brought to you by **CORE** rovided by Simorgh Research Repositor

Environmental Health Engineering and Management Journal 2020, 7(1), 15–22 http://ehemj.com

HE Environmental Health Engineering and Management Journal

Open Access

**Original** Article





doi 10.34172/EHEM.2020.03

# The effect of organic chelates and gibberellic acid on petroleum hydrocarbons degradation in the soil co-contaminated with Ni and crude oil under canola cultivation

Amir Hossein Baghaie<sup>1\*</sup>, Amir Daliri<sup>10</sup>

<sup>1</sup>Department of Soil Science, Arak Branch, Islamic Azad University, Arak, Iran

#### Abstract

**Background:** Soil remediation is one the important problem in environmental studies. Thus, this research was conducted to evaluate the effect of organic chelates and gibberellic acid  $(GA_3)$  on the degradation of crude oil in the soil co-contaminated with Ni and crude oil under canola cultivation.

**Methods:** For treatments, HEDTA and NTA chelates at rates of 0 and 2.5 mmol/kg soil and foliar GA<sub>3</sub> (0 (GA<sub>3</sub>(-) and 0.05 (GA<sub>3</sub>(+) mM) were used. In addition, the soil was polluted with Ni (0 and 100 mg Ni/kg soil) and crude oil at rates of 0, 2, and 4% (W/W). The plant used in this experiment was canola. The concentration of Ni in soil and plant was measured using atomic absorption spectroscopy (AAS). The concentration of total petroleum hydrocarbon (TPH) was measured using GC-mass. The mean differences were calculated according to the least significant difference (LSD) test.

**Results:** The greatest degradation of crude oil belonged to the non-Ni-polluted soil under cultivation of GA<sub>3</sub>-treated plant, while the lowest one was observed in the soil received the greatest level of HEDTA and NTA chelates. Applying 0.05 mM GA<sub>3</sub> foliar significantly increased the degradation of crude oil in soil and Ni in plant shoot by 12.1 and 8.3%, respectively. In addition, soil microbial respiration was also increased by 11.3%.

**Conclusion:** HEDTA, NTA, and GA<sub>3</sub> had a significant effect on the Ni phytoremediation efficiency and degradation of crude oil in soil that is a positive point in environmental pollution. However, the role of soil physico-chemical properties on the phytoremediation efficiency cannot be ignored. **Keywords:** Soil pollutants, Biodegradation, Environmental, Petroleum

**Citation:** Baghaie AH, Daliri A. The effect of organic chelates and gibberellic acid on petroleum hydrocarbons degradation in the soil co-contaminated with Ni and crude oil under canola cultivation. Environmental Health Engineering and Management Journal 2020; 7(1): 15–22. doi: 10.34172/ EHEM.2020.03.

### Introduction

Petroleum products are the most commonly used materials that are processed from crude oil in an oil refinery. Petroleum products or by-products are generally used in different industries, such as car and airplanes manufacturing (1,2). However, the use of petroleum compounds has been associated with environmental pollution that can be due to its toxicity, and it can cause soil and air pollution (3,4).

Organic pollutants, such as petroleum hydrocarbons, can enter the soil, air, and groundwater through the leakage from pipelines or underground storage tanks, illegal dumping and accidental spills, and as a result, contaminate the soil and air (5). Remediation of soil contaminated by petroleum hydrocarbons, such as crude oil, is generally difficult due to its high hydrophobicity and high sorption capacity. Thus, it is necessary to find a suitable way for the remediation of these components from the soil (6). Article History: Received: 18 September 2019 Accepted: 20 December 2019 ePublished: 4 February 2020

\*Correspondence to: Amir Hossein Baghaie Email: a-baghaie@iau-arak.ac.ir

However, in the industrialized areas, there is a synchronous effect of heavy metals and petroleum contamination, and these two types of pollution impede the remediation process of the contaminated soils (7).

Various technologies have been applied to remediate petroleum hydrocarbon-contaminated soil, such as bioremediation, volatilization, surfactant flushing, and chemical oxidation (8,9). However, these techniques are generally expensive and can lead to incomplete decomposition of contaminants (10). The efficiency of each method highly depends on the pollution type and soil chemical properties of each area (9). Among above-mentioned remediation methods, biodegradation (phytodegradation) is an environmentally friendly technique applicable for the remediation of soils contaminated by hydrocarbons (11). It should be noted that after bioremediation, the petroleum hydrocarbons convert into water, inorganic compounds, carbon dioxide,

<sup>© 2020</sup> The Author(s). Published by Kerman University of Medical Sciences. This is an open-access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

or to other simpler organic compounds (12).

On the other hand, applying organic chelates, such as ethylene diamine tetraacetic acid (EDTA), ethylene diamine diamine disuccinate, hydroxylethylene tetraacetic acid (HEDTA), and nitrilotriacetic acid (NTA) greatly contributes to the remediation of heavy metalscontaminated soils (13,14). However, their effect depends on the type of heavy metal, as Meers et al reported that the application of NTA (1.8 mmol/kg soil) chelate did not significantly increase the zinc (Zn), cadmium (Cd), and copper (Cu) concentration in plant compared to the control in sunflower. However, in the case of nickel (Ni), the uptake of Ni increased by 2.5-fold (15). Houshyar et al investigated the effectiveness of DTPA chelate on the Cd availability in the soils treated with sewage sludge and concluded that the application of DTPA chelate had significant effects on increasing the Cd uptake by corn (16). However, they did not consider the interaction effects of heavy metals. The positive role of applying organic chelate on increasing heavy metals uptake by plant was also reported by Baghaie et al (17).

Today, remediation of soils contaminated with petroleum hydrocarbons, such as crude oil, is also one of the major environmental problems (18). Due to the simultaneous effect of heavy metals contamination with petroleum hydrocarbons, finding a suitable way for the degradation of petroleum hydrocarbons with increasing heavy metals remediation seems necessary. Generally, serpentine rocks are the primary sources of Ni metal. In addition, it can be founded in Ni-contaminated areas due to various human activities, such as extraction of nickel ore, burning of fossil fuels, oil residues and sewage sludge and production of Ni-Cd batteries. Today, bioremediation is considered as one of the most successful methods for the degradation of petroleum hydrocarbons and heavy metals remediation (19). However, soil remediation efficiency has been impeded due to the slow growth of plants in these soils. Thus, applying a suitable strategy for enhancing plant growth in these soils can be helpful for the bioremediation process (20).

However, the use of plant growth regulators, such as gibberellic acid (GA<sub>3</sub>), could be effective in the remediation of contaminated soils via increasing plant growth. Chouychai et al investigated the effect of plant growth regulators on the phytoremediation of hexachlorocyclohexane-contaminated soil and concluded that the use of GA<sub>3</sub> can increase the phytoremediation efficiency (21). Shafigh et al investigated the phytoremediation of lead by corn in the Pb-polluted soil and concluded that the use of chelates with plant growth regulators can have an effective role in enhancing the remediation efficiency of the contaminated soils (22). Hence, despite of the positive effect of GA<sub>3</sub> application on increasing remediation efficiency, a practical solution should also be considered to increase the heavy metals availability in soil. Accordingly, the interaction effects of applying organic chelates and  $GA_3$  may be useful in the remediation of the contaminated soils. Thus, this research was conducted to evaluate the effects of organic chelates and  $GA_3$  on the petroleum hydrocarbons degradation in the soil co-contaminated with Ni and crude oil under canola cultivation.

#### Materials and Methods

To investigate the effect of HEDTA, NTA, and GA<sub>3</sub> on the petroleum hydrocarbons degradation in the Ni-treated soil, a non-Ni-polluted soil sample with low organic carbon was selected from the surface layer of soil (0-15 cm) around Pakal village, Arak, central of Iran. Physicochemical properties of the studied soil are shown in Table 1. This research was conducted as a factorial experiment in the layout of randomized complete block design. For treatments, HEDTA and NTA chelates at rates of 0 and 2.5 mmol/kg soil and foliar  $GA_3$  (0 ( $GA_3$ (-) and 0.05 ( $GA_3$ (+) mM) were used. In addition, the soil was polluted with Ni (0 and 100 mg Ni/kg soil) and crude oil at rates of 0, 2, and 4% (W/W). The plant used in this experiment was canola. The soil used in this experiment was polluted with Ni at rates of 0 and 100 mg Ni/kg soil and incubated for two weeks. Then, crude oil was added to the soil at rates of 0, 2, and 4% (W/W) and incubated for two weeks to reach equilibrium. Canola seedling surface was first sterilized in 15% H<sub>2</sub>O<sub>2</sub>, thoroughly washed in distilled water, and pre-germinated on moistened filter paper. After that, two canola seedlings were planted into each pot with five kg soil. GA<sub>3</sub> were sprayed on a canopy as foliar spray method 30 days after sowing (23). After 80 days, plants were harvested and the concentration of Ni in the soil was measured according to the Lindsay method (24). To measure the Ni concentration in the plant, samples were washed with deionized water, and then, dried in an oven at 60°C for 24 hours, and finally, digested in a mixture of HNO<sub>3</sub>/HClO<sub>4</sub> (85:15%, v/v) to determine the total concentration of Ni in the plant according to the Yang et al method (25). Afterwards, the Ni concentration in the plant was measured using atomic absorption spectroscopy (AAS). The basal soil microbial respiration was measured according to Besalatpour et al (26). For this purpose, 3 replicate soil samples for each treatment were incubated

Table 1. Selected physico-chemica	I properties of the studied soil
-----------------------------------	----------------------------------

Characteristic	Unit	Amount
Soil texture	-	Sandy loam
рН	-	7.3
EC	dS/m	1.2
Soil Pb availability	mg kg <sup>-1</sup>	ND
Soil Ni availability	mg kg <sup>-1</sup>	ND
Soil As availability	mg kg <sup>-1</sup>	ND
Organic carbon	%	0.1
CaCO <sub>3</sub>	%	9%

ND: Not detectable by atomic absorption spectroscopy.

for three days at 26°C in 250-mL glass containers closed with rubber stoppers. The evolved  $CO_2$  was trapped in a standard NaOH solution, and then, the excess of which in alkali was titrated with HCl (26). The total petroleum hydrocarbons (TPHs) in the soil were extracted from 30 g of the soil subsamples by soxhlet extractor using 150 mL of the mixture of dichloromethane and n-hexane (1:1, v/v) after 24 hours, and then, using the method applied in a study by Baghaie et al (27).

Ni translocation factor (TF) was calculated using the following formula (28):

$$TF=H_{shoot}/H_{root}$$
(1)

Where  $H_{shoot}$  and  $H_{root}$  are heavy metals concentrations in the plant shoot and root, respectively.

Statistical analyses were calculated according to the analysis of variance (ANOVA) procedure. The mean differences were calculated according to the least significant difference (LSD) test. Statistical significant value was considered at P=0.05.

# Results

The simple effect of applying HEDTA, NTA, and  $GA_3$  on increasing the soil Ni availability was significant (P = 0.05).

Table 2. Effects of the treatments on the soil Ni availability

The greatest soil Ni availability belonged to the Ni- and crude oil-polluted soil, which received the greatest level of HEDTA, NTA with the GA<sub>3</sub> foliar application, and the lowest one was observed in the soil received no pollutant (Table 2). Application of HEDTA in the Ni-polluted soil significantly (P=0.05) increased the soil Ni availability by 15.2%. Foliar GA<sub>3</sub> application had significant effect on increasing the soil Ni availability, as the application of 0.05 mM GA<sub>3</sub> had significantly increased the soil Ni availability (Table 2), which was contaminated with crude oil (4% W/W). Increasing soil contamination with crude oil had significant effect on increasing the soil Ni availability, as the greatest soil Ni availability belonged to the soil polluted with 4% (W/W) crude oil (Table 2).

The greatest root Ni concentration belonged to the plants cultivated in the Ni-polluted soil, which received the greatest level of organic chelate, such as HEDTA, simultaneous application of NTA with the GA3 foliar, while the lowest one was observed in the soil received no organic chelate (Table 3). Applying 2.5 mmol/kg soil HEDTA and NTA chelate in the Ni-polluted soil significantly increased the root Ni concentration by 18.4 and 12.1%, respectively.

The greatest shoot Ni concentration has belonged to the plants sprayed with GA<sub>3</sub> and cultivated in the soil that

	Chelate (mmol/kg soil)		Crude oil (%)			Crude oil (%)		
Ni Pollution (mg/kg soil)	Cherate (mr	noi/ kg soli)	0	2	4	0	2	4
	HEDTA	NTA		GA <sub>3</sub> (+)			GA <sub>3</sub> (-)	
	0	0	ND	ND	ND	ND	ND	ND
0	0	2.5	ND	ND	ND	ND	ND	ND
0	2.5	0	ND	ND	ND	ND	ND	ND
		2.5	ND	ND	ND	ND	ND	ND
100	0	0	18.3p	22.1	24.6i	17.8q	19.20	21.9m
	0	2.5	21.9m	25.2h	27.4e	20.4n	23.1k	24.8i
	2 5	0	24.2j	27.6e	29.4c	22.4	25.2h	28.1d
	2.5	2.5	27.1f	30.2b	31.3a	25.8g	27.4	29.4c

ND: Not detectable by the atomic absorption spectroscopy.

Note: Means with similar letters are not significant

Table 3. Effects of the treatments on root Ni concentration

	Chelate (mmol/kg soil)			Crude oil (%)		Crude oil (%)		
Ni Pollution (mg/kg soil)	Chelate (mi	moly kg soll)	0	2	4	0	2	4
	HEDTA	NTA		GA <sub>3</sub> (+)			GA <sub>3</sub> (-)	
	0	0	ND	ND	ND	ND	ND	ND
0	0	2.5	ND	ND	ND	ND	ND	ND
0	2.5	0	ND	ND	ND	ND	ND	ND
		2.5	ND	ND	ND	ND	ND	ND
100	0	0	82.81	85.1i	87.9g	78.1n	81.6m	83.1k
	0	2.5	84.6j	87.2g	90.4d	81.1m	83.2k	85.6i
	2.5	0	87.3g	91.3c	93.5b	84.4j	86.2h	88.1e
	2.5	2.5	91.4c	93.2b	95.1a	88.2e	90.4d	91.2c

ND: Not detectable by the atomic absorption spectroscopy. *Note*: Means with similar letters are not significant

received the greatest level of HEDTA and NTA chelates (Table 4), while the lowest that was observed in the soil received no organic chelate. Foliar application of GA<sub>3</sub> significantly increased the shoot Ni concentration that maybe related to the positive role of GA<sub>3</sub> on increasing plant resistance to abiotic stress such as Ni, and as a result, increasing Ni concentration in the plant. Based on the results of this study, applying 0.05 mM GA, as a plant growth regulator significantly increased plant biomass (data was not shown) and shoot Ni concentration by 8.3 and 12.2%, respectively. Similar results were observed for the positive effects of GA<sub>3</sub> application on increasing Ni TF value, as the greatest Ni TF value belonged to the GA<sub>3</sub>treated plants (Table 5).

NTA, can help increase the Ni TF value, as applying 2.5 mmol/kg soil HEDTA significantly increased the Ni TF value by 0.04 units in the soil under cultivation of GA<sub>3</sub>treated plant. Similar results for NTA chelate confirm the results of the present study clearly.

The greatest soil microbial respiration belonged to the soil treated with 4% (W/W) crude oil in non-Ni-polluted soil without receiving any organic chelates (Table 6), while the lowest one was observed in the soil received no petroleum hydrocarbons. Applying organic chelates had adverse effect on the soil microbial respiration, as applying 2.5

mmol/kg soil HEDTA in the Ni-polluted soil (100 mmol Ni/kg soil) significantly decreased the soil microbial respiration by 8.4%. The crude oil degradation in the soil showed a similar trend with the changes in soil microbial respiration (Table 7).

# Discussion

Applying organic chelates had a significant effect on increasing the heavy metal availability in the soil that maybe due to the role of organic chelate on the heavy metal solubility in the soil. Chen et al investigated the effects of HEDTA chelate on the heavy metals availability in the soil and concluded that applying this chelate has a positive role on increasing the availability of heavy metals in the soil, such as Ni, which is consistent with the results of this study (29). However, they did not investigate the role of soil chemical properties on the changes in the heavy metals availability in the soil. The results of a study by Evangelou et al about the role of organic chelate on increasing the heavy metal availability in soil confirm the results of this study clearly (30).

The positive role of foliar GA, application on increasing the soil Ni availability may be related to the role of plant growth regulators on increasing plant biomass (data were not shown) and may be increasing plant root exudate. The

 	 	oonoonaaaon	
			 _

Table 4. Effects of treatments on shoot Ni concentration

	Chelate (mmol/kg soil)			Crude oil (%)		Crude oil (%)		
Ni Pollution (mg/kg soil)	Chelate (mi	noi/kg soli)	0	2	4	0	2	4
	HEDTA	NTA		GA <sub>3</sub> (+)			GA <sub>3</sub> (-)	
	0	0	ND	ND	ND	ND	ND	ND
0	0	2.5	ND	ND	ND	ND	ND	ND
0	2.5	0	ND	ND	ND	ND	ND	ND
		2.5	ND	ND	ND	ND	ND	ND
100	0	0	33.9q	37.40	43.1	28.1s	32.6r	36.6p
	0	2.5	38.0n	41.9m	49.7i	34.1q	37.40	42.81
	2.5	0	47.1k	56.6e	63.6b	42.2m	47.4k	54.6g
	2.5	2.5	53.0h	59.6d	68.5a	48.5j	55.1f	62.0c

ND: Not detectable by the atomic absorption spectroscopy. Note: Means with similar letters are not significant

Table 5. Effects of treatments on the Ni TF value

	Chelate (mmol/kg soil)		Crude oil (%)			Crude oil (%)		
Ni Pollution (mg/kg soil)	Chelate (mi	noi/kg soli) –	0	2	4	0	2	4
	HEDTA	NTA		GA <sub>3</sub> (+)			GA <sub>3</sub> (-)	
	0	0	ND	ND	ND	ND	ND	ND
0	0	2.5	ND	ND	ND	ND	ND	ND
0	2.5	0	ND	ND	ND	ND	ND	ND
		2.5	ND	ND	ND	ND	ND	ND
	•	0	0.410	0.44m	0.49k	0.36q	0.40p	0.44m
100	0	2.5	0.451	0.48k	0.55g	0.42n	0.451	0.50i
100	2.5	0	0.54h	0.62d	0.68b	0.50i	0.55g	0.62d
	2.5	2.5	0.58f	0.64c	0.72a	0.55g	0.61	0.68b

ND: Not detectable by the atomic absorption spectroscopy. Note: Means with similar letters are not significant

	Chelate (mmol/kg soil)			Crude oil (%)			Crude oil (%)		
Ni Pollution (mg/kg soil)	Cherate (mi		0	2	4	0	2	4	
	HEDTA	NTA		GA <sub>3</sub> (+)			GA <sub>3</sub> (-)		
	0	0	54.1c	55.4b	57.1a	50.1h	52.2f	54.2c	
0	0	2.5	51.9g	53.2e	55.3b	47.8k	49.2i	51.6g	
	2.5	0	52.2f	53.9d	55.3b	45.1m	47.6k	49.3i	
		2.5	48.7j	51.4g	53.8d	40.4r	42.9p	44.8n	
	0	0	50.1h	52.2f	53.9d	44.8n	48.6j	51.4g	
100	0	2.5	48.6j	50.1h	51.9g	41.6q	43.20	46.81	
	2.5	0	45.1m	46.8	48.1j	39.2s	42.1p	44.8n	
	2.5	2.5	42.1p	44.8n	46.81	34.1v	35.1u	37.4t	

## Table 6. Effects of treatments on the soil microbial respiration

Note: Means with similar letters are not significant.

Table 7. Effects of treatments on the crude oil degradation in the soil

	Chelate (mmol/kg soil)			Crude oil (%)			Crude oil (%)		
Ni Pollution (mg/kg soil)	Chelate (mr	noi/kg soli) -	0	2	4	0	2	4	
	HEDTA	NTA		GA <sub>3</sub> (+)			GA <sub>3</sub> (-)		
	0	0	65.4d	67.2c	71.5a	62.3g	65.4d	68.1b	
0	0	2.5	63.3f	64.4e	67.4c	61.2h	63.1f	65.2	
0	2.5	0	63.1f	65.2d	68.4b	60.3i	62.4g	65.1d	
		2.5	61.2h	62.6g	64.1e	58.1j	61.2h	63.5f	
	0	0	55.4k	58.1j	65.9d	50.1m	55.3k	58.2j	
100	0	2.5	50.2m	52.91	55.4k	47.5p	49.4n	50.3m	
	2.5	0	50.1m	55.3k	58.1j	45.1r	47.2p	48.60	
	2.5	2.5	47.6p	50.1m	52.91	41.2t	43.4s	46.1q	

Note: Means with similar letters are not significant.

positive effects of plant root exudate on decreasing the soil pH, and thereby, increase the heavy metal availability in soil has been mentioned by researchers (31,32). Azimzadeh et al investigated the remediation of some heavy metals in the soil by corn and canola in single and mixed culture system and concluded that plant root exudate can increase the heavy metal availability in the soil (33). Generally, plant root exudate increases the availability of heavy metals in the soil, although heavy metals solubility varies depending on the type of the metal (31). A study by Hadi et alshowed that applying GA<sub>3</sub> and synthetic chelates had significant effect on increasing heavy metal uptake by plant, which is consistent with the results of this study (34).

A significant increase in the soil Ni availability was observed when the soil was polluted with petroleum hydrocarbon, which may be related to the role of organic carbon on increasing the heavy metal availability in the soil. Askary Mehrabadi et al investigated the phytoremediation of petroleum hydrocarbon and heavy metals using *Catharanthus roseus* and concluded that increasing the soil pollution by petroleum hydrocarbon could increase the heavy metal availability in the soil. In addition, they mentioned that the simultaneous contamination of soils with petroleum compounds and heavy metals had a negative effect on the plant growth (35). However, they introduced *Catharanthus roseus* as the phytoremediator of the petroleum-contaminated soil at low concentrations. Based on the results of this study, applying organic chelates had a positive effect on increasing the soil Ni availability, and thereby, increased Ni uptake by plant. Naghipour et al investigated the effect of EDTA and NTA on the heavy metal extraction from the sandy-loam-contaminated soils and concluded that applying these chelates had significant effects on heavy metal extraction from the soil, and thereby, heavy metal sorption by plants (36). In addition, they mentioned that using EDTA has higher efficiency in remediating the contaminated soils, in comparison with NTA, which is consistent with the results of the present study. Accordingly, applying 2.5 mmol/kg soil HEDTA compared to NTA chelate, significantly increased the Ni root concentration by 6.3% in Ni-polluted soil. It is mentioned that in order to decide whether artificial chelating agents can be applied, several important factors should be taken into account (36).

Soil polluted to crude oil pollution significantly increasing soil microbial respiration (Table 6). The remarkable point in this study is that the soil hydrocarbons act as a carbon source rather than pollutant factor. Based on the results of our studies, the greatest soil hydrocarbon degradation (Table 7) has belonged to the same treatment. Besalatpour el al reported the significant increases in soil microbial respiration in petroleum-contaminated soil and they concluded that the petroleum hydrocarbon can be used as a carbon sources rather than toxicity factor. However, they reported that the high concentration of petroleum can decrease the microbial activation (26). Based on the results of this study, increasing the crude oil pollution of soil to 4% (W/W) did not decrease the degradation of crude oil in the soil. However, the role of higher levels of crude oil pollution of soil on the petroleum hydrocarbon degradation in the soil needs to be investigated in the future studies. Franco et al reported a significant increase in microbial biomass in the crude oil-contaminated soil and concluded that this might be due to the use of oil hydrocarbon compounds by microorganisms as a carbon source (37), which is consistent with the results of this study.

Applying organic chelates, such as HEDTA and NTA, had adverse effects on the degradation of crude oil in the Ni-polluted soil that maybe due to the negative effect of heavy metals on the soil microbial activities (38). In fact, chelate application increased the heavy metals availability in the soil and reduced the activity of microorganisms to decompose petroleum hydrocarbons in the soil, although the Ni phytoremediation efficiency increased. Accordingly, application of 2.5 mmol/kg soil HEDTA chelate significantly decreased the soil microbial respiration and crude oil degradation by 11.6 and 8.9%, respectively, while the Ni TF value increased from 0.41 to 0.58 in the Ni-polluted soil (Table 5). Similar results were obtained for NTA chelate. Generally, soil microorganisms are among the first things that experience the negative impacts of pollutants, and their population and diversity can be used as an index to assess the degree of pollution in the environment (39). Although, the timing of the soil contamination with heavy metals can be a significant factor in the resistance of soil microorganisms to heavy metals, which are needed to be investigated in the future studies. Accordingly, Chander and Joergensen reported that the microbial population in long-term contaminated soils has better tolerance towards the application of organic chelate, compared to the short-term Zn-contaminated soil (40). Applying plant growth regulators such as  $GA_3$  (+), had significant effects on the petroleum hydrocarbons degradation in soil. Regardless of the amount of the Ni pollution in soil, the greatest and lowest degradation rates of crude-oil in soil belonged to the GA<sub>3</sub>(+) and GA<sub>3</sub>(-) treatments, respectively. The results of soil microbial activities showed the similar trends. The foliar application of GA<sub>3</sub> (mmol/kg) significantly increased the petroleum hydrocarbons degradation in the Ni-polluted soil (100 mg/kg soil) by 9.6%. However, increasing the soil contamination with Ni significantly decreased the crude oil degradation in soil. Regardless of the amount of crude oil in soil and GA<sub>3</sub> application, with increasing soil contamination with Ni from 0 to 100 mg Ni/kg soil, a significant decrease in the crude oil degradation in the soil and soil microbial activities was observed.

On the other hand, plant growth-promoting rhizobacteria such (PGPR) as GA<sub>3</sub> can increase plant biomass (data were not shown). The positive effect of PGPR on plant growth is providing the condition for plant growth via compounds that is synthesized by the microorganisms, for example phyto-hormones, or increase plant nutrients uptake (41). Shafigh et al investigated the effect of GA<sub>3</sub> on the Ni phytoremediation in the Ni-polluted soil and concluded that the application of GA<sub>3</sub> had a significant effect on increasing corn biomass, and thereby, increased Ni phytoremediation efficiency (42). Therefore, increasing plant biomass can increase plant resistance to heavy metals toxicity. On the other hand, increasing plant biomass can increase plant root exudate, and thereby, influence increasing soil microbial activities or petroleum hydrocarbon degradation. Generally, rhizoremediation has a great potential for the remediation of the soil contaminated with organic pollutants, such as petroleum hydrocarbons (43). The important point of this study is that although the greater Ni availability was observed in the soil under the cultivation of the GA<sub>3</sub>-treated plants, but due to the increased resistance of plant to abiotic stress (increased plant biomass), the microbial activities were also significantly increased, and as a result, the soil microbial respiration increased. Therefore, the use of plant growth hormone not only increased the Ni phytoremediation efficiency, but also increased the degradation of petroleum hydrocarbons in the soil, which is a positive point in environmental studies. However, the role of soil microbial activities on the heavy metal immobilization in soil cannot be ignored. Microorganisms, which are mostly prokaryotic, participate in redox reactions and change the valance of heavy metals, and thereby, change their activity, which can affect their mobility or toxicity (37). For example, microorganisms such as Rhizopus can absorb heavy metal ions in the soil and Thiobacillus can absorb heavy metal ions as well as inorganic ions, such as S, which combine with the metal ions to form a precipitate that can be separated from the soil (44).

## Conclusion

According to the results of this study, applying HEDTA and NTA chelates significantly increased the soil Ni availability, which has a negative effect on the degradation of crude oil in soil. However, using these chelates significantly increased the Ni uptake by plant. On the other hand,  $GA_3$  foliar application significantly increased the degradation of crude oil in soil that can be due to the positive role of microbial activities (microbial respiration) on the degradation of petroleum hydrocarbons in soil. Accordingly, it was concluded that in order to increase the degradation efficiency of petroleum hydrocarbons in soil, environmental management should be considered to increase the plant resistance to environmental pollutants, such as heavy metals. However, application of plant growth regulators, such as  $GA_3$ , can increase plant resistance to heavy metals toxicity via increasing plant biomass, which has a positive effect on increasing soil microbial activities, and thereby, increases the degradation of petroleum hydrocarbons in soil. However, the plant physiology, soil chemical properties, and type and amount of organic components has significant effects on the amount of petroleum hydrocarbon degradation, which should be considered in the future studies.

# Acknowledgements

The authors would like to appreciate Islamic Azad University, Arak Branch, for their assistance in analyzing samples.

### **Ethical issues**

The authors hereby certify that all data collected during the research are as expressed in the manuscript, and no data from the study has been or will be published elsewhere separately.

#### **Competing interests**

The authors have declared that they have no conflict of interests.

# Authors' contributions

All authors contributed in the data collection, analysis, and interpretation. All authors reviewed and approved the manuscript.

## References

- 1. Choromański P, Karwowska E, Łebkowska M. The influence of petroleum products on the methane fermentation process. J Hazard Mater 2016; 301: 327-31. doi: 10.1016/j. jhazmat.2015.09.011.
- 2. Maharaj R, St George A, Russel SN, Singh-Ackbarali D. The influence of recycled tyre rubber on the rheological properties of Trinidad lake asphalt and Trinidad petroleum bitumen. Int J Appl Chem 2009; 5(3): 181-91.
- Seeley ME, Wang Q, Bacosa H, Rosenheim BE, Liu Z. Environmental petroleum pollution analysis using ramped pyrolysis-gas chromatography-mass spectrometry. Org Geochem 2018; 124: 180-9. doi: 10.1016/j. orggeochem.2018.07.012.
- Wei H. Impacts of China's national vehicle fuel standards and subway development on air pollution. J Clean Prod 2019; 241: 118399. doi: 10.1016/j.jclepro.2019.118399.
- Mojiri A, Zhou JL, Ohashi A, Ozaki N, Kindaichi T. Comprehensive review of polycyclic aromatic hydrocarbons in water sources, their effects and treatments. Sci Total Environ 2019; 696: 133971. doi: 10.1016/j. scitotenv.2019.133971.
- Karthick A, Roy B, Chattopadhyay P. A review on the application of chemical surfactant and surfactant foam for remediation of petroleum oil contaminated soil. J Environ Manage 2019; 243: 187-205. doi: 10.1016/j. jenvman.2019.04.092.
- 7. Ma Y, Li X, Mao H, Wang B, Wang P. Remediation of hydrocarbon-heavy metal co-contaminated soil by

electrokinetics combined with biostimulation. Chem Eng J 2018; 353: 410-8. doi: 10.1016/j.cej.2018.07.131.

- Murray EW, Greenberg BM, Cryer K, Poltorak B, McKeown J, Spies J, et al. Kinetics of phytoremediation of petroleum hydrocarbon contaminated soil. Int J Phytoremediation 2019; 21(1): 27-33. doi: 10.1080/15226514.2018.1523870.
- Baoune H, Aparicio JD, Acuña A, El Hadj-Khelil AO, Sanchez L, Polti MA, et al. Effectiveness of the Zea mays-Streptomyces association for the phytoremediation of petroleum hydrocarbons impacted soils. Ecotoxicol Environ Saf 2019; 184: 109591. doi: 10.1016/j.ecoenv.2019.109591.
- Nishiwaki J, Kawabe Y, Komai T, Zhang M. Decomposition of gasoline hydrocarbons by natural microorganisms in Japanese soils. Geosciences 2018; 8(2): 35. doi: 10.3390/ geosciences8020035.
- Ite AE, Ibok UJ. Role of plants and microbes in bioremediation of petroleum hydrocarbons contaminated soils. International Journal of Environmental Bioremediation & Biodegradation 2019; 7(1): 1-19. doi: 10.12691/ijebb-7-1-1.
- Das N, Chandran P. Microbial degradation of petroleum hydrocarbon contaminants: an overview. Biotechnol Res Int 2011; 2011: 941810. doi: 10.4061/2011/941810.
- 13. Jiang M, Liu S, Li Y, Li X, Luo Z, Song H, et al. EDTAfacilitated toxic tolerance, absorption and translocation and phytoremediation of lead by dwarf bamboos. Ecotoxicol Environ Saf 2019; 170: 502-12. doi: 10.1016/j. ecoenv.2018.12.020.
- Ko CH, Yang BY, Chang FC. Enhancement of phytoextraction by Taiwanese chenopod and Napier grass by soapnut saponin and EDDS additions. Environ Sci Pollut Res Int 2019; 26(33): 34311-20. doi: 10.1007/s11356-019-04538-y.
- Meers E, Ruttens A, Hopgood MJ, Samson D, Tack FM. Comparison of EDTA and EDDS as potential soil amendments for enhanced phytoextraction of heavy metals. Chemosphere 2005; 58(8): 1011-22. doi: 10.1016/j. chemosphere.2004.09.047.
- Houshyar P, Baghaei A. Effectiveness of DTPA chelate on Cd availability in soils treated with sewage sludge. Journal of Water and Wastewater 2017; 28(4): 103-11. doi: 10.22093/ wwj.2017.45320. [In Persian].
- Baghaie AH. Effect of EDDS chelate on increasing Cd phytoremediation efficiency by corn (Maxima Cv) in a soil treated with municipal waste compost (a case study: Khomein Municipal Waste Compost). Journal of Environmental Health Engineering 2018; 5(4): 401-12. doi: 10.29252/jehe.5.4.401. [In Persian].
- Chikere CB, Mordi IJ, Chikere BO, Selvarajan R, Ashafa TO, Obieze CC. Comparative metagenomics and functional profiling of crude oil-polluted soils in Bodo West Community, Ogoni, with other sites of varying pollution history. Ann Microbiol 2019; 69(5): 495-513. doi: 10.1007/ s13213-019-1438-3.
- Abou-Elwafa SF, Amin AE, Shehzad T. Genetic mapping and transcriptional profiling of phytoremediation and heavy metals responsive genes in sorghum. Ecotoxicol Environ Saf 2019; 173: 366-72. doi: 10.1016/j.ecoenv.2019.02.022.
- 20. Kong Z, Wu Z, Glick BR, He S, Huang C, Wu L. Cooccurrence patterns of microbial communities affected by inoculants of plant growth-promoting bacteria during

phytoremediation of heavy metal-contaminated soils. Ecotoxicol Environ Saf 2019; 183: 109504. doi: 10.1016/j. ecoenv.2019.109504.

- Chouychai W, Kruatrachue M, Lee H. Effect of Plant growth regulators on phytoremediation of hexachlorocyclohexanecontaminated soil. Int J Phytoremediation 2015; 17(11): 1053-9. doi: 10.1080/15226514.2014.989309.
- 22. Shafigh M, Ghasemi-Fasaei R, Ronaghi A. Influence of plant growth regulators and humic acid on the phytoremediation of lead by maize in a Pb-polluted calcareous soil. Arch Agron Soil Sci 2016; 62(12): 1733-40. doi: 10.1080/03650340.2016.1170812.
- 23. Nizamani MR, Ansari MA, Siddiqui MA, Nizamani GS, Nizamani F, Naz M, et al. Effect of gibberellic acid on yield and yield attributes of canola (*Brassica napus* L.) varieties. Indian J Sci Res 2018; 6(8): 863-80.
- Lindsay WL, Norvell WA. Development of a DTPA soil test for zinc, iron, manganese, and copper. Soil Sci Soc Am J 1978; 42(3): 421-8. doi: 10.2136/sssaj1978.0361599500420 0030009x.
- 25. Yang Y, Ge Y, Zeng H, Zhou X, Peng L, Zeng Q. Phytoextraction of cadmium-contaminated soil and potential of regenerated tobacco biomass for recovery of cadmium. Sci Rep 2017; 7(1): 7210. doi: 10.1038/s41598-017-05834-8.
- Besalatpour AA, Hajabbasi MA, Khoshgoftarmanesh AH, Dorostkar V. Landfarming process effects on biochemical properties of petroleum-contaminated soils. Soil Sediment Contam 2011; 20(2): 234-48. doi: 10.1080/15320383.2011.546447.
- Baghaie AH, Keshavarzi M. The effect of montmorillonite nano-clay on the changes in petroleum hydrocarbon degradation and Cd concentration in plants grown in Cdpolluted soil. Avicenna J Environ Health Eng 2018; 5(2): 100-5. doi: 10.15171/ajehe.2018.13.
- Anning AK, Akoto R. Assisted phytoremediation of heavy metal contaminated soil from a mined site with *Typha latifolia* and *Chrysopogon zizanioides*. Ecotoxicol Environ Saf 2018; 148: 97-104. doi: 10.1016/j.ecoenv.2017.10.014.
- Chen H, Cutright T. EDTA and HEDTA effects on Cd, Cr, and Ni uptake by *Helianthus annuus*. Chemosphere 2001; 45(1): 21-8. doi: 10.1016/s0045-6535(01)00031-5.
- 30. Evangelou MW, Ebel M, Schaeffer A. Chelate assisted phytoextraction of heavy metals from soil. Effect, mechanism, toxicity, and fate of chelating agents. Chemosphere 2007; 68(6): 989-1003. doi: 10.1016/j. chemosphere.2007.01.062.
- 31. Kim S, Lim H, Lee I. Enhanced heavy metal phytoextraction by *Echinochloa crus-galli* using root exudates. J Biosci Bioeng 2010;109(1):47-50. doi:10.1016/j.jbiosc.2009.06.018.
- 32. Simón Solá MZ, Lovaisa N, Dávila Costa JS, Benimeli CS, Polti MA, Alvarez A. Multi-resistant plant growthpromoting actinobacteria and plant root exudates influence Cr (VI) and lindane dissipation. Chemosphere 2019; 222:

679-87. doi: 10.1016/j.chemosphere.2019.01.197.

- Azimzadeh Y, Shariatmadari H, Shirvani M. Remediation of some soil heavy metals by corn and canola in single and mixed culture system. Journal of Water and Soil 2013; 27(2): 406-14. doi: 10.22067/jsw.v0i0.24368. [In Persian].
- 34. Hadi F, Ali N, Ahmad A. Enhanced phytoremediation of cadmium-contaminated soil by *Parthenium hysterophorus* plant: effect of gibberellic acid (GA3) and synthetic chelator, alone and in combinations. Bioremediat J 2014; 18(1): 46-55. doi: 10.1080/10889868.2013.834871.
- 35. Askary Mehrabadi M, Amini F, Sabeti P. Evaluation of phytoremediation of petroleum hydrocarbon and heavy metals with using *Catharanthus roseus*. Iran J Plant Biol 2014; 6(3): 9-25. [In Persian].
- Naghipour D, Gharibi H, Taghavi K, Jaafari J. Influence of EDTA and NTA on heavy metal extraction from sandyloam contaminated soils. J Environ Chem Eng 2016; 4(3): 3512-8. doi: 10.1016/j.jece.2016.07.034.
- Franco IL, Contin M, Bragato G, De Nobili M. Microbiological resilience of soils contaminated with crude oil. Geoderma 2004; 121(1-2): 17-30. doi: 10.1016/j. geoderma.2003.10.002.
- 38. Cui J, Wang W, Peng Y, Zhou F, He D, Wang J, et al. Effects of simulated Cd deposition on soil Cd availability, microbial response, and crop Cd uptake in the passivationremediation process of Cd-contaminated purple soil. Sci Total Environ 2019; 683: 782-92. doi: 10.1016/j. scitotenv.2019.05.292.
- Mühlbachová G. Soil microbial activities and heavy metal mobility in long-term contaminated soils after addition of EDTA and EDDS. Ecol Eng 2011; 37(7): 1064-71. doi: 10.1016/j.ecoleng.2010.08.004.
- Chander K, Joergensen RG. Decomposition of Zn-rich *Arabidopsis halleri* Litter in low and high metal soil in the presence and absence of EDTA. Water Air Soil Pollut 2008; 188(1-4): 195-204. doi: 10.1007/s11270-007-9535-5.
- 41. Beneduzi A, Ambrosini A, Passaglia LM. Plant growthpromoting rhizobacteria (PGPR): their potential as antagonists and biocontrol agents. Genet Mol Biol 2012; 35(suppl 4): 1044-51. doi: 10.1590/s1415-47572012000600020.
- 42. Shafigh M, Ghasemi-Fasaei R, Ronaghi A. Influence of plant growth regulators and humic substance on the phytoremediation of nickel in a Ni-polluted soil. Journal of Water and Soil 2017; 31(1): 144-55. doi: 10.22067/jsw. v31i1.51798. [In Persian].
- Martin BC, George SJ, Price CA, Ryan MH, Tibbett M. The role of root exuded low molecular weight organic anions in facilitating petroleum hydrocarbon degradation: current knowledge and future directions. Sci Total Environ 2014; 472: 642-53. doi: 10.1016/j.scitotenv.2013.11.050.
- 44. Jin Y, Luan Y, Ning Y, Wang L. Effects and mechanisms of microbial remediation of heavy metals in soil: a critical review. Appl Sci 2018; 8(8): 1336. doi: 10.3390/app8081336.