

Complete nucleotide sequence of CHU: A Luz24likevirus infecting Pseudomonas aeruginosa and displaying a unique host range

Magill, D., Shaburova, O., Krylov, V., & Kulakov, L. (2015). Complete nucleotide sequence of CHU: A Luz24likevirus infecting Pseudomonas aeruginosa and displaying a unique host range. FEMS Microbiology Letters, 362(9), [fnv045]. DOI: 10.1093/femsle/fnv045

Published in:

FEMS Microbiology Letters

Document Version:

Peer reviewed version

Queen's University Belfast - Research Portal: Link to publication record in Queen's University Belfast Research Portal

Publisher rights © FEMS 2015.

This is a pre-copyedited, author-produced PDF of an article accepted for publication in FEMS Microbiology Letters following peer review. The version of record "Complete nucleotide sequence of phiCHU: A Luz24likevirus infecting Pseudomonas aeruginosa and displaying a unique host range, Damian J. Magill, Olga V. Shaburova, Elena N. Chesnokova, Elena A. Pleteneva, Victor N. Krylov, Leonid A. Kulakov is available online at: http://dx.doi.org/10.1093/femsle/fnv045

General rights

Copyright for the publications made accessible via the Queen's University Belfast Research Portal is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

The Research Portal is Queen's institutional repository that provides access to Queen's research output. Every effort has been made to ensure that content in the Research Portal does not infringe any person's rights, or applicable UK laws. If you discover content in the Research Portal that you believe breaches copyright or violates any law, please contact openaccess@qub.ac.uk.

1	Complete nucleotide sequence of phiCHU: A Luz24likevirus infecting Pseudomonas aeruginosa and
2	displaying a unique host range
3	Damian J Magill ¹ , Olga V Shaburova ² , Elena N Chesnokova ² , Elena A Pleteneva ² , Victor N Krylov ² , Leonid A Kulakov ¹
4	
5	¹ Queen's University Belfast, School of Biological Sciences, Medical Biology Centre, 97 Lisburn Road, Belfast BT9 7BL, Northern
6	Ireland
7	² Department of Microbiology, Laboratory for Genetics of Bacteriophages, I.I. Mechnikov Research Institute for Vaccines and Sera,
8	Moscow, Russia
9	
10	Keywords: bacteriophage; antibiotic resistance; phage therapy
11	
12	Correspondence: Leonid A Kulakov, Queen's University Belfast, School of Biological Sciences, Medical Biology Centre, 97 Lisburn
13	Road, Belfast BT9 7BL, Northern Ireland, Tel: +44 (0)28 9097 2799, Fax: +44 (0)28 9097 5877, Email: I.kulakov@qub.ac.uk
14	
15	

17	
17	
18	
19	
20	
21	
22	Abstract
23	A complete nucleotide sequence of the new Pseudomonas aeruginosa Luz24likevirus phiCHU was obtained. This virus was shown
24	to have a unique host range whereby it grew poorly on the standard laboratory strain PAO1, but infected 26 of 46 clinical isolates
25	screened, and strains harboring IncP2 plasmid pMG53. It was demonstrated that phiCHU has single strand interruptions in its
26	genome. Analysis of the phiCHU genome also suggested that recombination event(s) participated in the evolution of the leftmost
27	portion of the genome, presumably encoding early genes.
28	
29	
30	
31	

- *Pseudomonas aeruginosa* strains are opportunistic pathogens and a significant source of morbidity and mortality (e.g. in cystic
- 45 fibrosis patients). In the wake of growing antibiotic resistance, a resurgence of interest in bacteriophage therapy has taken place to
- 46 relieve the growing burden observed in healthcare systems (Burns *et al.* 2012; Weinstein *et al.* 2001).
- 47 P. aeruginosa bacteriophages (phages) are an extremely heterogeneous assemblage and in this work a novel phage of the
- 48 Luz24likevirus genus, phiCHU, was characterized. PhiCHU was isolated from a small pond in the Moscow locality and shown to

have a unique host range; it grew poorly on the standard P. aeruginosa laboratory strain PAO1, but infected efficiently a number of 49 clinical isolates resistant to other phages and lysed mucoid strains isolated from wound infections revealing itself to be a potential 50 therapeutic agent. How it can circumvent mucoidy is currently unknown. Table S1. shows a comparative analysis of phiCHU's host 51 range against known lytic phages phiKZ, EL, Lin68, PB1, 14/1, phiKF77, and phiKMV (Mesyanzhinov et al. 2002, Hertveldt et al. 52 2005, Krylov et al. 2007, Ceyssens et al. 2009, Kulakov et al. 1986, 1991, Lavigne et al. 2003). Of the 46 isolates tested phiCHU 53 grew well on 26, but exhibited turbid growth and varying plaque morphologies on a number of other strains. Our inability to isolate 54 lysogens from any strains led us to conclude that phiCHU behaves like a virulent phage, at least in these instances. The 55 Luz24likevirus genus has previously been reported as containing lytic phages apart from PaP3 (Tan et al. 2007). PhiCHU's 56 57 virulence and broad host range means that it is most likely suitable for use within therapeutic preparations. Importantly, phiCHU 58 demonstrated good growth on PAO38; a P. aeruginosa strain containing the IncP2 plasmid pMG53. Figure S1. demonstrates the 59 growth of phiCHU along with 8 other phages on *P. aeruginosa* strains. IncP2 plasmids confer a broad spectrum of traits to pseudomonads including, but not limited to, multiple forms of antibiotic resistance and metabolism of unusual carbon sources 60 (Jacoby et al. 1983). IncP2 being the most abundant plasmids found in nosocomial strains of P. aeruginosa (Hanson & Olsen, 61 1978) were also shown to confer resistance to many phages of *P. aeruginosa* through interference in their intracellular 62

63 development. This growth inhibition of different phages has been reported to be under the control of different loci within this plasmid

group (Freizon *et al.* 1989). We subsequently investigated whether pMG53 promotes efficient phiCHU growth, by conjugatively
 transferring this plasmid to PAO1. Upon carrying this out, PAO1 acquired sensitivity to phiCHU.

66

The need for phage genomics is paramount from a clinical perspective. Many phages can transfer bacterial genes by transduction, which poses a significant problem with respect to the potential dissemination of pathogenicity and resistance factors. Apt examples here would be the *P. aeruginosa* phages E79 and phiKZ (Morgan 1979, Dzhusupova 1982). Genomic analysis can help to elucidate whether these processes are likely to take place and therefore, whether a given phage is suitable for clinical use. Bacteriophage phiCHU particles were purified using isopycnic CsCl density gradient centrifugation and genomic DNA was extracted as described by Sambrook and Russell (2001). The phage was sequenced by the dideoxy method and both shotgun and primer walking on the whole genomic DNA was employed.

74

The genomic map of phiCHU is presented in Fig 1. phiCHU was shown to have a linear dsDNA genome of 45,626 bp with a GC content of 52.02%. 73 ORFs were predicted and annotated in Artemis (Rutherford *et al.* 2000) and functionality was assigned to 22 of these. Five ORFs encode proteins associated with genome replication, recombination and repair, 9 encode various structural proteins, 3 encode portal and terminase subunits, 2 encode the lysis machinery and the other 2 encode a putative gamma-glutamyl cyclotransferase and L-Glutamine-D-Fructose-6-Phosphate amidotransferase. All predicted genes lie in two bidirectionally transcribed units separated by a double intrinsic terminator; a structure indicative of the *Luz24likevirus* genus. Three tRNA genes (tRNA^{Asn}, tRNA^{Asp}, and tRNA^{Pro}) were predicted which are clustered at the extreme right of the genome. The genome is delineated
by 185 bp direct terminal repeats. This genomic organization and a high nucleotide homology of 94.79% to the *Luz24likevirus*vB_PaeP_C1-C14_Or (Its closest relative) (Accession: HE983844) demonstrates that phiCHU unequivocally belongs to the *Luz24likevirus* genus of the family *Podoviridae*.

85

Alignments of phiCHU and closely related *Luz24likeviruses* infecting *P. aeruginosa* with the Progressive Mauve algorithm (Darling *et al.* 2010) highlighted the presence of a gap of approximately 1.5 kb encompassing gp1 – gp4 (Fig. S2). This region shows a greater homology (94%) to different phages of the group (Luz24 and TL) (Ceyssens *et al.* 2008, Accession: NC_023583), which also exclusively infect *P. aeruginosa*. This finding suggests that the phiCHU genome may have evolved as a result of recombination between ancestors of vB_PaeP_C1-C14_Or and Luz24.

91

Knowledge of the nature of receptors utilized in phage adsorption is important for therapeutic applications. Investigations into tail
fibre proteins can yield useful information with respect to variations in host range. A number of authors have implicated the
presence of glycine rich regions in the identification of tail fibre proteins (Lucchini *et al.* 1999; Nilsson *et al.* 2000; Tetart *et al.* 1998).
20 of the 50 C-terminal residues of ORF 58 are glycines, thereby making this a likely candidate as one constituent of the tail fibre
complex. Variations in this ORF were investigated amongst *Luz24likeviruses* and it was found that phiCHU differed by only a single

97 residue from phiMR299-2 (Alemayehu *et al.* 2012). At this point (residue 267) phiMR299-2 like other *Luz24likeviruses*, possess the 98 isoteric residues serine or threonine (Fig S3). phiCHU however, possesses proline at this point. Molecular modelling and alignment 99 of phiCHU and phiMR299-2 putative tail fibres (Fig S4) demonstrated the extent to which this substitution alters protein structure 100 and therefore, this may represent one mechanism contributing to altered host specificity of phiCHU.

101

102 It was previously demonstrated that a P. putida phage tf, which is distantly related to Luz24likeviruses has localized single-strand

103 interruptions (nicks) in its genome (Glukhov *et al.* 2012). This genomic feature was not previously demonstrated for

104 Luz24likeviruses infecting P. aeruginosa. Here it was shown that denatured phiCHU DNA produces multiple bands on agarose gels,

105 which disappeared upon ligation (Fig. S5a). The use of total genomic DNA as a template in sequencing reactions permitted the

106 localization of nicks (Fig. 1 and Fig. S5b) and allowed the identification of the consensus associated with this feature (5' -

107 TACT/RTGMC – 3'). This proved to be the same consensus as previously reported in tf (Glukhov et al. 2012). 14 such sites were

108 reported for tf and 15 for the P. fluorescens phage UFV-P2 (Eller et al. 2014, Glukhov et al. 2012). phiCHU possesses 7 instances

109 of this consensus. The purpose of this enigmatic feature in phage genomes remains unknown.

110

111 In this work, a novel member of the Luz24likevirus genus of phages infecting P. aeruginosa was isolated and characterized.

112 phiCHU was shown to possess localized single-strand interruptions in its genome. It was also found that this phage exhibited a

unique host range, whereby it grew well on PAO1 only in the presence of a plasmid from the IncP2 group. In addition, we found
some evidence suggesting that recombination within disparate members of the genus contributed to the evolution of phiCHU.
Genbank Accession Number: KP233880
Acknowledgements
This research was funded by the School of Biological Sciences, Queens University Belfast. We are extremely grateful to Dr Anna
Kulakova for technical assistance.
Supporting Information
Additional supporting information can be found in the online version of this article:
Table S1. Sensitivity of clinical strains to phages.
Fig S1. Comparison of the sensitivity of <i>P. aeruginosa</i> strains to phages.
Fig S2. Progressive Mauve alignments of phiCHU with closely related Luz24likeviruses.
Fig S3. Sequence alignment of Luz24likevirus putative tail fibre proteins.

- 130 Fig S5a. Visualization of localized single-strand interruptions in phiCHU DNA.
- 131 **Figure S5b.** Localization of single-strand interruptions through sequencing on whole genomic DNA.
- 132 References
- 133 1. Alemayehu, D., Casey, P.G., McAuliffe, O., Guinane, C.M., Martin, J.G., Shanahan, F., Coffey, A., Ross, R.P. and Hill, C.,

134 (2012). Bacteriophages
MR299-2 and
NH-4 can eliminate Pseudomonas aeruginosa in the murine lung and on cystic

- 135 fibrosis lung airway cells. *MBio*, **3**(2), pp. e00029-12.
- 136
- 137 2. Burns, J.L., Gibson, R.L., McNamara, S., Yim, D., Emerson, J., Rosenfeld, M., Hiatt, P., McCoy, K., Castile, R., Smith, A.L.
- and Ramsey, B.W., (2001). Longitudinal assessment of *Pseudomonas aeruginosa* in young children with cystic fibrosis. *The Journal of infectious diseases*, **183**(3), pp. 444-452.
- 140
- 3. Ceyssens, P., Hertveldt, K., Ackermann, H., Noben, J., Demeke, M., Volckaert, G. and Lavigne, R., (2008). The intron-
- 142 containing genome of the lytic *Pseudomonas* phage LUZ24 resembles the temperate phage PaP3. *Virology*, **377**(2), pp. 233-
- 143 **238**.
- 144

145 4. Ceyssens, P., Miroshnikov, K., Mattheus, W., Krylov, V., Robben, J., Noben, J., Vanderschraeghe, S., Sykilinda, N.,

146 Kropinski, A.M. and Volckaert, G., (2009). Comparative analysis of the widespread and conserved PB1-like viruses infecting

147 Pseudomonas aeruginosa. Environmental microbiology, 11(11), pp. 2874-2883. 148 149 5. Darling, A.E., Mau, B. and Perna, N.T., (2010). progressive Mauve: multiple genome alignment with gene gain, loss and 150 rearrangement. PloS one, 5(6), pp. e11147. 151 152 6. DeLano, W.L. (2002) "The PyMOL molecular graphics system." 153 154 155 156 7. Dzhusupova, A.B., Plotnikova, T.G., and Krylov, V.N. (1982) "Detection of Transduction of Pseudomonas aeruginosa 157 Chromosome Markers by Virulent Bacteriophage
KZ in the Presence of Plasmid RMS148." *Genetika* **18** pp. 1799-1802. 158

159	8.	Eller, M.R., Vidigal, P.M., Salgado, R.L., Alves, M.P., Dias, R.S., Da Silva, C.C., DE Carvalho, A.F., Kropinski, A. and De
160		Paula, S.O., (2014). UFV-P2 as a member of the Luz24likevirus genus: a new overview on comparative functional genome
161		analyses of the LUZ24-like phages. BMC genomics, 15, pp. 7-2164-15-7.
162 163	9.	Freizon, E.V., Koplova, IuI., Cheremukhina, L.V., Krylov, V.N., (1989). The effect of the IncP-2-group plasmid on the growth
164		of Pseudomonas aeruginosa bacteriophages. Soviet Genetics, 25(7), pp. 751-758.
165 166		
167	10	Fuller, D.N., Rickgauer, J.P., Jardine, P.J., Grimes, S., Anderson, D.L. and Smith, D.E., (2007). Ionic effects on viral DNA
168		packaging and portal motor function in bacteriophage phi 29. Proceedings of the National Academy of Sciences of the
169		<i>United States of America,</i> 104 (27), pp. 11245-11250.
170		
171	11	. Glukhov, A.S., Krutilina, A.I., Shlyapnikov, M.G., Severinov, K., Lavysh, D., Kochetkov, V.V., McGrath, J.W., De Leeuwe, C.,
172		Shaburova, O.V. and Krylov, V.N., (2012). Genomic analysis of Pseudomonas putida phage tf with localized single-strand
173		DNA interruptions. <i>PloS one,</i> 7 (12), pp. e51163.
174		

175	12. Hansen, J.B. and Olsen, R.H., (1978). IncP2 group of <i>Pseudomonas</i> , a class of uniquely large plasmids. <i>Nature</i> , 274 (1), pp
176	715-717
177	
178	13. Hertveldt, K., Lavigne, R., Pleteneva, E., Sernova, N., Kurochkina, L., Korchevskii, R., Robben, J., Mesyanzhinov, V., Krylov,
179	V., and Volckaert, G., (2005). Genome comparison of Pseudomonas aeruginosa large phages. Journal of molecular biology.
180	354 (3), pp 536-545
181	
182	14. Jacoby, G.A., Sutton, L., Knobel, L. and Mammen, P., (1983). Properties of IncP-2 plasmids of Pseudomonas spp.
183	Antimicrobial Agents and Chemotherapy, 24(2), pp. 168-175.
184 185	15. Kelley, LA., and Sternberg, M. (2009) "Protein structure prediction on the Web: a case study using the Phyre server." Nature
186	<i>protocols</i> 4 (3) pp 363-371.
187	
188	16. Kindt, J., Tzlil, S., Ben-Shaul, A. and Gelbart, W.M., (2001). DNA packaging and ejection forces in bacteriophage.
189	Proceedings of the National Academy of Sciences of the United States of America, 98(24), pp. 13671-13674.
190	

191 17. Krieger, E., Keehyoung, J., Lee, J(1)., Lee, J(2)., Raman, S., Thompson, J., Tyka, M., Baker., M, and Karplus, K. (2009)

¹⁹² "Improving physical realism, stereochemistry, and side-chain accuracy in homology modeling: four approaches that

193 performed well in CASP8." *Proteins: Structure, Function, and Bioinformatics* **77**(S9) pp 114-122.

194

195	18. Krylov, V.N., Dela Cruz, DM., Hertveldt, K., and Ackermann, H.W., (2007). "phiKZ-like viruses", a proposed new genus of
196	movirus bacteriophages. Archives of virology, 152(10), pp. 1955-1959.
197 198	19. Kulakov, L.A., Balakshina, V.V., Morenkova, M.A., Zhulanova, E.I. and Boronin, A.M., (1991). Isolation and characterization
199	of Pseudomonas aeruginosa bacteriophage phikF77 mutants. Genetika, 27(11), pp. 1904-1911.
200	

201 20. Kulakov, L.A., Ksenzenko, V.N. and Boronin, A.M., (1986). Physical map of *Pseudomonas aeruginosa* phage phi kF77

202 genome. Localization of sites sensitive to restriction endonucleases. *Molekuliarnaia genetika, mikrobiologiia i*

203 *virusologiia,* **(10)**(10), pp. 13-16.

2	n	1
4	υ	4

204 205	21. Kulakov, L.A., Ksenzenko, V.N., Shlyapnikov, M.G., Kochetkov, V.V., Del Casale, A., Allen, C.C., Larkin, M.J., Ceyssens, P.
206	and Lavigne, R., (2009). Genomes of "phiKMV-like viruses" of Pseudomonas aeruginosa contain localized single-strand
207	interruptions. <i>Virology,</i> 391 (1), pp. 1-4.
208 209	22. Lavigne, R., Burkaltseva, M.V., Robben, J., Sykilinda, N.N., Kurochkina, L.P., Grymonprez, B., Jonckx, B., Krylov, V.N.,
210	Mesyanzhinov, V.V. and Volckaert, G., (2003). The genome of bacteriophage <i>qKMV</i> , a T7-like virus infecting Pseudomonas
211	<i>aeruginosa. Virology,</i> 312 (1), pp. 49-59.
212 213	23. Lucchini, S., Desiere, F. and Brussow, H., (1999). Comparative genomics of Streptococcus thermophilus phage species
214	supports a modular evolution theory. Journal of virology, 73(10), pp. 8647-8656.
215 216	24. Mesyanzhinov, V., Robben, J. Grymonprez, B., Kostyuchenko, V., Bourkaltseva, M., Sykilinda, N., Krylov, V., and Volckaert
217	G., (2002). The genome of bacteriophage phiKZ of <i>Pseudomonas aeruginosa</i> . J. Mol. Biol. 317 (1), pp. 1-19.
218 219	25. Morgan, A.F., 1979. Transduction of Pseudomonas aeruginosa with a mutant of bacteriophage E79. Journal of Bacteriology,
220	139 (1), pp. 137-140.
221	

222	26. Nakayama, K., Kanaya, S., Ohnishi, M., Terawaki, Y. and Hayashi, T., (1999). The complete nucleotide sequence of phi CTX,
223	a cytotoxin-converting phage of Pseudomonas aeruginosa: implications for phage evolution and horizontal gene transfer via
224	bacteriophages. Molecular microbiology, 31 (2), pp. 399-419.
225 226	27. Nilsson, N., Malmborg, A.C. and Borrebaeck, C.A., (2000). The phage infection process: a functional role for the distal linker
227	region of bacteriophage protein 3. Journal of virology, 74(9), pp. 4229-4235.
228 229	28. Rutherford, K., Parkhill, J., Crook, J., Horsnell, T., Rice, P., Rajandream, M.A. and Barrell, B., (2000). Artemis: sequence
230	visualization and annotation. Bioinformatics (Oxford, England), 16(10), pp. 944-945.
231 232	29. Sambrook, J and Russell, D.W., (2001). Molecular cloning: a laboratory manual Vol 1. pp 2.56-2.58. Cold spring harbor
233	laboratory press Cold Spring Harbor, New York:.
234 235	30. Tan, Y., Zhang, K., Rao, X., Jin, X., Huang, J., Zhu, J., Chen, Z., Hu, X., Shen, X. and Wang, L., (2007). Whole genome
236	sequencing of a novel temperate bacteriophage of P. aeruginosa: evidence of tRNA gene mediating integration of the phage
237	genome into the host bacterial chromosome. Cellular microbiology, 9(2), pp. 479-491.
238	

239 31. Tetart, F., Desplats, C. and Krisch, H., (1998). Genome plasticity in the distal tail fibre locus of the T-even bacteriophage:
 240 recombination between conserved motifs swaps adhesin specificity. *Journal of Molecular Biology*, 282(3), pp. 543-556.
 241
 242 32. Weinstein, R., Gaynes, R. and Edwards, J., (2005). Overview of nosocomial infections caused by gram-negative bacilli.

Clinical Infectious Diseases, **41**(6), pp. 848-854.



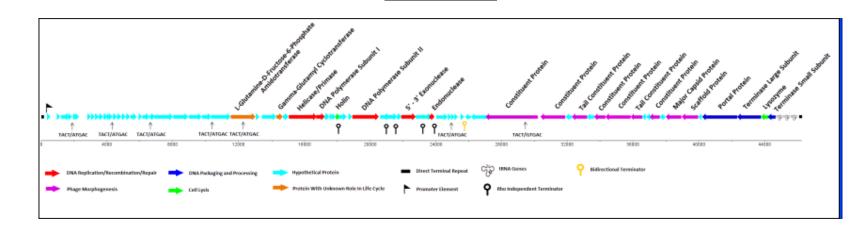


Figure 1. Genome map of the *P. aeruginosa* bacteriophage phiCHU.

- 247 Predicted ORFs are displayed as arrows indicating the direction of transcription. Functional annotations (if any) are displayed
- above ORFs and colour/symbol codes are presented at the bottom of the figure. Sequences associated with localized nicks are
- 249 displayed at their respective positions