



### Assessing role of lipid and polymer based delivery systems in inserts

Asiya Shaik\* and J Samreen Begum

Department of Pharmaceutics, Smt. Sarojini Ramulamma College of Pharmacy, Mahabubnagar, Telangana-509001, India.

#### ABSTRACT

Majority of gynaecological, proctologic diseases were treated through rectal or vaginal delivery of medications, which can bypass the first pass metabolism and can produce required local effect. Inserts are evolutionary pharmaceutical dosageform that are semi solid at room temperature but upon insertion in cavities of humans, melt and release the drug localised. The inserts are advantageous since they can be placed at scarce sites where blood perfusion is high that can enable for rapid absorption limiting many side effects. Ocular inserts represents the advanced technology in treating several ophthalmic diseases. Designing and development of optimum inserts is a challenge ever faced by Pharmaceutical researchers. In view of this, lipid and polymer based systems were additionally employed to improve the therapeutic efficiency of fore-said dosage forms. The review rationalises the importance of these inserts and their broad applications for multidisciplinary applications.

Keywords: Inserts; Suppositories; Ocular Inserts; Lipid; Polymers; Drug Delivery.

ISSN: Awaiting  
Research Article

#### Corresponding Author

Name: Asiya Shaik

Email: [asiyashaik99999@gmail.com](mailto:asiyashaik99999@gmail.com)

#### Article Info

Received on: 12-06-2020

Revised on: 29-09-2020

Accepted on: 06-10-2020

DOI: <https://doi.org/10.33974/ijrpps.v1i2.236>



Copyright© 2020, Asiya Shaik, et al. Assessing role of lipid and polymer based delivery systems in inserts, Production and hosting by Rubatosis Publications.

#### INTRODUCTION

Active pharmaceutical ingredients (API) can be administered through a variety of routes, the widely used one is the oral followed by parenteral route. [1,2] In spite of long history of use of medications in the rectal and vaginal suppositories/pessaries form, they are less commonly used and the global market is also limited due to patient incompliance.

On the other hand, suppositories have several benefits, especially in the treatment of gynaecological, proctological diseases.[3,4] API mix with various bases and this interaction can provide satisfactory pharmacokinetics with acceptable tolerance.

The eye as a portal for drug delivery is generally used for local therapy against systemic therapy to avoid

the risk of eye damage from high blood concentrations of the drug, which is not intended.[5] The exceptional anatomy and physiology of the eye make the organ impervious to the foreign substances, consequently posing the challenge to the formulator in overcoming of these protective layers without causing any permanent damage to the local tissues. Controlled release principles can be effectively applied in designing ocular inserts as an important approach in prolonging the drug residence time. Inserts are available in different shapes and sizes (Figure 1) [6]. In the present review, various methodologies for designing of insert such as suppositories/pessaries and ocular inserts were described in relation to several lipid/polymer based systems.

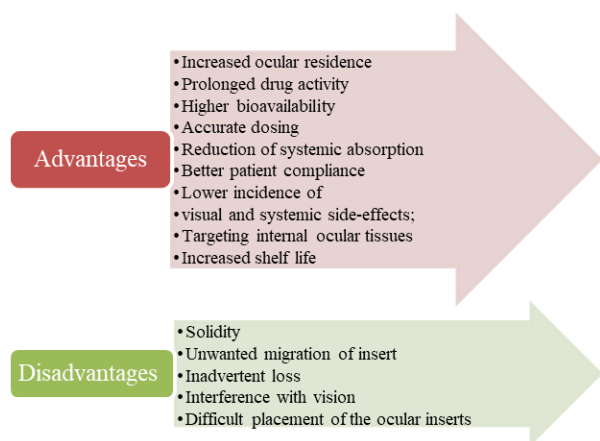


Figure 1: Various sizes and shapes of Suppositories

#### Types and designing of inserts

Suppositories are designed to deliver the medications through rectal and vaginal routes. Initially, they evolved as a more convenient form instead of liquid enema. Pessaries are frequently described as vaginal suppositories. These dosage forms were exhibited

several advantages- local effect on rectal mucosa; can be used effectively in pediatric, geriatric, unconscious patients; can also used for systemic benefit and avoids gastrointestinal irritation. Despite of all these, suppositories are few limitations such as patient acceptance, incomplete absorption and irritation to mucosa. Several local host factors may influence absorption in the rectum: the mucus layer, volume of rectal fluid, cell membrane, the tight junctions and the intracellular compartments are the local barriers for absorption. The suppository bases should have certain physico-chemical properties and significant effect on the uniformity and dosing of API [3]. According to United States Pharmacopeia, there are six main classes of bases like [7-10] Cocoa butter, Glycerin gelatin, Polyethylene glycol, Cocoa butter substitutes, Surfactant basis and tablet suppositories or inserts. In some other sources, classification was done on basis of melting dissolution. Ocular inserts are the solid or semisolid preparations designed for ophthalmic preparations and these were placed in lower fornix and less commonly in upper fornix and cornea. These are usually fabricated with polymeric vehicle containing drug. Several advantages and disadvantages of ocular inserts were given in Figure 2.



**Figure 2: Advantages and disadvantages of ocular inserts**

Polymers used for fabrication of ocular inserts; hydroxy propyl cellulose, poly vinyl pyrrolidone, hyaluronic acid, ethylene vinyl acetate, alginate chitosan, collagen, polypeptide, ethyl cellulose and cellulose acetate phthalate [11-16]. Ocular inserts are majorly classified into Insoluble ocular inserts, Soluble ocular inserts and Bioerodible ocular inserts.

#### Application of Lipid/polymer based systems

Many significant efforts have been tried for the potential application of lipid-based drug delivery systems (LBDS), to provide the required site specific controlled release of wide range of drugs and bioactive agents by improving their solubility. These systems includes several novel drug delivery systems like nanoparticles, microspheres, mucoadhesion [17-37], liposome and nanofibers etc [38-59]. Application of these systems and polymers to overcome the drawbacks

associated with inserts are studied. Several polymers like Chitosan, Alginate, PLGA, Polyvinylpyrrolidone, Zein ad Okra were used successfully. Additionally solid dispersion methodology is applied to enhance the solubility of Class II drugs [60-65]. Response surface methodology is applied to optimize the process parameters in the formulation of these systems [66-70]. Formulated LBDS are made into suppositories or insert to further enhance the residence time, thus finally improving the bioavailability.

#### Therapeutic areas

Inserts can be loaded with wide range of drugs like, Local Anesthetics, Protectants, Steroids, Astringents, Vasoconstrictors, Antiseptics, keratolytics, Anti-glaucoma, anti-bacterial and Anti-inflammatory etc.

#### CONCLUSION

Inserts were found to advantageous as it can eliminate several drawbacks associated with conventional drug delivery system. Application of lipid or polymer based systems, can further enhance the effectiveness of these systems in terms of residence time and bioavailability. However, polymer or lipid incompatibilities should be governed and careful preformulation studies need to be conducted. The safe and toxicological studies are strong advised for moving the product either to the preclinical or clinical trial phases. The importance of inserts lies in pediatric or geriatric patients indicating high compliance and minimal supervision since it is very safe to use.

#### REFERENCES

1. Rukhmakova O. A., Yarnykh T. G., Tykhonov O. I., Kotvitska A. A. (2018) Methodology for development of pediatric medicines for complex treatment of diseases of immune-dependent nature. *Journal of Global Pharma Technology*. 10 (12), 54-57.
2. Alhakamy, N. A., Ahmed, O. A. A., Kurakula, M., Caruso, G., Caraci, F., Asfour, H. Z., Alfarsi, A., Eid, B. G., Mohamed, A. I., Alruwaili, N. K., Abdulaal, W. H., Fahmy, U. A., Alhadrami, H. A., Eldakhakhny, B. M., & Abdel-Naim, A. B. (2020). Chitosan-based microparticles enhance ellagic acid's colon targeting and proapoptotic activity. *Pharmaceutics*, 12(7), 1-14. <https://doi.org/10.3390/pharmaceutics12070652>
3. Herasymova I. V., Tykhonov O. I., Shpychak O. S., Buryak M. V., Koval V. N. (2018) Pharmacological research of loravit suppositories. *Asian Journal of Pharmaceutics*. 12(1), 106-110
4. Alhakamy, N. A., Fahmy, U. A., Ahmed, O. A. A., Caruso, G., Caraci, F., Asfour, H. Z., Bakhrebah, M. A., Alomary, M. N., Abdulaal, W. H., Okbazghi, S. Z., Abdel-Naim, A. B., Eid, B. G., Aldawsari, H. M., Kurakula, M., & Mohamed, A. I. (2020). Chitosan coated microparticles enhance simvastatin colon targeting and pro-apoptotic activity. *Marine*

- Drugs*, 18(4), 226. <https://doi.org/10.3390/md18040226>.
5. Thakur RR et al,“ (2011) Modern Delivery Systems for Ocular Drug Formulations:A Comparative Overview W.R.T Conventional Dosage ”, International Journal of Research in Pharmaceutical and Biomedical Sciences, 2 (1):8-18.
  6. Hajare A., Mali S., Salunke S (2014). A rational approach to ocular drug delivery system: a overview. *WJPR*, 3(2):3324-3348.
  7. USP 35-NF 30. General Chapter Pharmaceutical Dosage Forms, The United States Pharmacopeial Convention, Rockville, MD, 2009.
  8. USP 31/NF 26. Inc. Chapter 1151, The United States Pharmacopeial Convention, Rockville, MD, 2008.
  9. USP 30/NF 25. Inc. NF monographs,The United States Pharmacopeial Convention, Rockville, MD, 2007.
  10. USP 29/NF 24. Inc. Front matter NF: Excipients, The United States Pharmacopeial Convention, Rockville, MD, 2007.
  11. Hasnain, M. S., Kiran, V., Kurakula, M., Rao, G. K., Tabish, M., & Nayak, A. K. (2020). Use of alginates for drug delivery in dentistry. In *Alginates in Drug Delivery* (pp. 387–404). Elsevier. <https://doi.org/10.1016/b978-0-12-817640-5.00015-7>
  12. Hasnain, M. S., Nayak, A. K., Kurakula, M., & Hoda, M. N. (2020). Alginate nanoparticles in drug delivery. In *Alginates in Drug Delivery* (pp. 129–152). Elsevier. <https://doi.org/10.1016/b978-0-12-817640-5.00006-6>
  13. Kurakula, M., Rao, G. K., Kiran, V., Hasnain, M. S., & Nayak, A. K. (2020). Alginate-based hydrogel systems for drug releasing in wound healing. In *Alginates in Drug Delivery* (pp. 323–358). Elsevier. <https://doi.org/10.1016/b978-0-12-817640-5.00013-3>
  14. Kurakula, M., & Rao, G. S. N. K. (2020). Type of Article: REVIEW Pharmaceutical Assessment of Polyvinylpyrrolidone (PVP): As Excipient from Conventional to Controlled Delivery Systems with a Spotlight on COVID-19 Inhibition. *Journal of Drug Delivery Science and Technology*, 102046.
  15. Naguib, G. H., Hassan, A. H., Al-Hazmi, F., Kurakula, M., Al-Dharrabh, A., Alkhalidi, H. M., Al-Ahdal, A. M., Hamed, M. T., & Pashley, D. H. (2017). Zein based magnesium oxide nanowires: Effect of anionic charge on size, release and stability. *Digest Journal of Nanomaterials and Biostructures*, 12(3), 741–749.
  16. Naguib, Ghada Hussein, Al-Hazmi, F. E., Kurakula, M., Abdulaziz Al-Dharrab, A., Mohamed Hosny, K., Mohammed Alkhalidi, H., Tharwat Hamed, M., Habiballah Hassan, A., Al-Mohammadi, A. M., Mohamed Alnowaiser, A., & Henry Pashley, D. (2018). Zein coated zinc oxide nanoparticles: Fabrication and antimicrobial evaluation as dental aid. *International Journal of Pharmacology*, 14(8), 1051–1059. <https://doi.org/10.3923/ijp.2018.1051.1059>
  17. Abdelhady, S., Honsy, K. M., & Kurakula, M. (2015). Electro Spun- Nanofibrous Mats: A Modern Wound Dressing Matrix with a Potential of Drug Delivery and Therapeutics. *Journal of Engineered Fibers and Fabrics*, 10(4), 155892501501000. <https://doi.org/10.1177/155892501501000411>
  18. Ahmed, O. A. A., Kurakula, M., Banjar, Z. M., Afouna, M. I., & Zidan, A. S. (2015). Quality by design coupled with near infrared in formulation of transdermal glimepiride liposomal films. *Journal of Pharmaceutical Sciences*, 104(6), 2062–2075. <https://doi.org/10.1002/jps.24448>
  19. Ahmed, S., Sarim Imam, S., Zafar, A., Ali, A., Aqil, M., & Gull, A. (2016). In vitro and preclinical assessment of factorial design based nanoethosomes transgel formulation of an opioid analgesic. *Artificial Cells, Nanomedicine and Biotechnology*, 44(8), 1793–1802. <https://doi.org/10.3109/21691401.2015.1102742>
  20. Andleeb, A., & Yar, M. (2020). Application of Electrospun Materials in Industrial Applications. *Electrospun Materials and Their Allied Applications*, 215–242. <https://doi.org/10.1002/9781119655039.ch8>
  21. Hosny, K. M., Aldawsari, H. M., Bahmdan, R. H., Sindi, A. M., Kurakula, M., Alrobaian, M. M., Aldryhim, A. Y., Alkhalidi, H. M., Bahmdan, H. H., Khallaf, R. A., & El Sisi, A. M. (2019). Preparation, Optimization, and Evaluation of Hyaluronic Acid-Based Hydrogel Loaded with Miconazole Self-Nanoemulsion for the Treatment of Oral Thrush. *AAPS PharmSciTech*, 20(7), 297. <https://doi.org/10.1208/s12249-019-1496-7>
  22. Kurakula, M., & A. Ahmed, T. (2015). Co-Delivery of Atorvastatin Nanocrystals in PLGA based in situ Gel for Anti-Hyperlipidemic Efficacy. *Current Drug Delivery*, 13(2), 211–220. <https://doi.org/10.2174/1567201813666151109102718>
  23. Kurakula, M., Ahmed, O. A. A., Fahmy, U. A., & Ahmed, T. A. (2016). Solid lipid nanoparticles for transdermal delivery of avanafil: optimization, formulation, in-vitro and ex-vivo studies. *Journal of Liposome Research*, 26(4), 288–296.

- <https://doi.org/10.3109/08982104.2015.1117490>
24. Kurakula, M., El-Helw, A. M., Sobahi, T. R., & Abdelaal, M. Y. (2015). Chitosan based atorvastatin nanocrystals: Effect of cationic charge on particle size, formulation stability, and in-vivo efficacy. *International Journal of Nanomedicine*, 10, 321–334. <https://doi.org/10.2147/IJN.S77731>
  25. Kurakula, M., & Koteswara Rao, G. S. N. (2020). Moving polyvinyl pyrrolidone electrospun nanofibers and bioprinted scaffolds toward multidisciplinary biomedical applications. *European Polymer Journal*, 136, 109919. <https://doi.org/10.1016/j.eurpolymj.2020.109919>
  26. Venkatesh, M., & Mallesh, K. (2013). Self-Nano Emulsifying Drug Delivery System (Snedds) for Oral Delivery of Atorvastatin- Formulation and Bioavailability Studies. *Journal of Drug Delivery and Therapeutics*, 3(3), 131–140. <https://doi.org/10.22270/jddt.v3i3.517>
  27. Naveen, N. R., Gopinath, C., & Kurakula, M. (2020). Okra-thioglycolic acid conjugate-synthesis, characterization, and evaluation as a mucoadhesive polymer. *Processes*, 8(3), 316. <https://doi.org/10.3390/pr8030316>
  28. Kurakula, M., Mohd, A. B., A, P. R., & Diwan, P. V. (2011a). Estimation of Prednisolone in Proliposomal formulation using RP HPLC method. *Int. J. Res. Pharm. Biomed. Sci.* 2011; 2: 663, 2(4), 1663–1669.
  29. Kurakula, M., Mohd, A. B., A, P. R., & Diwan, P. V. (2011b). Estimation of Prednisolone in Proliposomal formulation using RP HPLC method. *Int. J. Chem. Anal. Sci.* 2011; 2: 1193, 2(4), 1663–1669.
  30. Murali, V. P., Fujiwara, T., Gallop, C., Wang, Y., Wilson, J. A., Atwill, M. T., Kurakula, M., & Bumgardner, J. D. (2020). Modified electrospun chitosan membranes for controlled release of simvastatin. *International Journal of Pharmaceutics*, 584, 119438. <https://doi.org/10.1016/j.ijpharm.2020.119438>
  31. Kurakula, M., Sobahi, T. R., El-Helw, A., & Abdelaal, M. Y. (2014). Development and validation of a RP-HPLC method for assay of atorvastatin and its application in dissolution studies on thermosensitive hydrogel-based nanocrystals. *Tropical Journal of Pharmaceutical Research*, 13(10), 1681–1687. <https://doi.org/10.4314/tjpr.v13i10.16>
  32. Kurakula, M., Srinivas, C., Kasturi, N., & Diwan, P. V. (2012). Formulation and Evaluation of Prednisolone Proliposomal Gel for Effective Topical Pharmacotherapy. *International Journal of Pharmaceutical Sciences and Drug Research*, 4(1), 35. [www.ijpsdr.com](http://www.ijpsdr.com)
  33. NR Naveen, C Gopinath, DS Rao, (2018), A spotlight on thiolated natural polymers and their relevance in mucoadhesive drug delivery system, *Future J. Pharm. Sci.* 4 (1), 47-52
  34. NR Naveen, C Gopinath, DS Rao (2017), Isolation and assessment of natural mucoadhesive agent isolated from *Abelmoschus esculentus*, *Journal of Pharmacy Research* 11 (5), 438-443
  35. NR Naveen, TS Nagaraja, DR Bharathi, JNS Reddy (2013), Formulation Design and In Vitro Evaluation for Stomach Specific Drug Delivery System of Anti Retroviral drug–Acyclovir, *International Journal of Pharmacy and Life Sciences* 4 (3), 2506-2510.
  36. DRB P. Divya, N. Raghavendra Naveen, Snehalatha (2013), Optimization of cross linked tragacanth and comparison of drug Release rate profile with synthetic superdisintegrants on Metoclopramide orodispersible tablets, *International journal of pharmacy and life sciences* 4 (3), 2500-2505.
  37. NR Naveen, (2013) Design and characterization of sustained release matrix tablets of glimepiride by using synthetic and natural polymers, *International journal of drug discovery and herbal research* 3 (1), 573-578
  38. Mallesh, K., Pasula, N., & Kumar Ranjith, C. P. (2012). Piroxicam proliposomal gel: a novel approach for tropical delivery. *Journal of Pharmacy Research*, 5(3), 1755–1763.
  39. Patel, A., Hu, Y., Tiwari, J. K., & Velikov, K. P. (2010). Synthesis and characterisation of zein-curcumin colloidal particles. *Soft Matter*, 6(24), 6192.
  40. Podaralla, S., & Perumal, O. (2012). Influence of formulation factors on the preparation of zein nanoparticles. *AAPS PharmSciTech*, 13(3), 919–927.
  41. Regier, M. C., Taylor, J. D., Borczyk, T., Yang, Y., & Pannier, A. K. (2012). Fabrication and characterization of DNA-loaded zein nanospheres. *Journal of Nanobiotechnology*, 10, 44.
  42. Ren, W., Tian, G., Jian, S., Gu, Z., Zhou, L., Yan, L., et al. (2012). TWEEN coated NaYF<sub>4</sub>:Yb, Er/NaYF<sub>4</sub> core/shell upconversion nanoparticles for bioimaging and drug delivery. *RSC Advances*, 2(18), 7037.
  43. Saberi, A. H., Fang, Y., & McClements, D. J. (2013). Fabrication of vitamin E-enriched nanoemulsions: Factors affecting particle size using spontaneous emulsification. *Journal of Colloid and Interface Science*, 391, 95–102.
  44. Shukla, R., & Cheryan, M. (2001). Zein the industrial protein from corn. *Industrial Crops and Products*, 13, 171–192.

45. Tobitani, A., & Ross-Murphy, S. B. (1997). Heat-induced gelation of globular proteins. Model for the effects of time and temperature on the gelation time of BSA gels. *Macromolecules*, 30, 4845–4854.
46. Wang, Y., & Padua, G. W. (2010). Formation of zein microphases in ethanol–water. *Langmuir*, 26(15), 12897–12901.
47. Wang, Y., & Padua, G. W. (2012). Formation of zein spheres by evaporation-induced self assembly. *Colloid and Polymer Science*, 290(15), 1593–1598.
48. Wang, Q., Yin, L., & Padua, G. W. (2008). Effect of hydrophilic and lipophilic compounds on zein microstructures. *Food Biophysics*, 3(2), 174–181.
49. Wu, Y., Luo, Y., & Wang, Q. (2012). Antioxidant and antimicrobial properties of essential oils encapsulated in zein nanoparticles prepared by liquid–liquid dispersion method. *LWT — Food Science and Technology*, 48(2), 283–290.
50. Zhang, B., Luo, Y., & Wang, Q. (2011). Effect of acid and base treatments on structural, rheological, and antioxidant properties of  $\alpha$ -zein. *Food Chemistry*, 124(1), 210–220.
51. Zhang, Y., Niu, Y., Luo, Y., Ge, M., Yang, T., Yu, L. L., et al. (2014). Fabrication, characterization and antimicrobial activities of thymol-loaded zein nanoparticles stabilized by sodium caseinate–chitosan hydrochloride double layers. *Food Chemistry*, 142, 269–275.
52. Zhao, Y., Wang, Z., Zhang, W., & Jiang, X. (2010). Adsorbed Tween 80 is unique in its ability to improve the stability of gold nanoparticles in solutions of biomolecules. *Nanoscale*, 2(10), 2114–2119.
53. Zhong, Q., & Jin, M. (2009). Zein nanoparticles produced by liquid–liquid dispersion. *Food Hydrocolloids*, 23(8), 2380–2387.
54. Khan, Y., Durrani, S. K., Siddique, M., & Mehmood, M. (2011). Hydrothermal synthesis of alpha Fe<sub>2</sub>O<sub>3</sub> nanoparticles capped by Tween-80. *Materials Letters*, 65(14), 2224–2227.
55. Lai, L. F., & Guo, H. X. (2011). Preparation of new 5-fluorouracil-loaded zein nanoparticles for liver targeting. *International Journal of Pharmaceutics*, 404(1–2), 317–323.
56. Lee, S., Alwahab, N. S., & Moazzam, Z. M. (2013). Zein-based oral drug delivery system targeting activated macrophages. *International Journal of Pharmaceutics*, 454(1), 388–393.
57. Liang, H., Yang, Q., Deng, L., Lu, J., & Chen, J. (2011). Phospholipid-Tween 80 mixed micelles as an intravenous delivery carrier for paclitaxel. *Drug Development and Industrial Pharmacy*, 37(5), 597–605.
58. Malhotra, A., & Coupland, J. N. (2004). The effect of surfactants on the solubility, zeta potential, and viscosity of soy protein isolates. *Food Hydrocolloids*, 18(1), 101–108.
59. Matsushima, N., Danno, G. -i, Takezawa, H., & Izumi, Y. (1997). Three-dimensional structure of maize  $\alpha$ -zein proteins studied by small-angle X-ray scattering. *Biochimica et Biophysica Acta*, 1339, 14–22.
60. Vanitasagar, S., Srinivas, C., Subhashini, N. J. P., & Mallesh, K. (2012). Solid dispersion-a comparative study on the dissolution rate of aceclofenac. *International Journal of Pharmacy and Pharmaceutical Sciences*, 4(SUPPL.3), 274–278
61. Abdelhady, S., Honsy, K. M., & Kurakula, M. (2015). Electro Spun- Nanofibrous Mats: A Modern Wound Dressing Matrix with a Potential of Drug Delivery and Therapeutics. *Journal of Engineered Fibers and Fabrics*, 10(4), 155892501501000. <https://doi.org/10.1177/155892501501000411>
62. Raghavendra Naveen, N., Kurakula, M., & Gowthami, B. (2020). Process optimization by response surface methodology for preparation and evaluation of methotrexate loaded chitosan nanoparticles. *Materials Today: Proceedings*. <https://doi.org/10.1016/j.matpr.2020.01.491>
63. Kurakula, M., Naveen, N. R., & Yadav, K. S. (2020). Formulations for Polymer Coatings. *Polymer Coatings*, 415–443. <https://doi.org/10.1002/9781119655145.ch19>
64. Hu c., et al., (2015). Single and Multiple dose pharmacokinetics, Pharmacodynamic and Safety of the Novel Lipoprotein-Associated Phospholipase A2 Enzyme inhibitor Darapladip in Healthy chinese subjects: An Open Label Phase-1 Clinical Trial. *PLoS ONE* 10(10), 1–15.17.
65. Hu, Y., Yang, T., & Hu, X. (2012). Novel polysaccharides-based nanoparticle carriers prepared by polyelectrolyte complexation for protein drug delivery. *Polymer Bulletin*, 68, 1183–1199.
66. Muller, C., Rahmat, D., Sarti, F., Leithner, K., & Bernkop-Schnurch, A. (2012). Immobilization of 2-mercaptoethylamine on oxidized chitosan: a substantially mucoadhesive and permeation enhancing polymer. *Journal of Materials Chemistry*, 22(9), 3899–3908.
67. Palmberger, T. F., Albrecht, K., Loretz, B., & Bernkop-Schnurch, A. (2007). Thiolated polymers: evaluation of the influence of the amount of co-

- valently attached L-cysteine to poly(acrylic acid). *Eur J Pharm Biopharm*, 66(3), 405-412.
68. Kurakula, M., & Raghavendra Naveen, N. (2020). In situ gel loaded with chitosan-coated simvastatin nanoparticles: Promising delivery for effective anti-proliferative activity against tongue carcinoma. *Marine Drugs*, 18(4), 201. <https://doi.org/10.3390/md18040201>
69. Naveen, N.R., Gopinath, C., Rao, D.S. (2017) Design expert supported mathematical optimization of repaglinide gastroretentive floating tablets: In vitro and in vivo evaluation. *Future J. Pharm. Sci.* 3(2):140-147. (doi:10.1016/j.fjps.2017.05.003)
70. T Mallamma, DR Bharathi, RG Lakshmi, T Vyjayanthimala, J Nagasubbareddy, R Naveen (2014), Etoposide-loaded nanoparticles made from poly-e-caprolactone (PCL): formulation, characterization, in vitro drug release for controlled drug delivery system, *Int. J. Biopharm* 5, 5-12