# Selection of Suitable Diluents for Bacteriological Examination of Fishery Products

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For raw, iced and frozen samples of fish and prawn, significant difference was observed in total plate counts done with various diluents, the significance level ranging from 5% to 0.1%. For raw fish, N-saline, seawater and quarter strength Ringers' solution gave maximum total plate counts. In the case of iced-fish, n-saline yielded highest total plate counts. For frozen samples, however, peptone water and n-saline gave good recoveries. Trials with suitable ecombinations of diluents showed that though some of them were as good as the control, namely n-saline, none were superior in any way.

The primary objective in the quantitative assessment of microorganisms is to recover the surviving population and this in turn depends on the proper diluents used for obtaining manageable cell suspension. The suspending fluid is thought to exert some influence on microorganisms present in it. This is because a particular species or groups of organisms present in the food show varying degree of sensitivity to the inorganic ions due to hypo or hypertonic action. It has been shown that seawater possess a bactericidal effect on certain organisms such as E. coli (Carlucci & Pramer, 1959). On the other hand some marine bacteria show lytic tendencies when suspended in distilled water (Mac Leod, 1965)'

The effect of suspending fluid on the total viable population has been studied by many workers for a variety of materials. Butter-field (1932) studied the recovery of bacteria from riverwater using different diluents and observed better survival with phosphate buffer. Straka & Stokes (1957) claimed that the number of surviving bacteria after 20 min. in distilled water were reduced by 40–60% and in phosphate buffer 20–30% where as 0.1% peptone water permitted nearly 100% recovery even after 1 h. Sinnhuber & Lee (1964) while studying the microbioflora surviving radiation pasteurization of sea foods noted that 0.2% peptone water was superior to distilled water, 0.067 M phosphate buffer and 0.1% peptone water.

Later work by Lee & Harward (1970) claimed that Butterfield's phosphate buffer is superior to 0.2% peptone water for enumeration of microorganisms from frozen sea-foods and mixed vegetables. The present study was undertaken to determine the diluent effect on bacteria of fishery origin.

## Materials and Methods

Two types of fish, sardine (Sardinella longiceps) and mackerel (Rastrelliger kanagurta) and two species of prawns (Metapenaeus monoceros and Parapenaeopsis stylifera) were used in the study. Part of the sample was iced ( $0 \pm 1^{\circ}$ C) for 2 days and used as iced samples. Another part frozen and stored at -22°C was used as frozen samples.

Diluents: Diluents included in the study were

- 1. Distilled water
- 2. Seawater (full strength)
- 3. Normal saline (0.85 w/v Nacl)
- 4. Phosphate buffer (Butterfield, 1932 and ICMSF, 1978 Media No. 90)
- 5. Peptone water (0.1% peptone in dist. water)
- 6. Ringer's solution (Quarter strength, ICMSF (1978) media no. 97)

Additionally the following 3 combinations of these diluents were also tried.

## DILUENTS FOR BACTERIOLOGICAL EXAMINATION

- 1. Peptone saline (P.S.), (ICMSF, 1978 media no. 83)
- Peptone phosphate buffer (P.P.B.), (0.1% peptone in Butterfields phosphate buffer)

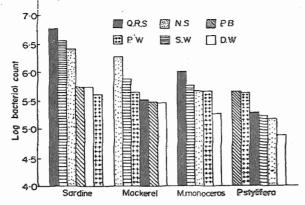


Fig. 1 Effect of single diluents of TPC of raw fish

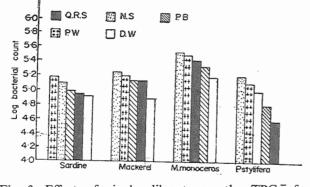


Fig. 3 Effect of single diluents on the TPC of frozen fish

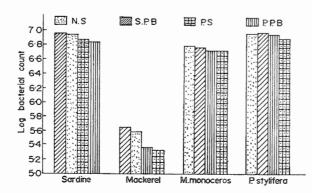


Fig. 5 Effect of combinations of diluents on the TPC of iced-fish

3. Phosphate buffered saline (P.B.S., (ICMSF, 1978 media no. 91).

Normal saline (N.S.) was used as control. Comparitive studies were made for raw, iced

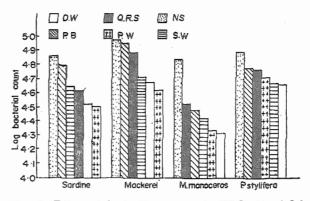
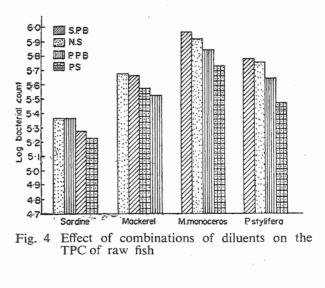


Fig. 2 Effect of single diluents on the TPC of iced-fish



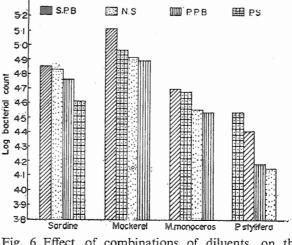


Fig. 6 Effect of combinations of diluents on the TPC of frozen fish

Raw fishIced fishFrozen fishSingleCombinationSingleCombinationDiluentSampleDiluentSampleDiluentSardineP<.001NS*P<.01NSP<.001NSP<.05NSMackerelP<.05NSP<.001NSP<.001NSP<.001NSM. monocerosP<.001NSP<.001NSP<.001NSP<.001NSM. styliferaP<.05NSP<.001NSP<.05NSP<.01NS*NS = Not significant difference (LSD) at 5% level and mean logarithmic bacterial count arranged in ascendingorder in different diluents for raw ized and frozen fish											
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DiluentSampleDiluentSampleDiluentSampleDiluentSampleDiluentSampleSardine $P < .001$ $NS^*$ $P < .01$ $NS$ $P < .01$ $NS$ $P < .001$ $NS$ $P < .05$ $NS$ $P < .05$ $NS$ Mackerel $P < .05$ $NS$ $P < .001$ $NS$ M. monoceros $P < .001$ $NS$ $P < .001$ $NS$ $P < .001$ $NS$ $P < .001$ $NS$ M. stylifera $P < .05$ $NS$ $P < .001$ $NS$ $P < .001$ $NS$ $P < .001$ $NS$ *NS = Not significant at 5%level. $Table 2$ .Least significant difference (LSD) at 5%level and mean logarithmic bacterial count arranged in ascending											
Sardine $P < .001$ $NS^*$ $P < .01$ $NS$ $P < .01$ $NS$ $P < .001$ $NS$ $P < .05$ $NS$ $P < .05$ $NS$ Mackerel $P < .05$ $NS$ $P < .001$ $NS$ M. monoceros $P < .001$ $NS$ M. stylifera $P < .05$ $NS$ $P < .05$ $NS$ $P < .001$ $NS$ $P < .01$ $NS$ $P < .01$ $NS$ *NS = Not significant at 5%level. $P < .01$ $NS$ $P < .001$ $NS$ $P < .05$ $NS$ Table 2. Least significant difference (LSD) at 5% level and mean logarithmic bacterial count arranged in ascending											
Mackerel M. monoceros $P < .05$ NS $P < .001$ $P < .001$ NS $P < .001$ $P < .001$ NS $P < .001$ $P > $											
M. monoceros       P<.001       NS       P<.05       NS       P<.001       P<.001       NS       P<.01       NS         M. stylifera       P<.05											
M. stylifera       P<.05       NS       P<.01       NS       P<.01       NS       P<.05       NS         *NS = Not significant at 5% level.       Table 2. Least significant difference (LSD) at 5% level and mean logarithmic bacterial count arranged in ascending       Table 2. Least significant difference (LSD) at 5%       Table 2. Least significant difference (LSD)       Table 2. Least significant difference											
*NS = Not significant at 5% level. <b>Table 2.</b> Least significant difference (LSD) at 5% level and mean logarithmic bacterial count arranged in ascending											
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order in different diluents, for raw iced and frozen fish											
Raw fish Iced fish Frozen fish											
Single Combined Single Combined Single Combined											
LSD at 5% level 0.4834 0.0734 0.1533 0.0245 0.0595 0.1522											
PW 5.6032 PS 5.2299 PW:4.5032 PPB:6.8542 QRS 4.9081 PS:4.6120											
PB 5.7089 PBS 5.2394 DW:4.5133 PS:6.8627 DW:4.9539 PPB:4.7608											
DW 5.7250 NS 5.3609 QRS:4.6197 PBS:6.9459 PB:4.9786 PBS:4.8343											
Sardine NS 6.4117 PPB 5.3613 SW:4.6437 NS:6.9401 NS:5.1777 NS:4.8545											
SW 6.5533 PB:4.7932											
QRS 6.7931 NS:4.8477											
LSD at 5% level 0.4154 0.0219 0.0822 0.1130 0.0624 0.0489											
PB 5.4606 PPB 4.5184 PW:4.6130 PS:5.3251 DW:4.8997 PPB:4.8909											
Mackerel DW 5.4719 PS 4.5258 DW:4.6735 PPB:5.3791 QRS:5.1110 PBS4.9124											
QRS 5.5006 PBS 5.6642 SW:4.7083 NS:5.5929 PB:5.1514											
PW 5.6263 QRS:4.8822 PS:4.9680											
SW 5.8530 NS 5.6739 PBS:5.6190 PW:5.2131 NS:5.0531											
NS 6.2472 PB:4.9471 NS:5.2619											
NS:4.9749											
LSD at 5% level 0.2314 0.0400 0.2927 0.0200 0.0821 0.0894											
DW:5.2331 PS:5.7210 DW:4.3117 PS:6.7236 DW:5.2105 PPB4.5328											
PW:5.6645 PPB:5.8289 PW:4.3375 PPB:6.7264 PB:5.3450 PBS:4.5435											
NS:5.6978 NS:5.9075 SW:4.4224 PBS:6.7707 QRS:5.4308 PS:4.6765											
<i>M. monoceros</i> SW:5.7428 PBS:5.9696 PB:4.4877 NS:6.7911 PW:5.4962											
QRS:5.9937 QRS:4.5378 NS:5.5491 NS:4.6994											
NS:4.8373											
LSD at 5% level 0.4156 0.0580 0.1026 0.0303 0.1384 0.2252											
DW:4.8717											
NS:5.1889 PS:5.4709 DW:4.6545 PS:6.8730 QRS:4.5853 PBS:4.1079											
SW:5.2022 PPB:5.6472 SW:4.6645 PPB:6.9359 PB:4.8018 PPB:4.1996											
SW:5.2022 NS:5.7573 PW:4.7070 NS:6.9524 DW:5.0040 NS:4.3988											
<i>P. stylifera</i> QRS:5.2811 PBS:5.7788 PB:4.7417 PBS:6.9532 PW:5.0970 PS:4.5252											
PW:5.6269 QRS:4.7610 NS:5.2056											
PB:5.6431 NS:4.8504											

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 Table 1. Summary of the level of significance of variance ratio of bacterial counts between diluents and between samples from the ANOVA

42 2 and frozen fish and depending on the bacterial flora expected in the products, some minor changes were made in the selection of diluents. For raw fish and iced ones which harbour marine flora, seawater was included in the study. For frozen fish which are expected to carry more terrestrial flora than marine, seawater was replaced by phosphate buffer.

Preparation of inoculum: To maintain uniformity of the sampling material 50 g of the muscle was dry-grinded into a paste aseptically and 10g lots of this material was transferred to 5 sterile blenders and homogenised with 90 ml of five diluents under study. Further decimal dilutions were prepared in 9 ml aliquots of the respective diluents.

The serial dilutions were then pour plated and incubated at room temperature  $(29 \pm 1^{\circ}C)$  for 48 h and bacterial counts estimated. Media and conditions used in this procedure were the same as that of Thampuran *et al* (1981).

### **Results and Discussion**

To make a comparitive study of the total plate counts (TPC) of raw, iced and frozen fish the data were analysed by analysis of variance after converting bacterial counts into log values. The level of significance of variance ratios 'between samples' and 'between diluents' of the raw iced and frozen fish and prawn are summerized in Table 1. In raw, iced and frozen, samples significant difference existed in bacterial count 'between the diluents' in single and in combination, the significance level ran-ging from 5 to 0.1%. 'Between samples', no significant difference was noted at 5% level. Two exceptions to this were mackerel (single diluent) and M. monoceros (Combination of diluents).

The least significant difference (LSD) at 5% level and the mean logarithmic counts in ascending order are presented in Table 2.

Figures 1 to 6 show the effect of diluent on bacterial counts estimated on raw iced and frozen samples of two types of fish and two species of prawns. The values used

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in the histograms represent the average of three trials.

For reaching the final conclusion, the overall average of the log values of the bacterial counts of the two types of fishes and two species of prawn were taken. This is given in Table 3. This shows that for

Table	3.	Average l	log	value c	of.	bacterial	count

Diluent	Raw fish		Iced	fish	Frozen fish	
	$S^*$	$\mathbb{C}^*$	S	$\mathbb{C}$	S	$\mathbb{C}$
D.W.	5.33		4.56		5.01	
S.W.	5.84		4.62		for an address of the	
N.S.	5.88		4.88		5.30	
P.W.	5.62		4.54		5.25	
P.B.	5.60		4.75		5.07	
Q.R.S.	5.89		4.70		5.01	
P.S.	-	5.24		6.45		4.69
P.B.S.		5.66		6.57		4.75
P.PB.		5.33		6.47		4.60
Control		5.67		6.57		4.60
(N.S.)						

S = Single diluentC = Combination of diluents

raw fish using single diluent, the highest recovery is obtained with Ringer's solution, followed by n-saline and seawater. Lowest count was obtained when distilled water was used. In the case of iced-fish, highest count was noted when n-saline was used. Frozen fish showed maximum count with n-saline and this was closely followed by peptone water and phosphate buffer.

The combinations of diluents were employed to see whether there was any synergistic effect in combining the ingredients of the single diluents. Table 3 indicates that for raw and iced fish, phosphate buffered saline came very near to the control n-saline in bacterial recovery. For frozen fish phosphate buffered saline showed a higher count than n-saline.

Thus it is clear that diluents exert a major role in the quantitative estimation of the bacteria. The variations in count obtained by using these diluents might be due to the difference in the background flora of the sample or due to sensitivity of species of the bacteria to these diluents. Hoadley & Cheng (1974) while studying the recovery of *Pseudomonas aeruginosa*, *Streptococcus faecalis* and *Escherichia coli* showed that tap water was highly toxic to all the strains. While recovery of *P. aeruginosa* was most successful when phosphate buffer was used, it had no effect on improving the count of *E. coli* and *S. faecalis*. Gray *et al* (1977) suggested that the diluents effect was ionic and not osmotic. Cations enhanced the protective property of the diluent and divalent cations were superior in this respect.

It can be concluded that in qualitative studies where sensitivity rather than reproducibility of the results is the deciding factor, the selection of a most suitable diluent is very essential. It should be noted that nsaline and phosphate buffer uniformly gave good results with raw, iced and frozen fish. Hence in routine works such as follow up studies in a processing or production line, a diluent such as saline or phosphate buffer can be used provided the composition of the diluent is definite and other parameters like media, incubation temperature and period are strictly followed.

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