

E. coli could not recognize the pili of any of the non-O1 *V. cholerae* suggests that there is no immunologic cross-reactivity between the pili of enterotoxigenic *E. coli* and those of *V. cholerae* non-O1.

This report clearly demonstrated the production of pili that were immunologically different from the pili (CFAs) of enterotoxigenic *E. coli* by some non-O1 strains of *V. cholerae*. Because pili of enterotoxigenic *E. coli* serve as colonization factors, by analogy we postulate that the pili of non-O1 *V. cholerae* may also play a role as colonization factors. This interesting possibility requires further study. It is also important to analyze the relationship between the pili of *V. cholerae* O1 reported by others [5, 9, 10] and those of *V. cholerae* non-O1 reported here.

TAKESHI HONDA, KESARA KASEMSUKSAKUL, TOMIAKI OGUCHI,
MITSUO KOHDA, TOSHIO MIWATANI

Research Institute for Microbial Diseases,
Osaka University, Yamadaoka, Suita,
Osaka, Japan

References

1. Morris JG Jr, Black RE. Cholera and other vibrioses in the United States. *N Engl J Med* 1985;312:343-50
2. Yamamoto K, Takeda Y, Miwatani T, Craig JP. Evidence that a non-O1 *Vibrio cholerae* produces enterotoxin that is similar but not identical to cholera enterotoxin. *Infect Immun* 1983;41:896-901
3. Honda T, Arita M, Takeda T, Yoh M, Miwatani T. Non-O1 *Vibrio cholerae* produces two newly identified toxins related to *Vibrio parahaemolyticus* haemolysin and *Escherichia coli* heat-stable enterotoxin. *Lancet* 1985;2:163-4
4. Gaastra W, de Graaf FK. Host-specific fimbrial adhesins of noninvasive enterotoxigenic *Escherichia coli* strains. *Microbiol Rev* 1982;46:129-61
5. Faris A, Lindahl M, Wadstrom T. High surface hydrophobicity of hemagglutinating *Vibrio cholerae* and other vibrios. *Current Microbiology* 1982;7:357-62
6. Honda T, Arita M, Miwatani T. Characterization of new hydrophobic pili of human enterotoxigenic *Escherichia coli*: a possible new colonization factor. *Infect Immun* 1984;43:959-65
7. Honda T, Khan MMA, Takeda Y, Miwatani T. Grouping of enterotoxigenic *Escherichia coli* by hydrophobicity and its relation to hemagglutination and enterotoxin productions. *FEMS Microbiol Lett* 1983;17:273-6
8. Miwatani T, Honda T. Colonization factors of human enterotoxigenic *Escherichia coli*. *Bifidobacteria Microflora* 1986;5:57-65
9. Ehara M, Ishibashi M, Watanabe S, Iwanaga M, Shimodori S, Naito T. Fimbriae of *Vibrio cholerae* O1: observation of fimbriae on the organisms adherent to the intestinal epithelium and development of a new medium to enhance fimbriae production. *Tropical Medicine* 1986;28:21-33
10. Tweedy JM, Park RWA, Hodgkiss W. Evidence for the presence of fimbriae (pili) on *Vibrio* species. *J Gen Microbiol* 1968;51:235-44

THE JOURNAL OF INFECTIOUS DISEASES • VOL. 157, NO. 1 • JANUARY 1988
© 1987 by The University of Chicago. All rights reserved. 0022-1899/88/5701-0040\$01.00

Immunoperoxidase Slide Test For Detecting Antibodies to *Borrelia burgdorferi*

COLLEAGUES—*Borrelia burgdorferi* has recently been established as the causative agent of Lyme disease [1, 2]. Determining circulating antibodies to *B. burgdorferi* followed by demonstrating the agent by culture or by histology has given evidence for the etiology of some dermatoses that were clinically defined long ago (erythema chronicum migrans, lymphadenosis benigna cutis, acrodermatitis chronica atrophicans). A causative role for *B. burgdorferi* is discussed for other disease entities [3]. So far, circulating antibodies to *B. burgdorferi* have been determined by immunofluorescence tests (IFT) and ELISA. Both methods require special technical equipment (fluorescence microscope, photometer) and experience in evaluating the results.

We developed an immunoperoxidase slide test (IPT) that is easily performed and read with a simple light microscope. The binding of specific IgG antibodies to smears of *B. burgdorferi* (strain B31) fixed on slides by acetone is visualized by subsequent incubation with peroxidase-conjugated antibody to human IgG followed by incubation with a chromogenic substrate. This method was com-

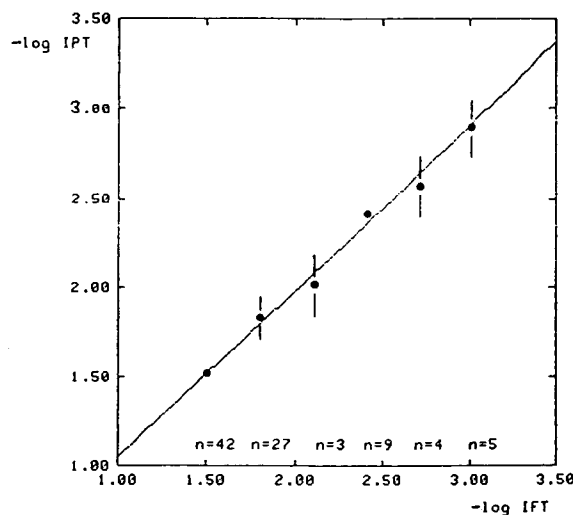


Figure 1. Correlation of titers of IgG antibody to *B. burgdorferi* in IPT and IFT. ●, — log of the mean titer for the no. of determinations indicated on figure; Bars represent ± 2 SD ($n = 90$, $r = .9805$, $P < .001$; $1.51 = -\log 1:32$, $1.81 = -\log 1:64$, $2.11 = -\log 1:128$, $2.41 = -\log 1:256$, $2.71 = -\log 1:512$, $3.01 = -\log 1:1024$).

Please address requests for reprints to Dr. J. Schmidli, Department of Dermatology, University of Bern, Inselspital, CH-3010 Bern, Switzerland.

pared with IFT by using both methods to test sera from 90 patients. Antibody titers obtained by using the two tests showed excellent correspondence (figure 1), with a correlation index (r) of .9805 ($P < .001$). In view of these almost identical results, the IPT looks promising as a simple screening assay suitable for every serological laboratory.

We have recently compared the titers of IgG and IgM antibodies to *B. burgdorferi* obtained by using IPT with those obtained by using ELISA and IFT [4].

J. SCHMIDLI, T. HUNZIKER, H. GERBER

*Department of Dermatology, University of Bern,
Inselspital, Bern, Switzerland*

References

1. Burgdorfer W, Barbour AG, Hayes SF, Benach JL, Grunwaldt E, Davis JP. Lyme disease—a tick-borne spirochetosis? *Science* 1982;**216**: 1317–9
2. Barbour AG, Burgdorfer W, Grunwaldt E, Steere AC. Antibodies of patients with Lyme disease to components of the *Ixodes dammini* spirochete. *J Clin Invest* 1983;**72**:504–15
3. Ruffli T, Lehner S, Aeschlimann A, Chamot E, Gigon F, Jeanneret J-P. Zum erweiterten spektrum zeckenübertragener spirochätosen. *Hautarzt* 1986;**37**:597–602
4. Schmidli J, Gerber H, Hunziker T, Aeschlimann A, Gern Lise, Kundi M, Stanek G. Immunoperoxidase slide test for the detection of antibodies against *Borrelia burgdorferi* in comparison to immunofluorescence test and ELISA. *Zentralbl Bakteriol Mikrobiol Hyg [A]* 1987 (in press)