

Utah State University

DigitalCommons@USU

Ecology Center Publications

Ecology Center

7-29-2020

What Is the Storage Effect, Why Should It Occur in Cancers, and How Can It Inform Cancer Therapy?

Anna K. Miller

H. Lee Moffitt Cancer Center and Research Institute

Joel S. Brown

H. Lee Moffitt Cancer Center and Research Institute

David Basanta

H. Lee Moffitt Cancer Center and Research Institute

Nancy Huntly

Utah State University

Follow this and additional works at: https://digitalcommons.usu.edu/eco_pubs

 Part of the [Ecology and Evolutionary Biology Commons](#)

Recommended Citation

Miller, A. K., Brown, J. S., Basanta, D., & Huntly, N. (2020). What Is the Storage Effect, Why Should It Occur in Cancers, and How Can It Inform Cancer Therapy? *Cancer Control*. <https://doi.org/10.1177/1073274820941968>

This Article is brought to you for free and open access by the Ecology Center at DigitalCommons@USU. It has been accepted for inclusion in Ecology Center Publications by an authorized administrator of DigitalCommons@USU. For more information, please contact digitalcommons@usu.edu.



What Is the Storage Effect, Why Should It Occur in Cancers, and How Can It Inform Cancer Therapy?

Cancer Control
Volume 27: 1-9
© The Author(s) 2020
Article reuse guidelines:
sagepub.com/journals-permissions
DOI: 10.1177/1073274820941968
journals.sagepub.com/home/ccx



Anna K. Miller, PhD¹ , Joel S. Brown, PhD¹, David Basanta, PhD¹, and Nancy Huntly, PhD²

Abstract

Intratumor heterogeneity is a feature of cancer that is associated with progression, treatment resistance, and recurrence. However, the mechanisms that allow diverse cancer cell lineages to coexist remain poorly understood. The storage effect is a coexistence mechanism that has been proposed to explain the diversity of a variety of ecological communities, including coral reef fish, plankton, and desert annual plants. Three ingredients are required for there to be a storage effect: (1) temporal variability in the environment, (2) buffered population growth, and (3) species-specific environmental responses. In this article, we argue that these conditions are observed in cancers and that it is likely that the storage effect contributes to intratumor diversity. Data that show the temporal variation within the tumor microenvironment are needed to quantify how cancer cells respond to fluctuations in the tumor microenvironment and what impact this has on interactions among cancer cell types. The presence of a storage effect within a patient's tumors could have a substantial impact on how we understand and treat cancer.

Keywords

storage effect, cancer, heterogeneity, coexistence, dormancy, quiescence, fluctuating environments

Received March 5, 2020. Received revised June 12, 2020. Accepted for publication June 19, 2020.

Introduction

Ecosystems in nature include coexisting species that compete for space, resources, and safety from predators.¹⁻⁴ Similarly, tumors exhibit microenvironmental heterogeneity that contains coexisting cancer cell types that experience variations in nutrients, toxic metabolites, and diverse types of normal cells.⁵ In nature, mechanisms of coexistence can explain the diversity of species within a community. Typical mechanisms include food-safety trade-offs (one species is the better competitor, while the other is better at avoiding predation), diet separation (each species has a subset of resources on which it is the more successful consumer), habitat selection (each species has a habitat within which it is more successful), and competition-colonization trade-offs (one species slowly outcompetes the other at a given spot, while the other is more successful at dispersing to unoccupied spots). Similar mechanisms likely explain some of the diversity of cancer cell types within and among a patient's tumors.⁶

Long underappreciated in ecology was a mechanism of coexistence now known as the *storage effect*.⁷⁻¹¹ Three ingredients can promote the coexistence of species by the storage effect: (1) temporal variability in environmental conditions that include periods favorable and unfavorable for survival and reproduction (we shall refer to these as good and bad), (2) the presence of 2 life-history states within each species (one

¹ Department of Integrated Mathematical Oncology, H. Lee Moffitt Cancer Center and Research Institute, Tampa, FL, USA

² Ecology Center & Department of Biology, Utah State University, Logan, UT, USA

Corresponding Author:

Anna K. Miller, Department of Integrated Mathematical Oncology, H. Lee Moffitt Cancer Center and Research Institute, USF Magnolia Drive, Tampa, FL 33612, USA.

Email: anna.miller@moffitt.org



conductive to proliferation during good periods, the other conducive to survival during bad periods), and (3) differences in how species perceive and invest effort to grow, or do not, in good and bad periods, reflecting some underlying trade-offs and adaptation to somewhat different environmental conditions.

Here, we posit that the storage effect may promote the coexistence of at least some cancer cell types within the tumors of some or many different cancers. To demonstrate the plausibility of the storage effect, we discuss how, as in ecosystems in nature, conditions (1) and (2) are universal properties of most tumor microenvironments. Next, we discuss why there may be trade-offs in the way cancer cells experience good and bad periods. We then discuss how the storage effect manifests in nature, followed by a simple mathematical model for cancer tumor cells, illustrating how the storage effect works. We conclude with a discussion of how knowledge of the presence of a storage effect in a patient's disease might inform therapies.

Temporal Variation in Nature and the Tumor Microenvironment

Very few, if any, ecological communities experience temporally constant environments. Migratory birds escape the harsh winter conditions of higher latitudes by migrating to destinations closer to the equator. Deciduous trees lose their leaves during dry and/or cold seasons. Year-to-year variation in temperature and precipitation may portend droughts or floods and hot spells and cold snaps. Fire, disease, or pestilence occurs episodically within ecosystems. A beaver damming a stream can raise water tables, drown surrounding terrestrial vegetation, and create a wetland. This wetland can revert when the beaver dam breaks or decays. Virtually, all living organisms experience predictable and unpredictable variability in environmental conditions that influence growth, reproduction, and survivorship.

Like natural ecosystems, cancer cells inhabiting a tumor microenvironment experience both regular and irregular temporal fluctuations. Blood flow changes due to unstable vasculature on time scales of minutes to hours.¹² This instability arises as cancer cells adapt to local hypoxic conditions by recruiting new blood vessels, which enable the delivery of nutrients and the removal of toxic metabolites so that cancer cells can survive when nutrients are low. However, as cancer cells induce the growth of new blood vessels, the vascular network becomes more irregular and disorganized, leading to unpredictable spatial and temporal variations in blood flow.¹³ Consequently, transient changes in perfusion lead to cycling hypoxia. Local fluctuations between hypoxia and reoxygenation affect cells adjacent to the poorly perfused blood vessels. This is in contrast to chronic hypoxia, which affects cells far away from vessels due to limited diffusion. Cycling hypoxia is associated with an increase in cell migration, metastatic potential, and resistance to treatments compared to chronic hypoxia.¹⁴

While changing patterns of blood flow likely create the greatest source of temporal variability in oxygen, pH, immune

infiltration, nutrients, and toxins, the architecture of cells within and around a tumor microenvironment also may change temporally. The rate of diffusion of nutrients (such as glucose and glutamine) toward a neighborhood of cancer cells and the diffusion of metabolites away (such as lactate, free-oxygen radicals, and pyruvate) may vary temporally as other neighborhoods of cancer cells block or unblock intracellular channels, as immune cells move in or out of an area, and as fibroblasts change extracellular matrices and/or the secretion of growth factors. Regardless of its source or regularity, coexisting cancer cells experience considerable temporal variability in opportunities and hazards that likely create good and bad periods in terms of their proliferation and survival rates, setting the potential for the storage effect to promote diversity of cancer cells within tumors.

Proliferative and Nonproliferative Life-History States in Nature and Cancer

In response to fluctuations that generate good and bad periods, many organisms have evolved different life-history states: one that is best at exploiting the good times and the other best at surviving through the bad times. Many single-celled protists have a proliferative state that allows the cell to feed, move, and proliferate.¹⁵ This state, however, may not be able to survive bad periods when their pool of water dries up or the environment offers only toxins and no nutrients. Such protists also have an encysted state. This state, although unable to proliferate, is highly resistant to desiccation, toxins, and nutrient deprivation. This state may be the only way the protist survives through bad periods. Having a fraction of the protist population in an encysted state means that some opportunities are lost during good periods, but survival is ensured during the bad. Baker's yeast is a classic example. The dry packet of encysted yeast can survive for years without an opportunity for growth and reproduction. Upon activation and favorable conditions, this yeast enters a feeding and proliferative state. Among species in nature, "quiescence" or "dormancy" refers to a range of cellular or organismal states characterized by slowed metabolism and relatively high resistance to hardships from their surrounding environment.¹⁶⁻²¹

In cancer, cells that are reversibly arrested in the G₀ phase of the cell cycle are referred to as quiescent, whereas "cell dormancy" typically refers to a long-term quiescence.²² Many cancer cells respond to hypoxia by becoming quiescent and upregulating autophagy to survive lower levels of nutrients and oxygen.¹³ Cancer cells may survive harsh environments (eg, hypoxia, low pH, toxins) by forming poly-aneuploid cancer cells (also referred to in the literature as polyploid giant cancer cells), a population of reversibly quiescent cells that form rapidly (within 72 hours) in response to environmental stress and later divide asymmetrically to repopulate a tumor with cells of normal ploidy that have increased resistance to chemotherapy.^{23,24} In their nonproliferative state, dormant or quiescent tumor cells are able to evade treatments that preferentially target highly proliferative cells. After treatment, these cells can

resume proliferation, which can result in tumor growth and relapse. Quiescent states are ubiquitous in cancer and can be associated with metastasis and relapse of cancerous growth.²⁵ As such, dormancy and quiescence challenge our ability to treat, control, or moderate cancer.

Some readers may see similarities between cancer stem cells (CSCs) and our description of quiescent and dormant cancer cells. The similarity does manifest when CSC refers to a subset of heterogeneous cancer cells that exhibit plasticity, drug resistance, and tumor-initiating capability. But, CSC concepts diverge from our use of quiescence and dormancy when CSCs are seen as a small population of undifferentiated, self-renewing cells that give rise to and sustain a population of terminally differentiated cells. Once fully differentiated, these cells are viewed as being nonproliferative even as they make up the bulk of the tumor. Cancer stem cells can undergo either symmetric or asymmetric cell division, where the former increases the CSC population and the latter promotes the differentiated population. For the storage effect, the proliferative state is vulnerable to harsh conditions while the nonproliferative quiescent state is not. Each state can phenotypically shift into the other.

In nature, quiescent and resistant states can have a powerful effect. They can allow populations to survive despite exposure to conditions that limit or preclude population growth. They can buffer populations against variation in favorability for growth on many time scales, including seasonal, annual, and multiannual; they can contribute to more diverse communities than would exist without them.^{9,10,26-28} Given how common dormancy/quiescence is in cancer, we expect to find similar ecological effects of these life-history stages for the cancer ecosystem. Quiescent or dormant states could maintain diverse cancer phenotypes that may proliferate, coexist, or displace each other over the course of time within the patient. For instance, disseminated cancer cells that stay dormant, sometimes for many years, and then reemerge as metabolically active cells that proliferate, often leading to the death of patients. These emergent metastases appear to result from enhanced cell lineage persistence through dormancy.^{23-25,29,30} The movement of cells into and out of dormancy, as the environment around them changes, could affect what clones co-occur in a tumor or in a patient by changing the ways in which cell-cell interactions such as competition play out at the level of populations of different cell types. The presence of quiescent and dormant, resistant states of cancer cells sets the potential for the storage effect to promote the diversity of cancer cells within tumors. These possibilities have clear relevance for cancer control.

Evidence for Population-Specific Behaviors and Trade-Offs in the Way Cancer Cells Experience Good and Bad Times

As in nature, where a year with plentiful but not excessive water would be expected to be generally a good year for

vegetation, cancer cells also would share many environmental needs and so experience good and bad times in part together. However, as with plants that differ in tolerance to low water availability or high temperatures, cancer cells differ in their sensitivities to hypoxia, low pH, immune infiltration, the absence of growth factors, and the stromal (created by normal cells) architecture of their microenvironment, generating potential for trade-offs that make some periods better for some cell clones and others better for other clones. The heterogeneity of cancer cells in many features, including their proliferative potential, hormone receptor expression, immunogenicity, sensitivity to drugs, motility, and angiogenic potential,³¹ enables cancer cells to have *population-specific environmental responses*, an essential component of the storage effect. For example, in stage 2 invasive breast cancers, cells on the tumor edge tend to have an acid-producing invasive, proliferative phenotype, whereas those in the hypoxic tumor core have a less proliferative phenotype.³² Tumors also have metabolic heterogeneity in the form of acid-resistant or glycolytic phenotypes.³³ Mathematical models of cancer evolution suggest that high nutrient variability (cycling hypoxia) gives a competitive advantage to cancer cells that have a higher rate of phenotypic transition, which promotes high levels of phenotypic heterogeneity.³⁴

Cancer cells, and even normal cells, have a wide range of behaviors associated with entering and exiting from nonproliferative states, which provide the *buffering life-history stage* that enables a storage effect. Cancer cells may remain in these states for times ranging from the very short (1 to several days) to the very long (decades, as observed for emergence of metastatic cancer growth after years of healthy remission).^{25,35} In fibroblasts, cells move deeper into quiescence following longer durations of serum starvation or contact inhibition and require a stronger growth stimulation to exit quiescence compared to cells in shallow quiescence.³⁶ Interactions between cancer cells and the tumor microenvironment can promote quiescence or dormancy. For example, bone-lining osteoblasts and endothelial cells on stable vasculature secrete factors that keep disseminated tumor cells dormant,³⁷ and adhesion of multiple myeloma cells to extracellular matrix components such as fibronectin leads to quiescence.³⁸

The time of exit from quiescence can vary within a clonal population, even when cultured in the same growth conditions.³⁹ Dormant cells awaken due to environmental changes. Bone is highly dynamic, undergoing constant remodeling, which can reawaken dormant cancer cells.⁴⁰ The growth of new blood vessels provokes sufficient change to the microenvironment to reactivate breast cancer cells.^{41,42} Age-related inflammation promotes the outgrowth of dormant cells in pancreatic ductal adenocarcinoma liver metastases.⁴³ Polyan euploid cancer cells emerge from quiescence after removal of chemotherapy, and they do so after a range of quiescence durations (Sarah Amend, PhD, email communication, February 2020). Drug resistance is associated with a heterogeneous population of nonproliferative cells, including drug-tolerant persisters, therapy-induced senescent cells, hypoxic drug-resistant,

and disseminated tumor cells. These populations differ in their propensities to remain in or exit the growth-arrested state.²²

A Simple Model of the Storage Effect in Cancer

Mathematical models have been useful to understand cancer biology and to help test what treatment strategies might improve patient outcomes over the standard of care. Many model types have been used, including game theory models,⁴⁴⁻⁵¹ agent-based models,^{33,52-57} and Lotka-Volterra-type dynamical models that consider cancer clonal cells as interacting populations.^{58,59} So far, very few of these models consider variation in the environment of cancer cells in time. It is unsurprising that temporal variation in the tumor environment has had little attention in dynamical models. This was long the case in models of natural ecosystems, and the data available for cancer tend to be static snapshots of a tumor. Radiographic and histologic images show cancer at a point in time. Although they reveal, and provide data for, modeling spatial heterogeneity in cancer, they do not capture or inform the potential variation in time that small-scale physiological studies indicate is there as well.^{12,14,60}

Stochastic dynamic models of interacting populations of cancer cells are a straightforward way to incorporate temporal environmental variation and are the class of models that have primarily been used to develop an understanding of the storage effect. The simple Lotka-Volterra-type population models so far used to study cancer are not sufficient to capture coexistence mechanisms such as the storage effect because they do not incorporate a nonproliferative life-history state nor do they incorporate species-specific demographic responses to a varying environment.^{3,61,62} Variation in the environment can be proxied as stochastic variation in demographic parameters, such as entry to and exit from quiescence and rates of growth, reproduction, or survival of cancer cells.

To show how coexistence can occur via the storage effect, we construct a simple discrete-time stochastic model that includes 2 species, each with 2 life-history stages (proliferative and quiescent) and stochastic variation in good and bad periods.

Illustrating the Storage Effect

The plausibility of the storage effect promoting the coexistence of different cell types within a tumor microenvironment emerges from (1) temporal variability in blood flow, immune infiltration, stromal architecture, and physical conditions and (2) the widespread occurrence of cell arrest, quiescence, and/or dormancy that allows cancer cells to exist in 2 states—proliferative and nonproliferative. Coexistence can occur if there is a trade-off in how 2 cell types experience good and bad periods.

To illustrate how storage effects can work, we propose a simple model that aims to keep assumptions to a minimum while containing the essential elements. We imagine that each cancer cell type has 2 stages: an arrested state and a

proliferative state. A fraction of cells, θ , leave the arrested state during each period. Those that enter the proliferative state have a mortality rate that depends on whether it is during a “good” or “bad” period. These periods could represent a microenvironment of a tumor where fluctuations in blood flow, hypoxia, and/or growth factors contribute to times of plenty and times of famine. Upon emerging from the arrested state, cells successfully survive to proliferate at fraction s . Let $s_g \gg s_b$ be these survival rates during good and bad periods, respectively.

The biology here is that if a cell attempts to be proliferative, it may lack key resources even as the cell commits to cell division. The consequence is cell mortality that would not have happened had the cell remained in an arrested state. For example, providing estrogen to breast cancer cells that otherwise have no other nutrients can cause mortality, as cells are tricked into trying to proliferate (Robert Gatenby, MD, email communication, February 2020). Those cells that survive produce a maximum number of daughter cells, f , that declines with competition from other cells in a proliferative state. We use a Ricker model to describe the competition between proliferative cells for resources. In the absence of competitors, a proliferative cell produces its maximum number of daughter cells; with competitors, the production of daughter cells declines exponentially with the density of other proliferative cells. We imagine that f can take on a positive value that represents the expected number of daughter cells that might accrue during a period sufficiently long to permit 1 or more rounds of cell division. At the end of the period, daughter cells enter the pool of cells in an arrested state.

Now let there be 2 cell types that are equal in all ways, except that they do not always experience good and bad periods in the same way. Although unlikely that 2 cell types would be identical, this assumption allows us to illustrate coexistence that only manifests because of the storage effect.

For cancer cell types, it is likely that what is good for one is, in part, good for the other type. However, differences in nutrient metabolism, sensitivity to growth factors, resistance to hypoxia, the ability for immune evasion, and/or physical conditions like pH mean that at times one cell type may experience a bad period while the other a good period. Let q be the probability of a good period, and let $0 \leq r \leq 1$ represent the degree to which the 2 cell types experience good and bad periods similarly. The term $(1 - r)$ then represents the probability that the 2 cell types are experiencing the type of period independently of each other. Thus, the probabilities of both experiencing good periods (p_{GG}), both experiencing bad periods (p_{BB}), type 1 experiencing good and type 2 experiencing bad (p_{GB}), and vice versa (p_{BG}) are given by:

$$\begin{aligned} p_{GG} &= rq + (1 - r)q^2 \\ p_{BB} &= r(1 - q) + (1 - r)(1 - q)^2 \\ p_{BG} &= p_{GB} = (1 - r)q(1 - q). \end{aligned}$$

As they must, p_{GG} , p_{BB} , p_{BG} , and p_{GB} sum to 1.

Let $x_k(t)$ be the number of cells of type k in an arrested state following time period t , where $k = 1$ or 2. The 2 species growth

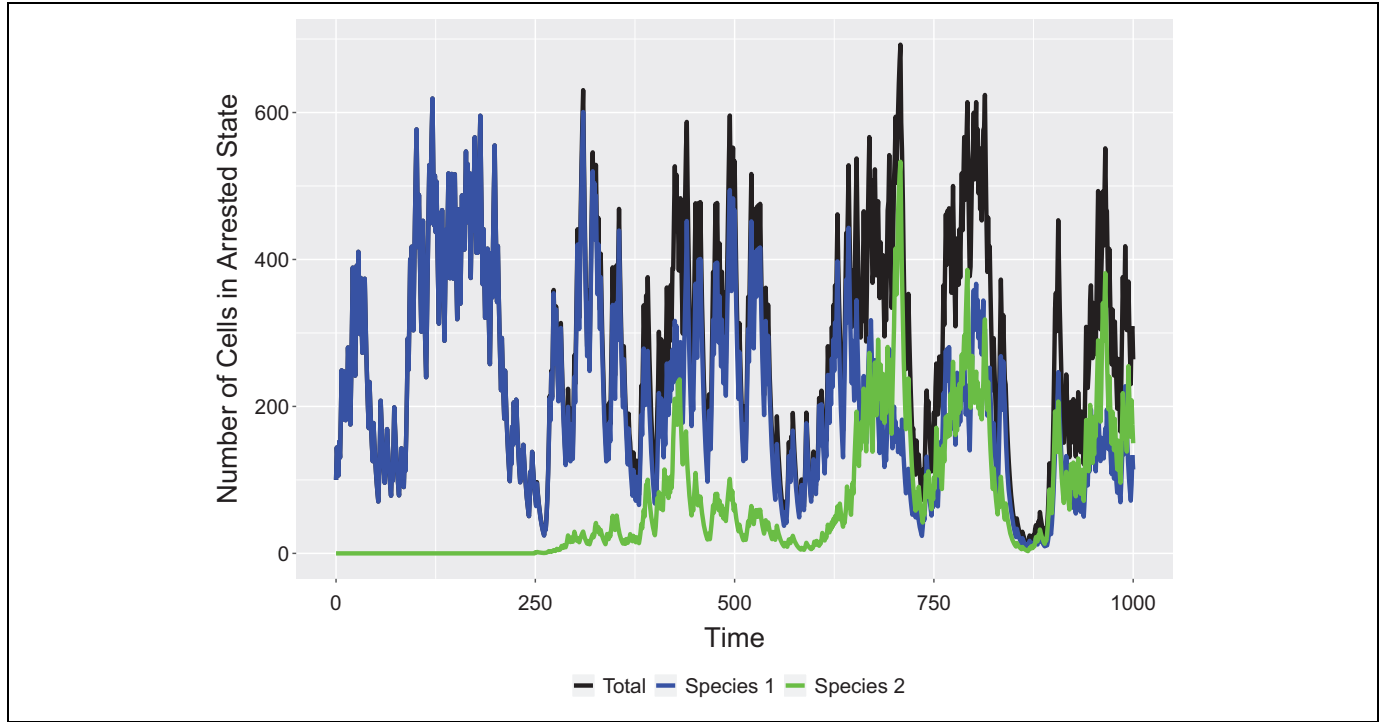


Figure 1. Invasion of resident population. Parameters: $s_g = 0.8$, $s_b = 0.1$, $R = 100$, $d = 0.05$, $\theta = 0.2$, $f = 5$, $q = 0.4$, $r = 0.8$, $x_1(0) = 100$, $x_2(0) = 0$, $x_2(250) = 1$.

equations take on the following form:

$$x_1(t+1) = (1-\theta)(1-d)x_1(t) + \theta \frac{s_{i1}x_1(t) + s_{j2}x_2(t)}{R} fe$$

$$x_2(t+1) = (1-\theta)(1-d)x_2(t) + \theta \frac{s_{i1}x_1(t) + s_{j2}x_2(t)}{R} fe$$

where $i, j = g$ or b , and d is the mortality rate of cells in an arrested state per period, and R scales the limits to growth by representing some measure of resources available to cells that are in a proliferative state.

The above model easily generates a coexistence via a storage effect. This can be seen by showing how either species 1 or 2 can invade a resident population of the other and that when together they settle on a dynamic equilibrium but with more or less temporal variation in the frequency of the 2 cell types within a microenvironment (Figure 1). When averaged over many microenvironments, the cell type frequencies might appear relatively stable across time. If a storage effect is operating within a tumor, then there may be surprisingly large fluctuations in cell composition at the level of small neighborhoods of cancer cells.

The storage effect is not possible in the absence of an arrested state. For $\theta = 1$, the 2 cancer cell types exist as a stochastic random walk with neither one having an advantage. An arrested state buffers a cell type when it experiences a bad period, while the other experiences a good period. Similarly, if $r = 1$, then a storage effect is not possible, as both cell types perceive good and bad years identically. The 2 will simply experience a stochastic random walk of population sizes with neither

experiencing an advantage when rare. Once $r < 1$ and $\theta < 1$, the coexistence of cell types by the storage effect will happen, but the strength of this will increase as r approaches 0. The advantage that a cell type gains when it is rare (thus driving it to increase in time at the expense of the other cell type) derives from the occurrence of periods where the rare species experiences a good and the resident cell type experiences a bad period, giving the rare species opportunity to realize the high growth potential of a good period without strong depression by competition. This is maximized when $r = 0$ and when $q = 0.5$ (good and bad periods are equally likely). Furthermore, as a long-term dynamic equilibrium, the average tumor burden increases as r goes from 1 to 0; and as q goes from 0 to 1 (Figure 2).

How Might Consideration of the Storage Effect Inform Cancer Therapy?

The existence of the storage effect contributing to diversity in cancer would have consequences for clinical treatments. It is recognized that intratumor and intertumor heterogeneity should inform cancer therapy.^{63,64} For instance, intratumor heterogeneity is established as a marker for poor prognosis. Additional therapeutic challenges arise if some of the diversity of cancer cell types involve storage effect dynamics. The presence of a storage effect presents a double challenge for therapy. First is the task of targeting quiescent cells, and second is the challenge of finding drugs effective on each or all of the cancer cell types.

Quiescence or dormancy now appears to be a major mechanism by which cancers evade drug therapies and initiate

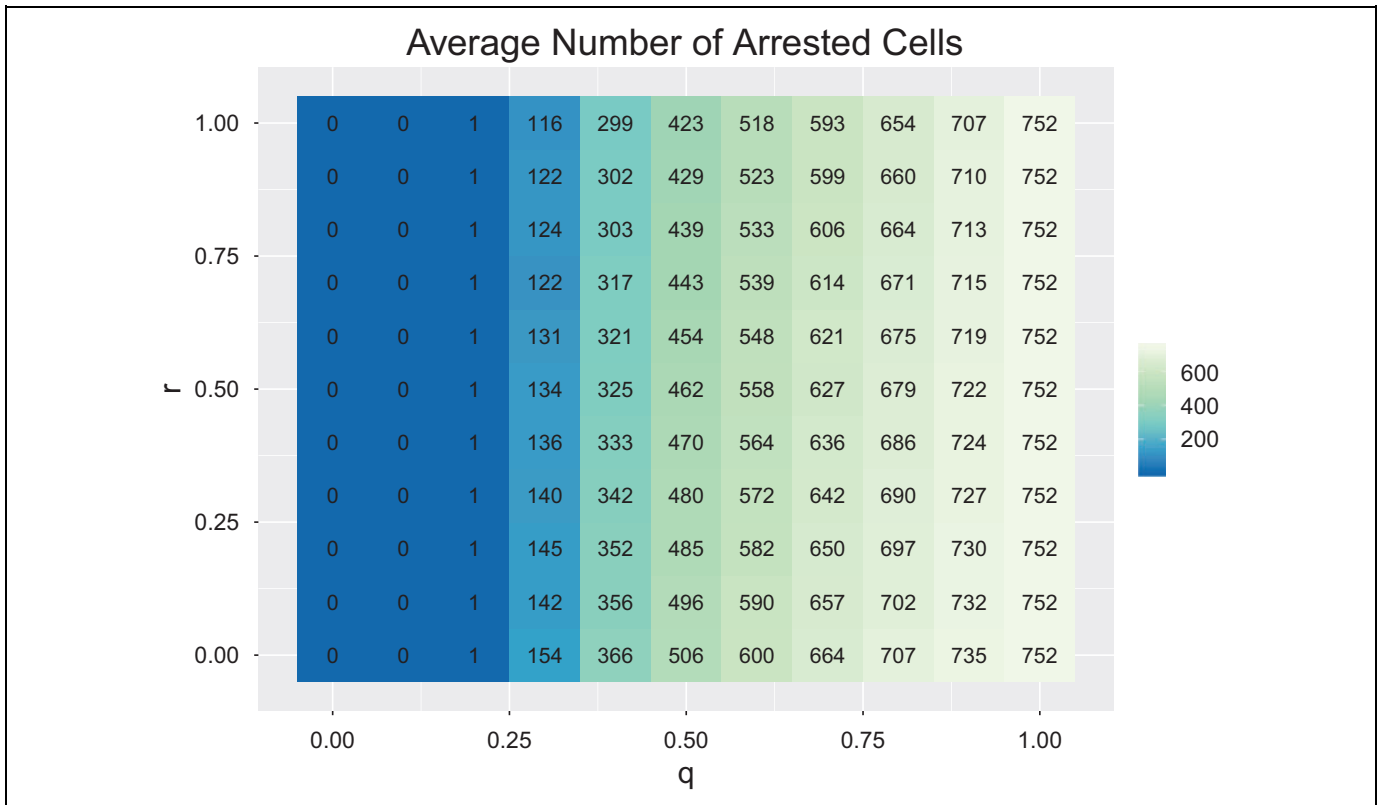


Figure 2. Average number of arrested cells as a function of q and r . Each combination represents an average of 10 simulations. Parameters: $s_g = 0.8$, $s_b = 0.1$, $R = 100$, $d = 0.05$, $\theta = 0.2$, $f = 5$, $x_1(0) = 100$, $x_2(0) = 100$, number of timesteps = 10^4 .

metastases,²³⁻²⁵ so there is great interest in whether quiescent or dormant cells can be targeted by therapies.⁶⁵ This article argues that we must also consider that there may be many coexisting clonal lineages of cancer cells that differ in physiologies of their nonproliferative and proliferative life-history states. If cancer cell types coexist via the storage effect, then one is not targeting just a single cell type, but a diversity of cell types with potentially differing susceptibilities to therapy. Longitudinally monitoring tumors and responses to therapy may require more frequent magnetic resonance imaging and other imaging techniques as noninvasive tools for identifying and tracking different tumor habitats based on vascularity, hypoxia, necrosis, and so on.^{66,67}

Pseudo-Resistance

Heterogeneity in quiescence could obscure detection of resistance to treatment through the phenomenon of *pseudo-resistance*, a situation where an otherwise treatment-sensitive population of cancer cells appears to be resistant. Pseudo-resistance could result from temporal variation that selects for cell lineages that maintain a high fraction of cells within a quiescent state. Many cytotoxic chemotherapies target proliferating cells that are actively dividing, and nonproliferative cells may be insensitive to these therapies. Those parts of a tumor that experience the greatest temporal variability may have the greatest fraction of nonproliferative cells. In this case, therapy may appear effective in eliminating tumor cells, but

continued efficacy may slow, as the remaining cancer clonal populations have lower fractions of cells in a proliferative state. The storage effect, by maintaining the coexistence of cancer clones that vary in their propensity for remaining quiescent or arrested, could exacerbate pseudo-resistance.

With pseudo-resistance, continuing the same drug regimen would continue to cull cancer cells as they emerge into a proliferative state. However, the fraction of cancer cells killed with each dosing would decline, generating the appearance of declining tumor burden, but without entirely eradicating the tumor. Prolonging the therapy may increase the likelihood of true resistance emerging from the surviving cancer cells and also may induce undue toxicities to the patient. The possibility that pseudo-resistance might reflect a storage effect would suggest novel therapeutic strategies. One approach might be to maintain the ongoing therapy, but reduce dosage or make it more intermittent (akin to maintenance therapy for patients in remission). Frequent and accurate measures of tumor burden might identify breakpoints in the rate of tumor burden decline, which would indicate the presence of different clones with different propensities to remain quiescent. A second therapeutic approach could be to include in tandem an intervention designed to amplify the rate at which cancer cells return from quiescence into a proliferative state. It might also be useful to begin with the first therapy and then add to it, rather than replace it with, additional therapies; if possible, the second therapy would be chosen for toxicity to quiescent cells.

The Storage Effect and Multidrug Therapies

The existence of a storage effect implies that additional data are needed to predict tumor progression. Knowing to what extent the variability that one sees in space also occurs in time at each spot becomes a valuable and critical piece of information. There is recognition of the need for serial histologies, but the destructive nature of histological sampling means exact resampling is impossible. Variable temporal dynamics means that local fluctuations could work against traditional therapies. An underlying storage effect would indicate that cell types differ in their dependence on or sensitivity to specific microenvironmental conditions that are fluctuating. Each of them could have an Achilles' heel. If one can identify or suggest the trade-off promoting a storage effect, then a background therapy targeting quiescent cells could be combined with a standard chemotherapy and a therapy known to target the specific attributes of the cancer cell types.

Caveats

As a first step, our focus here has been on how the storage effect could contribute toward the diversity of cancer cell types seen in a tumor. This would be in addition to other sources of intratumor heterogeneity and the many interactions and types of stromal cells such as fibroblasts, macrophages, and pericytes. Understanding the full community ecology of these diverse cell types and how the extracellular matrix, for instance, influences therapy resistance is important.^{68,69} From the vantage point of the storage effect, it would be of interest to know how these other members of the tumor microenvironment influence temporal and spatial fluctuations in key resources or hazards. If fibroblasts amplify (or dampen) temporal variability, then they may facilitate (or prevent) the coexistence of cancer cell types via the storage effect.

Even if the storage effect contributes to 2 coexisting cell types, identifying them poses challenges. First, each should have a proliferative and quiescent state, meaning that there should be 2 identifiably distinct proliferative cell types, 2 distinct quiescent types, and the correct proliferative types must be matched with their corresponding quiescent. This is not unlike the challenge of having 2 yeast species in the same sample where each yeast species has an active and an encysted form. Living biopsies or collections of circulating tumor cells may provide source material for single-cell culturing and the use of microfluidics to determine the presence of both life-history stages emerging from the single cell.^{70,71}

Summary

Here, we have described the diversity-promoting mechanism known as the storage effect and suggested that the conditions that make the storage effect relevant to ecosystems in nature also are found in cancer. Research is needed to better identify and quantify the features of cancer that characterize the storage effect. These features include population-specific responses to

the environment, which could give differential use of conditions in time by different cancer cell populations; and behaviors associated with entry into, exit from, and duration of quiescence in cancer cells, which could enable buffering of those populations to environmental fluctuations, allowing them to evade losses in bad times and realize strong growth in good times. The possibility that the storage effect is involved in the clonal ecological heterogeneity and quiescence/dormancy of cancer cells, both of which pose serious conundrums for cancer therapies, dictates that more attention should be paid to understanding the temporal dimensions of cancer. To do this, both the data and the models that inform our understanding of cancer must be expanded to better reveal and account for temporal variation. If temporal variation in environmental suitability for growth contributes significantly to population interactions within the cancer ecosystem, the implications to how we understand and treat cancer would be substantial.

Acknowledgments

The authors thank Drs Sarah Amend, Ryan Bishop, and Chris Whelan for helpful discussions on the content of this article.


Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The author(s) received no financial support for the research, authorship, and/or publication of this article: AKM and DB were funded by NIH/NCI 1U01CA202958-01. JSB was funded by NIH/NCI 1U54CA193489-01A1.

ORCID iD

Anna K. Miller  <https://orcid.org/0000-0002-3099-1648>

References

1. Warner RR, Chesson PL. Coexistence mediated by recruitment fluctuations: a field guide to the storage effect. *Am Nat.* 1985; 125(6):769-787. doi:10.1086/284379
2. Huntly N. Herbivores and the dynamics of communities and ecosystems. *Annu Rev Ecol Syst.* 1991;22(1):477-503.
3. Chesson P, Huntly N. The roles of harsh and fluctuating conditions in the dynamics of ecological communities. *Am Nat.* 1997; 150(5):519-553. doi:10.1086/286080
4. Chesson P. Species competition and predation. In: Leemans R, ed. *Ecological Systems.* Springer; 2013:223-256. doi:10.1007/978-1-4614-5755-8_13
5. Amend SR, Pienta KJ. Ecology meets cancer biology: the cancer swamp promotes the lethal cancer phenotype. *Oncotarget.* 2015; 6(12):9669-9678. doi:10.18632/oncotarget.3430
6. Kotler BP, Brown JS. Coexistence mechanisms in the community ecology of cancer: implications for cancer control. *Cancer Control.* 2020
7. Chesson PL, Huntly N. Community consequences of life-history traits in a variable environment. *Ann Zool Fennici.* 1988;25(1):5-16.

8. Chesson P, Huntly N. Short-term instabilities and long-term community dynamics. *Trends Ecol Evol (Amst)*. 1989;4(10):293-298. doi:10.1016/0169-5347(89)90024-4
9. Chesson P, Gebauer RLE, Schwinning S, et al. Resource pulses, species interactions, and diversity maintenance in arid and semi-arid environments. *Oecologia*. 2004;141(2):236-253. doi:10.1007/s00442-004-1551-1
10. Chesson P, Huntly NJ, Roxburgh SH, Pantastico-Caldas M, Facelli JM. The storage effect: definition and tests in two plant communities. In: Kelly CK, Bowler MG, Fox GA, eds. *Temporal Dynamics and Ecological Process*. Cambridge University Press; 2013:11-40. doi:10.1017/CBO9781139048170.003
11. Chesson PL, Warner RR. Environmental variability promotes coexistence in lottery competitive systems. *Am Nat*. 1981;117(6):923-943. doi:10.1086/283778
12. Saxena K, Jolly MK. Acute vs. chronic vs. cyclic hypoxia: their differential dynamics, molecular mechanisms, and effects on tumor progression. *Biomolecules*. 2019;9(8):339. doi:10.3390/biom9080339
13. Gillies RJ, Brown JS, Anderson ARA, Gatenby RA. Eco-evolutionary causes and consequences of temporal changes in intratumoural blood flow. *Nat Rev Cancer*. 2018;18(9):576-585. doi:10.1038/s41568-018-0030-7
14. Michiels C, Tellier C, Feron O. Cycling hypoxia: a key feature of the tumor microenvironment. *Biochim Biophys Acta*. 2016;1866(1):76-86. doi:10.1016/j.bbcan.2016.06.004
15. Alve E, Goldstein ST. Dispersal, survival and delayed growth of benthic foraminiferal propagules. *J Sea Res*. 2010;63(1):36-51. doi:10.1016/j.seares.2009.09.003
16. Cáceres CE. Dormancy in invertebrates. *Invertebr Biol*. 1997;116(4):371-383. doi:10.2307/3226870
17. Fenelon JC, Renfree MB. The history of the discovery of embryonic diapause in mammals. *Biol Reprod*. 2018;99(1):242-251. doi:10.1093/biolre/iou112
18. Lennon JT, Jones SE. Microbial seed banks: the ecological and evolutionary implications of dormancy. *Nat Rev Microbiol*. 2011;9(2):119-130. doi:10.1038/nrmicro2504
19. Baumgartner MF, Tarrant AM. The physiology and ecology of diapause in marine copepods. *Ann Rev Mar Sci*. 2017;9:387-411. doi:10.1146/annurev-marine-010816-060505
20. Tougeron K. Diapause research in insects: historical review and recent work perspectives. *Entomol Exp Appl*. 2019;167(1):27-36. doi:10.1111/eea.12753
21. Willis CG, Baskin CC, Baskin JM, et al. The evolution of seed dormancy: environmental cues, evolutionary hubs, and diversification of the seed plants. *New Phytol*. 2014;203(1):300-309. doi:10.1111/nph.12782
22. De Angelis ML, Francescangeli F, La Torre F, Zeuner A. Stem cell plasticity and dormancy in the development of cancer therapy resistance. *Front Oncol*. 2019;9:626. doi:10.3389/fonc.2019.00626
23. Amend SR, Torga G, Lin K-C, et al. Polyploid giant cancer cells: unrecognized actuators of tumorigenesis, metastasis, and resistance. *Prostate*. 2019;79(13):1489-1497. doi:10.1002/pros.23877
24. Pienta KJ, Hammarlund EU, Axelrod R, Brown JS, Amend SR. Poly-aneuploid cancer cells promote evolvability, generating lethal cancer. *Evol Appl*. 2020. doi:10.1111/eva.12929
25. Aguirre-Ghiso JA, Sosa MS. Emerging topics on disseminated cancer cell dormancy and the paradigm of metastasis. *Annu Rev Cancer Biol*. 2018;2(1):377-393. doi:10.1146/annurev-cancerbio-030617-050446
26. Cáceres CE. Temporal variation, dormancy, and coexistence: a field test of the storage effect. *Proc Natl Acad Sci U S A*. 1997;94(17):9171-9175. doi:10.1073/pnas.94.17.9171
27. Chesson P, Huntly N. Temporal hierarchies of variation and the maintenance of diversity. *Plant Species Biol*. 1993;8(2-3):195-206. doi:10.1111/j.1442-1984.1993.tb00070.x
28. Angert AL, Huxman TE, Chesson P, Venable DL. Functional tradeoffs determine species coexistence via the storage effect. *Proc Natl Acad Sci U S A*. 2009;106(28):11641-11645. doi:10.1073/pnas.0904512106
29. Aguirre-Ghiso JA. Models, mechanisms and clinical evidence for cancer dormancy. *Nat Rev Cancer*. 2007;7(11):834-846. doi:10.1038/nrc2256
30. Riggi N, Aguet M, Stamenkovic I. Cancer metastasis: a reappraisal of its underlying mechanisms and their relevance to treatment. *Annu Rev Pathol*. 2018;13:117-140. doi:10.1146/annurev-pathol-020117-044127
31. Marusyk A, Polyak K. Tumor heterogeneity: causes and consequences. *Biochim Biophys Acta*. 2010;1805(1):105-117. doi:10.1016/j.bbcan.2009.11.002
32. Lloyd MC, Cunningham JJ, Bui MM, et al. Darwinian dynamics of intratumoral heterogeneity: not solely random mutations but also variable environmental selection forces. *Cancer Res*. 2016;76(11):3136-3144. doi:10.1158/0008-5472.CAN-15-2962
33. Robertson-Tessi M, Gillies RJ, Gatenby RA, Anderson ARA. Impact of metabolic heterogeneity on tumor growth, invasion, and treatment outcomes. *Cancer Res*. 2015;75(8):1567-1579. doi:10.1158/0008-5472.CAN-14-1428
34. Ardaševa A, Gatenby RA, Anderson ARA, et al. Evolutionary dynamics of competing phenotype-structured populations in periodically fluctuating environments. *J Math Biol*. 2020;80(3):775-807. doi:10.1007/s00285-019-01441-5
35. Vallette FM, Olivier C, Lézot F, et al. Dormant, quiescent, tolerant and persister cells: four synonyms for the same target in cancer. *Biochem Pharmacol*. 2019;162:169-176. doi:10.1016/j.bcp.2018.11.004
36. Kwon JS, Everetts NJ, Wang X, et al. Controlling depth of cellular quiescence by an Rb-E2F network switch. *Cell Rep*. 2017;20(13):3223-3235. doi:10.1016/j.celrep.2017.09.007
37. Sosa MS, Bragado P, Aguirre-Ghiso JA. Mechanisms of disseminated cancer cell dormancy: an awakening field. *Nat Rev Cancer*. 2014;14(9):611-622. doi:10.1038/nrc3793
38. Meads MB, Gatenby RA, Dalton WS. Environment-mediated drug resistance: a major contributor to minimal residual disease. *Nat Rev Cancer*. 2009;9(9):665-674. doi:10.1038/nrc2714
39. Wang X, Fujimaki K, Mitchell GC, et al. Exit from quiescence displays a memory of cell growth and division. *Nat Commun*. 2017;8(1):321. doi:10.1038/s41467-017-00367-0
40. Lawson MA, McDonald MM, Kovacic N, et al. Osteoclasts control reactivation of dormant myeloma cells by remodelling the endosteal niche. *Nat Commun*. 2015;6:8983. doi:10.1038/ncomms9983

41. Croucher PI, McDonald MM, Martin TJ. Bone metastasis: the importance of the neighbourhood. *Nat Rev Cancer*. 2016;16(6):373-386. doi:10.1038/nrc.2016.44
42. Ghajar CM, Peinado H, Mori H, et al. The perivascular niche regulates breast tumour dormancy. *Nat Cell Biol*. 2013;15(7):807-817. doi:10.1038/ncb2767
43. Lenk L, Pein M, Will O, et al. The hepatic microenvironment essentially determines tumor cell dormancy and metastatic outgrowth of pancreatic ductal adenocarcinoma. *Oncoimmunology*. 2017;7(1):e1368603. doi:10.1080/2162402X.2017.1368603
44. Archetti M, Pienta KJ. Cooperation among cancer cells: applying game theory to cancer. *Nat Rev Cancer*. 2019;19(2):110-117. doi:10.1038/s41568-018-0083-7
45. Archetti M. Heterogeneity and proliferation of invasive cancer subclones in game theory models of the Warburg effect. *Cell Prolif*. 2015;48(2):259-269. doi:10.1111/cpr.12169
46. Gatenby RA, Vincent TL. An evolutionary model of carcinogenesis. *Cancer Res*. 2003;63(19):6212-6220.
47. Orlando PA, Gatenby RA, Brown JS. Cancer treatment as a game: integrating evolutionary game theory into the optimal control of chemotherapy. *Phys Biol*. 2012;9(6):065007. doi:10.1088/1478-3975/9/6/065007
48. Basanta D, Hatzikirou H, Deutsch A. Studying the emergence of invasiveness in tumours using game theory. *Eur Phys J B*. 2008;63(3):393-397. doi:10.1140/epjb/e2008-00249-y
49. Kaznatcheev A, Vander Velde R, Scott JG, Basanta D. Cancer treatment scheduling and dynamic heterogeneity in social dilemmas of tumour acidity and vasculature. *Br J Cancer*. 2017;116(6):785-792. doi:10.1038/bjc.2017.5
50. Basanta D, Gatenby RA, Anderson ARA. Exploiting evolution to treat drug resistance: combination therapy and the double bind. *Mol Pharm*. 2012;9(4):914-921. doi:10.1021/mp200458e
51. Basanta D, Scott JG, Fishman MN, et al. Investigating prostate cancer tumour–stroma interactions: clinical and biological insights from an evolutionary game. *Br J Cancer*. 2012;106(1):174-181. doi:10.1038/bjc.2011.517
52. Anderson ARA. A hybrid mathematical model of solid tumour invasion: the importance of cell adhesion. *Math Med Biol*. 2005;22(2):163-186. doi:10.1093/imammb/dqi005
53. Gallaher JA, Enriquez-Navas PM, Luddy KA, Gatenby RA, Anderson ARA. Spatial heterogeneity and evolutionary dynamics modulate time to recurrence in continuous and adaptive cancer therapies. *Cancer Res*. 2018;78(8):2127-2139. doi:10.1158/0008-5472.CAN-17-2649
54. Anderson ARA, Weaver AM, Cummings PT, Quaranta V. Tumor morphology and phenotypic evolution driven by selective pressure from the microenvironment. *Cell*. 2006;127(5):905-915. doi:10.1016/j.cell.2006.09.042
55. Basanta D, Strand DW, Lukner RB, et al. The role of transforming growth factor-beta-mediated tumor–stroma interactions in prostate cancer progression: an integrative approach. *Cancer Res*. 2009;69(17):7111-7120. doi:10.1158/0008-5472.CAN-08-3957
56. Rejniak KA, Anderson ARA. Hybrid models of tumor growth. *Wiley Interdiscip Rev Syst Biol Med*. 2011;3(1):115-125. doi:10.1002/wsbm.102
57. Araujo A, Cook LM, Lynch CC, Basanta D. An integrated computational model of the bone microenvironment in bone-metastatic prostate cancer. *Cancer Res*. 2014;74(9):2391-2401. doi:10.1158/0008-5472.CAN-13-2652
58. West JB, Dinh MN, Brown JS, et al. Multidrug cancer therapy in metastatic castrate-resistant prostate cancer: an evolution-based strategy. *Clin Cancer Res*. 2019;25(14):4413-4421. doi:10.1158/1078-0432.CCR-19-0006
59. Zhang J, Cunningham JJ, Brown JS, Gatenby RA. Integrating evolutionary dynamics into treatment of metastatic castrate-resistant prostate cancer. *Nat Commun*. 2017;8(1):1816. doi:10.1038/s41467-017-01968-5
60. Cárdenas-Navia LI, Mace D, Richardson RA, et al. The pervasive presence of fluctuating oxygenation in tumors. *Cancer Res*. 2008;68(14):5812-5819. doi:10.1158/0008-5472.CAN-07-6387
61. Turelli M. Niche overlap and invasion of competitors in random environments I. Models without demographic stochasticity. *Theor Popul Biol*. 1981;20(1):1-56. doi:10.1016/0040-5809(81)90002-2
62. Chesson P. Multispecies competition in variable environments. *Theor Popul Biol*. 1994;45(3):227-276. doi:10.1006/tpbi.1994.1013
63. Dagogo-Jack I, Shaw AT. Tumour heterogeneity and resistance to cancer therapies. *Nat Rev Clin Oncol*. 2018;15(2):81-94. doi:10.1038/nrclinonc.2017.166
64. Dzobo K, Senthebane DA, Thomford NE, et al. Not everyone fits the mold: intratumor and intertumor heterogeneity and innovative cancer drug design and development. *OMICS*. 2018;22(1):17-34. doi:10.1089/omi.2017.0174
65. Recasens A, Munoz L. Targeting cancer cell dormancy. *Trends Pharmacol Sci*. 2019;40(2):128-141. doi:10.1016/j.tips.2018.12.004
66. Napel S, Mu W, Jardim-Perassi BV, Aerts HJWL, Gillies RJ. Quantitative imaging of cancer in the postgenomic era: radio(geno)mics, deep learning, and habitats. *Cancer*. 2018;124(24):4633-4649. doi:10.1002/cncr.31630
67. Jardim-Perassi BV, Huang S, Dominguez-Viqueira W, et al. Multiparametric MRI and coregistered histology identify tumor habitats in breast cancer mouse models. *Cancer Res*. 2019;79(15):3952-3964. doi:10.1158/0008-5472.CAN-19-0213
68. Senthebane DA, Rowe A, Thomford NE, et al. The role of tumor microenvironment in chemoresistance: to survive, keep your enemies closer. *Int J Mol Sci*. 2017;18(7):1586. doi:10.3390/ijms18071586
69. Senthebane DA, Jonker T, Rowe A, et al. The role of tumor microenvironment in chemoresistance: 3D extracellular matrices as accomplices. *Int J Mol Sci*. 2018;19(10):2861. doi:10.3390/ijms19102861
70. Sant GR, Knopf KB, Albala DM. Live-single-cell phenotypic cancer biomarkers-future role in precision oncology? *NPJ Precis Oncol*. 2017;1(1):21. doi:10.1038/s41698-017-0025-y
71. Manak MS, Varsanik JS, Hogan BJ, et al. Live-cell phenotypic-biomarker microfluidic assay for the risk stratification of cancer patients via machine learning. *Nat Biomed Eng*. 2018;2(10):761-772. doi:10.1038/s41551-018-0285-z