

Nutritional conditions of jellyfish revealed by nucleic acid determinations- a case study on *Mnemiopsis leidyi*

Jamileh Javidpour\*, Catriona Clemmesen, Paul Rzepka

Marine Ecology, GEOMAR- Helmholtz Centre for Ocean Research Kiel, Germany

Email: [jjavid@geomar.de](mailto:jjavid@geomar.de)

Phone Nr.: +49431 6004408

Fax Nr.: +49431 6004402

Running Head: Gelatinous zooplankton nutrition

\*corresponding author

1 **Abstract**

2 Recent increases in reported jellyfish blooms along coastal areas have raised  
3 awareness on their ecological impacts on pelagic communities. Understanding their  
4 nutritional state that determines dietary statuses, energy adequacy and food  
5 inadequacy per capita in a population is mandatory to estimate their actual role and  
6 potential threat in marine ecosystems. In this study RNA and DNA measurements  
7 have been used to describe nutritional condition of the ctenophore *Mnemiopsis leidyi*  
8 under feeding and starvation conditions. A non-linear increase in DNA and RNA  
9 concentrations was observed in fed animals, whereas starved organisms represented a  
10 linear decrease in both nucleic acid concentrations. The observed DNA increase was  
11 not in accordance with the somatic growth and is likely attributed to sexual maturation  
12 stimulated under good nutritional condition. Although the RNA: DNA ratio showed a  
13 treatment effect, the same pattern of changes in fed and starved animals was observed.  
14 In this case study, correspondence plots and related analyses support the conclusion  
15 that nucleic acid determination on jellyfish has merit and is likely to provide  
16 complementary information on nutritional state of this group.

17 Key words:

18 gelatinous zooplankton, *Mnemiopsis leidyi*, RNA: DNA, nutritional condition

19

## 20 **Introduction**

21 Due to a global increase in abundance of gelatinous zooplankton, their future  
22 ecological role became a matter of concern for marine ecologists (Condon et al. 2012;  
23 Jackson 2008). One of the current challenges of marine ecologists is to determine the  
24 “in situ” eco-physiological and nutritional state of gelatinous zooplankton to gain a  
25 better understanding of physiological state and trophic interactions of this group.  
26 RNA:DNA ratios are one of the most used proxy in marine ecology that have been  
27 applied successfully as indicators of nutritional condition, growth and trophic  
28 interactions in fish and marine invertebrate (Chicharo and Chicharo 2008 and  
29 references there in, Koop et al. 2011). Principally RNA content provides information  
30 about the metabolic activity of the organism and varies with age, size, life stage or  
31 abiotic conditions. The amount of DNA is constant under changing environmental  
32 situations in all somatic tissue cells and reflects the cell number (Chicharo and  
33 Chicharo 2008; Dortch et al. 1983). Thus, a higher RNA: A DNA ratio reflects a better  
34 nutritional condition of an organism (Clemmesen 1994).

35 Impact of gelatinous plankton and magnitude of their predatory potential became  
36 highly conspicuous when the comb jelly *Mnemiopsis leidyi*, was introduced into the  
37 Black Sea, presumably via ballast water tanks, leading to a massive development of  
38 this species (Shiganova et al. 2001). *M. leidyi* is known as a generalist predator due to  
39 its high clearance rate on different types of food, high reproductive potential (self-  
40 fertilizing hermaphrodite, several 100 eggs/ ind per day under optimal conditions),  
41 wide tolerance limits for salinity (2-38) and temperature (0-32°C) and also high  
42 tolerance for low oxygen concentration (Costello et al. 2012; Purcell et al. 2001).

43 To our knowledge there is still no proof if nucleic acid measurements would be  
44 applicable to gelatinous carnivores especially to *M. leidyi*. The purpose of this first

45 study was to determine whether RNA and DNA comprise a measurable portion of the  
46 cellular mass in *M. leidy* and how their concentrations vary as a result of starvation  
47 and feeding traits.

#### 48 **Method**

49 Individuals of *M. leidy* were collected with “RV Polarfuchs” in the inner Kiel fjord  
50 (54°19,6' N; 10° 9,2' E) by means of a plankton net (HydroBios WP3 , 1000 µm mesh  
51 size and 80 cm diameter). The net was towed in a vertical profile over the whole water  
52 column from 18 m depth resulting in 120 healthy *M. leidy* of the same size ( $1 \pm 0.2$   
53 cm) to be selected for the experiment. Our experimental design was composed of one  
54 factor “nutritional condition” with two levels of (i) starvation and (ii) feeding. Due to  
55 the handling problems and space limitation, 60 individuals per each treatment were  
56 kept together in a 50 liter bucket that was filled with filtered sea water (5µm). *M.*  
57 *leidy* in the “feeding” treatment received daily copepods (copepodites and adults of  
58 *Acartia tonsa*), taken from our continuous standard culture, at a final concentration of  
59 2 mg Carbon per *M. leidy* per day over the experimental period. *M. leidy* in the  
60 “starvation” treatment did not receive any food. Salinity and temperature were kept  
61 similar to ambient conditions in the Kiel Fjord (salinity 17; temperature 12°C). Five  
62 individuals of *M. leidy* from each bucket were sampled for DNA and RNA  
63 measurements every 2 days over 14 days. To avoid possible bias by gut content,  
64 sampled individuals were kept in filtered seawater for 1-2 hours to allow for gut  
65 evacuation. After measurements of total length, sampled individuals were stored at -80  
66 °C and freeze dried prior to the nucleic acid analysis. Dry weight  $\pm 0.001 \mu\text{g}$  was  
67 quantified. DNA and RNA content of all individuals was determined by fluorescence  
68 technique described by Clemmesen 1993; Clemmesen 1988 and modified by Malzahn  
69 et al. 2003 and Belchier et al. 2004 eliminating some purification steps.

70 Statistical analysis- Our initial data exploration was carried out following the protocol  
71 described in (Zuur 2007; Zuur 2010) on response variables RNA, DNA and the ratio  
72 (RNA: DNA) and explanatory variables namely length, dry weight, time (date) and  
73 condition (treatment). Due to the experimental design one would expect a dependency  
74 structure as the same bucket is repeatedly sampled over time, potentially causing a  
75 'site' effect. The next step was thus to check for the “bucket” effect. The follow up  
76 analysis (Pearson correlations), showed that there is no inherent correlation between  
77 observations from different days ruling out a “site” effect. Pearson correlation  
78 coefficients analysis however showed a co-linearity between length and dry weight.  
79 This is expected as both variables indicate changes in growth. Additionally a negative  
80 relationship between dry weight and date was detected. Therefore we removed length  
81 and dry weight from the model and only considered date and treatment for the final  
82 analysis using the following three models:

83  $RNA \sim (Date + Treatment)$

84  $DNA \sim (Date + Treatment)$

85  $RATIO \sim (Date + Treatment)$

86 We have applied a generalized additive model (GAM) on DNA due to non-linear  
87 relationship between response and explanatory variables. Since we detected  
88 heterogeneity in the residuals obtained from the GAM model on RNA and the Ratio;  
89 we applied a Generalized Additive Mixed Model (GAMMs, Woods 2006) on these  
90 data sets. All statistical analyses were performed in the software R (Development Core  
91 Team, 2011). Our model validation did not show any heterogeneity patterns, serious  
92 deviation from normality, violation of independence (assessed by ACF plot) and  
93 finally no indication of autocorrelation in any of the models performed.

94 **Results**

95 Starved *M. leidyi* reduced their dry weight significantly by approximately 80% of the  
96 mean starting value over 14 days (from 3 to 0.5 mg ind<sup>-1</sup> in average,  $F_{(1, 8)} = 54.1$ ,  
97  $P < 0.01$ ), whereas the body length decreased by approx. 50% from 13.6 to 6.8 mm ( $F_{(1, 8)} = 24.3$   $P < 0.01$ ). In fed organisms however, neither the dry weight nor the body  
98 length showed a significant change over time ( $F_{(1, 8)} = 2.7$ ,  $P > 0.5$  and  $F_{(1, 8)} = 0.01$ ,  $P$   
99  $> 0.5$  respectively).

101 *DNA*- Our model output indicates a significant treatment effect on DNA  
102 concentrations ( $R = 0.6$ ,  $T = -7.296$ ,  $P < 0.001$ ). As can be seen from the Fig. 1 starved  
103 animals have a significantly lower amount of DNA compared to fed ones ( $F_{3,768} =$   
104  $15.996$ ,  $P < 0.001$ ). It is apparent from the Fig. 1 that a non-linear relationship between  
105 DNA and “date” in fed treatment exists. Under feeding conditions, DNA concentration  
106 increased up to day 10 when it reached its highest values of  $4.9 \pm 0.8 \mu\text{g ind}^{-1}$   
107 (mean $\pm$ SD). Thereafter it decreased over the last two sampling dates. In contrast, the  
108 smoother representing the relationship for the starved condition showed a negative and  
109 significant decrease of DNA over the experimental period ( $F_{1,0} = 4.188$ ,  $P = 0.04$ ).

110 *RNA*- Similar to the DNA pattern we observed a significant treatment effect on RNA  
111 concentrations ( $R = 0.81$ ,  $T = -9.153$ ,  $P < 0.001$ ). Fig. 2 presents the results obtained  
112 from GAMM model. RNA concentration in fed animals showed a significant non-  
113 linear changes over the experimental period ( $F_{2,136} = 2.947$ ,  $P = 0.05$ ) where RNA  
114 increased up to day 12 reaching highest value of  $22.6 \pm 12.4 \mu\text{g ind}^{-1}$  (mean $\pm$ SD) and  
115 then decreased. In starved animals however, RNA changed linearly over time with a  
116 significant decrease ( $F_{1,222} = 50.964$ ,  $P < 0.001$ ). Notice that the variation in the RNA  
117 concentrations in the feeding group is larger compared to the RNA concentrations  
118 found in starved animals ( $\sigma^2_1 = 45.61$  for fed animals;  $\sigma^2_2 = 11.86$  for starved animals).

119 *RNA:DNA*- The GAMM model revealed no significant interaction between  
120 “treatment” and “date”; therefore one smoother was capable in capturing the Ratio-  
121 date relationship. There was a significant effect of “treatment” on the Ratio ( $R = 0.75$ ,  
122  $T = -5.397$ ,  $P < 0.001$ ) where starving animals represented lower concentration than the  
123 fed animals. The effect of “date” alone was significantly different in both treatments  
124 ( $F_{2,832} = 15.72$ ,  $P < 0.001$ ) due to a decrease in the Ratio starting at day 8.

## 125 **Discussion**

126 The significant differences between DNA and RNA concentrations in starved and fed  
127 animals provides compelling evidence that this method can be applied to gelatinous  
128 zooplankton and specifically on *M. leidyi* as a measure of the nutritional condition.  
129 The most striking result emerging from our data is an increase of both DNA and RNA  
130 concentrations over the time of the experiment in fed animals, without any significant  
131 increase in dry weight or length. Given that every somatic cell contains the same  
132 amount of DNA, DNA concentrations can reflect cell numbers and the amount of  
133 RNA (relative to DNA) is a proxy for metabolic activity and consequently the  
134 nutritional condition of the individual (Dortch et al. 1983, Clemmesen 1994).  
135 Therefore an increase in DNA concentration often is reflected in an increase in  
136 somatic growth (Buckley et al. 1999, Chicharo & Chicharo 2008). Here, we observed  
137 a different and non-linear response where fed *M. leidyi* showed cell replication without  
138 increase in somatic growth. This resulted likely from the production of genital cells  
139 between days 6 to 10 under good nutritional conditions. Fed animals were performing  
140 reproductive growth in combination with an increased RNA content i.e. increased  
141 metabolic activity of their somatic cells, without an increase in somatic growth. This  
142 hypothesis is supported by previous experiments indicating that sufficient food supply  
143 (200 copepods  $l^{-1}$ ) can initiate egg production in ctenophores even in larval phases

144 (Jaspers et al. 2012; Martindale 1987; Reeve et al. 1989). Similar results have been  
145 observed for Washington clam where increase in DNA content was correlated with  
146 sexual maturation in male and female clams (Kim et al. 2005).

147 Under starved conditions, we observed a significant reduction in both dry weight and  
148 body length, which was fairly well reflected in DNA and RNA concentrations. *M.*  
149 *leidyi* appears to compensate a nutritional depletion by somatic degeneration. Here we  
150 observed a decrease of metabolic activities that starts at 4 days after starvation  
151 continues to decrease similar to results observed in larval fish indicative by a sharp  
152 reduction of RNA (Clemmesen 1994). This is in line with previous findings that *M.*  
153 *leidyi* is able to survive under starvation situations by reducing its body size (Anninsky  
154 et al. 2005) and might explain the ability of overwintering population.

155 Of all studied nucleic acid derived indices to determine the nutritional condition or the  
156 physiological state of many groups of animals, the RNA:DNA ratio has been the one  
157 mainly used. Although the RNA: DNA ratio in this study was significantly higher in  
158 the feeding group, the same pattern of changes in the ratio over time in both starved  
159 and fed organisms was observed. Some studies have shown better results when only  
160 RNA derived indices were used. This was specially the case when juvenile fish and  
161 invertebrates at a further developmental stage were analyzed and RNA concentrations  
162 and RNA: protein ratios were shown to be a better predictor of growth rate, than the  
163 RNA: DNA ratio (Foster et al. 1992, Rooker and Holt 1996, Houlihan et al. 1990).

164 One explanation for the lack of response in the RNA:DNA ratio is the change in cell  
165 size or number due to the different forms of tissue being formed with development and  
166 the different reaction patterns of the RNA:DNA ratio in dependence of developmental  
167 stage (Buckley et al. 1999) and the tissue type (Olivar et al. 2009). For *M. leidyi* this  
168 could be caused by the formation of gametes occurring already in an early stage of



169 development. Future studies should relate RNA to dry weight or protein content to  
170 evaluate if this is a better indicator, especially at times when gamete production  
171 occurs.

## 172 **Comments and recommendations**

173 Our finding is conclusive proof that nucleic acid derived indicators are able to describe  
174 the nutritional condition of gelatinous zooplankton and specifically *M. leidyi* and  
175 therefore can help understanding the physiological state which is needed for a better  
176 understanding of trophic interactions. To be able to apply this method directly to field  
177 collected organisms further laboratory experiments are needed to (i) calibrate these  
178 indicators in response to egg production and somatic growth rates (ii) characterize the  
179 reaction of *M. leidyi* to starvation on a detailed histological level and (iii) define  
180 thresholds and critical values for survival, like recently have been shown for larval  
181 fish (Meyer et al. 2012).

182

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### Figures and figure legends

Figure 1- Estimated smoothing curves generated for DNA concentration of *M. leidyi* under and (a) feeding and (b) starvation conditions. Fitted values obtained by GAM model. Dotted lines are 95% point-wise confidence bands.

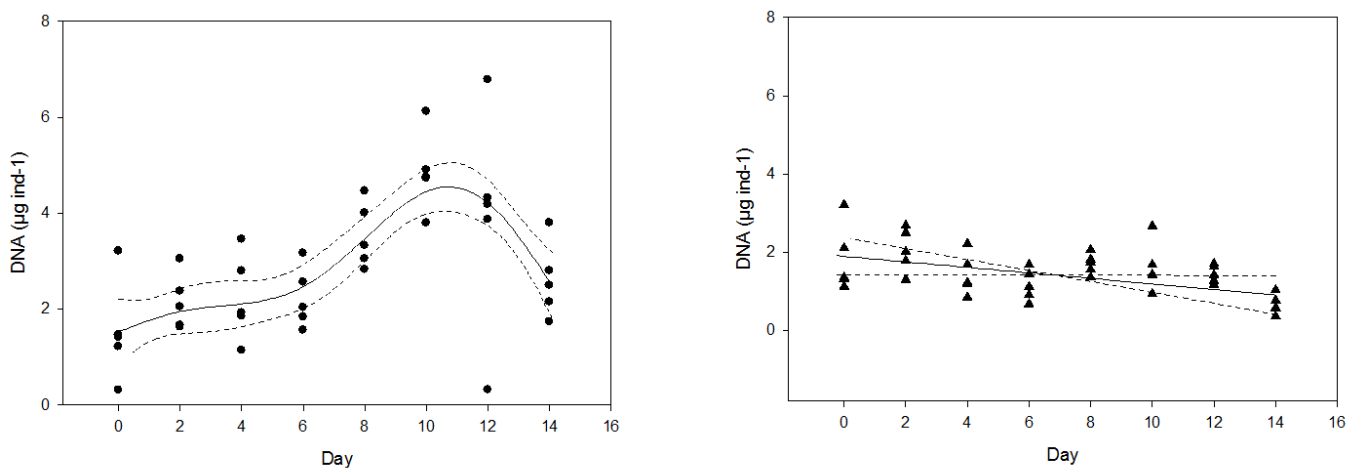


Figure 2- Estimated smoothing curves generated for RNA concentration of *M. leidyi* under and (a) feeding and (b) starvation conditions. Fitted values obtained by GAMM model. Dotted lines are 95% point-wise confidence bands.

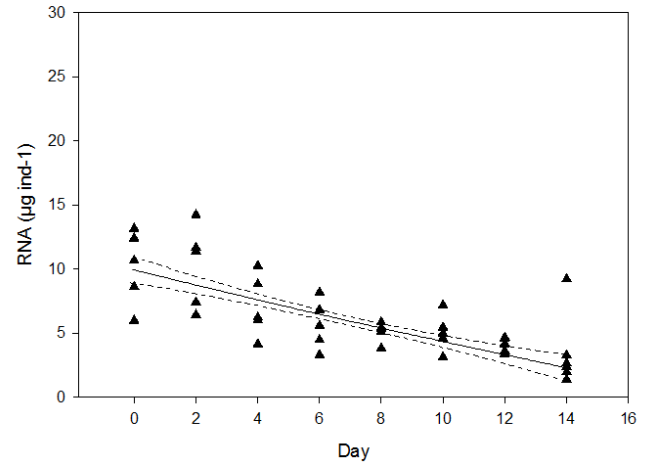
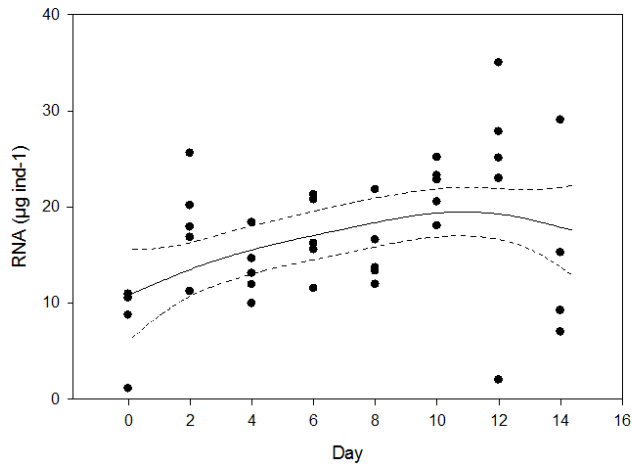


Figure 3- Estimated smoothing curves generated for RNA: DNA ratio of *M. leidyi* under and (a) feeding and (b) starvation conditions. Fitted values obtained by GAMM model. Dotted lines are 95% point-wise confidence bands.

