The effect of metformin vs placebo on sex hormones in CCTG MA.32 $\,$

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Abstract

Background: Metformin has been associated with lower breast cancer risk and improved outcomes in observational studies. Multiple biologic mechanisms have been proposed, including a recent report of altered sex hormones (SHs). We evaluated the effect of metformin on SHs in MA.32, a phase III trial of nondiabetic BC subjects randomized to metformin or placebo.

Methods: We studied the subgroup of post-menopausal hormone receptor negative BC subjects not receiving endocrine treatment who provided fasting blood at baseline and at 6 months after randomization. Sex hormone binding globulin (SHBG), bioavailable testosterone (BT) and estradiol levels were assayed using ECLIA (electrochemiluminescense immunoassay). Change from baseline to 6 months between study arms was compared using Wilcoxon sum rank tests and regression models.

Results: 312 women were eligible (141 metformin vs 171 placebo); the majority of subjects in each arm had T1/2, N0, HER2 negative BC and had received (neo)adjuvant chemotherapy. Mean age ± SD was 58.1±6.9 vs 57.5±7.9 years, mean BMI was 27.3±5.2 vs 28.9±6.4 kg/m² for metformin vs placebo respectively. Median estradiol decreased between baseline and 6 months on metformin vs placebo (-5.7 vs 0 pmol/L; p<0.001) in univariable analysis and after controlling for baseline BMI and BMI change (p<0.001). There was no change in SHBG or BT.

Conclusion: Metformin lowered estradiol levels, independent of BMI. This observation suggests a new metformin effect that has potential relevance to estrogen sensitive cancers.

Metformin has garnered attention as a potential anti-cancer agent across a range of cancers, including breast cancer (BC); potential effects on BC outcomes are being studied in CCTG MA.32, an ongoing Phase 3 adjuvant trial comparing metformin 850 mg bid vs placebo bid (each given for 5 years) in subjects receiving standard breast cancer therapy.¹ It has been postulated that metformin may impact BC directly (for example, via intra-tumoral LKB1 mediated AMPK activation leading to suppression of mTORC1 signaling) and/or indirectly (for example, via inhibition of hepatic gluconeogenesis with subsequent reduction in circulating insulin levels, reducing PI3K/Akt/mTOR signaling in cancer cells expressing the insulin receptor).² Data from neoadjuvant clinical trials have provided some support for both direct and indirect mechanisms.^{3,4} Recent research has suggested metformin may also act indirectly via an effect on sex hormones (SHs), although findings have been inconsistent.⁵⁻⁸

SHs are of relevance to both BC risk and prognosis, particularly in post-menopausal women. $^{9-12}$ In a case control study nested in the Women's Healthy Eating and Living Study (WHEL), BC patients who recurred had higher levels of estrogens than those who did not recur (22.7 vs 10.8 pg/mL; p = 0.05). The importance of SHs in hormone receptor positive BC is highlighted by the therapeutic effectiveness of aromatase inhibitors, which reduce estrogen production in postmenopausal women.

Campagnoli et al.^{5,6} studied the effect of metformin (1500 mg/day vs 1000 mg/day after a one-month run-in of 1000 mg/day) on SHs in 96 non diabetic post-menopausal BC patients (50% of whom were on tamoxifen) with pre-baseline testosterone above 0.28 ng/ml. Metformin reduced estradiol (-38%; p< 0.02) and free testosterone (-29%; p< 0.01); these differences remained statistically significant after controlling for baseline BMI and weight change.

Patterson et al.⁷ used a 2 X 2 factorial design to randomize 313 overweight or obese postmenopausal BC patients to metformin 1500mg/daily vs placebo and a lifestyle weight loss program vs control. Metformin (vs placebo) lowered estradiol (-10%, 95% CI, -18.5 to -1.5%) and testosterone (-9.5%, 95% CI, -15.2% to -3.8%) and increased sex hormone binding globulin (SHBG) (+7.5%; 95% CI, 2.4-12.6%) levels. However, estradiol appeared to be reduced only in those receiving both metformin and the lifestyle intervention.⁸ In a final study conducted in 382 overweight, glucose-intolerant patients without BC enrolled onto the Diabetes Prevention Program (DPP), ¹³ metformin had no impact on SHBG, estradiol, testosterone or dehydroepiandrosterone.

We investigated the SHs in postmenopausal women with hormone receptor negative BC (selected to avoid use of endocrine therapies that may impact SHs) enrolled onto the MA.32 trial. We also explored effects of the minor allele (C) of the rs11212617 single nucleotide polymorphism (SNP) that has been associated with greater metabolic response to metformin in type 2 diabetes, ¹⁴ and increased pathologic response to neoadjuvant metformin in early stage Her2 positive BC. ¹⁵

Methods

Design

We conducted a sub-study of patients enrolled onto the MA.32 randomized phase III clinical trial. The focus on SHs was not part of the original trial protocol; as evidence of an effect of metformin on SHs emerged, this work was approved by the trial steering committee as part of a priori plan to investigate potential mechanisms of metformin action. The primary objective of this sub-study was to compare change in levels of prespecified SHs (Estradiol, SHBG and Bioavailable Testosterone (BT) from baseline to 6

months between metformin and placebo arms; if a difference between study arms was found, secondary objectives were to explore two possible pathways of metformin action namely BMI and insulin change as well as the impact of the SNP rs11212617 on any SH changes.

Study Population

The Canadian Cancer Trials Group (CCTG) MA.32 Clinical Trial (Clinical Trials.gov identifier: NCT01101438 and EudraCT number: 2011-005230-18)1 is a phase III, double blind trial that randomly assigned 3649 non-diabetic patients with T1c-3 (any ER, PgR, HER2), N0-3, M0 BC who received standard treatment to receive metformin 850 mg po bid or placebo po bid for 5 years (including a 4-week ramp-up of one tablet per day) between 2010 and 2013. In May 2012, after 2382 women were enrolled, eligibility criteria were amended: those with T1cN0 disease had to have triple negative BC (ER negative, PgR negative, HER2 negative) to enter the trial and those with T2 NO BC were eligible only if they had at least one of the following risk factors: histologic grade III, presence of lymphovascular invasion, negative estrogen (ER) and progesterone (PgR) receptors, HER2 positivity, Oncotype Recurrence Score ≥ 25 or Ki-67 over 14%. Exclusion criteria included fasting glucose > 7.0 mmol/L (126 mg/dL), known diabetes or current use of diabetes medication, hypersensitivity to or intolerance of metformin, history of lactic acidosis, participation in trials of weight loss or exercise interventions, recurrence of BC or prior BC, excessive alcohol intake, or marked hepatic, kidney, or cardiac dysfunction.

All patients provided written informed consent to participate in the MA.32 clinical trial in keeping with approval by relevant institutional human subjects committees.

The SH sub-study was conducted in post-menopausal women (to avoid cyclical changes in endogenous hormone production) with hormone receptor negative BC, who were not receiving endocrine treatment (to avoid effects of hormonal agents used to treat BC). Post-menopausal was defined as prior bilateral oophorectomy or > 12 months since last menses without prior hysterectomy. Baseline blood was obtained before study treatment was initiated; patients were required to be on study treatment at the time of the sixmonth blood draw.

At baseline, information was collected on age and tumor characteristics (stage, histological type, immunohistochemical profile) and treatment. Height and weight were measured at baseline, and weight at six months. Body mass index (BMI) was calculated as weight/height² (kg/m²).

Blood Assays

The serum samples were collected into heparin tubes, aliquoted and stored at -80°C after an overnight fast of at least 12 hours. Paired specimens from each patient were retrieved, thawed and analyzed in the same batch by technicians blinded to patient treatment. The biologically most active estrogen (17ß Estradiol), SHGB and total testosterone levels were determined using competitive **ECLIA** (electrochemiluminescense immunoassay) on cobas e602 and insulin using Roche ECLIA, catalogue #12017547122, at a CAP/CLIA accredited clinical laboratory (Mount Sinai Services). Intra-assays coefficients of variability were 7%, 2.4%, 4.6.% and 3%, respectively. Albumin was assayed to allow calculation of BT from total testosterone, SHBG and albumin. The automated platform cobas e602 provides a lower detection limit for estradiol of 18.4 pmol/L and for total testosterone of 0.025 ng/mL. The measurement range for SHBG was 0.350 nmol/L to 200 nmol/L (defined as the limit of detection and the maximum of the master curve).

SNP Analysis

Genomic DNA was extracted from whole blood and samples were genotyped for the rs11212617 SNP (Chr11(GRCh38):g.108412434C>A) at The Centre for Applied Genomics, The Hospital for Sick Children, Toronto, Canada using a QIAsymphony magnetic bead DNA extractor (Qiagen, Germany) and a TaqMan PCR assay with duallabel MGB probes (Applied BiosystemsTM, ThermoFisher Scientific, Waltham, MA, USA).

Statistical Analysis

Patient and tumor characteristics at diagnosis were summarized. Since the distributions of the SHs were skewed, medians were used as measures of central tendency and 25th and 75th percentiles as measures of dispersion. Descriptive summaries were tabulated for baseline and change in the SH levels, where change was calculated as the six-month value minus the baseline value for each patient. Estradiol, testosterone and SHBG were all considered equally important main outcomes.

The pre-specified method of comparing the degree of change between arms was the Wilcoxon rank-sum test. Ten baseline and seven month 6 SHBG results (3%) were recorded as >200 nmol/L; these were replaced by 200 nmol/L. Overall, 25% of baseline and 33% of month 6 estradiol assay results were below the assay's limit of detection (LOD); change calculations, summary statistics and the Wilcoxon test were performed after replacing these results by half the LOD. Due to criticism of this method, ¹⁶ we

performed an additional sensitivity analysis using survival methods for left-censored data. The data can be analyzed using right-censored survival methods (product limit estimator and the log-rank test) by subtracting the left-censored observations from a large constant, a process called "flipping". With only one point of censoring (the LOD), the product-limit method simplifies to the construction of exceedance curves, where for a defined set of levels e* one calculates the proportion of patients in each arm whose estradiol at month 6 was higher than each of the e*.

For SHs with a statistically significant difference between arms, we explored two possible pathways of metformin action by fitting multivariable linear regression models controlling for baseline BMI and BMI change, and insulin change. This was done by log-transforming all continuous variables to reduce skewness and then fitting a model with SH change as dependent variable and with treatment status and baseline assay level as explanatory variables. Baseline and change in BMI, or insulin change, was then added to this model. We back-transformed the coefficient for treatment status to get the percent difference between the metformin and placebo arms. Finally, using a regression model for change that included an interaction term for treatment by SNP, coded as any C vs AA, we examined whether a metformin effect was restricted to patients with the C allele of the rs11212617 SNP. Reported p-values are nominal and not adjusted for multiple comparisons.

A priori power calculations indicated we had 80% power with 150 patients per arm to detect differences in change between the arms of 12%, 14% and 20% for SHBG, testosterone and estradiol respectively, using a two-sided alpha of 0.05 without adjustment for multiple comparisons.

Results

Patient Population

Three hundred and twelve women were eligible based on post-menopausal status, ER/PgR negative invasive BC not receiving hormonal treatment and availability of blood at baseline and 6 months (the latter on study treatment), 141 on metformin and 171 on placebo arm (Fig. 1 - Consort Diagram). Patient and tumor characteristics are shown in Table 1. At baseline mean age (± standard deviation) was 58.1±6.9 vs 57.5±7.9 years, mean weight 72.6±14.1 vs 76.6±17.5 kg and mean BMI 27.3±5.2 vs 28.9±6.4 kg/m² in metformin vs placebo arms, respectively. Combining clinical stage (in patients receiving neoadjuvant therapy) and pathologic stage, the majority of patients had T1 or T2 (39.0% T1 and 54.6% T2 in the metformin arm, 42.7% T1 and 46.8% T2 in the placebo arm), and node negative breast cancer (66.7% and 61.4% in the metformin and placebo arms, respectively). HER2 amplification was observed in 22.7% and 15.8% of the women in the metformin and placebo arms respectively. Ninety-eight percent of women had received neoadjuvant or adjuvant chemotherapy.

Baseline SHs measurements

Estradiol levels were below the assay's lower detection limit of 18.4 pmol/L in 24.1% of metformin and 26.2% of placebo patients, Chi-square test p=0.68. As noted above, in the primary analysis, the estradiol levels of these cases were set to half the lower detection limit, 9.2 pmol/L.

Baseline SH measurements are shown in Table 2. At baseline the median estradiol was 32.2 vs 33.3 pmol/L, SHBG 76.4 vs 72.8 nmol/L and BT 0.02 vs 0.03 nmol/L for metformin vs placebo, respectively.

Change in SHs at six months

In the main analysis, the median change between baseline and month 6 in estradiol was 5.7 pmol/L in the metformin arm vs 0 in the placebo arm (Wilcoxon test p<0.001, Table 2). The supplementary analysis supported this result - Figure 2 shows that the estradiol exceedance probabilities were consistently lower in the metformin than the placebo group (log-rank p<0.001), for example 56.9% vs 71.0% had estradiol levels over 20 pmol/L, 19.7% vs 42.6% had estradiol levels over 40 pmol/L and 8.0% vs 23.1% had estradiol levels over 60 pmol/L, respectively. The multivariable regression model estimated that month 6 estradiol was approximately 30.1% (95% CI, 19.0% to 39.7%) lower in the metformin vs placebo groups. When adjustment for baseline BMI and BMI change was added to the basic model, the reduction in estradiol in metformin vs placebo subjects was 25.7% (95% CI, 13.2% to 36.4%) and when adjustment for change in insulin was added to the basic model this was 30.1% (95% CI, 18.7% to 39.8%). In addition, examination of the interaction term in the regression model did not find evidence that the reduction in estradiol associated with the metformin arm was affected by the C allele of the SNP rs11212617 (Figure 3).

In contrast to estradiol, median changes in SHBG and BT were similar in the metformin vs placebo arms (SHBG -5.9 vs. -5.9 nmol/L, p=0.43; BT 0 vs 0 nmol/L, p=0.24), as observed in Table 2.

Discussion

Our observation of a statistically significant decrease in estradiol between baseline and month 6 in the metformin arm as compared to placebo is consistent with the work published by Campagnoli et al.,^{5,6} who studied a selected postmenopausal non-diabetic BC population (50% of whom were receiving tamoxifen) who were required to have

high baseline testosterone levels. In that study, a decrease in estradiol (-38%; p< 0.02) and in testosterone (-29%, p< 0.02) was seen in those receiving metformin 1500 mg/day (close to the 1700 mg/day administered in MA.32) vs metformin 1000 mg/day. The 30% reduction in estradiol we identified was similar to that seen by Campagnoli et al. Our population differs from that studies by Campagnoli et al. in that our subjects were not selected for high baseline levels of testosterone - this difference in entry criteria may account for our failure to identify changes in SHBG or BT with metformin. These observations suggest that the estradiol change we observed is independent of testosterone or of a potential effect of metformin on liver synthesis of SHBG.

Patterson et al.,⁷ reported reductions in estradiol, testosterone and SHBG in overweight or obese BC patients receiving metformin, however, it was not clear whether the small reductions in estradiol that were observed (-10%, 95% CI -18.5 to -1.5%) in those receiving metformin were due to co-administration of the lifestyle-based weight loss intervention. Small changes were also seen in testosterone and SHBG. As noted above, metformin had no impact on SHBG, estradiol, testosterone and dehydroepiandrosterone in 382 overweight, glucose-intolerant patients enrolled onto the Diabetes Prevention Program.¹³

Our study is the first to report the independent effect of metformin on estradiol in a placebo-controlled trial without co-intervention (tamoxifen or lifestyle intervention). The reduction we observed (approximately 30%) is substantial and of potential clinical relevance in breast cancer, and possibly in women with hormone receptor positive BC, although we did not study whether similar effects would have occurred in women receiving hormonal therapies for their BC. Our findings are also potentially relevant to other estrogen-sensitive cancers, notably endometrial cancer for which observational data suggest strong associations of metformin with both risk and

prognosis. Should beneficial effects of metformin be seen in our primary efficacy analysis in hormone receptor positive BC, we plan to investigate effects of metformin on SHs, including estradiol, in this population. The independence of the observed reduction in estradiol from baseline BMI, BMI change and insulin change suggests it did not occur as a result of loss of fat mass, nor is it associated with an insulin effect.

The mechanism by which metformin lowered estradiol remains unclear. Preclinical data have suggested that metformin may inhibit aromatase activity, potentially accounting for our observed reduction and also suggesting an additional mechanism of anti-cancer action of metformin. Both ER positive breast cancer cells and breast adipose stromal cells exhibited reductions in aromatase mRNA levels in response to metformin treatment via mechanisms involving the suppression of promoter (PII) and P1.3 specific transcripts as well as activation of AMPK. 18,19

Strengths of our study include its conduct in the setting of a placebo controlled randomized trial in carefully selected postmenopausal women (thereby excluding menstrual cycle variability in sex hormones) who were not receiving hormonal therapy (thereby excluding potential confounding by these treatments). Limitations include the use of a non-highly sensitive estradiol assay – just under 30% of estradiol assays yielded results below the lower detection limit. In our primary analyses we assigned an estradiol level of half the lower detection limit. We performed an additional sensitivity analysis to generate exceedance curves that provided results similar to those obtained in our primary analysis. The similarity of our findings using two different methods of analysis, one designed specifically for censored values, reduces the likelihood that use of a more sensitive assay we would have led to different findings. Furthermore, the selection criteria used in this sub-study, particularly the requirement that subjects be on study medication for the 6 month blood draw, may have led to some imbalances

between study arms. Additionally, the p-values reported are not adjusted for multiple comparisons, thus the possibility of false positives cannot be excluded.

In conclusion, metformin lowered estradiol levels, independent of BMI and insulin in non-diabetic post-menopausal women with ER and PR negative BC enrolled onto MA.32 trial. This observation suggests a new mechanism of metformin action that may be relevant in breast and other estrogen mediated cancers.

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Notes

Role of the funder: The funders had no role in the design of the study; the collection, analysis, and interpretation of the data; the writing of the manuscript; and the decision to submit the manuscript for publication.

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Table 1: Patient and tumor characteristics at baseline

Characteristics	Metformin	Placebo	
Characteristics	n=141	n=171	
Age, years mean (SD)	58.1 (± 6.9)	57.5 (± 7.9)	
Weight, kg mean (SD)	72.6 (± 14.1)	76.6 (± 17.5)	
BMI, kg/m² mean (SD)	27.3 (± 5.2)	28.9 (± 6.4)	
Post-menopausal, n (%)	141 (100)	171 (100)	
Receptor status, n (%)			
ER/PR negative	141 (100)	171 (100)	
Her2 status, n (%)			
Her2 positive	32 (22.7)	27 (15.8)	
Her2 negative	109 (77.3)	144 (84.2)	
Any (neo)adjuvant chemotherapy, n (%)			
Yes	139 (98.6)	168 (98.2)	
No	2 (1.4)	3 (1.8)	
T stage, n (%)			
T1	55 (39.0)	73 (42.7)	
T2	77 (54.6)	80 (46.8)	
T3	9 (6.4)	18 (10.5)	
N stage, n (%)			
N0	94 (66.7)	105 (61.4)	
N1	34 (24.1)	40 (23.4)	
N2	8 (5.7)	18 (10.5)	
N3	5 (3.5)	8 (4.7)	

Table 2: Baseline Sex Hormones measurements and change from baseline to month 6 on metformin vs placebo arms.

Sex Hormones	Baseline Median (Q1, Q3)		Change ¹		
			Median (Q1, Q3)		
	Metformin	Placebo	Metformin	Placebo	P^2
	n=140	n=170	n=136	N=168	
Estradiol pmol/L	32.2 (19.0, 45.5)	33.3 (9.2, 56.4)	-5.7 (-18.6,0)	0 (-12.6, 14.7)	< 0.001
BT nmol/L	0.02 (0.01, 0.05)	0.03 (0.02,0.06)	0 (-0.01, 0.01)	0 (-0.01, 0.02)	0.02
SHBG nmol/L	76.4 (60.9, 112)	72.8 (48.8, 105)	-5.9 (-15.6, 3.0)	-5.9 (-17.4, 1.5)	0.43

^{*}Q1 and Q3 are the 25th and 75th percentiles, SHBG - Sex Hormone Binding Globulin, BT - Bioavailable Testosterone

¹ Change calculated with-in patient as month 6 value minus baseline value

² P-value from two-sided Wilcoxon rank sum test comparing change between the two arms

Figure titles and legends

Figure1: Consort Diagram.

Figure 2: Exceedance curves for estradiol at 6 months, by study arm. The curves

give the proportion of patients whose estradiol was higher than any chosen cut-off on

the x-axis, for example, 20% of metformin patients versus 43% of placebo patients had

estradiol scores over 40 pmol/L.

Figure 3: Linear regression model for change in estradiol, adjusted for baseline

estradiol, with treatment (metformin vs placebo) and SNP (Any C vs AA) as

explanatory variables. ¹ E_{AnyC}/E_{AA} denotes the ratio of estradiol levels at month 6 for the

two SNP levels, adjusted to the same baseline estradiol. ² Main effects model: log(month 6

estradiol) - log(baseline estradiol) as outcome and treatment status, SNP status and baseline

assay level as covariates. The p-value is for the null hypothesis of no SNP effect, which in terms

of back-transformed results is that the ratio $E_{AnyC}/E_{AA}=1$. ³ Interaction model: A treatment

status-by-SNP status interaction was added to the main effects model. The p-value is for the

null hypothesis of no interaction effect, which in terms of back-transformed results is that

 E_{AnyC}/E_{AA} in the metformin arm equals E_{AnyC}/E_{AA} in the placebo arm.



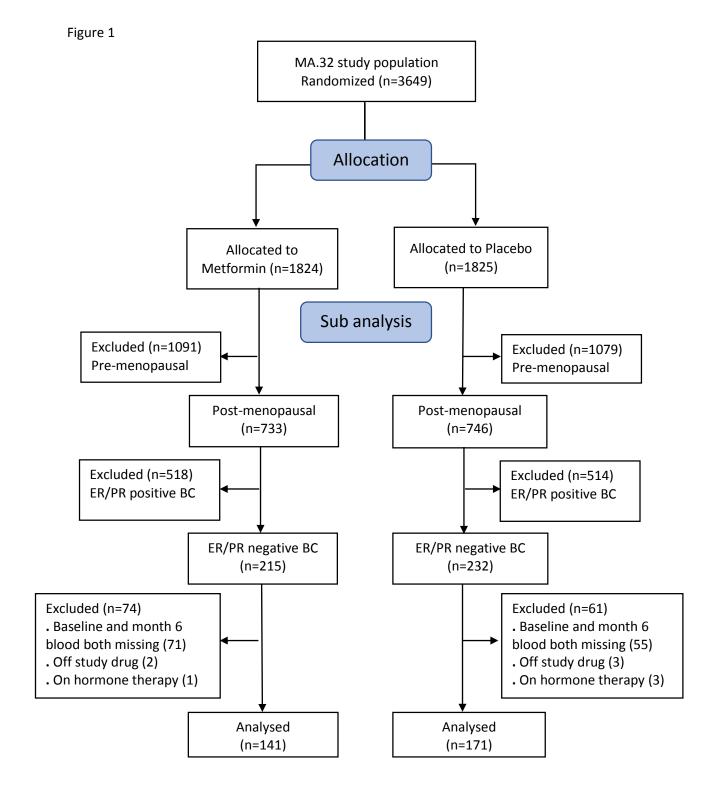


Figure 2

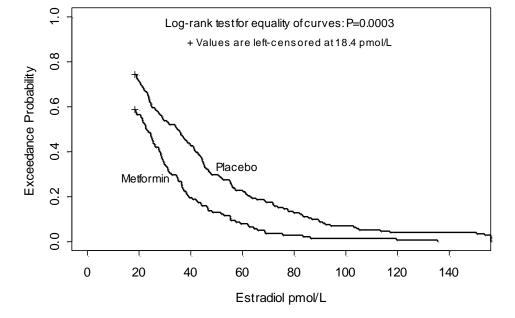


Figure 3

Regression model for estradiol with an overall SNP effect ²						
E _{AnyC} /E _{AA} ratio ² in both arms combined						
	1.00 (0.85 – 1.18)					
Regression model for estradiol that allows a different SNP effect by study arm ³						
E _{AnyC} /E _{AA} ratio ¹	E _{AnyC} /E _{AA} ratio					
in	in	Ratio of two				
metformin arm	placebo arm	treatment arm ratios	P^3			
0.94	1.05	0.90 (0.64 - 1.25)	0.51			