# we are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists



125,000 International authors and editors 140M



Our authors are among the

TOP 1%





WEB OF SCIENCE

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

## Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected. For more information visit www.intechopen.com



#### Chapter

## Algae Essential Oils: Chemistry, Ecology, and Biological Activities

Mohamed El Hattab

#### Abstract

This chapter focuses on the essential oils and volatile fractions of seaweed. It includes an introduction to the essentials and volatile fractions and the main chemical classes found. This part is completed by a presentation of the fundamental aspects of biodiversity and the chemodiversity of the marine environment followed by the taxonomy and systematics of marine macroalgae. The heart of this chapter concerns the chemistry of volatile products extracted from marine algae. It reports the specificities of the marine natural products chemistry in comparison to that of terrestrial organisms. The description of volatile compounds in seaweed is divided into two parts, the first reports the common compounds identified in main volatile fractions and the second cover the specific volatile components. These include C11 hydrocarbons, sulfur compounds, and halogenated hydrocarbons. These latter are playing a very important role in communication and chemical defense. The last part includes aspects of chemical ecology and biological activities of volatile products.

**Keywords:** essential oils, marine algae, C11 hydrocarbons, sulfur compounds, halogenated sesquiterpenes, chemical ecology, biological activities

#### 1. Introduction

The origin of the distillation methods is an invention attributed to the Arab alchemists and to the Persian scientist Avicenna (980-1037) with the establishment of the steam distillation process. Avicenna invented a setup to prepare essential oils and aromatic waters. Essential oils, sometimes called quintessence, are a very complex mixture of volatile compounds produced by the secondary metabolism in various plant organs (flowers, fruits, seeds, leaves, etc.) and algae. According to ISO and AFNOR standards, essential oils are defined as volatile composition obtained from raw materials by steam distillation and/or by cold expression from citrus peels (known as essences) [1]. The definition of an essential oil excludes other volatile fractions obtained by steam distillation and/or hydrodistillation from the crude extract resulted from solvent extraction, supercritical fluid extraction, solventand water-free microwave extraction, ultrasound-accelerated solvent extraction, solid-phase microextraction, and headspace extraction. The chemical composition of essential oils and volatile fraction could be quite similar. Moreover, it should be pointed out the clear difference between the physical and chemical properties of essential oils and fixed or fatty oils. The fixed oils contain mainly triglycerides, esters composed of three saturated fatty acids linked to glycerol, characterized by

high boiling and low volatility. The chemical composition of essential oils is principally composed of terpenes derived from the mevalonate and methylerythritol pathways [2]. Monoterpenes and sesquiterpenes are commonly the main contributor group of compounds identified in several essential oils [3]. Moreover, some essential oils contain other chemical classes, such as phenols (derived from shikimic acid pathway); the saturated and unsaturated fatty acids, acting as biosynthetic precursors; alkanes; and, more rarely, nitrogen and sulfur derivatives [4]. The essential oils play an important role in the allelopathic interaction of plants. They are involved in defense and signaling processes [5] and attraction of pollinating insects [6]. They constitute an important raw material source for the pharmaceutical, food, cosmetics, and perfume industries [7]. The essential oils of different plants exhibit a broad spectrum of biological activities. They show antibacterial activities attributed, in some cases, to the presence of phenolic compounds [8]. The literature reports also the excellent antioxidant [9], anti-inflammatory [10], and cancer chemoprotective activities [11].

#### 2. Marine biodiversity and chemodiversity

More than 70% of the Earth's surface are oceans and seas. It is not surprising to affirm that the marine environment is characterized by an important biodiversity in comparison to terrestrial organisms. In 2010, 230,000 marine species were listed [12]. Consequently, with the increase of biological space (biodiversity), more novel metabolites (high chemodiversity), involved in ecological interactions, are produced in order to ensure easy adaptation of the species [13, 14]. Furthermore, the chemodiversity of the marine ecosystem has no equivalent in terrestrial environment. The large groups of the sea organisms, such as red algae and soft corals, are known to produce a great variety of quite unique secondary metabolites, such as highly halogenated terpenes, definitely due to the high halogen concentration of the sea water, and acetogenins from *Laurencia* (Rhodophyta) [15, 16], toxic polyketide from sponges [17], and prostaglandins from the gorgonian corals [18, 19].

#### 3. Systematics and taxonomy of macroalgae

It was the French botanist Joseph Pitton de Tournefort (1656–1708) who grouped the species into genera and then the Swedish naturalist Carl von Linné (1707–1778), founder of systematics (or taxonomy), who classified the organisms into increasingly large groups: species, genera, families, orders, classes, phylum (or phyla), and kingdoms. Algae, according to Feldmann and Chadefaud [20, 21], are classified into six branches differentiated by the nature of the pigments, the nature and situation of carbohydrate reserves, and the presence or absence, number, and arrangement of flagella:

- Pyrrophycophyta: unicellular marine or freshwater algae
- Euglenophycophyta: unicellular freshwater algae rich in organic matter
- Chrysophycophyta: most are single-celled; freshwater and sea water
- Chlorophycophyta: green algae; single or multi-cell; marine, freshwater, and terrestrial environments

- Phaeophycophyta: brown algae; always multicellular and almost exclusively marine
- Rhodophycophyta: red algae; mainly multicellular and mostly marine

#### 3.1 Phaeophyceae (or Fucophyceae)

There are about 2000 species (in 265 genera) of brown algae [22], and less than 1% are known from freshwaters (3–7 genera) [23]. The brown color is due to Fucoxanthin (carotenoid pigment) and in some species to the presence of tannins (phenolic compounds).

#### 3.2 Chlorophyta

There are estimated to be at least 600 genera with 10,000 species within the green algae [24] recognized inhabiting mostly in the water's surface of the calmer seas. They are characterized by the presence of chloroplasts with two envelope membranes, stacked thylakoids, and chlorophyll a and b. In their fundamental biochemistry (photosynthetic pigments, storage polysaccharides, etc.), the Chlorophyta resemble the higher plants [24].

#### 3.3 Rhodophyta

They are primarily marine in distribution sometimes inhabiting the deep water, with less than 3% (150 species from 20 genera) of the over 6500–10,000 species occurring in truly freshwater habitats [25]. The red algae are characterized by eukaryotic cells, with the complete absence of flagellar structures, food reserves of starch, presence of phycobilins, chloroplasts without stacked thylakoids, and no external endoplasmic reticulum.

#### 4. Chemistry of marine algae volatile compounds

The fragrances of terrestrial plants have aroused human interest since antiquity; they were related to spiritual and civilizational aspects. It is not surprising that the first research work on odorous volatile products was carried out on aromatic plants. Phytochemists have quickly associated the odors emanating from trees and shrubs to terpenes (notably monoterpenes), spices to phenols and derivatives, and fruits and flowers to aldehydes, esters, and ketones. The smell connected with marine flora are much less familiar. Unlike the wide number of terrestrial odoriferous plants, relatively few marine seaweeds possess an attractive odor. Although the natural products chemistry of terrestrial organisms was known before the nineteenth century, the one of the marine derived is more recent, and it has only emerged over the past 75 years. This is due to the complexity to access the marine environment. The marine natural products had become an important subdiscipline of natural products chemistry, which has experienced a particular craze which has led to the isolation and characterization of thousands of secondary metabolites belonging to original chemical skeletons without equivalent in the terrestrial environment.

Historically, volatile oils of terrestrial plants were used in Chinese [26] and Egyptian civilizations [27–29] few centuries ago, whereas the first works on the isolation of volatile products of marine algae were carried out, on the brown alga *Fucus* [30] and the red algae *P. fastigiata* and *P. nigrescens* [31] when the seaweeds are exposed to air, at the beginning of the 1930s, followed later by the Katayama

researches in 1951–1961 [32] and Moore prior to 1966 [33]. The volatile organic compounds in marine algae, as in plants and fungi, released into the seawater, are involved in the chemical communications process; these compounds play an important role as either pheromones or allelochemicals for communication and interaction with the surrounding environment [34, 35]. The species produce the volatile organic compounds in closed relation to their physiology; the algae must adapt to abiotic stresses of their ecosystem [36]. The volatile components of marine algae contain a mixture of chemical classes such as terpenes, hydrocarbons, fatty acids, esters, alcohols, aldehydes, ketones [37–41], C11-hydrocarbons [33, 42], polyphenols and derivatives [43, 44], and halogenated [45] and sulfur compounds [46, 47]. The distinctive ocean smell is due to the presence of terpenes, but particularly, to a fraction of acyclic and cyclic non-isoprenoid C11-hydrocarbons acting as pheromones and playing an important role in the chemical communication [48], it seems to be most abundant in brown algae of the genus Dictyopteris [33]. As for terrestrial plants, the monoterpenes identified in algae such as linalool, citral, geraniol, and terpinolene, 1,8-cineole,  $\alpha$ -pinene and  $\beta$ -pinene, and eugenol and isoeugenol could be valued in perfumery. While the disagreeable odor is related to amines and halogenated, sulfurous, and other specific compounds [49], the dimethyl sulfide, mainly distributed in Chlorophyta and in some Rhodophyta [50], has a very unpleasant odor molecule. It results from the enzymatic cleavage of dimethyl-2-carboxyethylsulfonium hydroxide, from the green algae species (*E. intestinalis* and *A. centralis*) [51].

#### 4.1 Common volatile organic compounds of macroalgae

#### 4.1.1 Hydrocarbons and oxygenated hydrocarbons

The alkanes and alkenes are common compounds in the majority of volatile fraction and essential oils of marine macroalgae. The chemical composition reveals the presence of the linear and branched saturated hydrocarbons from  $C_7$  to  $C_{36}$  [37, 52–54], the unsaturated hydrocarbons from  $C_8$  to  $C_{19}$  with the presence of 1 [37, 52–54] to 4 degrees of unsaturation [55] in the volatile fraction obtained by several extraction techniques. We also noted the presence of mono- and di-alcohol of  $C_4$ – $C_{18}$  [37, 52–54, 56, 57]. Some short-chain ( $C_6$ ,  $C_9$ ) and middle-chain ( $C_{10}$ ) aliphatic aldehydes are formed in marine algae from fatty acids ( $C_{20}$ ), whereas they are formed from  $C_{18}$  in higher plants [58–60]. Also, it has been reported that long-chain aldehydes ( $C_{14}$ ,  $C_{17}$ ) of the green alga *U. pertusa* [61, 62] are formed by decomposition of fatty acids through the corresponding 2-hydroperoxy acid; this later are encountered in a variety of marine algae [63, 64].

In addition to aldehydes, the ketone compounds were commonly reported in the aroma composition of algae [65]; the presence of  $\beta$ -ionone and 6-methyl-5hepten-2-one which are formed via the oxidative cleavage of carotenoids such as lycopene and phyotene was mentioned [66].  $\beta$ -ionone, present in several essential earth oils, is a powerful odorant for the perfume industry. 6-methyl-5-hepten-2-one, in addition to its pleasant fragrant note, is often used as an intermediary in the synthesis of several monoterpenes highly valorized in perfumery. In addition, other simple ketone (C<sub>6</sub>—C<sub>19</sub>) compounds such as maltol [53], octan-3-one [57], nonacosan-2-one [67], and undeca-1,4-dien-3-one [42] are identified in the volatile fractions of algae. Saturated fatty acids from C<sub>3</sub> to C<sub>18</sub> and their ester derivatives have also been identified in the chemical composition of volatile algae fractions [37]. Unsaturated fatty acids and their corresponding esters, in particular Eicosa-5,8,11,14-methyltetraenoate and Eicosa-5,8,11,14,17-methyl-pentaenoate [42], are usually found; this is probably related to their implications in biosynthetic processes of other metabolites.

#### 4.1.2 Amine compounds

The amine compounds have been described several times in marine algae [68–70]; the small amine molecules such as methyl amine, dimethylamine, ethylamine, and propylamine were found in algae [71]. The volatile amines in algae result from decarboxylation of amino acids [71]. Although present in brown and green algae, the amine compounds were especially found in red algae.

#### 4.1.3 Halogen compounds

The volatile halogen compounds are rare in terrestrial plants, but quite habitual in marine algae because of the presence of chlorine and bromine ions at a high concentration in seawater. The red algae possess the highest abundance of halogenated organic compounds, which are found as terpenoid, phenols, carbonyl compounds, and fatty acid-derived metabolites [45]. They were produced in marine algae and emitted into the atmosphere; the highest amounts of brominated compounds released were done by L. saccharina [72]. Chemical investigations of marine algae have shown the presence of 2-bromophenol, 2,4-dibromophenol, and 2,4,6tribromophenol in numerous red, green, and brown algae. It has been reported the biosynthesis of bromophenols in U. lactuca via the bromoperoxidases in the presence of precursors such as phenol, 4-hydroxybenzoic acid, and 4-hydroxybenzyl alcohol [73]. The bromoperoxidases are involved in the biosynthesis of brominated alkanes, such as CHBr<sub>3</sub>, CH<sub>2</sub>Br<sub>2</sub>, CHClBr<sub>2</sub>, and others in several marine organisms, among them, the red alga Asparagopsis sp. [74]. The biosynthesis of organohalogens has known enormous interest as reported in several literature review [75–77]. As indicated for bromocompounds, the iodoperoxidases are responsible of the production of iodinated compounds in marine algae [78-80]. The chemical investigation of 29 macroalgae species reveals their release of volatile iodocompounds iodoethane, 1-iodopropane, 2-iodopropane, 1-iodo-2-methylpropane, 1-iodobutane, 2-iodobutane, diiodomethane, and chloroiodomethane [81]; it has reported that diiodomethane was the main iodinated compound released by brown macroalgae [82].

#### 4.1.4 Terpenoid compounds

Terpenes, or terpenoids, are a large and diverse class of plant secondary metabolites, produced by numerous varieties of plants and algae from isoprene building blocks; they play a major ecological role, most notably in defense against plant-feeding insects and herbivores [82]. However, some terpenoids are involved in primary metabolism, such as stability of cell membranes and photosynthesis. The terpenes display enormous structural diversity, are the main constituents of essential oils of terrestrial plants and seaweeds [83], and are characterized by their pleasant strong odor. The terpenoids are biosynthesized mainly via two pathways, the mevalonate pathway and the MEP pathway. The chemical screening of volatile fraction and/or essential oils of algae reveals the presence of high content of monoterpenes and sesquiterpenes and rarely diterpenes [42]. The most significant acyclic monoterpenes found in algae are myrcene (1), ocimene (2), geranial (3), neral (4), citronellol (5), and geraniol (6) (**Figure 1**). Moreover, the most odoriferous compounds identified in algae are included in the acyclic group of monoterpenes [84].

Likewise, the most common monocyclic algae volatile oil is 1,8-cineole (8) [84], while  $\alpha$ -pinene (9) and  $\beta$ -pinene (10) are the most commonly reported of bicyclic monoterpenes (**Figure 2**) [84, 85]. Sesquiterpenes from marine macroalgae constitute a large group, compared to monoterpenes, of secondary metabolites [86]; some of them are halogenated [87]. Some of the algae sesquiterpenes act as

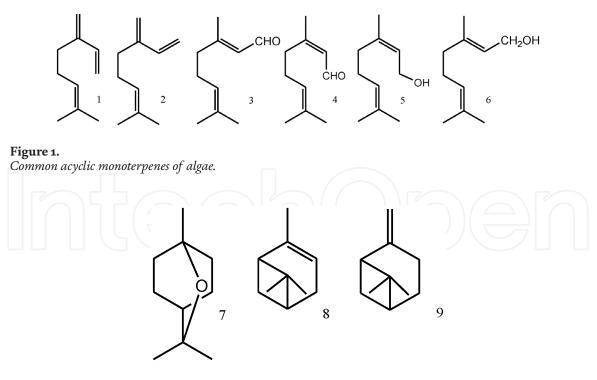


Figure 2.

Most representative monocyclic and bicyclic monoterpenes of algae.

semiochemicals, chemical defense agents, and/or pheromones. They may be acyclic, cyclic, or bicyclic, including several original structures. Among all marine macroalgae, the genus *Laurencia* (red algae) is the most potent source of sesquiterpenes.

The most common sesquiterpenes reported in marine algae (10–53) are grouped in **Table 1** and illustrated in **Figure 3**. The only diterpene and triterpene described as volatile compounds are, respectively, phytol and squalene. Phytol is a degradation product of chlorophyll and the precursor of vitamin E. The squalene is via the epoxy squalene, the biosynthetic precursors of triterpenes and steroids.

#### 4.2 Specific volatile compounds of macroalgae

#### 4.2.1 Odoriferous C11 hydrocarbons from brown algae (Phaeophyta)

The brown algae produce a variety of volatile derivatives whose chemical nature and biological function are different from those of red algae. They are hydrocarbons with 11 carbon atoms without halogens which can be classified according to their chemical structure into four groups [94]: (a) derivatives of cyclopropane, (b) derivatives of cyclopentene, (c) derivatives of cycloheptadiene, and (d) acyclic olefins. The only volatile hydrocarbon with eight carbon atoms identified in brown algae is fucoserratene. These metabolites, which are known in all the species of *Phaeophyceae*, are not specific to an order or a family. They have been isolated from diverse groups of brown algae (e.g., the *Zonaria*, *Desmarestia*, *Dictyota*, *Ectocarpus*, *Laminaria*, and *Fucus*); it appears to be most abundant in brown algae of the genus *Dictyopteris* [95].

They are involved in the reproduction process of the alga; they are sex pheromones. To date, it has been revealed that these algal pheromones are involved at least in three well-defined ecological interactions [96]: (i) synchronization of the mating of male and female cells by the controlled release of male spermatozoids, (ii) enhancement of the mating efficiency by attraction, and (iii) chemical defense of the plant due to the presence of high amounts of pheromones within and release from the thalli into the environment. Furthermore, the relationship between

N°	Compounds	Species and References	N°	Compounds	Species and References
10	(E)-Farnesene	D.m. <sup>[88]</sup> D. d. <sup>[89]</sup>	32	Germacrene D	D. m. <sup>[42]</sup> D. div. <sup>[96]</sup>
11	Hexahydro Farnesyl- acetone	(B. f., C. m., C. g., C.e., P. d., L. p., L. c.) <sup>[90]</sup> C. v. <sup>(91)</sup>	33	Sativene	D. m. <sup>[42]</sup>
12	$\alpha$ -Cubebene	D. m. <sup>[42]</sup> D. d. <sup>[89]</sup>	34	Zonarene	D. m. <sup>[42]</sup>
13	β-Cubebene	D. m. <sup>[42]</sup> D. d. <sup>[89]</sup>	35	Vulgarol	D. m. <sup>(42)</sup>
14	α-Copaene	D. m. <sup>[42]</sup> D. d. <sup>[89]</sup>	36	4,4a,5,6,7,8-hexahydro-5- methyl-8-(1-methylethyl)- 2(3H)naphtalenone	D. m. <sup>[42]</sup>
15	(-)-Copaene	D. Div. <sup>[92]</sup>	37	1,2,3,4,4a,7-hexahydro- 1,6-dimethyl-4-(1-methyl ethyl) Naphtalène	D. m. <sup>[42]</sup>
16	δ-Cadinene	D. m. <sup>[42]</sup> Z. m. <sup>[93]</sup> D. d. <sup>[89]</sup>	38	Epi-bicyclo- sesqui- phellandrene	<i>D. m</i> . <sup>[42]</sup>
17	γ-Cadinene	D. div. <sup>[92]</sup> D. d. <sup>[89]</sup>	39	Cyclosativene	D. d. <sup>[89]</sup>
18	α-Cadinol	P. t. <sup>[94]</sup> U. p. <sup>[95]</sup>	40	α-Ylangene	D. d. <sup>[89]</sup>
19	δ-Cadinol	Z. m. <sup>[93]</sup> D. div. <sup>[92]</sup>	41	γ-Muurolene	D. d. <sup>[89]</sup>
20	1,10-di-epi- cubebol	D. m. <sup>[42]</sup>	42	β-Cadinene	D. d <sup>[89]</sup>
21	Cubenol	D. div. <sup>[42]</sup> , D. p. <sup>[96]</sup> , P.t. <sup>[94]</sup> , U. p. <sup>[95]</sup>	43	Ar-curcumene	D. d. <sup>[89]</sup>
22	Epi-Cubenol	Z. m. <sup>[93]</sup>	44	Bicyclogermacrene	D. d. <sup>[89]</sup>
23	Cadalene	D. div. <sup>[92]</sup>	45	Epizonarene	D. d. <sup>[89]</sup>
24	α-Calacorene	D. m. <sup>[42]</sup>	46	α-Muurolene	D. d. <sup>[89]</sup>
25	β-Bourbonene	D. m. <sup>[42]</sup> , D. d. <sup>[89]</sup>	47	δ-Selinene	D. d. <sup>[89]</sup>
26	Azulene	D. m. <sup>[42]</sup>	48	trans-γ-Bisabolene	H. f. <sup>[89]</sup>
27	Axenol	D. m. <sup>[42]</sup>	49	trans-Cadina-1,4-diene	D. d. <sup>[89]</sup>
28 Aromadendrene		D. m. <sup>[42]</sup> D. d. <sup>[89]</sup>	50	α-Cadinene	D. d. <sup>[89]</sup>
29	α-Amorphene	D. m. <sup>[42]</sup> D. d. <sup>[89]</sup>	51	Germacrene B	D. d. <sup>[89]</sup>
30	Albicanol	D. m. <sup>[42]</sup>	52	Dendrolasin	H. f. <sup>[89]</sup>
31	β-Elemene	D. d. <sup>[96, 92]</sup>	53	τ-Muurolol	D. d. <sup>[89]</sup>

Abbreviation: D. m., Dictyopteris membranacea; D. d., Dictyota dichotoma; B. f., Bangia fuscopurpurea; C. m., Cystoseira mediterranea; C. g., Callithamnion granulatum; C. e., Cystoseira elegans; P. d., Polysiphonia denudata; L. p., Laurencia papillosa; L. c., Laurencia coronopus; C. v., Cladophora vagabunda; D. div., Dictyota divaricata; Z. m., Zostera marina; P. t., Pyropia tenera; U. p., Ulva pertusa; D. p., Dictyota prolifera; H. p., Halopteris filicina.

#### Table 1.

Most common sesquiterpenes of macroalgae [88–93].

structures of pheromones and the taxonomic classifications of algae are still not established. Until now, a series of 12 (54–65) hydrocarbons and epoxides (**Figure 4**) have been characterized, and more than 50 stereoisomers are known within the pheromone bouquets of more than 100 different species of brown algae [48, 96–99].

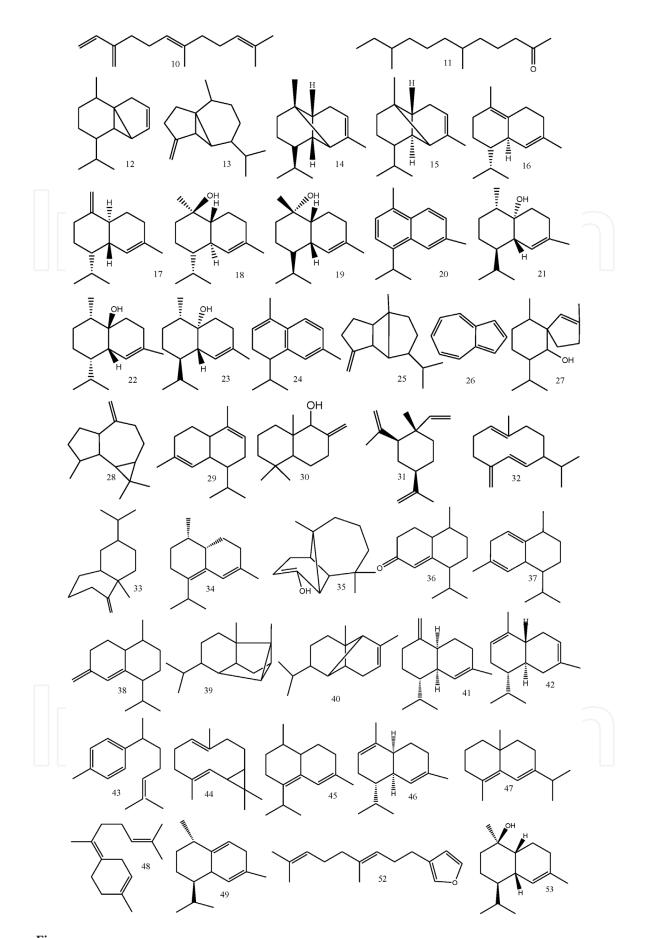
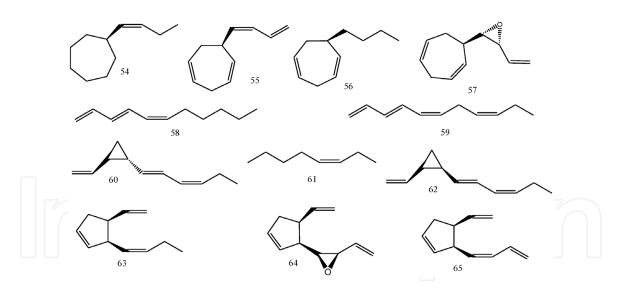


Figure 3. Common sesquiterpenes described in volatile oil of marine algae.

Moreover, the presence of C11 hydrocarbons is not only limited to marine brown algae. The same compounds have been reported in cultures of diatoms [100], the volatile fraction released during blooms of microalgae in freshwater lakes [101] and,



**Figure 4.** *Pheromones of brown algae.* 

inquisitively, in higher plants [102, 103]. **Table 2** reports the pheromones described in **Figure 4**, the algae from which they are derived, as well as their attraction or release activities. In comparison to the number of brown algae species, the chemodiversity of pheromones is relatively limited, so, the semiochemical activity of the same molecule is noted in more than one species. Female gametes secrete a mixture of products, not just one pheromone and depending on species; released pheromones are either optically pure or enantiomeric mixtures.

N°	Pheromones	Release/	Algal species
		Attraction	
		(R/A)	
54	Ectocarpene	A	Scytosiphon sp. E. fasciculatus, A. tricularis. S. rigidula
55	Desmarestene	A/R	D. acculeata, D. viridis
		A	C. spongiosus, D. firma
56	Dictyotene	A	D. dichotoma , D. diemensis, D. prolifera
57	Lamoxirene	A/R	L. angustata, L. sinclari
			L. japonica, L. digitata, L. hyperborea
			L. sacharina, Pleurophydus, A. classiforia, A. esculenta
			A. marginata, E. radiata, E. arborea, P. california, U.
			pinnatifida, D. reticulata, L. variegata, L. littoralis, M.
			integrifolia, M. pyrifera, N. luetkeana, P. porra, A.
			cribrosum, C. triplicata, H. sessile, K. gyrata
58	Cystophorene	A	C. siliquosa.
59	Finavarrene	R	P. callitricha
60	Hormosirene	A	H. banksii, X. chondrophylla, X. gladiata, D. antarctica D. potatorum, D. willana, C. peregrina, C. bullosa, A.
			mirabilis, M. simplex, S. lomentaria.
61	Fucco- serratene	A	F. serratus, F. spiralis, F. vesiculosus.
62	Pre-	А	E. siliculosus.
	ectocarpene		
63	Multifidene	A/R	C. multifida, Z. angustata, C. tomentosa
64	Caudoxirene	R	P. caudata, D. foeniculaceus.
65	Viridene	A	S. phinneyi
		R/A	D. virdis

**Table 2.**C11 and C8 pheromone activities from marine brown algae.

#### Essential Oils - Bioactive Compounds, New Perspectives and Applications

However, it has been verified that the biological activity is associated with a single constituent which may not be the major product. These by-products sometimes play a role of modulator of response of the gametes, and in general, they do not have a determined biological function [94].

#### 4.2.2 Sulfur compounds in the genus Dictyopteris

The organic sulfur compounds are widespread in terrestrial and marine plants [104]. Due to the relatively high sulfate concentration in seawater, and the particularly high sulfide concentration in anoxic environments, it was expected that many sulfides would occur in the marine environment [104]. They are reported in few taxa and act as chemical defenses against herbivores [105]. As part of this single group, some *Dictyopteris* species (*Phaeophyceae*, *Dictyotales*) are acknowledged to produce considerable amounts of sulfur-containing compounds (**Figure 5**); many of them were found in *D. polypodioides* [106]. Among the first seaweeds discovered to produce organic sulfur compounds were the Hawaiian brown algae

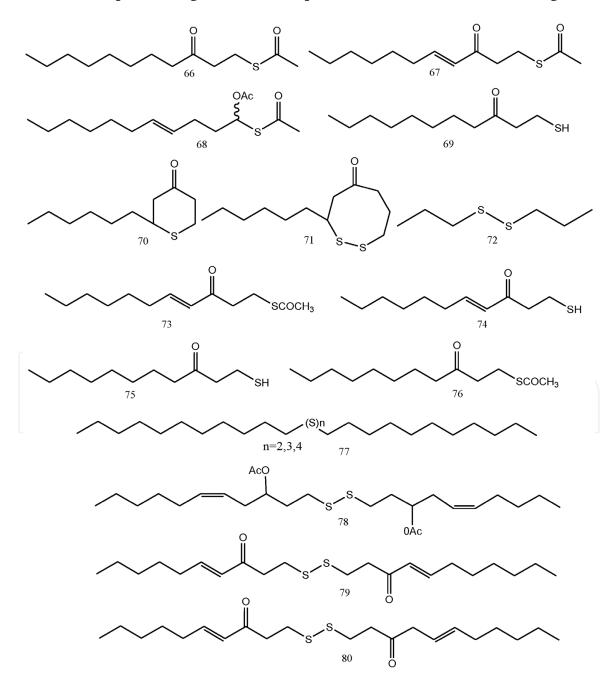


Figure 5. Sulfur compounds of the genus Dictyopteris.

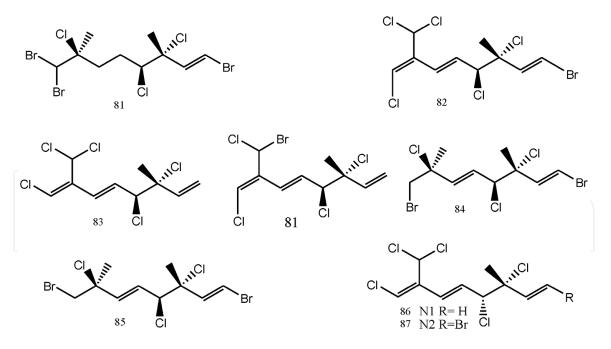
*D. plagiogramma* and *D. australis* [107]. Eight compounds containing a C11 unit attached to a sulfur atom with oxygen substituent at C-3 have been isolated and characterized [47]; most of these compounds appear to be biosynthetically related to C11 hydrocarbon pheromones and may originate from oxidative degradation of highly unsaturated eicosanoids (arachidonic acid) via oxygenated intermediates. The 1-undecen-3-ol, present in essential oils from *Dictyopteris* spp., may represent the common precursor to both classes of C11 compounds [95, 107]. The C11 sulfur metabolites seem to be restricted to the *Dictyopteris* genus.

#### 4.2.3 Halogenated terpenes from red algae (Rhodophyta)

As noted previously, the halogenated compounds are common in the marine environment. They are formed among diverse species such as bacteria, sponges, molluscs, algae, and several marine worms. Among all marine algae, the Rhodophyta class possesses a privileged biosynthetic pathway for organohalogen compounds. A huge number of organohalogens have been isolated from most genera of Rhodophyta [108, 109]. The genus Laurencia is the most prolific source of sesquiterpenes among all marine macroalgae, most notably, the halogenated sesquiterpenes belonging to a variety of chemical skeletons including chamigrane, bisabolane, laurane, snyderane, and brasilane along with some rearranged derivatives [110, 111]. Inquisitively, bromine is the most occurring halogen in marine natural products, despite that its concentration in seawater is lower than that of chlorine. To the best of our knowledge, the isolation of halogenated monoterpenes is limited to three families of marine red algae, the *Plocamiaceae* and *Rhizophyllidaceae* [112, 113], and *Ceramiaceae* [114]. The chemical structure of Rhodophyta monoterpenes is characterized by multiple halogen substitutions (chlorine and bromine) and by uncommon carbon cycle structures in the case of cyclic compounds. All halogenated acyclic seaweed monoterpenes appear to be derived from the halogenation of myrcene or ocimene [114]. As indicated in the rich bibliography dedicated to this purpose [45, 113, 115–117], the almost majority of halogenated terpenoids (monoterpenes, sesquiterpenes, and diterpenes) described in red algae are isolated from crude solvent extracts. Monoterpenes, even halogenated, are characterized by high volatility; they are the main constituents of essential oils and volatile fractions. The selective supercritical fluid extraction, by adjusting time and pressure, of Santa Cruz P. cartilagineum [118] has allowed the isolation of eight halogenated monoterpenes (81–87) (Figure 6).

The same species collected along the central coast of Chile [119] conduct to the isolation of eight monoterpenes (88–95), four of which are based on the 1-(2-chlororovinyl)-2,4,5-trichloro-1,5-dimethylcyclohexane skeleton (**Figure** 7). As in the genus *Plocamium*, the chemical study of the genera *Portieria* [120], *Ochtodes* [121], and *Microcladia* [114, 122] has led to the isolation of over 100 of acyclic, cyclic, and tetrahydrofuran halogenated monoterpenes. A large number of halogenated sesquiterpenes, more than monoterpenes, were described in red algae especially in the genus *Laurencia* (Ceramiales). Although the sesquiterpenes are also volatile compounds, we describe in this paragraph only the ones reported in the chemical composition of essential oils and volatile fraction of red algae.

The first brominated sesquiterpene (**Figure 8**) ketone spirolaurenone (96), chamigrane skeleton, was described in the essential oil of *L. glandulifera* (Japan) in 1970 [123], followed by the 10-Bromo-7-chamigren-2-one (97) in the same species [124]. The preintricatol (98), found in *L. gracilis* [125], seem to be the precursor of halogenated sesquiterpenes of chamigrene type. The Puertitols A (99) and B (100) were isolated from *L. obtusa* [126] as well as the metabolites (101) and (102) from *L. caespitosa* [127]. An important halosesquiterpene characteristic of the family



**Figure 6.** *Monoterpenes isolated Santa Cruz* Plocamium cartilagineum.

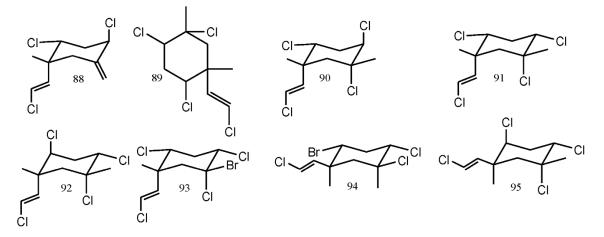


Figure 7. Monoterpenes isolated from Plocamium cartilagineum (Chile).

Rhodomelaceae is elatol (103); it is isolated from *L. elata* [128] and from several other species of *Laurencia* [129]. An exhaustive literature review has described the chemical structure data and biological activities of the halogenated sesquiterpenes of red algae [130, 131].

#### 4.2.4 Ecology

The volatile compounds play an important role in the inter- and intraspecies chemical communication in marine algae. They act as pheromones [97] or allelochemicals, chemical defenses against herbivores [132, 133], and inhibition of bacterial and fungal biofilms [134]. The genus *Dictyopteris* produce a high amount of C11 hydrocarbons, some of which act as pheromones that stimulate gamete release or attract sperm during sexual reproduction [96]. The first male-attracting metabolite was elucidated as ectocarpene (54) [135] which shows a moderate activity at 10 mM. A subsequent study revealed that the real pheromone used by the female gamete was pre-ectocarpene (62) which is active at 5 pM. In fact, the alga produces pre-ectocarpene which undergoes a thermal rearrangement (Cope rearrangement)

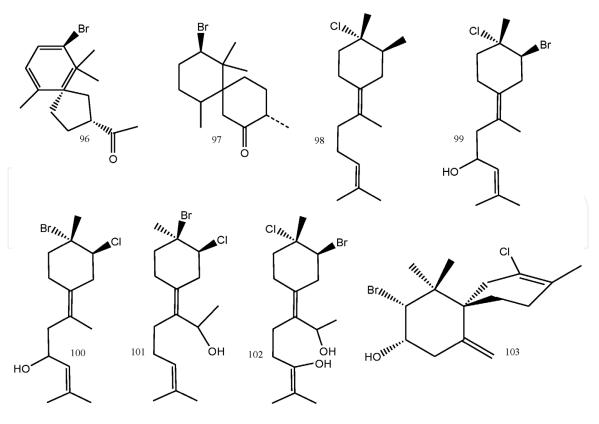


Figure 8. Halogenated sesquiterpenes from red the genus Laurencia.

to lead to ectocarpene [136]. The Cope rearrangement occurs between the time of the releasing and attraction of the pheromone; the sigmatropic transformation serves as a natural control mechanism for deactivation of the pheromone [136]. The genus *Dictyopteris* produces significant amounts of C11-sulfur compounds which are involved in chemical defense [137].

In green algae, the volatile compounds, such as (Z)-8- heptadecane, act also as allochemicals [61]. In the genus *Caulerpa*, the caulerpenyne is the most abundant cytotoxic sesquiterpene produced by *C. taxifolia* and *C. racemosa* [138, 139]. It is involved either in the chemical defense of the plant against herbivore or within the framework of interspecific competition as antifeedant and/or antifouling activities [140]. In red algae, the halogenated organic compounds are produced, probably, to be involved in the defense system against microorganism infection [141], herbivore attack [141], space competitors [142], and harmful fouling by different types of epiphytes [142].

#### 4.2.5 Biological activities

There are several reports of secondary metabolites, among them are numerous volatile compounds, derived from macroalgae which exhibit a broad range of biological activities such as antibiotics [40, 143].

The essential oil of *D. membranacea* has shown a strong antibacterial activity against *Staphylococcus aureus* and *Agrobacterium tumefaciens*, which is translated by an MIC of 1519 µg/mL [106]. The volatile oil of *P. pavonica* possesses a moderate antimicrobial activity against *Staphylococcus aureus* and *Candida albicans* [144]; antifungal against *Macrophomina phaseolina*, *Rhizoctonia solani*, and *Fusarium solani* [145]; cytotoxicity against KB cells [146]; and antitumor activity against lung and human carcinoma cell lines [147]. On the other hand, the volatile of *H. clathratus* showed a pronounced antimicrobial activity against *S. cerevisiae* compared with Canesten as reference material [148].

The cytotoxicity is the most common activity observed for halogenated organic compounds isolated from the family Rhodomelaceae. A large number of these compounds were shown to be cytotoxic to a wide range of cancer cell lines [115].

Among many of the halogenated sesquiterpenes evaluated for their in vitro cytotoxic effects against HeLa and HEP-2 cancer cell lines, and against nontumoral VERO cells, during both lag- and log-phase cell growth [149], elatol (103) turned out the most active compound with IC50 values of 4.1 and 1.3  $\mu$ M to HeLa, 2.4 and 2.0  $\mu$ M to HEP-2, and 2.3 and 25.0  $\mu$ M to VERO cells, in lag- and log-phase, respectively [150]. Further studies were carried on the evaluation of the cytotoxicity against several tumor cell lines of chamigrane [150] and Laurane- and Cuparane-type sesquiterpenes and were found to display a wide range of potency levels [151, 152]. Other activities of halosesquiterpenes such as antibacterial activity [153], antifungal activity [154], and antiviral activity [155] were investigated and conducted to promising results.

#### 5. Conclusion

Essential oils from terrestrial plants have been known for a very long time. They have been applied in different domain, particularly in aromatherapy. Essential oils from seaweed are much more recent. The fragrant note of marine origin is becoming more and more interesting among perfumers, the species of the genus *Dictyopteris* and *Dictyota* could be considered as the best example. This importance is related to the great biodiversity and chemodiversity of the marine environment compared to the terrestrial environment. The chemical composition of essential oils and volatile fractions of macroalgae contains compounds usually found in terrestrial essential oils. These include hydrocarbons, oxygenated hydrocarbons, terpenes, and fatty acids. However, they contain specific products such as halogenated products, C11 hydrocarbons, sulfur compounds, and halogenated terpenes. The specific compounds play a very important role on the chemical ecology; they are involved in defense mechanisms and chemical communication. The volatile fractions of algae show a broad spectrum of biological activity, such as antibacterial, antifungal, anticancer, and antibiotic activities.

#### Acknowledgement

This work is supported by the General Directorate of Scientific Research and Technological Development (DGRSDT)—Algeria.

#### **Author details**

Mohamed El Hattab Laboratory of Natural Products Chemistry and Biomolecules, Chemistry Department, Faculty of Sciences, University of Blida 1, Algeria

\*Address all correspondence to: elhattab@univ-blida.dz

#### IntechOpen

© 2020 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

#### References

[1] Abdul Aziz ZA, Ahmad A, Mohd Setapar SH, Karakucuk A, Azim MM, Lokhat D, et al. Essential oils: Extraction techniques, pharmaceutical and therapeutic potential – A review. Current Drug Metabolism. 2018;**19**:1-12. DOI: 10. 2174/1389200219666180723144850

[2] Hirsch AH, Diederich F. The non-Mevalonate pathway to isoprenoid biosynthesis: A potential source of new drug targets. Chimia. 2008;62(4):
226-230. DOI: 10.2533/chimia.2008.226

[3] Hüsnü Can Baser K, Buchbauer G.Handbook of Essential Oils: ScienceTechnology and Applications. 2nd ed.Boca Raton London: Taylor and Francis;2016

[4] Bakkali F, Averbeck S, Averbeck D, Idaomar M. Biological effects of essential oils—A review. Food and Chemical Toxicology. 2008;**46**:446-475. DOI: 10.1016/j.fct.2007.09.106

[5] Vallet M, Strittmatter M, Murúa P, Lacoste S, Dupont J, Hubas C, et al. Chemically-mediated interactions between macroalgae, their fungal endophytes, and protistan pathogens. Frontiers in Microbiology. 2018;**9**(3161):13. DOI: 10.3389/fmicb. 2018.03161

[6] Cseke LJ, Kaufman PB,
Kirakosyan A. The biology of essential oils in the pollination of flowers.
Natural Product Communications.
2007;2:1317-1336. DOI:
10.1177/1934578X0700201225

[7] Sarkic A, Stappen I. Essential oils and their single compounds in cosmetics—A critical review. Cosmetics. 2018;5(11): 1-21. DOI: 10.3390/cosmetics5010011

[8] Golkar P, Moattar F. Essential oil composition, bioactive compounds, and antioxidant activities in Iberis Amara L. Natural Product Communications. 2019;**40**(5):1-8. DOI: 10.1177/1934578X19846355

[9] Koh K, Pearce A, Marshman G, Finlay-Jones J, Hart P. Tea tree oil reduces histamine-inducedsk in inflammation. The British Journal of Dermatology. 2002;**147**:1212-1217. DOI: 10.1046/ j.1365-2133.2002.05034.x

[10] Maruyama N, Sekimoto Y, Ishibashi H, Inouye S, Oshima H, Yamaguchi H, et al. Suppression of neutrophil accumulation in mice by cutaneous application of geranium essentialoil. Journal of Inflammation. 2005;**2**:1-21. DOI: 10.1186/1476-9255-2-1

[11] Edris AE. Pharmaceutical and therapeutic potentials of essential oils and their individual volatile constituents: A review. Phytotherapy Research. 2007;**21**:308-323. DOI: 10.1002/ptr.2072

[12] Bouchet P. The magnitude of marine biodiversity. In: Duarte CM, editor. The Exploration of Marine Biodiversity.Scientific and Technological Challenges.Bilbao: Fundacion BBVA; 2006.pp. 31-62. ISBN: 84-96515-27-3

[13] Becerra JX, Noge K, Venable DL. Macroevolutionary chemical escalation in an ancient plant-herbivore arms race. Proceedings of the National Academy of Sciences. 2009;**106**(43):18062-18066. DOI: 10.1073/pnas.0904456106

[14] Becerra JX. Synchronous
coadaptation in an ancient case
of herbivory. Proceedings of the
National Academy of Sciences.
2003;100(22):12804-12807. DOI: 10.1073/
pnas.2133013100

[15] Aydogmus Z, Imre S, Ersoy L,
Wray V. Halogenated secondary metabolites from *Laurencia obtusa*.
Natural Product Research. 2004;**18**(1):43-49. DOI: 10.1080/1057563031000122086 [16] Wanke T, Philippus AC,
Zatelli GA, Vieira LFO, Lhullier C,
Falkenberg M. C15 acetogenins from the
Laurencia complex: 50 years of research
An overview. Revista Brasileira de
Farmacognosia. 2015;25:569-587. DOI:
10.1016/j.bjp.2015.07.027

[17] Epifanio RDA, Pinheiro LS, Alves NC. Polyketides from the marine sponge plakortis angulo spiculatus. Journal of the Brazilian Chemical Society. 2005;**16**(6B):1367-1371. DOI: 10.1590/S0103-50532005000800010

[18] Di Costanzo F, Di Dato V, Ianora A, Romano G. Prostaglandins in marine organisms: A review. Marine Drugs. 2019;**17**:428. DOI: 10.3390/md17070428

[19] Imbs AB. Prostaglandins and oxylipins of corals. Russian Journal of Marine Biology. 2011;**37**:325-334. DOI: 10.1134/S1063074011050075

[20] Chandler CJ, Wilts BD, Brodie J, Vignolini S. Structural color in marine algae. Advanced Optical Materials. 2017;5:11. DOI: 10.1002/ adom. 201600646

[21] Pereira L, Neto JM. Marine Algae Biodiversity, Taxonomy, Environmental Assessment, and Biotechnology. Boca Raton: CRC Press Taylor & Francis Group; 2015. p. 397

[22] Van den Hoek C, Mann DG, Jahns HM. Algae. An Introduction to Phycology. Cambridge U.K.: Cambridge Univ. Press; 1995. ISBN: 0521304199 – 0521316871

[23] Silberfeld T, Rousseau F,
de Reviers B. An updated classification of Brown algae (*Ochrophyta*, *Phaeophyceae*). Cryptogamie Algologie.
2014;35(2):117-156. DOI: 10.7872/crya. v35.iss2.2014.117

[24] Fang L, Leliaert F, Zhang Z-H, Penny D, Zhong B-J. Evolution of the Chlorophyta: Insights from chloroplast phylogenomic analyses. Journal of Systematics and Evolution. 2017;55(4):322-332. DOI: 10.1111/ jse.12248

[25] Guiry MD, Guiry GM. AlgaeBase.World-wide Electronic Publication.National University of Ireland, Galway;2017. Available from: http://www.algaebase.org. [Accessed: 20 May 2017]

[26] Gnatta JR, Kurebayashi LFS, Turrini RNT, Silva MJP. Aromatherapy and nursing: Historical and theoretical conception. Revista da Escola de Enfermagem da U.S.P. 2016;**50**(1):127-133. DOI: 10.1590/ S0080-623420160000100017

[27] Aboelsoud NH. Herbal medicine in ancient Egypt. Journal of Medicinal Plant Research. 2010;4(2):82-86. DOI: 10.5897/JMPR09.013

[28] Farag RS, Daw ZY, Hewedi FM, El-Baroty GSA. Antimicrobial activity of some Egyptian spice essential oils. Journal of Food Protection. 1989;**52**(9):665-667. DOI: 10.4315/0362-028X-52.9.665

[29] Viuda-Martos M, Mohamady M, Fernández-López J, El Razik KA, Omer E, Pérez-Alvarez J, et al. In vitro antioxidant and antibacterial activities of essentials oils obtained from Egyptian aromatic plants. Food Control. 2011;**22**:1715-1722. DOI: 10.1016/j.food cont.2011.04.003

[30] Heavisides E, Rouger C, Reichel AF, Ulrich C, Wenzel-Storjohann A, Sebens S, et al. Seasonal variations in the Metabolome and bioactivity profile of *Fucus vesiculosus* extracted by an optimised, pressurised liquid extraction protocol. Marine Drugs. 2018;**16**:503. DOI: 10.3390/md16120503

[31] Battley EH. Advances in microbial ecology. Volume 11. K. C. Marshall. The Quarterly Review of Biology. 1992;**67**(2):209. DOI: 10.1086/417584

[32] Katayama T. Volatile Constituents. In: Lewin RA, editor. Physiology and Biochemistry of Algae. New York: American Press; 1962. pp. 467-472

[33] Moore RE. Volatile compounds from marine algae. Accounts of Chemical Research. 1977;**10**:40-47. DOI: 10.1021/ ar50110a002

[34] Dudavera N, Negre F, Nagegowda DA, Orlova I. Plant volatiles: Recent advances and future perspectives. Critical Reviews in Plant Sciences. 2006;**25**:417-440. DOI: 10.1080/07352680600899973

[35] Wiesemeier T, Hay M, Pohnert G. The potential role of wound activated volatile release in the chemical defence of the brown alga *Dictyota dichotoma*: Blend recognition by marine herbivores. Aquatic Sciences. 2007;**69**:403-412. DOI: 10.1007/s00027-007-0889-y

[36] Kajiwara T, Akakabe Y, Matsui K, Kodama K, Koga H, NagakuraT. (+)-(3S, 4S)-3-butyl-4-vinylcyclopentene in brown algae of the genus Dictyopteris. Phytochemistry. 1997;45(3):529-532. DOI: 10.1016/S0031-9422(96)00884-9

[37] Sugisawa H, Nakamura K, Tamura H. The aroma profile of the volatiles in marine green algae (*Ulva pertusa*). Food Reviews International. 1990;**6**(4):573-589. DOI: 10.1080/87559129009540893

[38] Kajiwara T, Hatanaka A, Tanaka Y, Kawai T, Ishihara M, Tsuneya T, et al. Specificity of the enzyme system producing long chain aldehydes in the green alga, *Ulva pertusa*. Phytochemistry. 1989;**28**(2):636-639. DOI: 10.1016/0031-9422(89)80070-6

[39] Kawasaki W, Matsui K, Akakabe Y,
Itai N, Kajiwara T. Volatiles from *Zostera marina*. Phytochemistry.
1998;47(1):27-29. DOI: 10.1016/ S0031-9422(97)88555-X [40] Ozdemir G, Horzum Z, Sukatar A, Karabay-Yavasoglu N. Antimicrobial activities of volatile components and various extracts of Dictyopteris membranaceae and *Cystoseira barbata* from the coast of Izmir, Turkey. Pharmaceutical Biology. 2006;**44**(3):183-188. DOI: 10.1080/13880200600685949

[41] Giogios I, Grigorakis K, Nengas I, Papasolomontos S, Papaioannoub N, Alexis MN. Fatty acid composition and volatile compounds of selected marine oils and meals. Journal of the Science of Food and Agriculture. 2009;**89**:88-100. DOI: 10.1002/jsfa.3414

[42] El Hattab M, Culioli G, Piovetti L, Chitour SE, Valls R. Comparison of various extraction methods for identification and determination of volatile metabolites from the brown alga *Dictyopteris membranacea*. Journal of Chromatography. A. 2007;**1143**(1-2):1-7. DOI: 10.1016/j.chroma. 2006.12.057

[43] De Carvalho LR, Roque NF.
Fenóishalogenados e/ou sulfatados de macroalgasmarinhas. Quimica Nova.
2000;23(6):757-764. DOI: 10.1590/ s0100-40422000000600009

[44] Fleury BG, Kelecom A, Pereira RC, Teixeira VL. Polyphenols, terpenes and sterols in Brazilian dictyotales and fucales (*Phaeophyta*). Botanica Marina. 1994;**37**(5):457-462. DOI: 10.1515/ botm.1994.37.5.457

[45] Kladi M, Vagias C, Roussis V. Volatile halogenated metabolites from marine red algae. Phytochemistry Reviews. 2004;**3**(3):337-366. DOI: 10.1515/ botm.1994.37.5.457

[46] Roller P, Kalfred A, Moore RE. Isolation of S-(3-oxoundecyl) thioacetate, bis-(3-oxoundecyl) disulphide, (–)-3-hexyl-4,5dithiacycloheptanone, and S-(trans-3-oxoundec-4-enyl) thioacetate from Dictyopteris. Chemical Communications. 1971;**273**:503-504. DOI: 10.1039/C29710000503 [47] Schnitzler I, Boland W, Hay ME. Organic sulfur compounds from Dictyopteris spp. deter feeding by an herbivorous amphipod (*Ampithoe longimana*) but not by an herbivorous sea urchin (*Arbacia punctulata*). Journal of Chemical Ecology. 1998;**24**(10):1715-1732. DOI: 10.1023/ A:1020876830580

[48] Boland W, Müller DG. On the odor of the Mediterranean seaweed *Dictyopteris membranacea*: New C11 hydrocarbons from marine brown algae. Tetrahedron Letters. 1987;**28**(3):307-310. DOI: 10.1016/ S0040-4039(00)95714-9

[49] Mouritsen OG. Seaweeds Edible, Available and Sustainable. London: The University of Chicago Press, Ltd.; 2013

[50] Haas P. The liberation of methyl sulphide by seaweed. The Biochemical Journal. 1935;**29**:1297-1299. DOI: 10.1042/ bj0291297

[51] Challenger F, Simpson MI. Studies on biological methylation. Part XII. A precursor of the dimethyl sulphide evolved by *Polysiphonia fastigiata*. Dimethyl-2-carboxyethylsulphonium hydroxide and its salts. Journal of the Chemical Society. 1948;**1948**:1591-1597. DOI: 10.1039/JR9480001591

[52] Karabay-Yavasoglu NU, Sukatar A, Ozdemir G, Horzum Z. Antimicrobial activity of volatile components and various extracts of the red alga. Janiarubens. Phytotherapy Research. 2007;**21**:153-156. DOI: 10.1002/ptr.2045

[53] Yamamoto M, Baldermann S, Yoshikawa K, Fujita A, Mase N, Watanabe N. Determination of volatile compounds in four commercial samples of Japanese green algae using solid phase microextraction gas chromatography mass spectrometry. Scientific World Journal. 2014;**2014**:1-8. DOI: 10.1155/2014/289780

[54] Terezinha M, Neta SL, Narain N. Volatile components in seaweeds. Marine Biology and Oceanography. 2018;**2**(2):195-201. DOI: 10.31031/ EIMBO.2018.02.000535

[55] Kajiwara T, Yoshikawa H, Matsui K, Hatanaka A, Kawai T, Ishihara T, et al.
Specificity of the enzyme system producing long chain aldehydes in the green alga, *Ulva pertusa*.
Phytochemistry. 1989;28(2):636-639.
DOI: 10.1016/0031-9422(89)80070-6

[56] Beauchêne D, Grua-Priol J, Lamer T, Demaimay M, Quémeneur F. Concentration by pervaporation of aroma compounds from *Fucus serratus*. Journal of Chemical Technology and Biotechnology. 2000;**75**(6): 451-458. DOI: 10.1002/1097-4660(200006)75:6<451: AID-JCTB231>3.0.CO;2-U

[57] Le Pape MA, Grua-Priol J, Prost C, Demaimay M. Optimization of dynamic headspace extraction of the edible red algae *Palmaria palmata* and identification of the volatile components. Journal of Agricultural and Food Chemistry. 2004;**52**(3):550-556. DOI: 10.1021/jf030478x

[58] Boonprab K, Matsui K, Akakabe Y, Yotsukura N, Kajiwara T. Hydroperoxyarachidonic acid mediated n-hexanal and (Z)-3- and (E)-2-nonenal formation in *Laminaria angustata*. Phytochemistry. 2003;**63**(6):669-678. DOI: 10.1016/S0031-9422(03)00026-8

[59] Boonprab K, Matsui K, Akakabe Y, Yoshida M, Yotsukura N, Chirapart A, et al. Formation of aldehyde flavor (n-hexanal, 3Z-nonenal and 2E-nonenal) in the brown alga, *Laminaria angustata*. Journal of Applied Phycology. 2006;**18**(3-5):409-412. DOI: 10.1007/s10811-006-9038-6

[60] Akakabe Y, Matsui K, Kajiwara T. 2,4-Decadienals are produced via (R)-11-HPITE from arachidonic acid in marine green alga *Ulva conglobata*. Bioorganic & Medicinal Chemistry.

2003;**11**:3607-3609. DOI: 10.1016/ S0968-0896(03)00364-X

[61] Akakabe Y, Kajiwara T. Bioactive volatile compounds from marine algae: Feeding attractants. Journal of Applied Phycology. 2008;**20**:661-664. DOI: 10.1007/978-1-4020-9619-8\_26

[62] Akabe Y, Matsui K, Kajiwara T.
Enantioselective α-hydroperoxylation of long chain fatty acids which crude enzyme of marine green alga *Ulva pertusa*. Tetrahedron Letters.
1999;40:1137-1140. DOI: 10.1016/S0040-4039(98)02547-7

[63] Akakabe Y, Matsui K, Kajiwara T.  $\alpha$ -Oxidation of long-chain unsaturated fatty acids in the marine green alga *Ulva pertusa*. Bioscience, Biotechnology, and Biochemistry. 2000;**64**:2680-2681. DOI: 10.1271/bbb.64.2680

[64] Akakabe Y, Matsui K, Kajiwara T. Enantioselective 2- hydroperoxylation of long-chain fatty acids in marine green algae. Fisheries Science. 2001;**67**:328-332. DOI: 10.1046/j.1444-2906.2001.00235.x

[65] Vilara EG, O'Sullivan MG, Kerry JP, Kilcawleya KN. Volatile compounds of six species of edible seaweed: A review. Algal Research. 2020;**45**:101740. DOI: 10.1016/j.algal.2019.101740

[66] Firouzi J, Gohari A, Rustaiyan A, Larijani K, Saeidnia S. Composition of the essential oil of *Nizamuddin zanardinii*, a Brown alga collected from Oman gulf. Journal of Essential Oil Bearing Plants. 2013;**16**(5):689-692. DOI: 10.1080/0972060X.2013.862072

[67] Püttmann W. Thermodesorptiongas chromatography-mass spectrometric analysis of biological materials for potential molecular precursors of the constituents of the crude oils. Journal of Chromatography. 1991;**552**:325-336. DOI: 10.1016/S0021-9673(01)95949-7

[68] Steiner M, Hartmann T. The occurrence and distribution of volatile

amines in marine algae. Planta. 1968;**79**(2):113-121. DOI: 10.1007/ BF00390154

[69] Percot A, Yalçin A, Aysel V,
Erdugan H, Dural B, Guven KC.
β-Phenylethylamine content in marine algae around Turkish coasts. Botanica
Marina. 2009;52:87-90. DOI: 10.1515/
BOT.2009.031

[70] Barwell CJ. Pharmacologicallyactive amines in some marine algae and algal food products. Journal of Home & Consumer Horticulture. 2008;**1**(1): 77-82. DOI: 10.1300/J280v01n01\_04

[71] Takaoka M, Ando Y. Essential oil of seaweeds. I. Composition of the oil of *Dictyopteris divaricata*. Nippon Kagaku Kaishi (1921-1957). 1951;**72**:999-1003. DOI: 10.1246/nikkashi1948.72.999

[72] Flodin C, Whitfield F.
4-Hydroxybenzoic acid: A likely precursor of 2,4,6-tribromophenol in *Ulva lactuca*. Phytochemistry.
1999;**51**(2):249-255. DOI: 10.1016/S0031-9422(98)00754-7

[73] Flodin C, Whitfield F. Biosynthesis of bromophenols in marine algae.
Water Science and Technology.
1999;40(6):53-58. DOI: 10.1016/
S0273-1223(99)00537-5

[74] Marshall RA, Hamilton JTG, Dring MJ, Harper DB. Do vesicle cells of the red alga Asparagopsis (Falkenbergiastage) play a role in bromocarbon production? Chemosphere. 2003;**52**:471. DOI: 10.1016/S0045-6535(03)00197-8

[75] Garson MJ. The biosynthesis of marine natural products. Chemical Reviews. 1993;**93**:1699-1733. DOI: 10.1021/cr00021a003

[76] Moore BS. Biosynthesis of marine natural products: Microorganisms (part a). Natural Product Reports.
2005;22:580-593. DOI: 10.1039/ b404737k [77] Moore BS. Biosynthesis of marine natural products: Macroorganisms
(part B). Natural Product Reports.
2006;23:615-629. DOI: 10.1039/ b508781n

[78] Le Blanc C, Colin C, Cosse A, Delage L, La Barre S, Morin P, et al. Biochimie. 2006;**88**:1773-1785. DOI: 10.1016/j.biochi.2006.09.001

[79] Moore RM. Methyl halide production and loss rates in sea water from field incubation experiments. Marine Chemistry. 2006;**101**:213-219. DOI: 10.1016/j.marchem.2006.03.003

[80] Giese B, Laturnus F, Adams FC, Wiencke C. Release of volatile iodinated C1–C4 hydrocarbons by marine macroalgae from various climate zones. Environmental Science & Technology. 1999;**33**(14):2432-2439. DOI: 10.1021/ es980731n

[81] Carpenter LJ, Malin G, Liss PS, Kupper FC. Novel biogenic iodinecontaining trihalomethanes and other short-lived halocarbons in the coastal East Atlantic. Global Biogeochemical Cycles. 2000;**14**:1191-1204. DOI: 10.1029/2000GB001257

[82] Michelozzi M. Defensive roles of terpenoid mixtures in conifers. Acta Botanica Gallica. 1999;**146**(1):73-84. DOI: 10.1080/ 12538078.1999.10515803

[83] Kumari S, Pundhir S, Priya P, Jeena G, Punetha A, Chawla K, et al. EssOilDB: A database of essential oils reflecting terpene composition and variability in the plant kingdom. Database. 2014;**2014**:1-14. DOI: 10.1093/ database/bau120

[84] Naylor S, Hanke FJ, Manes LV, Crews P. Chemical and biological aspects of marine monoterpenes. Fortschritte der Chemie Organischer Naturstoffe. 1983;**44**:189-241. DOI: 10.2307/2107215

[85] Marmulla R, Harder J. Microbial monoterpene transformations–a review.

Frontiers in Microbiology. 2014;**5**:346-359. DOI: 10.3389 /fmicb.2014.00346

[86] Soares AR. Extraction, isolation, and identification of Sesquiterpenes from Laurencia species. In: Natural Products from Marine Algae Methods and Protocols. Vol. 1308. New York: Humana Press; 2015. pp. 225-240

[87] Ji NY, Li XM, Li K, Ding LP, Gloer JB, Wang BG. Diterpenes, sesquiterpenes, and a C15-acetogenin from the marine red alga *Laurencia mariannensis*. Journal of Natural Products. 2007;**70**(12):1901-1905. DOI: 10.1021/np070378b

[88] Jerković I, Marijanović Z, Roje M, Kuś PM, Jokić S, ČozÏ-Rakovac R. Phytochemical study of the headspace volatile organic compounds of fresh algae and seagrass from the Adriatic Sea (single point collection). PLoS One. 2018;**13**(5):1-13. DOI: 10.1371/journal. pone.0196462

[89] Kamenarska Z, Ivanova A, Stancheva R, Stoyneva M, Stefanov K, Dimitrova-Konaklieva S, et al. Volatile compounds from some Black Sea red algae and their chemotaxonomic application. Botanica Marina. 2006;**49**(1):47-56. DOI: 10.1515/ BOT.2006.006

[90] Elenkov I, Georgieva T,
Hadjieva P, Dimitrova-Konaklieva S,
Popov S. Terpenoids and sterols in *Cladophora vagabunda*. Phytochemistry.
1995;**38**(2):457-459. DOI:
10.1016/0031-9422(94)00704-W

[91] Yamada K, Tan H, Tatematsu H, Ojiva M. Dictyoprolene and neodictyoprolene, two new odoriferous compounds from the brown alga *Dictyopteris prolifera*: Structures and synthesis. Tetrahedron. 1986;**42**(14):3775-3780. DOI: 10.1016/ S0040-4020(01)87531-1

[92] Kajiwara T, Kashibe M, Matsui K, Hatanaka A. Volatile compounds and

long-chain aldehydes formation in conchocelis filaments of a red alga, *Porphyra tenera*. Phytochemistry. 1990;**29**(7):2193-2195. DOI: 10.1016/0031-9422(90)83036-Z

[93] Fujimura T, Kawai T, Shiga M, Kajiwara T, Hatanaka A. Long-chain aldehyde production in thalli culture of the marine green alga *Ulva pertusa*. Phytochemistry. 1990;**29**(3):745-747. DOI: 10.1016/0031-9422(90)80011-5

[94] Maier I, Muller DG. Sexual pheromones in algae. The Biological Bulletin. 1986;**170**:145-175. DOI: 10.2307/1541801

[95] Moore RE. Volatiles compounds from marine algae. Accounts of Chemical Research. 1977;**10**:40-47. DOI: 10.1021/ar50110a002

[96] Boland W. The chemistry of gamete attraction: Chemical structures, biosynthesis, and abiotic degradation. Proceedings of the National Academy of Sciences of the United States of America. 1995;**92**:37-43. DOI: 10.1073/pnas.92.1.37

[97] Pohnert G, Boland W. The oxylipin chemistry of attraction and defense in brown algae and diatoms. Natural Product Reports. 2002;**19**:108-122. DOI: 10.1039/a806888g

[98] Fink P. Ecological functions of volatile organic compounds in aquatic systems. Marine and Freshwater Behaviour and Physiology. 2007;**40**:155-168. DOI: 10.1080/10236240701602218

[99] Amsler CD, Fairhead VA. Defensive and sensory chemical ecology of brown algae. Advances in Botanical Research. 2006;**43**:1-91. DOI: 10.1016/ S0065-2296(05)43001-3

[100] Derenbach JB, Pesandeo D. Investigations into a small fraction of volatile hydrocarbons: III. Two diatom cultures produce ectocarpene, a pheromone of brown algae. Marine Chemistry. 1986;**19**:337-432. DOI: 10.1016/0304-4203(86)90054-X

[101] Juttner F, Wurster K. Evidence of ectocarpene and dictyopterenes a and C in the water of a freshwater lake. Limnology and Oceanography. 1984;**29**:1322-1324. DOI: 10.4319/ lo.1984.29.6.1322

[102] Boland W, Jaenicke L, Brauner A. Vinyl olefines and sesquiterpenes in the root oil of *Senecio isatideus*. Zeitschrift für Naturforschung. 1982;**37C**:5-9. DOI: 10.1515/znc-1982-1-202

[103] Wang Y, Li X, Jiang Q, Sun H, Jiang J, Chen S, et al. GC-MS analysis of the volatile constituents in the leaves of 14 composite plants. Molecules. 2018;**23**:166. DOI: 10.3390/ molecules23010166

[104] Faulkner J. Interesting aspects of marine natural products chemistry. Tetrahedron. 1977;**33**:1421-1443. DOI: 10.1016/0040-4020(77)88001-0

[105] Hay ME, Duffy JE, Fenical W, Gustaíson K. Chemical defense in the seaweed *Dictyopteris delicatula*: Differential effects against reef fishes and amphipods. Marine Ecology Progress Series. 1988;**48**:185-192. DOI: 10.3354/meps048185

[106] Riad N, Zahi MR, Trovato E, Bouzidi N, Daghbouche Y, Utczas M, et al. Chemical screening and antibacterial activity of essential oil and volatile fraction of *Dictyopteris polypodioides*. Microchemical Journal. 2020;**152**:104415. DOI: 10.1016/j. microc.2019.104415

[107] Moore RE. Chemotaxis and the odor of seaweed. Lloydia.
1976;**39**:181-190. DOI: 10.1016/ B978-0-12-505050-0.50014-8

[108] Blunt JW, Copp BR, Munro MHG, Northcote PT, Prinsep MR. Marine natural products. Natural Product Reports. 2003;**20**:1-48. DOI: 10.1039/ b207130b

[109] Faulkner DJ. Marine naturalproducts. Natural Product Reports.2001;18:1-49. DOI: 10.1039/B009029H

[110] Li XD, Ding W, Miao FP, Ji NY. Halogenated chamigrane sesquiterpenes from *Laurencia okamurae*. Magnetic Resonance in Chemistry. 2012;**50**(2): 174-177. DOI: 10.1002/mrc.2870

[111] Kim SK. Handbook of Marine
Macroalgae Biotechnology and
Applied Phycology, 1st ed. Chichester,
West Sussex: Wiley-Blackwell John
Wiley& Sons; 2012. p. 592. DOI:
10.1002/9781119977087

[112] Coll JC, Wright AD. Tropical marine algae. I. New halogenated monoterpenes from Chondrococus hornemannii (Rhodophyta, Gigartinales, Rhizophyllidaceae). Australian Journal of Chemistry. 1987;**40**:1893-1900. DOI: 10.1071/CH9871893

[113] Cikoš MA, Jurin M, Rakovac RC,
Jokic S, Jerkovic I. Update on monoterpenes from red macroalgae:
Isolation, analysis, and bioactivity.
Marine Drugs. 2019;17:537. DOI:
10.3390/md17090537

[114] Crews P, Ng P, Kho-Wiseman E, Pace C. Halogenated monoterpenes of the red alga Microcladia.
Phytochemistry. 1976;15:1707-1711.
DOI: 10.1016/S0031-9422(00)97461-2

[115] Wang BW, Gloer JB, Ji N-Y, Zhao J-C. Halogenated organic molecules of Rhodomelaceae origin: Chemistry and biology. Chemical Reviews. 2013;**113**:3632-3685. DOI: 10.1021/ cr9002215

[116] Cabrita MT, Vale C, Pilar RA. Halogenated compounds from marine algae. Marine Drugs. 2010;**8**:2301-2317. DOI: 10.3390/md8082301 [117] Kamada T, Phan CS, Vivian Shi-Ting Sien VST, Vairappan CS. Halogenated chamigrane sesquiterpenes from *Bornean Laurencia* majuscula. Journal of Applied Phycology. 2018;**30**:3373-3378. DOI: 10.1007/s10811-018-1452-z

[118] Gao D, Okuda R. Supercritical fluid extraction of halogenated monoterpenes from the red alga *Plocamium cartilagineum*. Journal of AOAC International. 2001;**84**:1313-1331. PMID: 11601448

[119] San-Martin A, Negrete R, Rovirosa J. Insecticide and acaricide activities of polyhalogenated monoterpenes from Chilean *Plocamium cartilagineum*. Phytochemistry.
1991;**30**:2165-2169. DOI:
10.1016/0031-9422(91)83607-M

[120] Wright AD, König GM, Sticher O.
Five new monoterpenes from the marine red alga *Portieria hornemannii*.
Tetrahedron. 1991;47:5717-5724. DOI: 10.1016/S0040-4020(01)86524-8

[121] Paul VJ, McConnell OJ, Fenical W.
Cyclic monoterpenoid feeding deterrents from the red marine alga *Ochtodes crockeri*. The Journal of Organic Chemistry. 1980;45:3401-3407. DOI: 10.1021/jo01305a006

[122] Wise ML, Rorrer GL, Polzin JJ, Croteau R. Biosynthesis of marine natural products: Isolation and characterization of a myrcene synthase from cultured tissues of the marine red alga *Ochtodes secundiramea*. Archives of Biochemistry and Biophysics. 2002;**400**(1):125-132. DOI: 10.1006/abbi.2002.2780

[123] Suzuki M, Kurosawa E, Irie T. Spirolaurenone, a new sesquiterpenoid containing bromine from *Laurencia glandulifera* Kützing. Tetrahedron Letters. 1970;**11**:4995-4998. DOI: 10.1016/S0040-4039 (00)89329-6

[124] Suzuki M, Kurosawa E, Irie T. Three new sesquiterpenoids containing

bromine, minor constituents of *Laurencia glandulifera* Kützing. Tetrahedron Letters. 1974;**15**:821-824. DOI: 10.1016/S0040-4039(01)82342-X

[125] Konig M, Wright D. New C15 Acetogenins and Sesquiterpenes from the red alga Laurencia sp. cf. *L. gracilis*. Journal of Natural Products. 1994;**57**:477. DOI: 10.1021/np50106a006

[126] Vazquez JT, Chang M, Nakanishi K, Martin JD, Martin VS, Perez R. Puertitols: Novel sesquiterpenes from *Laurencia obtusa*. Structure elucidation and absolute configuration and conformation based on circular dichroism. Journal of Natural Products. 1988;**51**(6):1257-1260. DOI: 10.1021/ np50060a036

[127] North M, Fernandez JJ, Padilla A.
Bisabolane halogenated sesquiterpenes from Laurencia. Phytochemistry.
1992;**31**:326-327. DOI: 10.1016/ 0031-9422(91)83065-S

[128] Sims JJ, Lin GHY, Wing RM. Marine natural products X elatol, a halogenated sesquiterpene alcohol from the red alga *Laurencia elata*. Tetrahedron Letters. 1974;**15**:3487-3490. DOI: 10.1016/S0040-4039(01)91944-6

[129] Lhullier C, Donnangelo A, Caro M, Palermo JA, Horta PA. Isolation of elatol from *Laurencia microcladia* and its palatability to the sea urchin Echinometralucunter. Biochemical Systematics and Ecology. 2009;**37**:254-259. DOI: 10.1016/j.bse.2009.04.004

[130] Al-Massarani SM. Phytochemical and biological properties of Sesquiterpene constituents from the marine red seaweed Laurencia: A review. Natural Products Chemistry and Research. 2014;**2**(5):1-13. DOI: 10.4172/2329-6836.1000147

[131] Dembitsky VM, Tolstikov GA. Natural halogenated sesquiterpenes from marine organisms. Chemistry for Sustainable Development. 2004;**12**:1-12 [132] Hay ME, Piel J, Boland W, Schnitzler I. Seaweed sex pheromones and their degradation products frequently suppress amphipod feeding but rarely suppress sea urchin feeding. Chemoecology. 1998;8(2):91-98. DOI: 10.1007/PL00001809

[133] Pelletreau K, Muller-Parker G. Sulfuric acid in the phaeophyte alga *Desmarestia mundadeter* feeding by the sea urchin *Strongylocentrotus droebachiensis*. Marine Biology. 2002;**141**:1-9. DOI: 10.1007/s00227-002-0809-6

[134] Bakus GJ, Targett NM, Schulte B. Chemical ecology of marine organisms: An overview. Journal of Chemical Ecology. 1986;**12**:951-987. DOI: 10.1007/ BF01638991

[135] Muller DG, Jaenicke L, Donike M, Akintobi T. Sex attractant in a brown alga - chemical structure. Science.
1971;171:1132. DOI: 10.1126/science.
171.3976.1132

[136] Ponhert G, Boland W. Pericyclic reactions in nature: Synthesis and cope rearrangement of thermolabile bis-alkenylcyclopropanes from female gametes of marine brown algae (Phaeophyceae). Tetrahedron Letters. 1997;**53**(40):13681-13694. DOI: 10.1016/ S0040-4020(97)00886-7

[137] Schnitzler I, Pohnert G, Hay ME, Boland W. Chemical defense of brown algae (Dictyopteris spp.) against the herbivorous amphipod Ampithoelongimana. Oecologia. 2001;**126**(4):515-521. DOI: 10.1007/ s004420000546

[138] Valls R, Artaud J, Amade P,
Vincente N, Piovetti L. Determination of caulerpenyne, a toxin from the green alga *Caulerpa taxifolia* (Caulerpaceae).
Journal of Chromatography.
A. 1994;663(1):114-118. DOI: 10.1016/0021-9673(94)80502-4

[139] Dumay O, Pergent G, Pergent-Martini C, Amade P. Variations in caulerpenyne contents in *Caulerpa taxifolia* and *Caulerpa racemosa*. Journal of Chemical Ecology. 2002;**28**(2):343-352. DOI: 10.1023/a:1017938225559

[140] Paul VJ, Fenical W. Chemical defense in tropical green algae, order Caulerpales. Marine Ecology Progress Series. 1986;**34**:157-169. DOI: 10.3354/ meps034157

[141] Goodwin KD, North WJ, Lidstrom ME. Production of bromoform and dibromomethane by Giant kelp: Factors affecting release and comparison to anthropogenic bromine sources. Limnology and Oceanography. 1997;**42**(8):1725-1734. DOI: 10.4319/ lo.1997.42.8.1725

[142] Dworjanyn SA, De Nys R, Steinberg PD. Localisation and surface quantification of secondary metabolites in the red alga *Delisea pulchra*. Marine Biology. 1999;**133**:727-736. DOI: 10.1007/s002270050514

[143] Scheuer PJ. Some marineecological phenomena: Chemical basisand biomedical potential. Science.1990;248:173-177. DOI: 10.1126/science.2183350

[144] Kamenarska Z, Gasic MJ, Zlatovic M, Rasovic A, Sladic D, Kljajic Z, et al. Chemical composition of the brown alga *Padina pavonia* (L.) Gaill. From the Adriatic Sea. Botanica Marina. 2002;**45**:339-345. DOI: 10.1515/ BOT.2002.034

[145] Sultana V, Ehteshamul-H S, Ara J, Athar M. Comparative efficacy of brown, green and red seaweeds in the control of root infecting fungi and okra. International Journal of Environmental Science and Technology. 2005;2(2):129-132. DOI: 10.1007/BF03325866

[146] Ktari L, Guyot M. A cytotoxic oxysterol from the marine Red Sea alga

*Padina pavonica* (L.) Thivy. Journal of Applied Phycology. 1990;**11**:511-513. DOI: 10.1023/A:1008162624027

[147] Awad NE, Selim MA, Metawe HM, Matloub AA. Cytotoxic xenicane diterpenes from the brown alga *Padina pavonia* (L.) Gaill. Phytotherapy Research. 2008;**22**:1610-1613. DOI: 10.1002/ptr.2532

[148] Awad NE, Motawe HM, Selim MA, Matloub AA. Volatile constituents of the brown algae *Padina pavonia* (L.) Gaill. And *Hydroclathrus clathratus* (C. Agardh) Howe and their antimicrobial activity. Medicinal and Aromatic Plant Science and Biotechnology. 2009;**3**(1):12-15

[149] Dias T, Brito I, Moujir L, Paiz N, Darias J, Cueto M. Cytotoxic sesquiterpenes from *Aplysia dactylomela*. Journal of Natural Products. 2005;**68**:1677-1679. DOI: 10.1021/ np050240y

[150] Juagdan EG, Kalidindi R,
Scheuer P. Two new chamigranes from an hawaiian red alga, *Laurencia cartilaginea*. Tetrahedron.
1997;53(2):521-528. DOI: 10.1016/ S0040-4020(96)01002-2

[151] Kladi M, Vagias C, Furnari G, MoreauD, RoussakisC, RoussisV. Cytotoxic cuparene sesquiterpenes from *Laurencia microcladia*. Tetrahedron Letters. 2005; **46**:5723-5726. DOI: 10.1016/j. tetlet.2005.06.076

[152] Kladi M, Vagias C, Papazafir P, Furnari G, Serio D, Roussis V. New sesquiterpenes from the red alga *Laurencia microcladia*. Tetrahedron. 2007;**63**:7606-7611. DOI: 10.1016/j. tet.2007.05.051

[153] Vairappan CS. Potent antibacterial activity of halogenated metabolites from Malaysian red algae, *Laurencia majuscula* (Rhodomelaceae, Ceramiales). Biomolecular Engineering.

2003;**20**:255-259. DOI: 10.1016/ S1389-0344(03)00067-4

[154] Kurata K, Amiya T. Bis(2,3,6tribromo-4,5-dihydroxybenzyl)ether from the red alga, *Symphyocladia latiuscula*. Phytochemistry. 1980;**19**:141-142. DOI: 10.1016/0031-9422(80)85032-1

[155] Kimura J, Kamada N, Tsujimoto Y. Fourteen Chamigrane derivatives from a red alga, *Laurencia nidifica*. Bulletin of the Chemical Society of Japan. 1999;**72**:289-292. DOI: 10.1246/ bcsj.72.289

