undetectable substance that can induce CFA on canids. Further research with larger number of individuals, probably with revised doses, will be performed to confirm these preliminary results.

491

492 **Conflict of interest**

493

We declare that none of the authors of this manuscript has any actual or potential conflict of interest including any financial, personal or other relationships with other people or organizations.

497

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499

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506 References

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- Avery, M.L., Pavelka, M.A., Bergman, D.L., Decker, D.G., Knittle, C.E., Linz, G.M., 1995.
 Aversive conditioning to reduce raven predation on California least tern eggs.
 Colon. Waterbirds 131–138.
- 511 Avery, M.L., Humphrey, J.S., Primus, T.M., Decker, D.G., McGrane, A.P., 1998. 512 Anthraquinone protects rice seed from birds. Crop Prot. 17(3), 225–230.
- 513 Bergstrom, B.J., 2017. Carnivore conservation: shifting the paradigm from control to
- 514 coexistence. J. Mammal. 98(1), 1–6.
- 515 Booze, T.F., Oehme, F.W., 1985. Metaldehyde toxicity: a review. Vet. Hum. Tox. 27(1),

516 11–19.

- 517 Bryant, P.A., Boakes, R.A., McGregor, I.S., 1993. Taste-potentiated odor aversion 518 learning based on amphetamine. Physiol. Behav. 54(2), 393–398.
- Budavari, S.E., 1996. The Merck Index: An Encyclopaedia of Chemicals, Drugs, and
 Biologicals, 12th ed. Merck & Co, Whitehouse Station.
- 521 Bures, J., 1998. Ethology, physiological psychology, and neurobiology of CTA, in: Bures,
- J., Bermudez–Rattoni, F., Yamamoto, T. (Eds.), Conditioned Taste Aversion.
 Oxford University Press, Oxford, UK, pp. 1–13.
- 524 Burns, R.J., 1980. Evaluation of conditioned predation aversion for controlling coyote 525 predation. J. Wildl. Manage. 44(4), 938–942.
- 526 Cagnacci, F., Massei, G., Cowan, D.P., Delahay, R.J., 2005. Can learned aversion be used
- to control bait uptake by Eurasian badgers? Appl. Anim. Behav. Sci. 92, 159–
 168.
- 529 Conover, M.R., 1989. Potential compounds for establishing conditioned food aversions
 530 in raccoons. Wildl. Soc. Bull. 17, 430–435.
- 531 Conover, M.R., 1990. Reducing mammalian predation on eggs by using a conditioned
 532 taste aversion to deceive predators. J. Wildl. Manage. 54, 360–365.
- Conover, M.R., Lyons, K.S., 2003. Reducing or delaying egg depredation by punishing
 free-ranging predators for opening eggs. Appl. Anim. Behav. Sci. 83(3), 177–
 185.
- Corwin, R.M., Green, S.P., Keefe, T.J., 1989. Dose titration and confirmation tests for
 determination of cesticidal efficacy of epsiprantel in dogs. American journal of
 veterinary research, 50(7), 1076-1077.
- 539 European Commission, 2007.Commission Recommendation of 18 June 2007 on 540 guidelines for the accommodation and care of animals used for experimental

- 541and other scientific purposes.http://eur-lex.europa.eu/legal-542content/EN/TXT/?uri=uriserv:OJ.L_.2007.197.01.0001.01.ENG (Accessed April5432015).
- Cotterill, J., Massei, G., Cowan, D.P., 2006. Masking the taste of the conditioned taste
 aversion agent Levamisole using an ion-exchange resin, for practical application
 in wildlife management. Pest Manage. Sci. 62 (2), 120–125.
- Cowan, D.P., Reynolds, J.C., Gill, E.L., 2000. Reducing predation through conditioned
 taste aversion, in: Gosling, L.M., Sutherland, W.J. (Eds.), Behaviour and
 Conservation. Cambridge University Press, New York, NY, pp. 281–299.
- Cox, R., Baker, S.E., Macdonald, D.W., Berdoy, M., 2004. Protecting egg prey from
 Carrion Crows: the potential of aversive conditioning. Appl. Anim. Behav. Sci.
 87, 325–342.
- 553 Dimmick, C.R., Nicolaus, L.K., 1990. Efficiency of conditioned aversion on reducing 554 predation by crows. J. Appl. Ecol. 27, 200–209.
- 555 Dueser, R.D., Martin, J.D., Moncrief, N.D., 2018. Pen trial of estrogen-induced
 556 conditioned food aversion to eggs in raccoons (*Procyon lotor*). Appl. Anim.
 557 Behav. Sci. 201, 93–101.
- 558 EMEA, 2015. Committee for Veterinary Medicinal Products: CVMP assessment report 559 for NexGard. European Agency for the Evaluation of Medicinal Products, 560 London (GB) (Accessed May 2015)
- 561 Fenwick, S., Mikulka, P.J., Klein, S.B., 1975. The effect of different levels of pre-562 exposure to sucrose on the acquisition and extinction of a conditioned 563 aversion. Behav. Biol. 14, 231–235.
- 564 Forbes, J.M., 1998. Dietary awareness. Appl. Anim. Behav. Sci. 57(3), 287–297.

- 565 Frohberg, H., 1984. Results of toxicological studies on praziquantel. Arzneimittel-566 Forschung, 34(9B), 1137–1144.
- 567 Frohberg, H., Schulze, M.S., 1981. Toxicological profile of praziquantel, a new drug 568 against cestode and schistosome infections, as compared to some other 569 schistosomicides. Arzneimittel-Forschung, 31(3a), 555–565.
- 570 Garcia, J., Hankins, W.G., Rusiniak, K., 1974. Behavioral regulation of the milieu interne 571 in man and rat. Science. 185, 824–831.
- Gentle, M., Massei, G., Saunders, G., 2004. Levamisole can reduce bait monopolization
 in wild red foxes *Vulpes vulpes*. Mamm. Rev. 34(4), 325–330.
- 574 Gentle, M., Massei, G., Quy, R., 2006. Diversity of diet influences the persistence of 575 conditioned taste aversion in rats. Appl. Anim. Behav. Sci. 97(2), 303–311.
- 576 Gill, E.L., Whiterow, A., Cowan, D.P., 2000. A comparative assessment of potential
- 577 conditioned taste aversion agents for vertebrate management. Appl. Anim.
 578 Behav. Sci. 67(3), 229–240.
- 579 Gordon, C.E., Eldridge, D.J., Ripple, W.J., Crowther, M.S., Moore, B.D., Letnic, M., 2017.
- 580 Shrub encroachment is linked to extirpation of an apex predator. J. Anim. Ecol.
 581 86(1), 147–157.
- 582 Gupta, R.C., 2012. Chapter 53 Metaldehyde, in: Gupta, R.C, (Ed.), Veterinary 583 Toxicology: Basic and Clinical Principles. 2nd Ed. Academic Press, London. Pp. 584 624–628.
- 585 Gustavson, C.R., Garcia, J., Hankins, W.G., Rusiniak, K.W., 1974. Coyote predation 586 control by aversive conditioning. Science 184, 581–583.

- 587 Gustavson, C.R., Gustavson, J.C., Holzer, G.A., 1983. Thiabendazole-based taste 588 aversions in dingoes (*Canis familiaris dingo*) and New Guinea wild dogs (*Canis* 589 *familiaris hallstromi*). Appl. Anim. Ethol. 10, 385–388.
- Hamdani, J., Moës, A. J., & Amighi, K., 2003. Physical and thermal characterisation of
 Precirol[®] and Compritol[®] as lipophilic glycerides used for the preparation of
- 592 controlled-release matrix pellets. Int. J. Pharm. 260(1), 47-57.
- Hayes, W.J., Jr., E.R. Laws, Jr., 1991. Handbook of Pesticide Toxicology. Vol. 3. Classes
 of Pesticides. Academic Press Inc., New York.
- Kalat, J.W., Rozin, P., 1973. Learned safety as a mechanism in long-delay taste-aversion
 learning in rats. J. Comp. Physiol. Psychol. 83, 198–207.
- 597 Kukanich, B., 2008. A review of selected systemic antifungal drugs for use in dogs and 598 cats. Vet. Med. 103, 41–50.
- Lanusse, C.E., Alvarez, L.I., Sallovitz, J.M., Mottier, M.L., Sanchez-Bruni, S.F., 2009.
 Antinematodal drugs, in: Riviere, J.E., Papich, M.G. (Eds.), Veterinary

601 Pharmacology and Therapeutics. Wiley-Blackwell, pp. 1053–1094.

- 602 Lee, C.C., Russell, J.Q., Minor, J.L., 1978. Oral toxicity of ferric 603 dimethyl-dithiocarbamate (ferbam) and tetramethylthiuram disulfide (thiram) in rodents. J. Toxicol. Environ. Health. 4(1), 93–106. 604
- 605 Lopez-Antia, A., Ortiz-Santaliestra, M.E., Mateo, R., 2014. Experimental approaches to
- test pesticide-treated seed avoidance by birds under a simulated diversification
 of food sources. Sci. Total Environ. 496, 179–187.
- Lynn, R.C., 2009. Antiparasitic drugs, in: Bowman DD (Ed), Georgi's parasitology for
 veterinarians. Elsevier, pp. 254–294.

- 610 Macdonald, D.W., Sillero-Zubiri, C., 2004. The biology and conservation of wild canids.
- 611 Oxford University Press, Oxford, UK.
- Mackenzie, C.D., 2016. The safety of pyrantel, oxantel and morantel, in: Marchiondo,
- A.A., (Ed.), Pyrantel Parasiticide Therapy in Humans and Domestic Animals.
 Cambridge Press, Cambridge, MA, USA.
- Maita, K., Tsuda, S., Shirasu, Y., 1991. Chronic toxicity studies with thiram in Wistar rats
 and beagle dogs. Toxicol. Sci. 16(4), 667–686.
- Maguire, G.S., Stojanovic, D., Weston, M.A., 2009. Conditioned taste aversion reduces
 fox depredation on model eggs on beaches. Wildl. Res. 36 (8), 702–708.
- Margalida, A., Mateo, R., 2019. Illegal killing of birds in Europe continues. Science 363
 (6432), 1161.
- Mašić, I., Ilić, I., Ibrić, S., Parojčić, J., & Đurić, Z., 2012. An investigation into the effect of formulation variables and process parameters on characteristics of granules obtained by in situ fluidized hot melt granulation. Int. J. Pharm. 423(2), 202-
- Massei, G., Cowan, D.P., 2002. Strength and persistence of conditioned taste aversion
 agents in rats: evaluation of eleven potential compounds. Appl. Anim. Behav.
 Sci. 75, 249–260.
- Massei, G., Lyon, A.J., Cowan, D.P., 2002. Conditioned taste aversion can reduce egg
 predation by rats. J. Wildl. Manag. 66(4), 1134–1140.
- Massei, G., Lyon, A.J., Cowan, D.P., 2003a. Levamisol can induce conditioned taste
 aversion in foxes. Wildl. Res. 30, 633–637.
- Massei, G., Lyon, A.J., Cowan, D.P., 2003b. Potential compounds for inducing
 conditioned taste aversion in ferrets. New Zeal. J. Zool. 30, 95–100.

634	Mikulka, P.J., Klein, S.B., 1977. The effect of CS familiarization and extinction procedure
635	on the resistance to extinction of a taste aversion. Behav. Biol. 19, 518–522.
636	Miller, D.B., Miller, L.L., 1983. Bupropion, d-amphetamine, and amitriptyline-induced
637	conditioned taste aversion in rats: dose effects. Pharmacol. Biochem. Behav.
638	18(5), 737–740.
639	Mitchell, D., 1976. Experiments on neophobia in wild and laboratory rats: A
640	reevaluation. J. Comp. Physiol. Psychol. 90(2), 190–197.
641	Mueller, R.S., 2007. Treatment of Fungal Infections, in: Mueller, R.S., (Ed.),
642	Dermatology for the Small Animal Practitioner. Teton New Media, Jackson, WY,
643	USA.
644	Nicolaus, L.K., Nellis, D.W., 1987. The first evaluation of the use of conditioned taste
645	aversion to control predation by mongooses upon eggs. Appl. Anim. Behav. Sci.
646	17, 329–346.
647	Nicolaus, L.K., Farmer, P.V., Gustavson, C.R., Gustavson, J.C., 1989a. The potential of
648	estrogen-based conditioned aversion in controlling depredation: A step closer
649	toward the "magic bullet". Appl. Anim. Behav. Sci. 23(1), 1–14.
650	Nicolaus, L.K., Herrera, J., Nicolaus, J.C., Dimmick, C.R., 1989b. Carbachol as a
651	conditioned taste aversion agent to control avian depredation. Agric. Ecosyst.
652	Environ. 26(1), 13–21.
653	Nicolaus, L.K., Herrera, J., Nicolaus, J.C., Gustavson, C.R., 1989c. Ethinyl estradiol and

- 654 generalized aversions to eggs among free-ranging predators. Appl. Anim. 655 Behav. Sci. 24(4), 313–324.
- NIIRDN., 1990. Drugs in Japan (Ethical Drugs). Japan Pharmaceutical Information
 Center (Ed.), Yakugyo Jiho Co., Ltd., Tokyo, Japan. pp 983

- Nielsen, S., Travaini, A., Vassallo, A.I., Procopio, D., Zapata, S.C., 2015. Conditioned
 taste aversion in the grey fox (*Pseudalopex griseus*), in Southern Argentine
 Patagonia. Appl. Anim. Behav. Sci. 163, 167–174.
- Nolte, D.L., Barnett, J.P., 2000. A repellent to reduce mouse damage to longleaf pine
 seed. Int. Biodeterior. Biodegrad. 45,169–74.
- Norbury, G., O'Connor, C., Byrom, A., 2005. Conditioned food aversion to eggs in
 captive-reared ferrets, *Mustela furo*: a test of seven potential compounds.
 Appl. Anim. Behav. Sci. 93(1), 111–121.
- O'Donnell, S., Webb, J.K., Shine, R., 2010. Conditioned taste aversion enhances the
 survival of an endangered predator imperiled by a toxic invader. J. Appl. Ecol.
 47, 558–565.
- O'Neil, M.J, Smith, A., Heckelman, P.E., Obenchain, J.R., Gallipeau, J.A.R., D'Arecca,
 M.A., 2001. The Merck Index. Merck & Co. Inc., Whitehouse Station, NJ (USA).
- 671 Pitts, N.E., Migliardi, J.R., 1974. Antiminth (pyrantel pamoate): The clinical evaluation

of a new broad-spectrum anthelminthic. Clinical pediatrics, 13(1), 87–94.

- Revusky, S.H., Bedarf, E.W., 1967. Association of illness with prior ingestion of novel
 foods. Science 155, 219–220.
- 675 Reynolds, J.C., Tapper, S.C., 1996. Control of mammalian predators in game 676 management and conservation. Mammal rev. 26, 127–156.
- 677 Robinson, M., Hook, F., Everson, K.E., 1976. Efficacy of oxantel pamoate and pyrantel
- 678 pamoate in combination against Trichuris vulpis, Ancylostoma caninum and
 679 Toxocara canis in dogs. Aust. Vet. Pract. 6, 173–176

- Robinson, H.J., Stoerk, H.C., Graessle, O.E., 1965. Studies on the toxicologic and
 pharmacologic properties of thiabendazole. Toxicol. Appl. Pharmacol. 7(1), 53–
 63.
- Robinson, H.J., Phares, H.F., Graessle, O.E., 1978. The toxicological and antifungal
 properties of thiabendazole. Ecotoxicol. Environ. Saf. 1(4), 471–476.
- Sayre, R.W., Clark, L., 2001. Effect of primary and secondary repellents on European
 starlings: an initial assessment. J. Wildl. Manage. 65, 461–469.
- Schafer, E.W., 1972. The acute oral toxicity of 369 pesticidal, pharmaceutical and other
 chemicals to wild birds. Toxicol. Appl. Pharmacol. 21(3), 315–330.
- 689 Scholz, H., Schultes, E., 1973a. Report on an acute oral safety evaluation of the 690 anthelmintic HOE 881 in mice. Frankfurt am Main (Germany): Hoechst-Roussel.
- 691 Scholz, H., Schultes, E., 1973b. Report on an acute oral safety evaluation of the 692 anthelmintic HOE 881 in rats. Frankfurt am Main (Germany): Hoechst-Roussel.
- Semel, B., Nicolaus, L.K., 1992. Estrogen-based aversion to eggs among free-ranging
 raccoons. Ecol. Appl. 2(4), 439–449.
- Shukla, D., Chakraborty, S., Singh, S., & Mishra, B., 2011. Lipid-based oral
 multiparticulate formulations–advantages, technological advances and
 industrial applications. Expert Opin. Drug Deliv. 8(2), 207–224.
- Symoens, J., De Cree, J., Van Bever, W., Janssen, P., 1979. Levamisole, in: Goldberg,
 N.E., (Ed.), Pharmacological and Biochemical Properties of Drug Substances.
 Vol. 2, American Pharmaceutical Association Academy of Pharmaceutical
 Sciences, Washington DC, pp. 407–464.

- Ternent, M.A., Garshelis, D.L., 1999. Taste-aversion conditioning to reduce nuisance
 activity by black bears in a Minnesota military reservation. Wildl. Soc. Bull. 27,
 704 720–728.
- Tettenborn, D., 1972. Acute toxicity and local tolerance of clotrimazole. Summary of
 test results. Arzneimittel-Forschung, 22(8), 1272.
- 707 Underwood, A. J. (1994). On beyond BACI: sampling designs that might reliably detect
 708 environmental disturbances. Ecol. Appl. 4(1), 3–15.
- US EPA; Pesticide Fact Sheet. Spinosad. New Chemical/First Food Use (Cotton). 1997.
- 710 Washington, DC: USEPA, Off. Prev. Pest. Tox. Sub. (7501C).
 711 http://www.epa.gov/opprd001/factsheets/spinosad.pdf
- 712 Wallach, A.D., Dekker, A.H., Lurgi, M., Montoya, J.M., Fordham, D.A., Ritchie, E.G.,
- 2017. Trophic cascades in 3D: network analysis reveals how apex predators
 structure ecosystems. Methods Ecol. Evol. 8(1), 135–142.
- 715 Walther, F.M., Allan, M.J., Roepke, R.K., Nuernberger, M.C., 2014. Safety of fluralaner
- chewable tablets (Bravecto TM), a novel systemic antiparasitic drug, in dogs
 after oral administration. Parasit. Vectors 7, 87.
- Webb, J.K., Brown, G.P., Child, T., Greenlees, M.J., Phillips, B.L., Shine, R., 2008. A
 native dasyurid predator (common planigale, *Planigale maculata*) rapidly learns
 to avoid a toxic invader. Austral Ecol. 33(7), 821–829.
- Werner, S.J., Linz, G.M., Tupper, S.K., Carlson, J.C., 2010. Laboratory efficacy of
 chemical repellents for reducing blackbird damage in rice and sunflower crops.
- 723 J. Wildl. Manag. 74(6), 1400–1404.
- Yasuda, Y., Kihara, T., Nishimura, H., 1981. Effect of ethinyl estradiol on development
 of mouse fetuses. Teratology 23(2), 233–239.

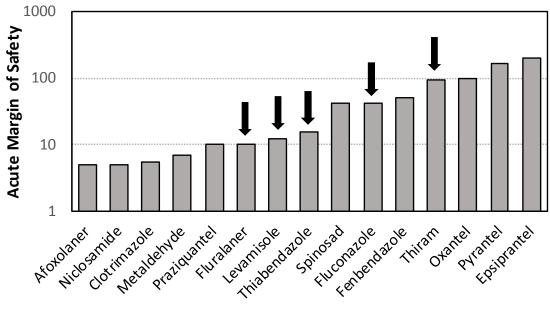
- 726 Ziegler, J.M., Gustavson, C.R., Holzer, G.A., Gruber, D., 1982. Anthelmintic-based taste
- aversion in wolves. Appl. Anim. Ethol. 9, 373–377.

Table 1. List of chemicals selected as potential CTA-inducing compounds, doses used and effective dose on dogs and LD₅₀ for rats. Oral doses in mg/kg of body weight.

Substance	Main use	Odour	Taste	CTA studies	Acute oral LD ₅₀		Potential	Ref.
					Rat	Dog	CTA dose in canids	
Niclosamide	Anthelmintic	Unknown	Unknown	No	>1000-2500	>6000	500	а
Epsiprantel	Anthelmintic	No	No	No	>5000	>200	25	b, c
Pyrantel (pamoate)	Anthelmintic	No	No	No	>5000	>690	30	d, e
Oxantel	Anthelmintic	Unknown	Unknown	No	980	170	10	f, g
Praziquantel	Anthelmintic	Weak	Yes (bitter)	No	2000-3000	>200	200	h, i
Fenbendazole	Anthelmintic	Weak	No	No	>10000	500	200	j, k
Levamisole	Anthelmintic	Yes	Yes (bitter)	Yes (canids, mustelids, rodents)	480	Unknown	50 (40-80)	l, d
Thiabendazole	Anthelmintic/Antifungal	No	No	Yes (canids, bears, rodents)	3100	Unknown	200	m, n
Fluconazole	Antifungal	No	No	No	1271	300	30-70	0
Clotrimazole	Fungicide	No	Weak (metal)	Yes (rodents)	708	>2000	140	р
Thiram	Fungicide	No	No	No	3700-4000	Unknown	40	q, r
Afoxolaner	Insecticide	Unknown	Unknown	No	>1000	Unknown	120-200	s
Fluralaner	Insecticide	Unknown	Unknown	No	>2000	Unknown	200-300	t
Spinosad	Insecticide	Yes (rancid)	Yes (bitter)	No	>3738	Unknown	90	u
Metaldehyde	Mollusquicide	Yes (mint)	No	No	690-927	500	100	v, w

^a Hayes et al., 1991; ^b Corwin et al., 1989; ^c Lynn, 2009; ^d Lanusse et al., 2009; ^e Pitts and Migliardi, 1974; [†] Mackenzie, 2016; ^g Robinson et al., 1976; ^h Frohberg and Schulze, 1981; ⁱ Frohberg 1984; ^j Scholz and Schultes, 1973a; ^k Scholz and Schultes 1973b; ^l Symoens et al., 1979; ^m Robinson et al., 1965; ⁿRobinson et al., 1978; ^o NIIRDN 1990; ^p Tettenborn, 1972; ^q Lee et al. 1978; ^r Maita et al., 1991; ^s EMEA, 2015; ^t Walther et al., 2014; ^u US EPA, 1997; ^v Booze and Oehme, 1985; ^w Gupta, 2012.

Figure 1. Acute margin of safety (MOS) in a logarithm scale of the substances that can be used to induce conditioned taste aversion in canids. MOS was calculated as the ratio between LD50 in rat and the potential dose to induce CTA in animals (canids if this information was available). Substances with a MOS above 10 were considered good candidates as CTA inducers if other requirements were also fulfilled. The substances finally used in the experimental tests with penned dogs are marked with arrows.



Substances

Figure 2. Proportion of wet food rejected (untransformed data) by treatment and control dogs across four 30-min two-choice pre-conditioning and four 30-min two-choice post-conditioning taste aversion tests. Error bars represent standard errors of the mean. ** indicate differences with p < 0.01.

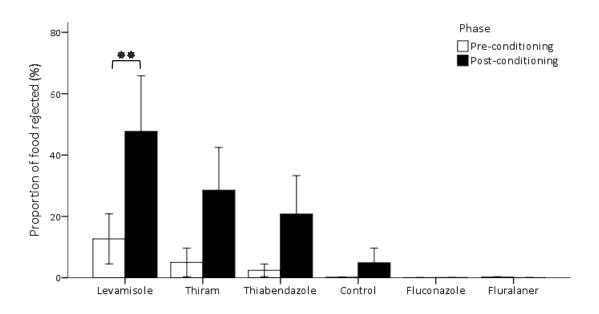


Figure 3. Latency time to start eating the wet food (seconds) by treatment and control dogs across four 30-min two-choice pre-conditioning and four 30-min two-choice post-conditioning taste aversion tests. Error bars represent standard errors of the mean. ** indicate differences with p < 0.01.

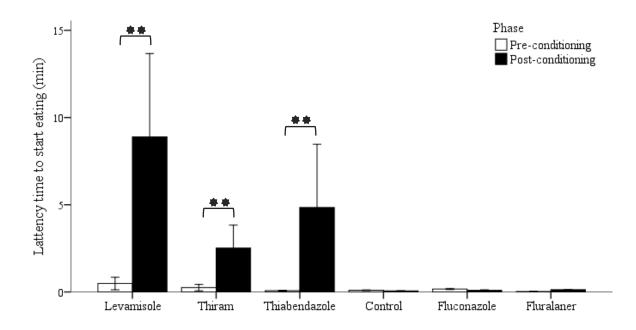


Figure 4. Time spent eating all wet food (minutes) by treatment and control dogs across four 30-min two-choice pre-conditioning and four 30-min two choice post-conditioning taste aversion tests. Error bars represent standard errors of the mean. * and ** indicate differences with p < 0.05 and p < 0.01, respectively.

