



THE UNIVERSITY *of* EDINBURGH

Edinburgh Research Explorer

## Plasma proteome responses in salmonid fish following immunization

**Citation for published version:**

Bakke, FK, Monte, MM, Stead, DA, Causey, DR, Douglas, A, Macqueen, D & Dooley, H 2020, 'Plasma proteome responses in salmonid fish following immunization', *Frontiers in Immunology*.  
<https://doi.org/10.3389/fimmu.2020.581070>

**Digital Object Identifier (DOI):**

[10.3389/fimmu.2020.581070](https://doi.org/10.3389/fimmu.2020.581070)

**Link:**

[Link to publication record in Edinburgh Research Explorer](#)

**Document Version:**

Publisher's PDF, also known as Version of record

**Published In:**

Frontiers in Immunology

**General rights**

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

**Take down policy**

The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact [openaccess@ed.ac.uk](mailto:openaccess@ed.ac.uk) providing details, and we will remove access to the work immediately and investigate your claim.





# Plasma Proteome Responses in Salmonid Fish Following Immunization

Fiona K. Bakke<sup>1</sup>, Milena M. Monte<sup>1</sup>, David A. Stead<sup>2</sup>, Dwight R. Causey<sup>1</sup>, Alex Douglas<sup>1</sup>, Daniel J. Macqueen<sup>3\*†</sup> and Helen Dooley<sup>4\*†</sup>

<sup>1</sup> School of Biological Sciences, University of Aberdeen, Aberdeen, United Kingdom, <sup>2</sup> Aberdeen Proteomics, The Rowett Institute, University of Aberdeen, Aberdeen, United Kingdom, <sup>3</sup> The Roslin Institute and Royal (Dick) School of Veterinary Studies, University of Edinburgh, Edinburgh, United Kingdom, <sup>4</sup> Department of Microbiology and Immunology, Institute of Marine and Environmental Technology (IMET), University of Maryland School of Medicine, Baltimore, MD, United States

## OPEN ACCESS

### Edited by:

Monica Hongroe Solbakken,  
University of Oslo, Norway

### Reviewed by:

Kim Dawn Thompson,  
Moredun Research Institute,  
United Kingdom  
Bo Peng,  
Sun Yat-sen University, China

### \*Correspondence:

Daniel J. Macqueen  
daniel.macqueen@roslin.ed.ac.uk  
Helen Dooley  
hdooley@som.umaryland.edu

<sup>†</sup>These authors have contributed  
equally to this work

### Specialty section:

This article was submitted to  
Comparative Immunology,  
a section of the journal  
Frontiers in Immunology

**Received:** 07 July 2020

**Accepted:** 21 September 2020

**Published:** 08 October 2020

### Citation:

Bakke FK, Monte MM, Stead DA,  
Causey DR, Douglas A, Macqueen DJ  
and Dooley H (2020) Plasma  
Proteome Responses in Salmonid Fish  
Following Immunization.  
*Front. Immunol.* 11:581070.  
doi: 10.3389/fimmu.2020.581070

Vaccination plays a critical role in the protection of humans and other animals from infectious diseases. However, the same vaccine often confers different protection levels among individuals due to variation in genetics and/or immunological histories. While this represents a well-recognized issue in humans, it has received little attention in fish. Here we address this knowledge gap in a proteomic study of rainbow trout (*Oncorhynchus mykiss*, Walbaum), using non-lethal repeated blood sampling to establish the plasma protein response of individual fish following immunization. Six trout were immunized with adjuvanted hen egg-white lysozyme (HEL) and peripheral blood sampled at ten time points from day 0 to day 84 post-injection. We confirm that an antigen-specific antibody response to HEL was raised, showing differences in timing and magnitude among individuals. Using label-free liquid chromatography-mass spectrometry, we quantified the abundance of 278 plasma proteins across the timecourse. As part of the analysis, we show that this approach can distinguish many (but not all) duplicated plasma proteins encoded by paralogous genes retained from the salmonid-specific whole genome duplication event. Global variation in the plasma proteome was predominantly explained by individual differences among fish. However, sampling day explained a major component of variation in abundance for a statistically defined subset of 41 proteins, representing 15% of those detected. These proteins clustered into five groups showing distinct temporal responses to HEL immunization at the population level, and include classical immune (e.g. complement system members) and acute phase molecules (e.g. apolipoproteins, haptoglobins), several enzymes and other proteins supporting the immune response, in addition to evolutionarily conserved molecules that are as yet uncharacterized. Overall, this study improves our understanding of the fish plasma proteome, provides valuable marker proteins for different phases of the immune response, and has implications for vaccine development and the design of immune challenge experiments.

**Keywords:** plasma, salmonid, immunity, proteome, liquid chromatography-mass spectrometry, trout, immunoglobulin M

## INTRODUCTION

Vaccination plays a critical role in protecting humans and other animals from infectious diseases (e.g. (1–4)). While a perfect vaccine will impart full protection to every member of a targeted population, in reality this is limited by the inherent variability among individuals, including genetics and distinct environmental and/or immunological histories. Indeed, it is well recognized that humans show high variability in immune phenotypic parameters (e.g. serum protein levels) and responses (e.g. cytokine induction), which can largely be explained by non-heritable environmental influences (5, 6). However, the extent of individual-level variation in the immune response remains poorly defined in most species. This includes fishes used in aquaculture, an industry that plays a crucial role in global food provision and economic security, yet remains threatened by infectious disease outbreaks (e.g. (7)).

Vaccination is applied broadly in aquaculture (4), and there remains a need to develop new and improved vaccines for many species and diseases, along with less labor-intensive modes of administration (3). An important part of the development pipeline is vaccine efficacy testing, which can be assessed directly *via* the extent of protection (e.g. using disease challenge tests), or indirectly, by studying changes in immune parameters, for example by confirming an antigen-specific antibody response (8, 9) or recording changes in the expression of immune molecules (e.g. (10, 11)). Such work usually relies on the terminal sampling of tissues from large numbers of fish, at different time points, to adequately capture the response dynamic. However, underlying such studies is the assumption that individuals in a population will show similar immune responses (12). A step towards testing this assumption in salmonid species was the development of methods for repeated non-lethal sampling of blood from the same fish (12, 13). This approach can help elucidate immunological variation between individuals, either in their constitutive state or following immunization/disease challenge.

The current study tests the hypothesis that rainbow trout, a commercially important salmonid fish, shows a high degree of individual variation in response to immunization. By combining non-lethal blood sampling with high-throughput proteomics, we sought to document global changes in plasma protein abundance in individual fish following a common immunization protocol. The plasma proteome provides a strong index of biological status (14), including in fish (e.g. (15, 16)), and can be analysed using liquid chromatography-mass spectrometry (LC-MS) using small blood samples (e.g. (17)). Recent work has applied such approaches in salmonids (18), including to document changes in plasma protein abundance during sea water adaptation in rainbow trout (19). Such approaches require a comprehensive protein database for the target species and enable simultaneous identification and quantification of hundreds to thousands of proteins per experiment (20–23).

Our study design involved the immunization of a small cohort of rainbow trout with adjuvanted hen egg-white lysozyme (HEL) followed by sequential blood sampling of every individual at ten points across a 12-week timecourse.

After verifying an antigen-specific antibody response was generated by each individual, high-throughput LC-MS proteomics was used to monitor changes in plasma protein abundance in the same animals. Our analyses revealed the extent of individual versus population-wide responses to the same immunization protocol, as well as providing a detailed characterization of a salmonid plasma proteome.

## MATERIALS AND METHODS

### Immunization Protocol and Plasma Sampling

All procedures were conducted following the UK Home Office 'Animals and Scientific Procedures Act 1986; Amendment Regulations 2012' on animal care and use, with prior ethical approval from the University of Aberdeen's Animal Welfare and Ethical Review Body (AWERB). Rainbow trout ( $245.6 \pm 0.64$  g;  $27.6 \pm 0.19$  cm; mixed sex) were purchased from Almondbank Hatchery, Perthshire. The size of fish was chosen to ensure we could collect enough plasma per animal to perform technical optimization and data acquisition without exceeding 10% blood volume taken from any individual per month (13). The fish were maintained in 1-metre diameter tanks containing continuously circulating fresh water at  $14 \pm 1^\circ\text{C}$  in the aquarium facility of the School of Biological Sciences, University of Aberdeen, and were fed twice daily with standard commercial pellets (EWOS, Scotland). Fish were sedated with 2-phenoxyethanol (Sigma Aldrich) prior to any experimental procedure and daily monitoring undertaken for the duration of the experiment. Following PIT tagging fish were allowed 1 week to recover prior to immunization. Six fish were immunized intraperitoneally (IP) with hen egg-white lysozyme (HEL) emulsified in an equal volume of Complete Freund's Adjuvant (CFA), and one control fish was immunized with CFA only. HEL is a model antigen that has been used for immunization studies in several other vertebrate species (e.g. (24–26)). We can be certain that our study animals have no prior exposure to this antigen and know from other species the response to HEL is T-dependent and therefore a good proxy for the response we would wish to raise when developing an effective vaccine. Blood samples were drawn on days 0, 7, 14, 21, 28, 35, 42, 56, 70, and 84 post-immunization into syringes containing 100  $\mu\text{l}$  porcine heparin reconstituted to 1,000 U/ml in PBS. Plasma was aliquoted into low protein binding tubes, flash frozen, and stored at  $-80^\circ\text{C}$  prior to analysis.

### Measurement of Antigen-Specific IgM

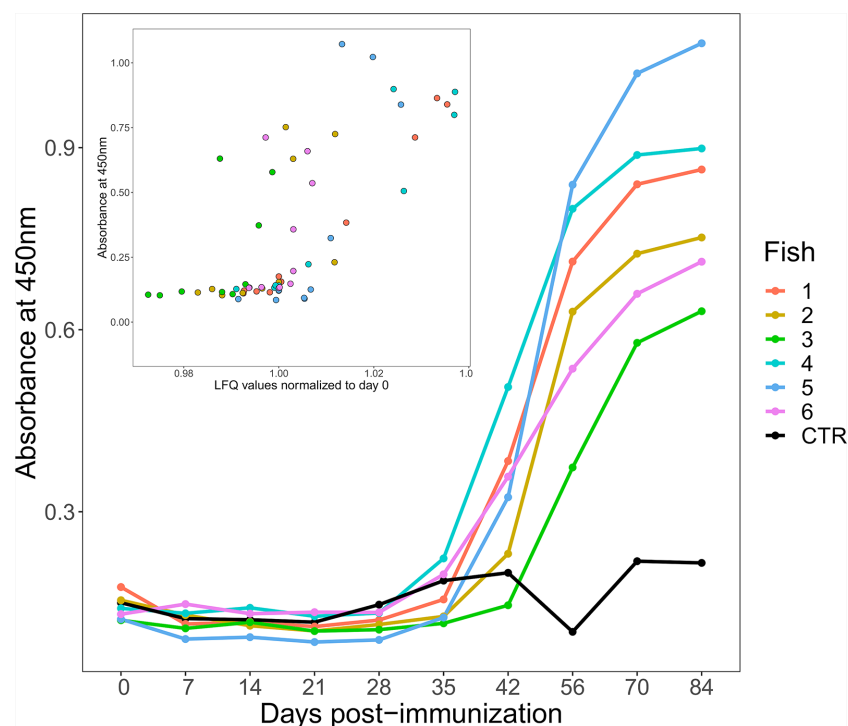
ELISA plates were coated for 48 h at  $4^\circ\text{C}$  with 100  $\mu\text{l}$ /well of HEL diluted in 0.05 M carbonate-bicarbonate (pH 9.6) buffer to 10  $\mu\text{g}/\text{ml}$ . Wells were emptied, washed once with 200  $\mu\text{l}$  PBS containing 0.05% Tween20 (PBST), and blocked with 200  $\mu\text{l}$  3% fat-free milk solution (MPBS) for 3 h at room temperature. Plasma samples were pre-diluted 1/10 in PBS then three-fold dilutions performed so that titration curves could be obtained for each sample. Wells were washed with PBST x3, plasma samples

added at 100  $\mu$ l/well then incubated overnight at 4°C. Wells were washed three times with PBST then 4C10 anti-salmonid IgM monoclonal supernatant (27), diluted 1:1 in PBS, was added at 100  $\mu$ l/well and the plates incubated overnight at 4°C. After three washes with PBST, anti-mouse HRP-conjugated antibody (Sigma Aldrich), diluted 1:1000 in MPBS, was added at 100  $\mu$ l/well and incubated for 2 h at room temperature. Following three further washes with PBST, binding was detected by the addition of 100  $\mu$ l/well TMB solution; the reaction was stopped by the addition of 50  $\mu$ l/well 1M H<sub>2</sub>SO<sub>4</sub>, and the plate read at 450 nm. Data were normalized across the plates, and titration curves plotted for each timepoint for each animal. To illustrate the data more easily we picked a dilution where signal was subsaturated for all samples, in this case the 1/270 dilution, and used this to plot absorbance at 450nm against time (as shown in **Figure 1**). Plasma from our CFA-only immunized animal was used as the negative control. The following technical controls were also performed; plasma samples on an irrelevant target (i.e. ELISA wells both coated and blocked with MPBS), PBS in place of plasma, and monoclonal antibodies raised against irrelevant targets in place of the 4C10 monoclonal.

## LC-MS Analysis

Plasma samples were prepared for proteomic analysis at Aberdeen Proteomics (University of Aberdeen) for the same n=6 trout individuals assayed by ELISA (as above). 1  $\mu$ l of plasma was

diluted with 99  $\mu$ l 50mM ammonium bicarbonate and proteins were reduced in 2mM dithiothreitol for 25 min at 60°C, S-alkylated in 4mM iodoacetamide for 30 min at 25°C in the dark, digested with porcine trypsin (sequencing grade, Promega) overnight at 37°C, then freeze-dried. The protein pellets were dissolved in 40  $\mu$ l 0.1% TFA and desalted using ZipTip  $\mu$ -C18 stage tips (Merck Millipore) following the manufacturer's instructions. The eluted peptide solutions were dried and dissolved in 10  $\mu$ l LC-MS loading solvent (98 parts UHQ water: 2 parts acetonitrile: 0.1 parts formic acid). Samples were transferred to a 96-well microtitre plate ready for injection into an UltiMate 3000 RSLCnano LC system (Thermo Scientific Dionex) coupled to a Q-Exactive Hybrid Quadrupole Orbitrap MS system (Thermo Scientific). The LC was configured for pre-concentration onto a PepMap RSLC C18 50  $\mu$ m x 25 cm column (Thermo Scientific P/N ES802) fitted to an EASY-Spray ion source (Thermo Scientific). The loading pump solvent was UHQ water: acetonitrile: formic acid (98: 2: 0.1) at a flow rate of 10  $\mu$ l/min; nano pump solvent A was UHQ water: formic acid (100: 0.1); nano pump solvent B was acetonitrile: UHQ water: formic acid (80: 20: 0.1). The LC gradient was programmed to increase the proportion of solvent B from 3% to 10% between 5 and 15 min, from 10% to 40% between 15 and 95 min, from 40% to 80% from 95 to 100 min and hold for 10 min before re-equilibration of the nano-column in 3% solvent B for 25 min. MS data acquisition was started at 10 min into the LC gradient, 5 min after switching the flow through the pre-column, and continued for a total of 100 min.



**FIGURE 1** | Changes in antigen-specific IgM levels following immunization of six rainbow trout (coloured lines) to HEL-CFA as compared to the CFA-only control fish (black line). The data shown are representative of duplicate technical replicates. The insert shows a positive correlation between antigen-specific and total IgM levels estimated by ELISA and label free proteomics, respectively (Spearman's Rho = 0.71, P < 0.001).

## Computational Proteomics

MS data were uploaded to MaxQuant v1.5.3.30 (28) together with a reference database containing all protein sequences ( $n=71,625$ ) predicted in the current rainbow trout RefSeq genome assembly (NCBI accession: GCA\_002163495.1) (29). Despite representing a comprehensive annotation of proteins, we noted a poor representation of immunoglobulin (Ig) M (IgM) proteins. As we were interested in quantifying total IgM in addition to antigen-specific IgM, we supplemented the reference database with 317 *O. mykiss* IgM proteins downloaded from the NCBI nr protein database. The Andromeda peptide search engine within MaxQuant (21) was used to match the mass spectra of all detected peptides against the reference protein database, using the label-free quantification method. Digestion type was set to “trypsin” and two missed cleavages were permitted to maximize peptide detection if tryptic digestion was incomplete. Variable modifications of methionine oxidation and N-terminal acetylation were allowed. To increase quantitative accuracy, the Fast LFQ option was not selected. “Match between runs” was used to maximize peptide detection; any features with retention times within 0.7 min and mass within the mass tolerance of peptides identified during previous runs (30) were assigned the same peptide identity. The false discovery rate was set at 0.01, using a target-decoy based search applied at both peptide and protein group levels, which calculated the probability that identified peptides and proteins were false hits (31).

The Andromeda platform within MaxQuant generated protein groups and majority protein groups (MPGs) comprising all proteins sharing at least 50% of their peptides (31). This additional filter increases confidence in the identifications, and only MPGs were considered in subsequent statistical analyses. Proteins identified as contaminants and false positives were removed. Hereafter, where possible we use the term protein in exchange of MPG, as it provides a more intuitive biological description. Protein abundance values for each sample were generated using the label-free quantification (LFQ) method (32).

## Statistical Analyses

Among 604 unique proteins initially identified by MaxQuant (Table S1), we retained those with abundance (LFQ) values in at least seven out of ten samples across the post-immunization timecourse for all six individuals (278 MPGs; Table S2). Our rationale for this cut-off was to filter the analysis to only consistently measured proteins, allowing reliable downstream inferences. After filtering, just 130 missing values remained in the dataset (0.78% of 16,680 data-points) across individuals. LFQ values for the remaining proteins were  $\log_2$ -transformed and missing values imputed using a random forest-based method (33). Subsequent statistical analyses were performed on transformed imputed values using Minitab v.19 and R v3.6.2.

Principal component analyses (PCA) were performed in R using the *prcomp* function and visualized in *ggplot2* v3.2.1 (34). One-way analysis of variance (ANOVA) was performed in Minitab on abundance levels for each of the 278 proteins separately, using sampling day as a fixed factor. The Anderson-Darling test was performed to verify that the residuals were normally distributed and

Levene's test used to assess homogeneity of variance. We recorded the  $R^2$  value for each analysis, defining the proportion of variance in protein abundance explained by sampling day. We classified proteins of interest as markers for population responses to immunization as those showing a Benjamini-Hochberg corrected P-value  $\leq 0.05$  (corrected at the level of 278 separate analyses), indicating an effect of sampling day. The relationship between P-value and  $R^2$  is shown in Figure S1, where a Benjamini-Hochberg corrected P-value  $\leq 0.05$  corresponds to  $R^2 > 0.34$ . The ANOVA data are provided in Table S3. For 41 proteins meeting the cut-off, differences in abundance across days were assessed using Tukey's test. Hierarchical clustering analysis was performed in PermutMatrix (35) using abundance data for the 41 proteins, in addition to the mean total IgM abundance (mean  $\log_2$  transformed imputed values for all fish, at each time point, for proteins in the dataset annotated as IgM) and antigen-specific Ig titres assessed by ELISA. Clustering and seriation were based on Pearson's correlation coefficient dissimilarity z-scores. The multiple-fragment heuristic seriation method was used with complete linkage (furthest neighbour) aggregation to obtain hierarchical clusters.

## Annotation of Trout Plasma Proteins

We used STRING (36) to annotate the 278 proteins in the filtered rainbow trout plasma proteome set on the basis of identifiable orthologues in human, testing for statistical enrichment in protein-protein interaction (PPI) networks, done using default parameters, and recording enrichment ( $P < 0.0001$ ) for Biological Processes within the Gene Ontology (37) and Reactome Pathway (38) frameworks (Table S4). We also used the GO FEAT (39) webserver to summarize the annotation of GO Biological Processes for the same protein set.

## Analysis of Duplicated Proteins

The common ancestor to salmonid fishes underwent a whole genome duplication (WGD) event (Ss4R) ~88–103 million years ago (40) and consequently, approximately half of the functional genes are found in duplicated pairs (41, 42). Duplicated proteins retained from Ss4R share identities ranging from ~75% to >99% (41). Previous LC-MS studies in salmonids have not tested how analysis platforms using the protein group approach handle the presence of duplicated proteins. We thus aimed to establish how MaxQuant organizes duplicated proteins into MPGs using the current dataset on the basis of protein similarity (Table 1; Figure S2; Tables S5, S6).

The analysis was performed using the unfiltered high-confidence 604 MPGs and compared the number of MPGs containing duplicated proteins (“scenario i”) with the number of duplicated proteins classified into unique MPGs (“scenario ii”). The analysis was informed by BLASTp searches (43) using one representative protein per MPG as the query versus all other proteins within the full set of MPGs. For each MPG, we checked if the proteins present were translated from distinct NCBI RefSeq genes (i.e. duplicated genes), as opposed to representing isoforms of the same gene. For MPGs solely comprised of proteins translated from the same gene, BLASTp was used to record any duplicated proteins within other MPGs (cut-off: >70%



**TABLE 1** | Comparison of amino acid sequence identity and shared peptide content between Ss4R and other duplicates classified into the same or different MPGs.

	% Similarity Mean	% Similarity S.D.	% Similarity Range	% Shared peptides Mean	% Shared peptides S.D.	% Shared peptides Range
<i>Duplicates in same MPG (Scenario i)</i>						
Ss4R duplicates	95.28	4.65	80.34 - 100	92.91	11.36	46.47 - 100
Other duplicates	94.66	7.17	73.21 - 100	91.12	12.93	53.13 - 100
<i>Duplicates in different MPGs (Scenario ii)</i>						
Ss4R duplicates	90.40	6.10	70.20 - 98.58	34.49	28.76	0 - 100
Other duplicates	92.16	5.89	77.34 - 98.86	43.65	33.43	0 - 100

identity/coverage). We used ClustalW sequence alignment (44) within BioEdit (v.7.0.5) (45) to align putative duplicated proteins within MPGs and between MPGs. The alignments were trimmed to the shortest protein present, and the mean pairwise identity between duplicated proteins was recorded. Chromosomal locations of genes encoding duplicated proteins were recorded to record duplications resulting from Ss4R (after (29, 42)). Finally, the number of detected peptides shared by duplicated proteins was recorded.

## Phylogenetic Analysis

Phylogenetic analyses were used to clarify the evolution of several proteins detected in the trout plasma proteome. For the analysis of apolipoprotein proteins we used human A-I, A-IV, and E protein sequences (NCBI accession numbers: AAA35545, AAA96731, and NP\_000032, respectively) as BLASTp queries (43) against the NCBI nr protein database and collected the highest scoring homologues ( $e$ -value  $<1e^{-10}$ ) from coelacanth (*Latimeria chalumnae*), spotted gar (*Lepisosteus oculatus*), zebrafish (*Danio rerio*), northern pike (*Esox lucius*), Atlantic salmon (*Salmo salar*) and rainbow trout. We also performed phylogenetic analysis for two plasma proteins with ‘uncharacterized’ annotations (XP\_021436350 and XP\_021423950). For this analysis, the trout proteins were used as the query for BLASTp searches against the following taxa in NCBI: invertebrates, jawless fish, Chondrichthyes, Teleostei, Amphibia, Reptilia, Aves, and Mammalia. Hits for these ‘uncharacterized’ proteins were filtered to proteins showing  $>40/30\%$  sequence coverage/identity to the query ( $e$ -value  $<1e^{-04}$ ). BLAST data are provided in **Table S7**.

For each analysis, sequence alignment was performed using Mafft v.7 with default parameters (46) and trees were built using the IQ-TREE maximum likelihood method (47, 48), which estimated and employed the best fitting amino acid substitution model (49). Ultra-fast bootstrapping (50) was used to generate branch support values. Consensus trees were visualised and rendered in Mega X (51). All used sequence alignments are given in **Supplementary Data 1**.

## RESULTS AND DISCUSSION

### Antigen-Specific and Total IgM Response

ELISA demonstrated that an antigen-specific IgM response was generated by all six fish injected with HEL-CFA, however the timing and magnitude of the response varied considerably between individuals (**Figure 1**). While antigen-specific IgM levels began to

increase in four of the six HEL-immunized fish between 28 and 35 days post-immunization, fish 2 and fish 3 did not show antigen-specific IgM binding appreciably above that of the CFA-only control fish even at day 42 (**Figure 1**). While all six HEL-immunized fish showed antigen-specific IgM levels significantly (3–5 fold) higher than the CFA-only control by day 84, there was an approximately 2-fold difference in ELISA signal between some of the antigen exposed individuals (**Figure 1**).

In mammals, primary antigen exposure causes a rapid increase in both total and antigen-specific IgM levels. This contrasts with sharks (i.e. cartilaginous fish), where total IgM levels remain relatively constant and it is only the monomeric antigen-specific portion that increases in concentration upon immunization (26). We were curious to establish if the HEL-specific IgM responses observed in trout were coupled to changes in total IgM, as in mammals, or were largely uncoupled, as in sharks. As the IgM proteins in our database were present as 11 distinct, closely related proteins, we took their average values at each time point to approximate the total IgM response. Antigen-specific IgM response and total IgM levels were significantly correlated for each of our 60 samples (Spearman’s Rho = 0.71,  $P < 0.001$ ; **Figure 1**).

### Overview of Trout Plasma Protein Functions

Biological processes and molecular pathways overrepresented among the 278 trout plasma proteins are shown in **Table S4**. These plasma proteins are highly biologically connected, showing 8.42 times more protein-protein interactions than expected by chance ( $P < 1.0e^{-16}$ ; visualized in **Figure S3**). The overrepresented Biological Processes (GO framework) and Reactome pathways reflect the large number of proteins from, or involved in the regulation of, the complement system (e.g. 24 annotated with Reactome term ‘complement cascade’,  $P = 1.83e^{-30}$ ), diverse immune functions (e.g. 53 annotated with Reactome term ‘immune system’,  $P = 8.88e^{-17}$ , 29 annotated with GO term ‘humoral immune response’,  $P = 4.88e^{-23}$ , and 23 annotated with GO term ‘adaptive immune response’,  $P = 1.52e^{-15}$ ), a range of blood, blood clotting/coagulation and platelet functions (e.g. 34 annotated with Reactome term ‘hemostasis’,  $P = 7.78e^{-19}$ ), in addition to metabolic functions (e.g. 5 annotated with Reactome term ‘gluconeogenesis’,  $P = 8.14e^{-05}$ ). GO FEAT analysis highlighted similar terms, but also emphasised representation of proteins involved in lipid transport and metabolism, largely apolipoproteins (see later section).

## Analysis of Duplicated Proteins

In LC-MS proteomics, it is common to collapse proteins that share identical or highly similar peptides into single protein groups. This method aims to capture protein isoforms derived from the same gene, but also creates potential for proteins translated from highly similar duplicated genes to be grouped together. Logically, this situation will arise most commonly for proteins sharing high similarity, such as those translated from gene duplicates retained from Ss4R (41, 42). On these grounds, we predicted that the classification of duplicated proteins into distinct MPGs or a common MPG by MaxQuant would be a product of paralogous sequence divergence.

Among the 604 MPGs available for analysis, 46.5% were comprised of proteins without identifiable duplicates in the dataset (**Figure S2**). Among the remaining 286 MPGs, 31% included duplicated proteins translated from distinct genes (“scenario i”), whereas for the other 69%, any duplicated proteins in the dataset were in unique MPGs (“scenario ii”) (**Tables S5, S6**). Under scenarios i and ii, ~70% and ~85% of the proteins were retained from the Ss4R event, respectively (**Figure S2**). On average, duplicated proteins classified into the same MPG shared higher amino acid sequence identity and a higher percentage of shared peptides than duplicated proteins classified into two or more MPGs (**Table 1**). Under both scenarios we observed a significant positive correlation between the proportion of shared peptides and % identity between duplicated proteins (**Figure S4**; Pearson’s  $R=0.62$  and  $0.70$ ;  $P=0.006$  and  $P<0.0001$  for scenario i and ii, respectively). MaxQuant classified many duplicated proteins sharing >95% amino acid level into distinct MPGs, despite sharing >50% common peptides (**Figure S4**). Overall, these data indicate that MaxQuant often distinguishes protein duplicates across a range of sequence divergence levels relevant to the detection of Ss4R duplicates (40, 41). However, the results advocate for a careful approach when drawing conclusions about duplicated proteins using the protein group concept in LC-MS proteomics, requiring a protein-by-protein investigation of the make-up of protein groups.

## Individual-Level Variation Dominates the Plasma Proteome

We used PCA to visualise groupings among the 60 sampled plasma proteomes according to fish individual and the ten post-immunization time points (**Figure 2**). We did this twice, firstly with the overall plasma protein abundance levels (**Figure 2A**), and secondly using the same values normalized to day 0 values (i.e. day 7/day 0, day 14/day 0, etc.) in order to remove the effect of variability in starting abundances, thus focusing on the response dynamic (**Figure 2B**). For the first analysis, PC1 and PC2 together explained around a third of the total variation and showed largely fish-specific groupings (**Figure 2A**). For the second analysis, a similar amount of total variation was explained by PC1 and PC2, and while there was a more substantial overlap among fish individuals (**Figure 2B**), this was not clearly explained by differences among days. Overall, a substantial proportion of total variation in our dataset is

explained by overarching differences in plasma protein abundance and responses among the six trout individuals.

## Proteins Showing Similar Population Responses to Immunization

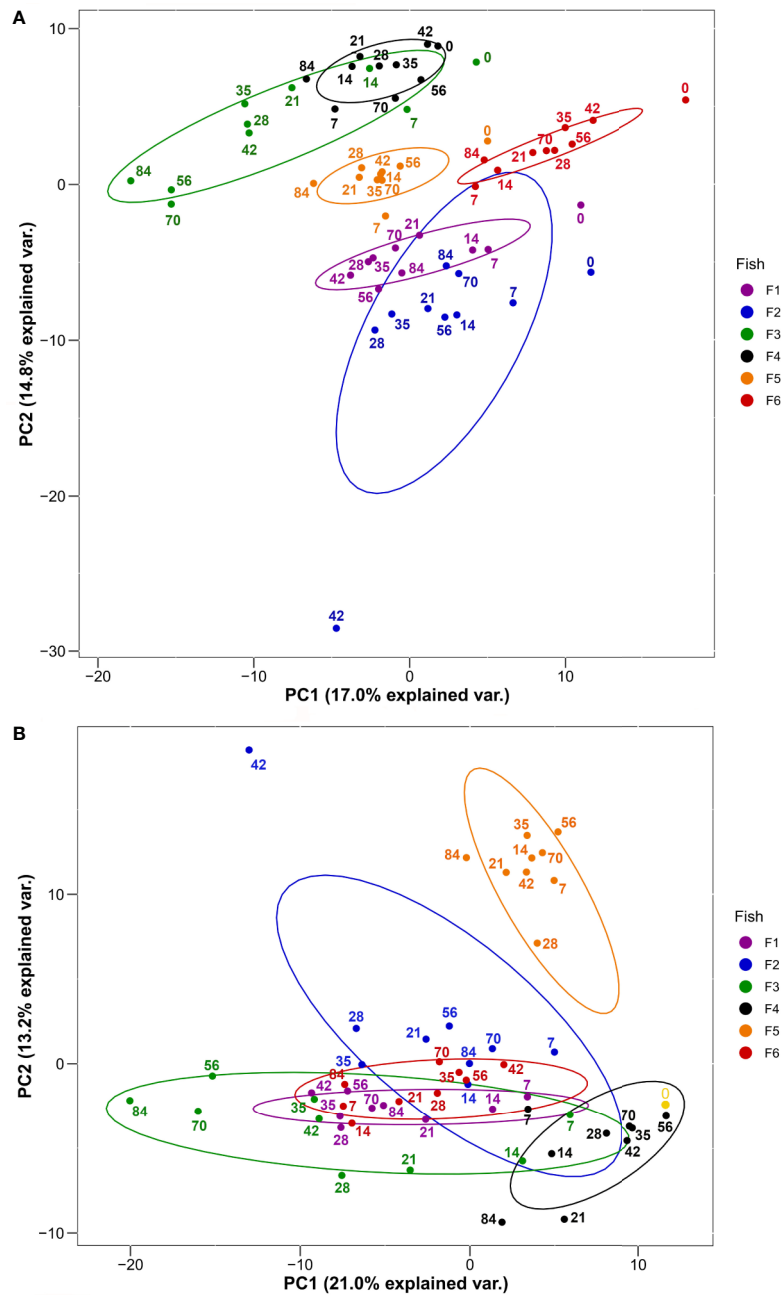
We next classified proteins according to the extent of variation in abundance levels explained by sampling day using one-way ANOVA (**Table S3**). We recorded  $R^2$  to describe the variance explained by differences among sampling days. The relationship between  $P$ -value and  $R^2$  is shown in **Figure S4**, which highlights the cut-off we employed (corrected  $P \leq 0.05$ ) to identify 41 proteins showing an overall population effect of sampling day. These proteins represent approximately 15% of all detected plasma proteins, and within our dataset are characterized by the most similar overall abundances within days, and most similar responses to immunization across the six individuals between days. We grouped these proteins according to commonalities in their average response to immunization using hierarchical clustering, also including mean total Ig and antigen-specific IgM levels (**Figure 3**). This revealed two major clades (“A” and “B”), separating proteins showing highest abundances during the early or late part of the timecourse (**Figure 3**). In the following sections, we outline the responses and characteristics of proteins grouped into five sub-clusters, two within clade A (clusters A1 and A2) and three within clade B (clusters B1, B2, and B3).

### Cluster A1: IgM and Neuropilin-2

Mean total IgM and antigen-specific IgM clustered closely, together with a single IgM heavy chain protein (**Figure 3**), which showed large variation between fish individuals, such that abundance levels were not significantly different between any time points (**Figure 4A**). The other protein in this cluster, neuropilin-2, showed a significantly higher abundance at day 56 compared to days 14–21 (**Figure 4B**). Neuropilin-2 is a transmembrane glycoprotein that is abundantly expressed in immune cells, and supports multiple immune functions, including antigen presentation (52, 53).

### Cluster A2: Diverse Molecules With Immune Functions; From Complement to Metabolic Enzymes

Members of the A2 cluster showed lowest abundance during the earliest days in the timecourse (**Figure 3**) and included complement system proteins annotated as C1q-like, C3, C6, and factor H (**Figure 5**). For instance, a C3 protein (XP\_021469459.1), which is a central component of the complement system, showed significantly higher levels on days 21–84 than day 0 (**Figure 5A**). Similarly, complement C6, part of the membrane attack complex (54), was significantly higher on days 28–70 than day 0 (**Figure 5B**). Similar abundance profile changes were observed for complement C1q-like protein 2 and two distinct factor H proteins (**Figure S5**). Several additional proteins with known immune functions were present in cluster A2, including C type lectin receptor B (**Figure 5C**), a pattern recognition receptor, precerebellin-like protein, a known acute

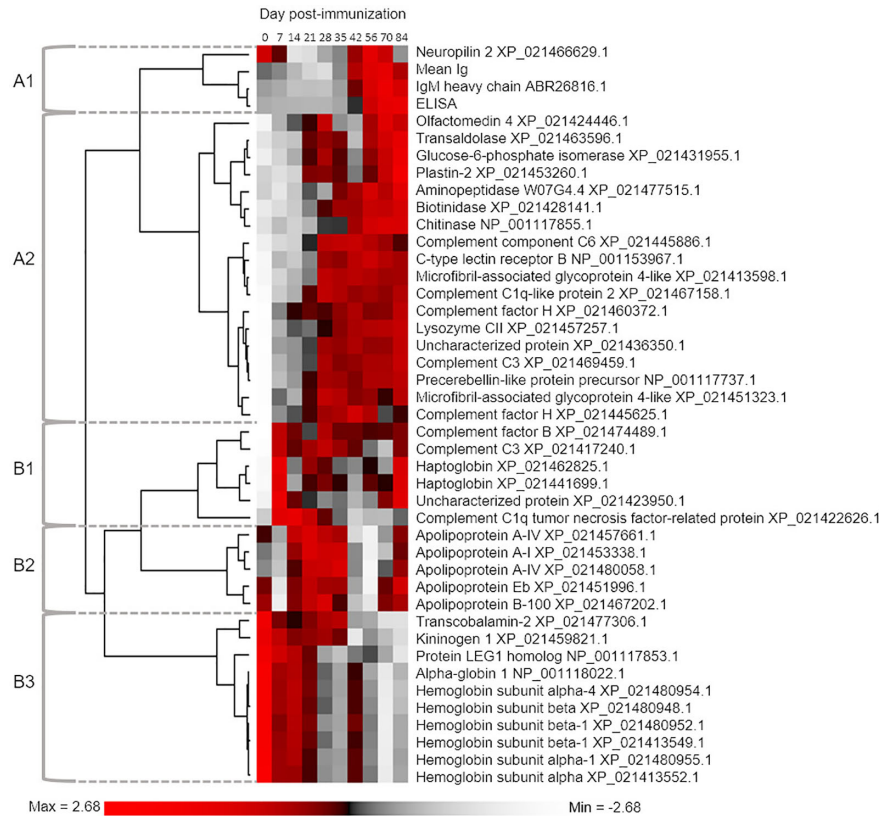


**FIGURE 2** | PCA of 278 plasma proteins derived from the six HEL-immunized fish shown in **Figure 1** using **(A)**  $\log_2$ -transformed imputed values and **(B)**  $\log_2$ -transformed imputed values normalized to Day 0 values. Normalized Day 0 value common to all fish is shown in yellow. Ellipses are 95% confidence intervals around the centroid. Sampling days are shown by numbering within the plots.

phase protein showing regional similarity to C1q (**Figure 5D**) (55, 56) and the antibacterial enzyme lysozyme (57) (**Figure 5E**). Two separate microfibril-associated glycoprotein 4 proteins also grouped within cluster A2 (**Figure S5**), and may act as soluble pathogen recognition molecules (58). Cluster A2 also included a protein annotated ‘uncharacterized’ (XP\_0214236350; detailed further below) that showed significantly higher abundance at days 21-84 compared to day 0 (**Figure 5**).

Within cluster A2, glucose-6-phosphate isomerase (GPI) levels increased steadily during the timecourse, and reached peak levels at day 84, which was significantly higher than days 0-42 (**Figure 6A**). While best known for its role in glycolysis and gluconeogenesis, in mammals a secreted form of GPI (“neuroleukin”) induces maturation of B cells into plasma cells, thereby increasing Ig production (59). Thus, the increasing levels of GPI may have been supporting the development of the





**FIGURE 3** | Hierarchical clustering analysis used to group proteins according to similar mean abundance profiles across the sampling timecourse. The 41 included proteins were defined by One-way ANOVA defining an overall population effect of sampling day (see **Table S3**). For comparison, we also included data for mean total Ig levels and antigen-specific IgM levels. The five defined clusters are referred to in the main text.

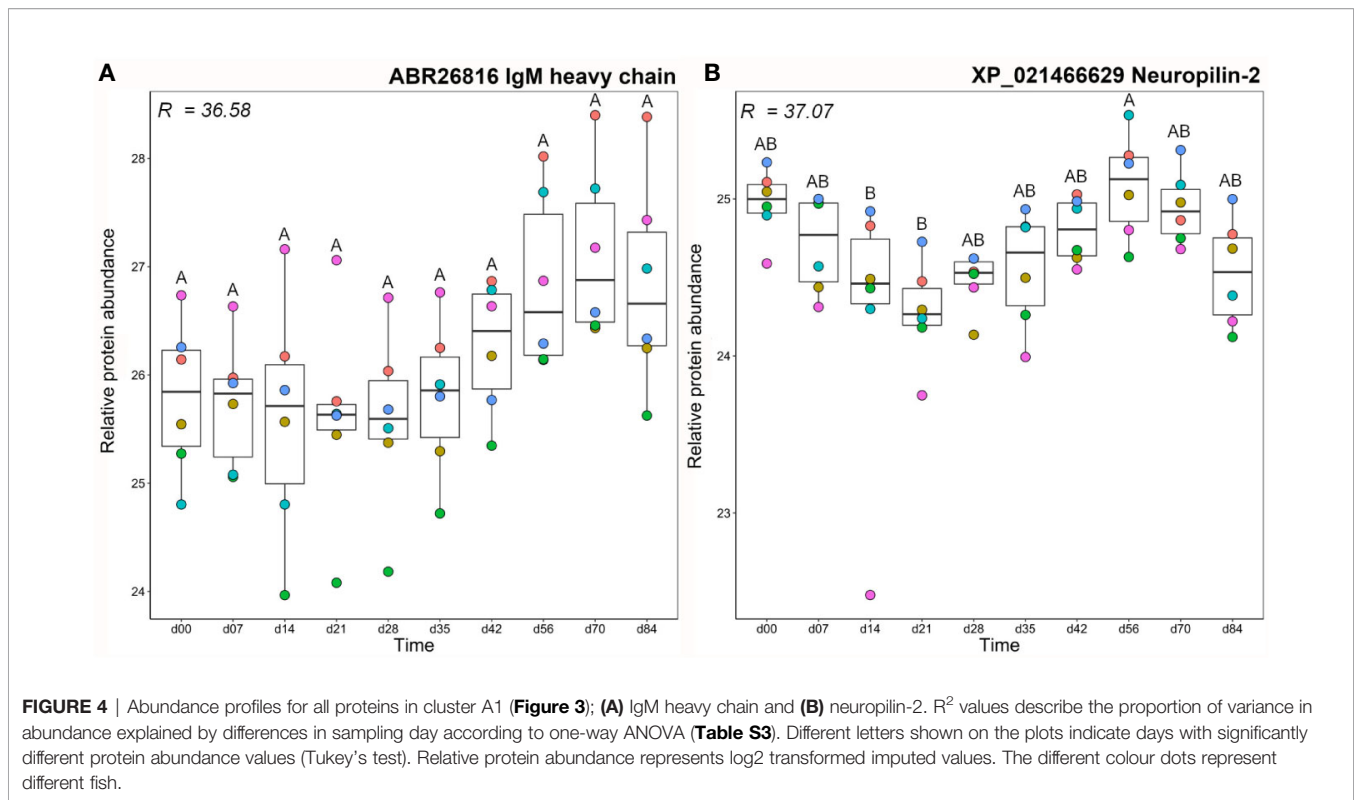
observed antigen-specific IgM response. Like GPI, plastin-2 (LCP1 or L-plastin) increased during the timecourse to show highest levels at day 84, with days 70 and 84 each showing significantly higher levels than days 0–14 (**Figure 6B**). Plastin-2 is an actin-regulating protein expressed specifically in leucocytes, and in humans is among the most abundant of all monocyte and T-cell proteins, with key immune functions including lymphocyte migration and T-cell activation (60, 61). The metabolic enzyme transaldolase showed a similar abundance profile (**Figure S5**), and is involved in the pentose phosphate pathway; its increasing levels during the time course may help support the high metabolic demands of an adaptive immune response (62).

Olfactomedin-4 showed a significant increase in abundance at days 28 and days 56–84 compared to day 0 (**Figure S5**). The secreted form of olfactomedin-4 can interact with many proteins, such as NOD1/NOD2, lectins, cadherins, and cathepsins, all of which regulate immune functions (63). The enzymes chitinase (**Figure 6C**), biotinidase (**Figure S5**), and aminopeptidase W07G4.4 (**Figure S5**), all increased steadily in abundance across the timecourse, with chitinase and biotinidase reaching significantly higher levels than day 0 by days 35–42. Chitinase has well established roles in innate immune protection,

providing antifungal and antihelminthic activity (64). While no direct immune functions have been attributed to biotinidase, this enzyme may support immune function indirectly, e.g. biotin is an essential cofactor for enzyme functioning in many cell types, including maturing and proliferating lymphocytes (65, 66). Upregulation of biotinidase should increase the supply of biotin, perhaps supporting the production of antigen-specific Ig in the adaptive phase.

### Cluster B1: Including Haptoglobins and Complement Proteins

Cluster B1 contained proteins that tended to show highest levels from day 7 onwards (**Figure 3**). This included two complement proteins, a second C3 molecule (XP\_021417240) along with factor B (Bf), an integral component of the alternative pathway C3 convertase (67). Both molecules showed a dramatic increase in abundance between day 0 and 7 and remained at elevated levels across the remainder of the timecourse (**Figures 7A, B**). Cluster B1 contained two haptoglobin (Hp) molecules, also showing large increases in abundance from day 0 to 7, consistent with Hp being an acute-phase protein, before dropping back to levels not significantly different from 0 between days 14–70, and increasing again on day 84



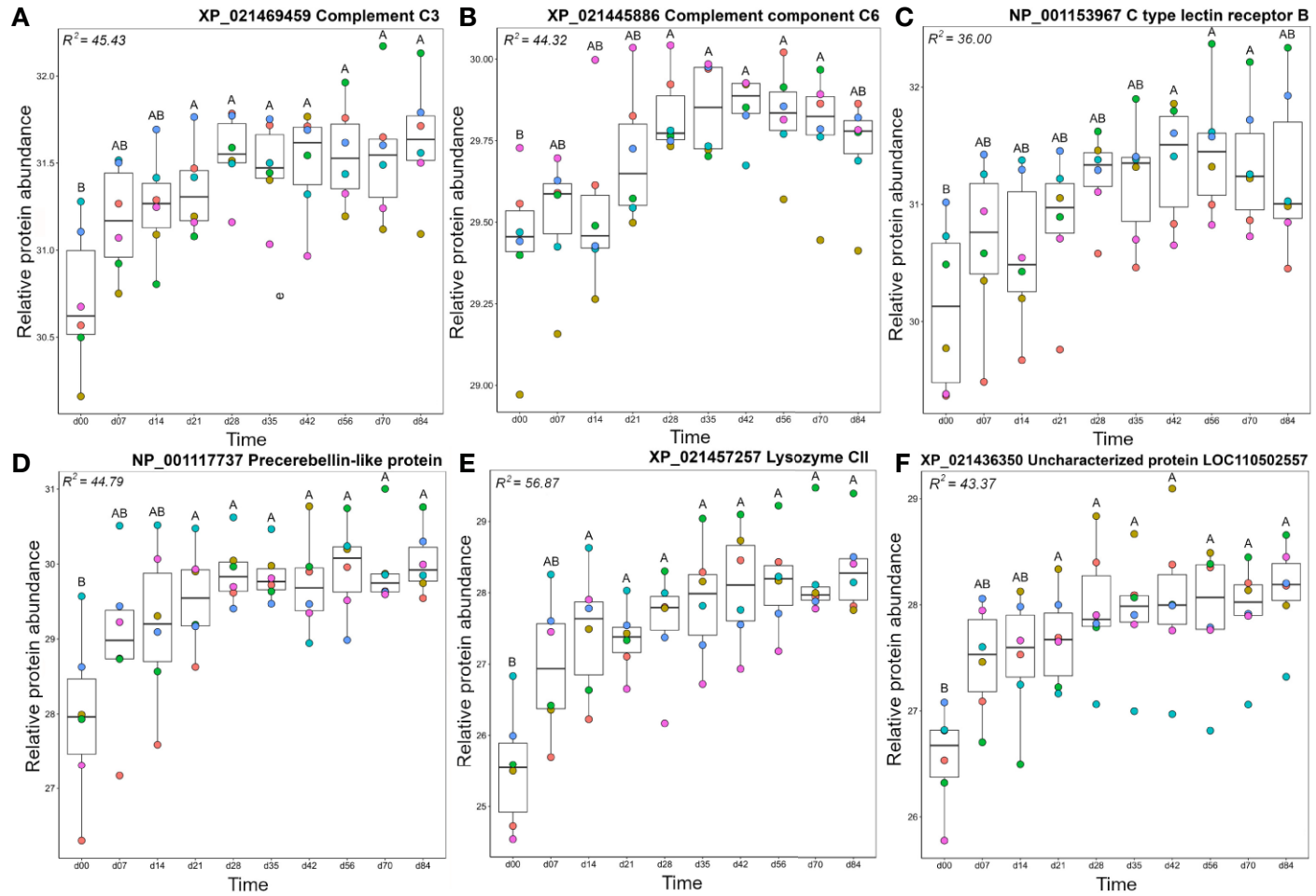
(Figures 7C, D). Hp is best known for sequestering haemoglobin released by haemolysis in mammals (68). However, it has recently been shown that Hp is a divergent member of the complement-activating MASP family (69) and possesses various immunoregulatory functions (70, 71). The adipokine C1q TNF-related protein (CTPR3) showed a similar mean profile, but with larger variation among individuals, and only showed a significant difference in abundance between days 14 and 42 (Figure 7E). Given the wide-ranging effects ascribed to CTPR3, including roles in metabolism and inflammation (reviewed in (72)), it is difficult to predict the consequences of these changes. A second uncharacterized protein (XP\_0211423950; detailed further below) was contained in cluster B1. This protein increased significantly in abundance between day 0 and day 7, dropped to a slightly lower level from days 14–70, then increased in abundance again at day 84 (Figure 7F), an abundance profile remarkably similar to the Hp proteins, with which it clustered (Figure 3).

## Cluster B2: Apolipoproteins

Cluster B2 was comprised of five apolipoproteins (Figure 3), among 17 unique apolipoproteins detected in our dataset (Table S2). Each of the five apolipoproteins showed related changes in abundance during the timecourse, but with distinct levels of individual variation within days (Figure 8). Two of these proteins showed among the highest  $R^2$  values reported in our dataset (Table S3; Figures 8A, B). Apolipoprotein A-I (XP\_021453338) showed highest levels at days 21–35 (significantly higher than days 0–7 and days 42–70)

and lowest levels at day 56 (significantly lower than all days except days 42 and 70) (Figure 8A), which was highly similar to apolipoprotein A-IV (Figure 8B). A distinct protein annotated as apolipoprotein A-I (XP\_021480058) showed a similar profile, but with greater individual variation within days, showing a significantly lower abundance at day 56 compared to days 21–35 (Figure 8C). Apolipoprotein B-100 was unique in showing a significant decrease from day 0 to day 7, before significantly increasing by day 21, and significantly decreasing again by day 56 (Figure 8D). Apolipoprotein Eb showed significantly lower abundance at day 56 compared to days 21–35 and day 84 (Figure 8E).

The similar annotations and abundance profiles of multiple apolipoprotein A family members led us to question their evolutionary relationships. A phylogenetic analysis was performed for all annotated apolipoprotein A proteins in our dataset, within a broader background of orthologs from additional vertebrate species (Figure 9). Note, we included the apolipoprotein E family in this analysis as the sister group to the apolipoprotein A family, but excluded apolipoprotein B molecules, which are more evolutionarily divergent (73). This analysis provides a different, and more well supported evolutionary history of apolipoprotein AI and IV than existing scenarios (73). Specifically, our tree strongly supports a scenario where the ancestor to ray-finned fish and lobe-finned fish had separate genes encoding three distinct proteins related to apolipoprotein AI and IV, all of which are retained in coelacanth (Figure 9). Included among these three proteins, humans retain apolipoprotein AI and IV while ray-finned fish



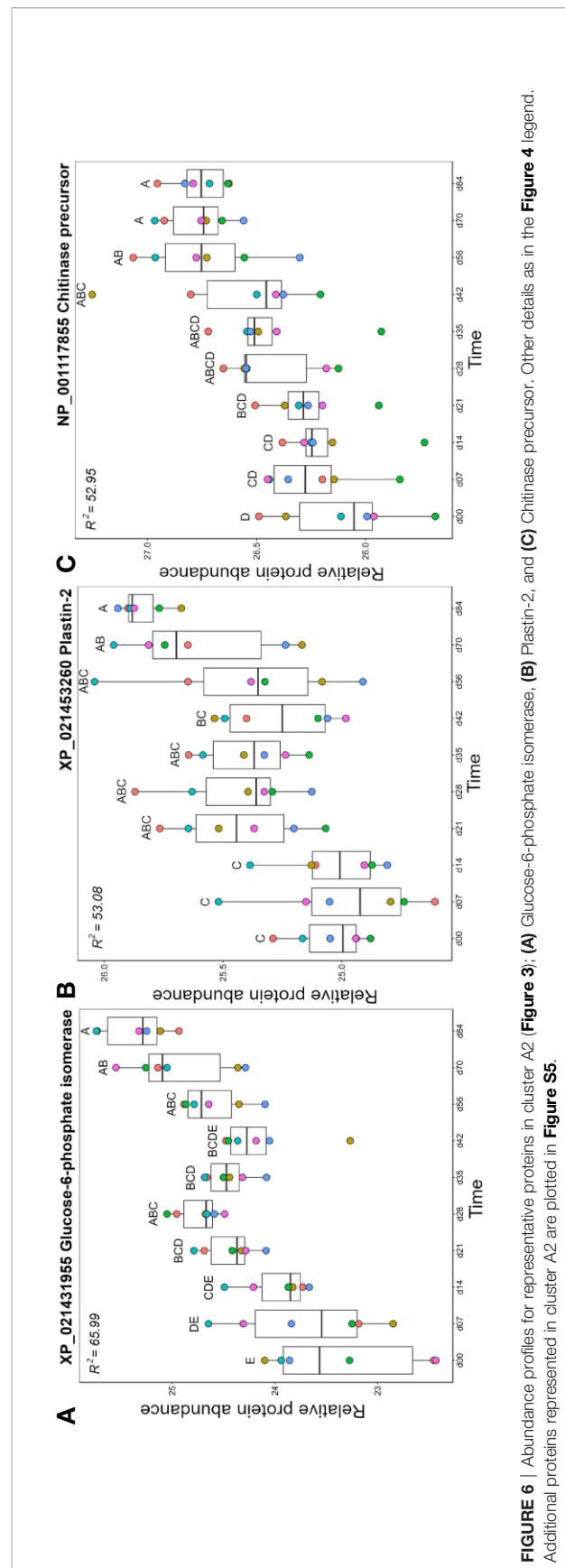
**FIGURE 5** | Abundance profiles for representative proteins in cluster A2 (**Figure 3**); **(A)** complement C3, **(B)** complement C6, **(C)** C type lectin receptor B, **(D)** precerebellin-like protein, **(E)** Lysozyme CII, and **(F)** uncharacterized protein XP\_021436350. Other details as in the **Figure 4** legend. Additional proteins represented in cluster A2 are plotted in **Figure S5**.

only retain orthologues of AI (**Figure 9**). However, ray-finned fish and coelacanth retain a distinct paralogue of apolipoprotein AI, absent in humans (**Figure 9**). Three rainbow trout representatives of this clade are grouped within Cluster B2, with two representing salmonid-specific duplicates (i.e. XP\_021453338 and XP\_021457661) (**Figure 9**) showing highly correlated abundance changes (**Figure 3**). The third protein (XP\_021480058) is a more ancient paralogue that arose in the common ancestor to salmonids and zebrafish after the split from spotted gar, which is consistent with the teleost-specific WGD event (74). The current annotation of apolipoprotein A-like proteins in salmonids is thus not supported by phylogenetic analysis, and future work might expand the phylogenetic analysis of the apolipoprotein A family to achieve an evolutionarily-supported nomenclature.

When interpreting the changes in abundance of apolipoproteins in our data, it is important to note that while classically associated with lipid transport and metabolism (75) these molecules also modulate innate and adaptive immune processes (reviewed in (76)). Recent data from amphioxus (77) suggest at least some of these immune roles are evolutionarily ancient. Apolipoprotein B-100 has been shown to inhibit *Staphylococcus aureus* colonization (78), while class A apolipoproteins can bind and neutralize bacterial LPS (79) and have bactericidal (80–82) and viricidal (83) activity in teleosts, including salmonids. They also kill pathogens indirectly, acting as immunostimulants to enhance respiratory burst responses in innate immune cells (84). Class A and E apolipoproteins also modulate many adaptive immune processes. Generally these immune functions tend towards regulation, e.g. inhibiting the production of pro-inflammatory cytokines by disrupting contact between stimulated T cells and monocytes (85), stimulating regulatory T cell expansion (86), and inhibiting T cell activation through the downregulation of antigen-presentation by dendritic cells (87). Thus, the complex changes in abundance observed across the timecourse may reflect the multifaceted regulatory roles of rainbow trout apolipoproteins at different phases of the immune response.

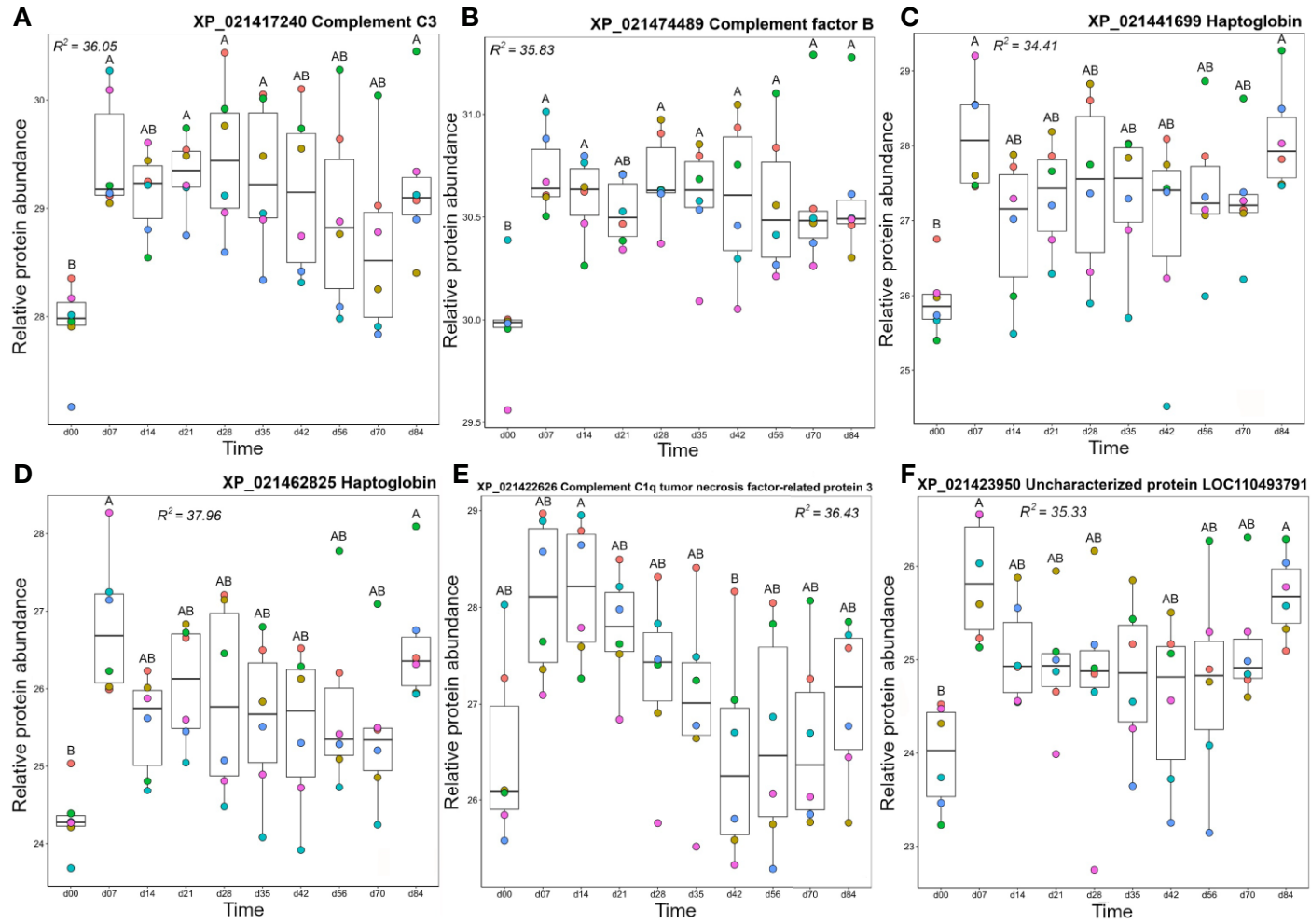
### Cluster B3: Including Haemoglobin and Proteins With Blood Functions

Proteins in cluster B3 showed highest levels on day 0, then decreased in abundance over the timecourse (**Figure 10, Figure S6**). Seven haemoglobin (Hb) proteins fell within this cluster and reached lowest levels on day 70. The presence of Hb in plasma is due to red blood cell (RBC) lysis, occurring biologically *in vivo* or during sample processing *ex vivo*. The drop in abundance of Hb subunits may simply reflect improvement in our sampling and processing technique over the experiment, however, there are several biological processes that could cause or contribute to this drop. For example, repeated blood sampling could reduce RBC counts and consequently reduce free Hb. However, Collet and colleagues (12) recorded no significant drop in haematocrit using the same approach with more regular bleeding (taken every 4 days). While we did not measure haematocrit, the fish in our study were sampled every 7 days using the same volume criteria

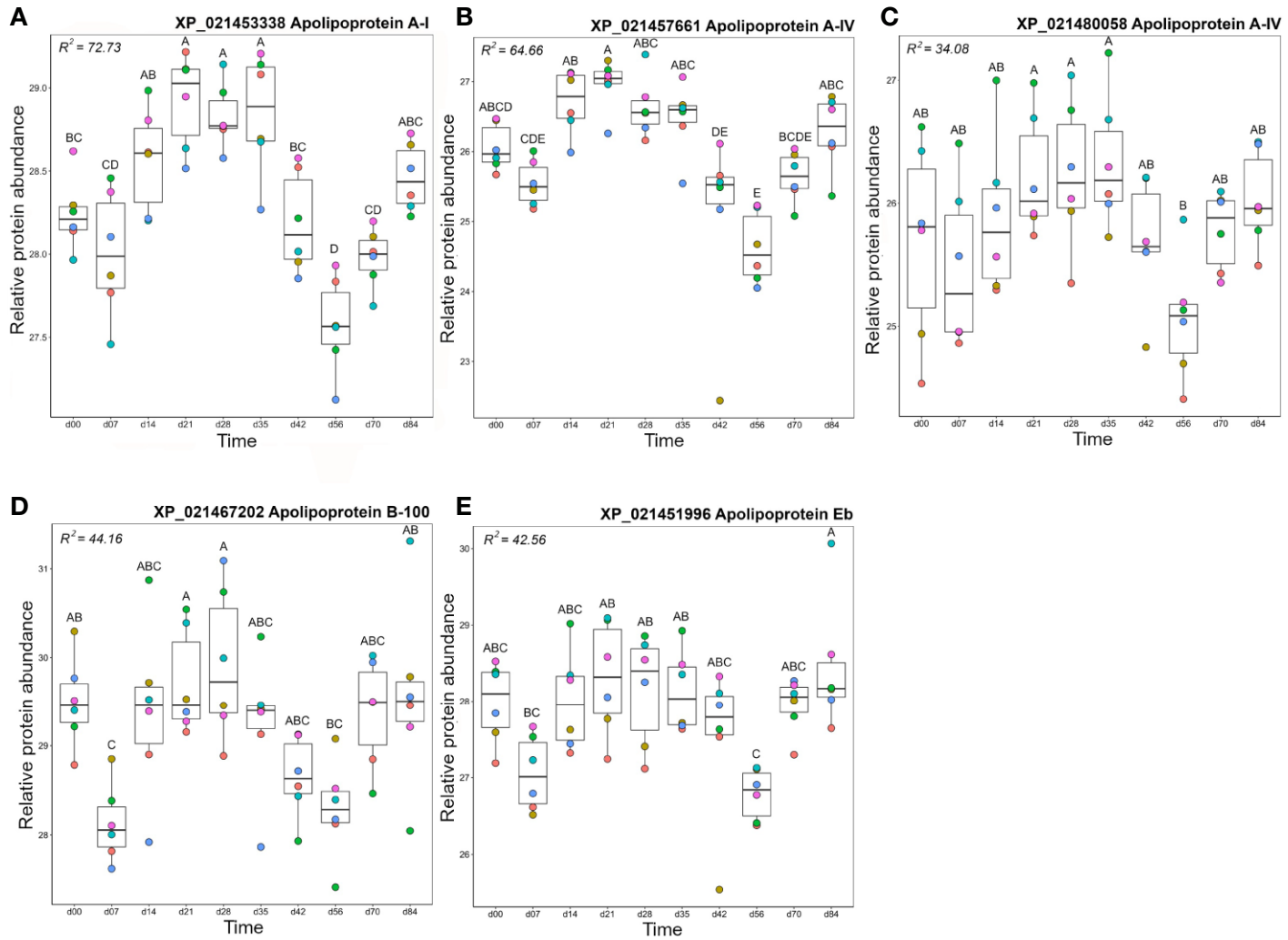


**FIGURE 6** | Abundance profiles for representative proteins in cluster A2 (**Figure 3**): **(A)** Glucose-6-phosphate isomerase, **(B)** Plastin-2, and **(C)** Chitinase precursor. Other details as in the **Figure 4** legend. Additional proteins represented in cluster A2 are plotted in **Figure S5**.

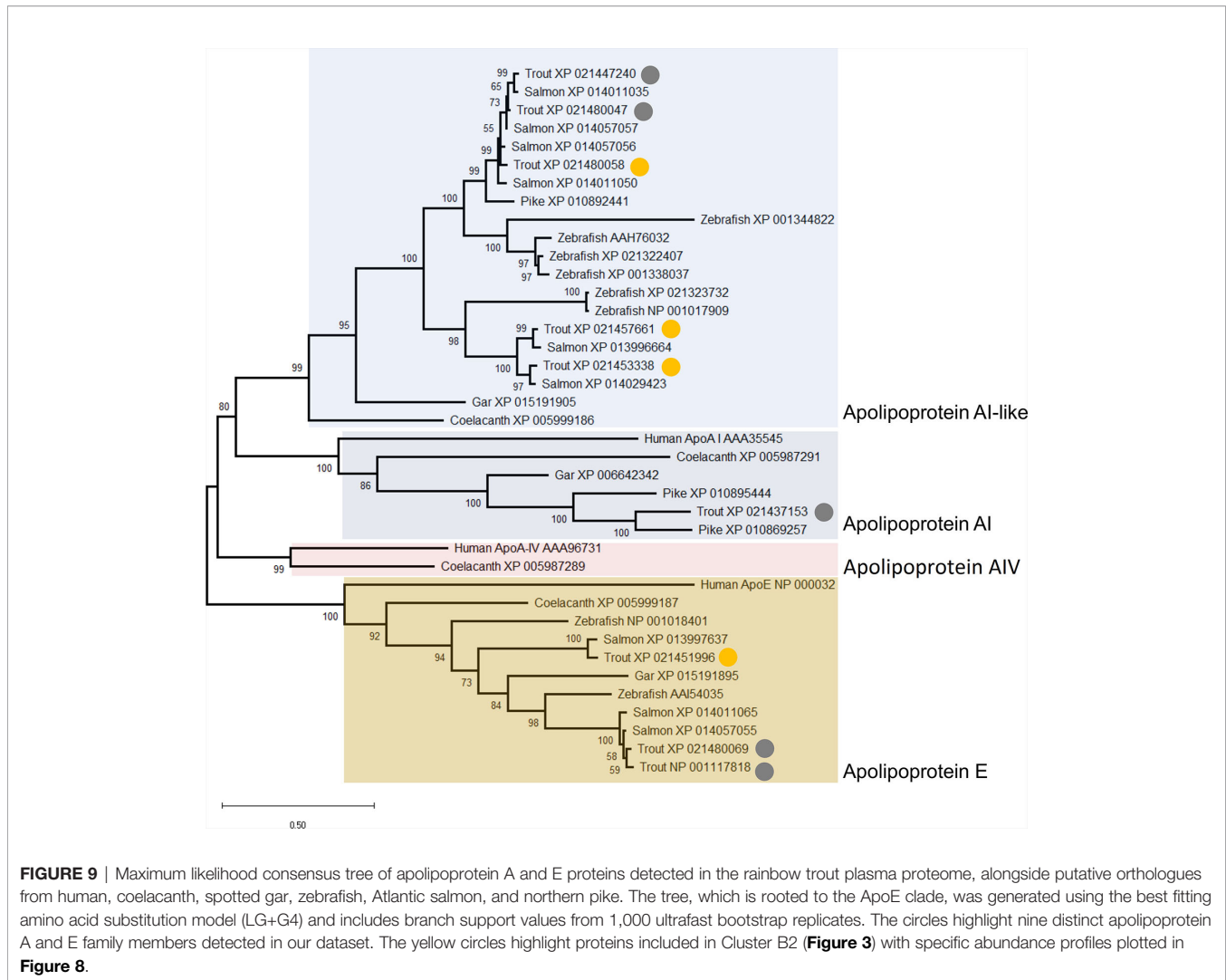




**FIGURE 7** | Abundance profiles for all proteins in Cluster B1 (**Figure 3**); **(A)** complement C3, **(B)** complement factor B, **(C)** haptoglobin, **(D)** haptoglobin, **(E)** complement C1q tumor necrosis factor-related protein 3, and **(F)** uncharacterized protein XP\_021423950. Other details as in the **Figure 4** legend.



**FIGURE 8** | Abundance profiles for all proteins in Cluster B2 (**Figure 3**); **(A)** apolipoprotein A-I, **(B)** apolipoprotein A-IV, **(C)** apolipoprotein A-IV, **(D)** Apolipoprotein B-100, and **(E)** apolipoprotein Eb. Other details as in the **Figure 4** legend.



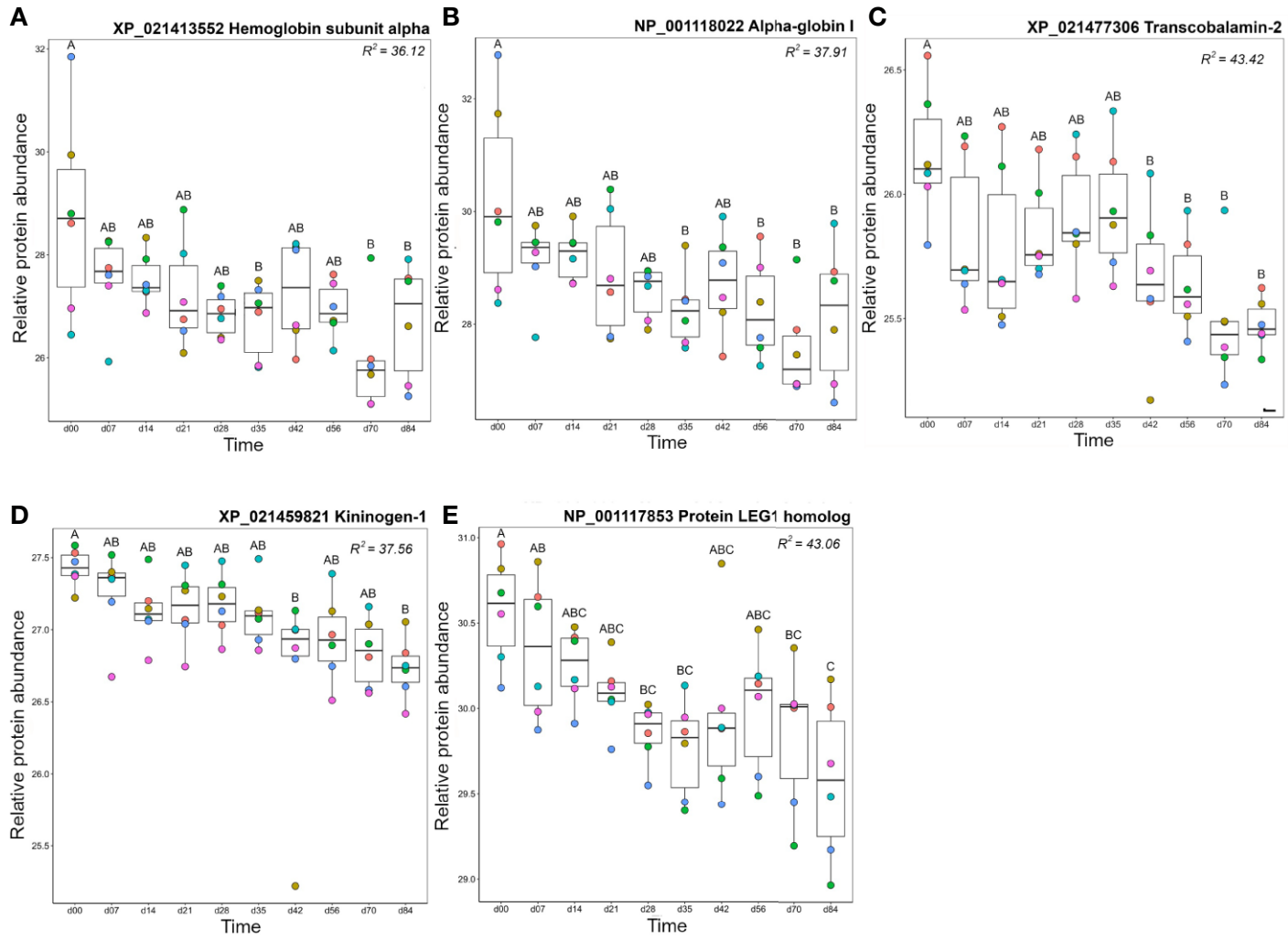
(<10% blood volume from each animal in any 4-week period). Thus, it would be surprising if reduced Hb was caused by reduced haematocrit. Replacement of removed RBCs with newer, more robust, cells could partly explain the drop in free Hb. Finally, there is known to be a slow turn-over of RBCs *in vivo* due to formation of the alternative pathway C3 convertase (formed from fragments of C3 and Bf) of the complement system on their surface. It is worth noting that free Hb levels drop around the time that the levels of one C3 and Bf decrease, while the two molecules of factor H, a regulatory protein which accelerates the decay of the C3 convertase and thus would protect host RBCs from lysis, show an increase. Further study is required to ascertain which, if any, of these mechanisms explain our data.

Transcobalamin-2, which is required to bind and transport vitamin B12 for red blood cell production (88, 89), also decreased over the timecourse, with highest abundance levels between day 0 and day 35, dropping at day 42 and remaining at this level until day 84 (Figure 10C). Kininogen-1 is a multi-functional protein with a major role in blood clotting and blood pressure regulation

but is also a mediator of immune inflammation (90). Kininogen-1 levels decreased steadily over the experimental timecourse such that there was a significant drop between day 0 and day 84 (Figure 10D). Protein LEG1 homolog, a structural protein necessary for liver development and function (91), also decreased in abundance over the study timecourse, showing lower levels at days 70–84 than at day 0 (Figure 10E).

### Evolutionary Conservation of “Uncharacterized” Trout Plasma Proteins

The presence of two proteins annotated as ‘uncharacterized’ within the group of 41 proteins included in Figure 3 (Figures 5F, 8F), led us to question their annotations and evolutionary origins. We determined if these molecules show homology to proteins in other taxa using BLASTp (Table S7). Despite not being detected in any mammal, both proteins are conserved across multiple vertebrate classes (Table S7). The protein in cluster A2 (XP\_021436350) is a 22 kDa cysteine-rich glycoprotein containing no known conserved domains,



**FIGURE 10** | Proteins in Cluster B3 (Figure 3); (A) hemoglobin subunit alpha, (B) alpha-globin I, (C) transcobalamin-2, (D) kininogen-1, and (E) protein LEG1 homolog. Other details are as in the Figure 4 legend.



conserved in teleosts, amphibians and reptiles (**Figure 11A**). The protein in cluster B1 (XP\_021423950) is a 24 kDa cysteine-rich protein with no known conserved domains, and conserved in cartilaginous fish, birds, and teleosts (**Figure 11B**). The identification of conserved uncharacterized proteins showing temporal changes in abundance highly correlated to known immune molecules highlights a strength of the “discovery” approach enabled by LC-MS proteomics; specifically, the scope for revealing novel molecules with putative immune functions, which clearly warrant further attention.

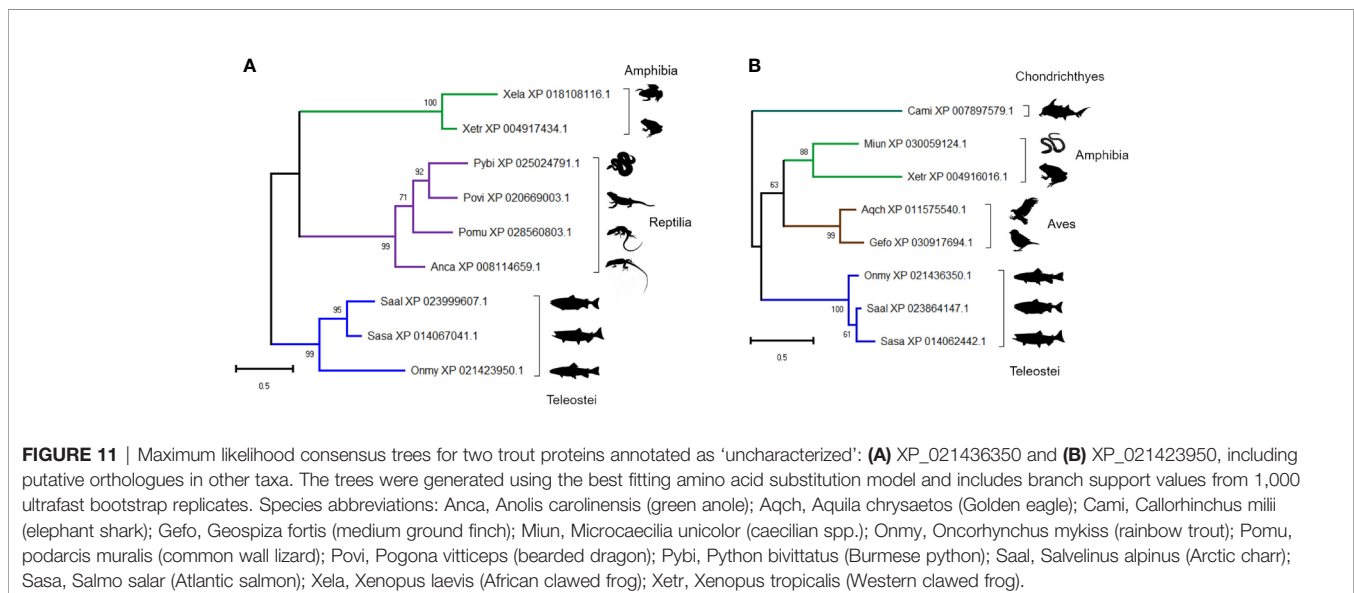
## CONCLUSIONS, PERSPECTIVES, AND CAVEATS

This is the first study to apply label-free proteomics to quantify long-term changes in plasma protein abundance in a salmonid fish following immunization, capturing different phases in the immune response. By keeping samples from each individual separate, we were able to explore the variation in humoral response between individuals as well as population-level responses. It is well-established that vaccination does not induce comparable protective responses in individual fish (92–94), however the extent of individual variation we observed here was nevertheless surprising. For example, although all fish produced antigen-specific IgM, the kinetics and magnitude of the response varied markedly between individuals. Importantly, now we have proved antigen-specific IgM levels as measured by antigen-binding ELISA correlate highly with total IgM as measured by LC-MS/MS, future studies would require much smaller plasma volumes (i.e. only 1  $\mu$ l of plasma is required for LC-MS analysis compared to  $\geq 10$   $\mu$ l for ELISA) thus permitting study of immune responses in much smaller fish (e.g. fingerlings of usual vaccination size).

Mirroring the data on the IgM response, we found that the overall plasma proteome response to HEL immunization was

predominantly explained by differences between fish individuals. Due to the small number of individuals in our study we did not attempt to explore the factors contributing to this variation in response. However, given the importance of consistent, population-wide protection to the eventual success of aquaculture vaccines, this is certainly something that should be explored in future studies. The power of our study comes from our ability to compare each time point from an individual animal to its own day 0 sample. We cannot rule out that some changes in protein abundance are linked to the repeated sampling protocol rather than immune challenge, the globin proteins being a possible example. However, it seems unlikely this would be the case for all the proteins showing similar abundance profiles across fish individuals, especially as many have previously established immune functions. Following additional validation, at least some of these molecules may provide useful new immune biomarkers that can be utilized in future vaccine trials in salmonids. Of note is our discovery of two uncharacterized plasma proteins with putative immune functions. Our phylogenetic analyses indicate these proteins are absent from placental mammals, including humans and mice, so likely would not have been identified using standard comparative approaches. It will be interesting to discover what role these new molecules play in immune protection in trout and, where present, other species. As yet we are unable to correlate any specific proteins with high production of antigen-specific antibody, however, as data from other proteomic studies is accrued and new markers are utilized, we are confident such information will eventually be forthcoming.

Finally, our study highlights how LC-MS is an enabling technology for the study of non-mammalian species where large sequence datasets are often available, but species-specific monoclonal antibodies are in short supply. Indeed, the power of LC-MS is illustrated by our ability to differentiate proteins with high amino acid identity, such as those expressed from recently



duplicated genes. These may have different functional roles/response kinetics but would be difficult to raise discriminatory antibodies against. Even if such antibodies were available, the research effort and sample volumes required to monitor abundance changes of >250 plasma proteins renders the use of classical methods such as ELISA or Western blots impractical. However, we also recognize there is room to further improve our approach; for example, although we detected over 600 unique proteins in our samples, only 278 proteins were detected in at least seven out of ten samples for all six individuals and thus retained following filtering. This issue could perhaps be overcome through application of an isobaric labelling strategy, whereby the peptides generated following trypsin treatment of each sample are tagged with chemical groups of identical mass but differing in their distribution of stable heavy isotopes. Tagged samples are pooled then analyzed simultaneously by MS. While tagging kits can be costly, such strategies should reduce the number of missing observations and consequentially improve the number of unique proteins retained for subsequent analyses. Alternatively, another recently developed label free MS method called sequential window acquisition of all theoretical mass spectra (SWATH) can be used to increase precision across the detectable proteome [e.g. (95)]. The application of such methods would also facilitate the detection of important but low abundance proteins, such as cytokines, that were not picked up during this study. Even with such limitations, the in-depth information obtained *via* LC-MS studies such as ours will undoubtedly improve our understanding of fish immune responses and the immunological variation between individuals, hopefully accelerating the testing of new aquaculture vaccines and improved administration methods.

## DATA AVAILABILITY STATEMENT

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/**Supplementary Material**.

## REFERENCES

- Crane M, Hyatt A. Viruses of fish: an overview of significant pathogens. *Viruses* (2011) 3:2025–46. doi: 10.3390/v3112025
- Centers for Disease Control and Prevention (CDC). *A CDC framework for preventing infectious diseases: Sustaining the essentials and innovating for the future* (2011). Available at: <https://www.cdc.gov/ddid/docs/ID-Framework.pdf> (Accessed May 17, 2020).
- Plant KP, Lapatra SE. Advances in fish vaccine delivery. *Dev Comp Immunol* (2011) 35:1256–62. doi: 10.1016/j.dci.2011.03.007
- Gudding R, Van Muiswinkel WB. A history of fish vaccination: science-based disease prevention in aquaculture. *Fish Shellfish Immun* (2013) 35:1683–8. doi: 10.1016/j.fsi.2013.09.031
- Brodin P, Jovic V, Gao T, Bhattacharya S, Lopez Angel CJ, Furman D, et al. Variation in the human immune system is largely driven by non-heritable influences. *Cell* (2015) 160:37–47. doi: 10.1016/j.cell.2014.12.020
- ter Horst R, Jaeger M, Smeekens SP, Oosting M, Swertz MA, Li Y, et al. Host and environmental factors influencing individual human cytokine responses. *Cell* (2016) 167:1111–24. doi: 10.1016/j.cell.2016.10.018
- Stentiford GD, Sritunyaluksana K, Flegel TW, Williams BAP, Withyachumnarnkul B, Itsathitphaisarn O, et al. New paradigms to help solve the global aquaculture disease crisis. *PLoS Pathog* (2017) 13:e1006160. doi: 10.1371/journal.ppat.1006160
- Ballesteros NA, Rodriguez Saint-Jean S, Perez-Prieto SII. Food pellets as an effective delivery method for a DNA vaccine against infectious pancreatic necrosis virus in rainbow trout (*Oncorhynchus mykiss*, Walbaum), Fish. Shellfish. *Immunol* (2014) 37:220–8. doi: 10.1016/j.fsi.2014.02.003
- Hoare R, Jung S-J, Ngo TPH, Bartie KL, Thompson KD, Adams A. Efficacy of a polyvalent injectable vaccine against *Flavobacterium psychrophilum* administered to rainbow trout (*Oncorhynchus mykiss* L.). *J Fish Dis* (2018) 42:229–36. doi: 10.1111/jfd.12919
- Skov J, Chettri JK, Jaafar RM, Kania PW, Dalgaard I, Buchmann K. Effects of soluble immunostimulants on mucosal immune responses in rainbow trout immersion-vaccinated against *Yersinia ruckeri*. *Aquaculture* (2018) 492:237–46. doi: 10.1016/j.aquaculture.2018.04.011
- Wangkahart E, Secombes CJ, Wang T. Dissecting the immune pathways stimulated following injection vaccination of rainbow trout (*Oncorhynchus mykiss*) against enteric redmouth disease (ERM). *Fish Shellfish Immun* (2019) 85:18–30. doi: 10.1016/j.fsi.2017.07.056

## ETHICS STATEMENT

The animal study was reviewed and approved by UK home office and University of Aberdeen's Animal Welfare and Ethical Review Body (AWERB).

## AUTHOR CONTRIBUTIONS

Study conception and design: DM and HD. Animal work: MM and HD. Proteomics lab work: DS. Proteomic data analysis: FB, DC, AD. Data interpretation: FB, DM, and HD. Drafted figures and tables: FB and DM. Drafted manuscript: FB, DM, and HD. All authors contributed to the article and approved the submitted version.

## FUNDING

This work was supported by the Biotechnology and Biological Sciences Research Council (BBSRC) grant numbers: BB/M010996/1, BB/M026345/1, BBS/E/D/20002174, and BBS/E/D/10002071.

## ACKNOWLEDGMENTS

Our thanks to Prof. Chris Secombes (University of Aberdeen) for the 4C10 anti-salmonid IgM mAb used in our ELISAs and for his valuable intellectual contributions during the planning of this project. We also gratefully acknowledge the supervisory support given by Prof. Sam Martin (University of Aberdeen) to FB.

## SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fimmu.2020.581070/full#supplementary-material>

12. Collet B, Urquhart K, Monte M, Collins C, Perez SG, Secombes CJ, et al. Individual monitoring of immune response in Atlantic salmon *Salmo salar* following experimental infection with Infectious Salmon Anaemia Virus (ISAV). *PLoS One* (2015) 10:e0137767. doi: 10.1371/journal.pone.0137767
13. Monte M, Urquhart K, Secombes CJ, Collet B. Individual monitoring of immune responses in rainbow trout after cohabitation and intraperitoneal injection challenge with *Yersinia ruckeri*. *Fish Shellfish Immun* (2016) 55:469–78. doi: 10.1016/j.fsi.2016.05.041
14. Anderson NL, Anderson NG. The Human Plasma Proteome: History, character, and diagnostic prospects. *Mol Cell Proteomics* (2002) 1:845–67. doi: 10.1074/mcp.R200007-MCP200
15. Veiseth-Kent E, Grove H, Færgestad EM, Fjæra SO. Changes in muscle and blood plasma proteomes of Atlantic salmon (*Salmo salar*) induced by crowding. *Aquaculture* (2010) 309:272–9. doi: 10.1016/j.aquaculture.2010.09.028
16. Braceland M, Bickerdike R, Tinsley J, Cockerill D, McLoughlin MF, Graham DA, et al. The serum proteome of Atlantic salmon, *Salmo salar*, during pancreas disease (PD) following infection with salmonid alphavirus subtype 3 (SAV3). *J Proteomics* (2013) 94:423–36. doi: 10.1016/j.jprot.2013.10.016
17. Babaei F, Ramalingam R, Tavendale A, Liang Y, Yan LSK, Ajuh P, et al. Novel blood collection method allows plasma proteome analysis from single zebrafish. *J Proteome Res* (2013) 12:1580–90. doi: 10.1021/pr3009226
18. Nynja J, Arnold G, Fröhlich T, Ciereszko A. Proteomic identification of rainbow trout blood plasma proteins and their relationship to seminal plasma proteins. *Proteomics* (2017) 17:1600460. doi: 10.1002/pmic.201600460
19. Morro B, Doherty MK, Balseiro P, Handeland SO, MacKenzie S, Sveler H, et al. Plasma proteome profiling of freshwater and seawater life stages of rainbow trout (*Oncorhynchus mykiss*). *PLoS One* (2020) 15:e0227003. doi: 10.1371/journal.pone.0227003
20. Aebersold R, Mann M. Mass spectrometry-based proteomics. *Nature* (2003) 422:198–207. doi: 10.1038/nature01511
21. Cox J, Neuhauser N, Michalski A, Scheltema RA, Olsen JV, Mann M. Andromeda: a peptide search engine integrated into the MaxQuant environment. *J Proteome Res* (2011) 10:1794–805. doi: 10.1021/pr101065j
22. Causey DR, Pohl MAM, Stead DA, Martin SAM, Secombes CJ, Macqueen DJ. High-throughput proteomic profiling of the fish liver following bacterial infection. *BMC Genomics* (2018) 19:719. doi: 10.1186/s12864-018-5092-0
23. Causey DR, Kim J-H, Stead DA, Martin SAM, Devlin RH, Macqueen DJ. Proteomic comparison of selective breeding and growth hormone transgenesis in fish: unique pathways to enhanced growth. *J Proteomics* (2019) 192:114–24. doi: 10.1016/j.jprot.2018.08.013
24. Tasumi S, Velikovskiy CA, Xu G, Gai A, Wittrup KD, Flajnik MF, et al. High-affinity lamprey VLRa and VLRb monoclonal antibodies. *PNAS* (2009) 31:12891–6. doi: 10.1073/pnas.0505379103
25. De Genst E, Silence K, Decanniere K, Conrath K, Loris R, Kinne R, et al. Molecular basis for the preferential left recognition by dromedary heavy-chain antibodies. *PNAS* (2006) 103:4586–91. doi: 10.1073/pnas.0505379103
26. Dooley H, Flajnik MF. Shark immunity bites back: Affinity maturation and memory response in the nurse shark, *Ginglymostoma cirratum*. *Eur J Immunol* (2005) 35:936–45. doi: 10.1002/eji.200425760
27. Thuvander A, Fossum C, Lorenzen N. Monoclonal antibodies to salmonid immunoglobulin: characterization and applicability in immunoassays. *Dev Comp Immunol* (1990) 14:415–23. doi: 10.1016/0145-305X(90)90034-C
28. Cox J, Mann M. MaxQuant enables high peptide identification rates, individualized p.p.b.-range mass accuracies and proteome-wide protein quantification. *Nat Biotechnol* (2008) 26:1367–72. doi: 10.1038/nbt.1511
29. Pearce DE, Barson NJ, Nome T, Gao G, Campbell MA, Abadía-Cardoso A, et al. Sex-dependent dominance maintains migration supergene in rainbow trout. *Nat Ecol Evol* (2019) 3:1731–42. doi: 10.1038/s41559-019-1044-6
30. Beer LA, Pengyuan L, Ky B, Barnhart KT, Speicher DW. Efficient Quantitative comparisons of plasma proteomes using label-free analysis with MaxQuant. In: D Greening, R Simpson, editors. *Methods in molecular biology*, vol. 1619. New York, NY: Humana Press (2017). p. 339–52. doi: 10.1007/978-1-4939-7057-5\_23
31. Tyanova S, Temu T, Cox J. The MaxQuant computational platform for mass spectrometry-based shotgun proteomics. *Nat Protoc* (2016) 11:2301–2319. doi: 10.1038/nprot.2016.136
32. Cox J, Hein MY, Luber CA, Paron I, Nagaraj N, Mann M. Accurate proteome-wide label-free quantification by delayed normalization and maximal peptide ratio extraction, termed MaxLFQ. *Mol Cell Proteomics* (2014) 13:2513–26. doi: 10.1074/mcp.M113.031591
33. Stekhoven DJ, Bühlmann P. MissForest - non-parametric missing value imputation for mixed-type data. *Bioinformatics* (2012) 28:112–8. doi: 10.1093/bioinformatics/btr597
34. Wickham H. *ggplot2: elegant graphics for data analysis*. New York: Springer (2016).
35. Caraux G, Pinloche S. PermutMatrix: a graphical environment to arrange gene expression profiles in optimal linear order. *Bioinformatics* (2005) 21:1280–1. doi: 10.1093/bioinformatics/bti141
36. Szklarczyk D, Gable AL, Lyon D, Junge A, Wyder S, Huerta-Cepas J, et al. STRING v11: protein-protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets. *Nucleic Acids Res* (2018) 47:607–13. doi: 10.1093/nar/gky1131
37. Ashburner M, Ball CA, Blake JA, Botstein D, Butler H, Cherry M, et al. Gene ontology: tool for the unification of biology. *Nat Genet* (2000) 25:25–29. doi: 10.1038/75556
38. Joshi-Tope G, Gillespie M, Vastrik I, d'Eustachio P, Schmidt E, de Bono B, et al. Reactome: a knowledgebase of biological pathways. *Nucleic Acids Res* (2005) 33:D428–32. doi: 10.1093/nar/gki072
39. Araujo FA, Barh DB, Silva A, Guimarães L, Ramos RTJ. GO FEAT: a rapid web-based functional annotation tool for genomic and transcriptomic data. *Sci Rep* (2018) 8:1–4. doi: 10.1038/s41598-018-20211-9
40. Macqueen DJ, Johnston IA. A well-constrained estimate for the timing of the salmonid whole genome duplication reveals major decoupling from species diversification. *Proc R Soc B* (2014) 281:20132881. doi: 10.1098/rspb.2013.2881
41. Berthelot C, Brunet F, Chalopin D, Juanchich A, Bernard M, Noël B, et al. The rainbow trout genome provides novel insights into evolution after whole-genome duplication in vertebrates. *Nat Commun* (2014) 5:1–10. doi: 10.1038/ncomms4657
42. Lien S, Koop BF, Sandve SR, Miller JR, Kent MP, Nome T, et al. The Atlantic salmon genome provides insights into rediploidization. *Nature* (2016) 533:200–5. doi: 10.1038/nature17164
43. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. Basic local alignment search tool. *J Mol Biol* (1990) 215:403–10. doi: 10.1016/S0022-2836(05)80360-2
44. Thompson JD, Higgins DG, Gibson TJ. CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Res* (1994) 22:4673–80. doi: 10.1093/nar/22.22.4673
45. Hall T. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symp Ser* (1999) 41:95–8. doi: 10.14601/Phytopathol\_Mediterr-14998u1.29
46. Katoh K, Rozewicki J, Yamada KD. MAFFT online service: multiple sequence alignment, interactive sequence choice and visualization. *Brief Bioinform* (2019) 20:1160–1166. doi: 10.1093/bib/bbx108
47. Nguyen LT, Schmidt HA, von Haeseler A, Minh BQ. IQ-TREE: A fast and effective stochastic algorithm for estimating maximum-likelihood phylogenies. *Mol Biol Evol* (2015) 32:268–74. doi: 10.1093/molbev/msu300
48. Trifinopoulos J, Nguyen L-T, von Haeseler A, Minh BQ. W-IQ-TREE: a fast online phylogenetic tool for maximum likelihood analysis. *Nucleic Acids Res* (2016) 44:W232–W5. doi: 10.1093/nar/gkw256
49. Kalyaanamoorthy S, Minh BQ, Wong TKF, von Haeseler A, Jermin LS. ModelFinder: fast model selection for accurate phylogenetic estimates. *Nat Methods* (2017) 14:587–9. doi: 10.1038/nmeth.4285
50. Hoang DT, Chernomor O, von Haeseler A, Minh BQ, Vinh LS. UFBoot2: Improving the ultrafast bootstrap approximation. *Mol Biol Evol* (2018) 35:518–22. doi: 10.1093/molbev/msx281
51. Kumar S, Stecher G, Li M, Knyaz C, Tamura K. MEGA X: Molecular Evolutionary Genetics Analysis across Computing Platforms. *Mol Biol Evol* (2018) 35:1547–9. doi: 10.1093/molbev/msy096
52. Roy S, Bag AK, Singh RK, Talmadge JE, Batra SK, Datta K. Multifaceted role of neuropilins in the immune system: potential targets for immunotherapy. *Front Immunol* (2017) 8:1228. doi: 10.3389/fimmu.2017.01228



53. Schellenburg S, Schulz A, Poitz DM, Muders MH. Role of neuropilin-2 in the Immune System. *Mol Immunol* (2017) 90:239–44. doi: 10.1016/j.molimm.2017.08.010
54. Boshra H, Li J, Sunyer O. Recent advances on the complement system of teleost fish. *Fish Shellfish Immun* (2005) 20:239–62. doi: 10.1016/j.fsi.2005.04.004
55. Gerwick L, Reynolds WS, Bayne CJ. A precerebellin-like protein is part of the acute phase response in rainbow trout, *Oncorhynchus mykiss*. *Dev Comp Immunol* (2017) 24:597–607. doi: 10.1016/S0145-305X(00)00016-1
56. Raida MK, Buchmann K. Innate immune response in rainbow trout (*Oncorhynchus mykiss*) against primary and secondary infections with *Yersinia ruckeri* O1. *Dev Comp Immunol* (2009) 33:35–45. doi: 10.1016/j.dci.2008.07.001
57. Saurabh S, Sahoo PK. Lysozyme: an important defence molecule of fish innate immune system. *Aquac Res* (2008) 39:223–39. doi: 10.1111/j.1365-2109.2007.01883.x
58. Niu D, Peatman E, Liu H, Lu J, Kucuktas H, Liu S, et al. Microfibrillar-associated protein 4 (MFAP4) genes in catfish play a novel role in innate immune responses. *Dev Comp Immunol* (2011) 35:568–79. doi: 10.1016/j.dci.2011.01.002
59. Gurney ME, Apatoff BR, Spear GT, Baumel MJ, Antel JP, Bania MB, et al. Neuroleukin: a lymphokine product of lectin-stimulated T cells. *Science* (1986) 234:574–81. doi: 10.1126/science.3020690
60. Morley SC. The actin-bundling protein L-plastin supports T-cell motility and activation. *Immunol Rev* (2013) 256:48–62. doi: 10.1111/imr.12102
61. Freely M, O'Dowd F, Paul T, Kashanin D, Davies A, Kelleher D, et al. L-plastin regulates polarization and migration in chemokine-stimulated human T lymphocytes. *J Immunol* (2012) 188:6357–70. doi: 10.4049/jimmunol.1103242
62. Ganeshan K, Chawla A. Metabolic Regulation of Immune Responses. *Annu Rev Immunol* (2014) 32:609–34. doi: 10.1146/annurev-immunol-032713-120236
63. Liu W, Rodgers GP. Olfactomedin 4 expression and functions in innate immunity, inflammation, and cancer. *Cancer Metastasis Rev* (2016) 35:201–12. doi: 10.1007/s10555-016-9624-2
64. Lee CG, Da Silva CA, Dela Cruz CS, Ahangari F, Ma B, Kang M-J, et al. Role of chitin and chitinase/chitinase-like proteins in inflammation, tissue remodelling and injury. *Annu Rev Physiol* (2011) 73:479–501. doi: 10.1146/annurev-physiol-012110-142250
65. Zemljeni J, Mock DM. Utilization of biotin in proliferating human lymphocytes. *J Nutr* (2000) 130:335S–7S. doi: 10.1093/jn/130.2.335S
66. Báez-Saldaña A, Díaz G, Espinoza B, Ortega E. Biotin deficiency induces changes in subpopulations of spleen lymphocytes in mice. *Am J Clin Nutr* (1998) 67:431–7. doi: 10.1093/ajcn/67.3.431
67. Li X-P, Sun L. A teleost complement factor Ba possesses antimicrobial activity and inhibits bacterial infection in fish. *Dev Comp Immunol* (2017) 71:49–58. doi: 10.1016/j.dci.2017.01.021
68. Wicher KB, Fries E. Haptoglobin, a hemoglobin-binding plasma protein, is present in bony fish and mammals but not in frog and chicken. *Proc Natl Acad Sci* (2006) 103:4168–73. doi: 10.1073/pnas.0508723103
69. Redmond AK, Ohta Y, Criscitiello MF, Macqueen DJ, Flajnik MF, Dooley H. Haptoglobin is a divergent MASP family member that neofunctionalized to recycle hemoglobin via CD163 in mammals. *J Immunol* (2018) 201:2483–2491. doi: 10.4049/jimmunol.1800508
70. Arredouani M, Matthijs P, Van Hoeyveld E, Kasran A, Baumann H, Ceuppens JL, et al. Haptoglobin directly affects T cells and suppresses T helper cell type 2 cytokine release. *Immunology* (2003) 108:144–51. doi: 10.1046/j.1365-2567.2003.01569.x
71. Arredouani MS, Kasran A, Vanoirbeek JA, Berger FG, Baumann H, Ceuppens JL. Haptoglobin dampens endotoxigen-induced inflammatory effects both *in vitro* and *in vivo*. *Immunology* (2005) 114:263–71. doi: 10.1111/j.1365-2567.2004.02071.x
72. Li Y, Wright GL, Peterson JM. C1q/TNF-related protein 3 (CTRP3) Function and regulation. *Compr Physiol* (2017) 7:863–78. doi: 10.1002/cphy.c160044
73. Otis JP, Zeituni EM, Thierer JH, Anderson JL, Brown AC, Boehm ED, et al. Zebrafish as a model for apolipoprotein biology: comprehensive expression analysis and a role for ApoA-IV in regulating food intake. *Dis Model Mech* (2015) 8:295–309. doi: 10.1242/dmm.018754
74. Jaillon O, Aury J-M, Brunet F, Petit J-L, Stange-Thomann N, Mauceli E, et al. Genome duplication in the teleost fish *Tetraodon nigroviridis* reveals the early vertebrate proto-karyotype. *Nature* (2004) 431:946–57. doi: 10.1038/nature03025
75. Dominiczak MH, Caslake MJ. Apolipoproteins: metabolic role and clinical biochemistry applications. *Ann. Clin Biochem* (2011) 48:498–515. doi: 10.1258/acb.2011.011111
76. Kaji H. High-density lipoproteins and the immune system. *J Lipids* (2013) 2013:684903. doi: 10.1155/2013/684903
77. Wang W, Qu Q, Chen J. Identification, expression analysis, and antibacterial activity of Apolipoprotein A-I from amphioxus (*Branchiostoma belcheri*). *Comp Biochem Physiol B Biochem Mol Biol* (2019) 238:110329. doi: 10.1016/j.cbpb.2019.110329
78. Peterson MM, Mack JL, Hall PR, Alsup AA, Alexander SM, Sully EK, et al. Apolipoprotein B is an innate barrier against invasive *Staphylococcus aureus* infection. *Cell Host Microbe* (2008) 4:555–66. doi: 10.1016/j.chom.2008.10.001
79. Wurfel MM, Kunitake ST, Lichenstein H, Kane JP, Wright SD. Lipopolysaccharide (LPS)-binding protein is carried on lipoproteins and acts as a cofactor in the neutralization of LPS. *J Exp Med* (1994) 180:1025–35. doi: 10.1084/jem.180.3.1025
80. Concha MII, Smith VJ, Castro K, Bastias A, Romero A, Amthauer RJ. Apolipoproteins A-I and A-II are potentially important effectors of innate immunity in the teleost fish *Cyprinus carpio*. *Eur J Biochem* (2004) 271:2984–90. doi: 10.1111/j.1432-1033.2004.04228.x
81. Villarreal F, Bastias A, Casado A, Amthauer R, Concha M. Apolipoprotein A-I, an antimicrobial protein in *Oncorhynchus mykiss*: evaluation of its expression in primary defence barriers and plasma levels in sick and healthy fish. *Fish Shellfish Immun* (2007) 23:197–209. doi: 10.1016/j.fsi.2006.10.008
82. Johnston LD, Brown G, Gauthier D, Reece K, Kator H, van Veld P. Apolipoprotein A-I from striped bass (*Morone saxatilis*) demonstrates antibacterial activity *in vitro*. *Comp Biochem Phys B* (2008) 151:167–75. doi: 10.1016/j.cbpb.2008.06.011
83. Wei J, Gao P, Zhang P, Guo M, Xu M, Wei S, et al. Isolation and function analysis of apolipoprotein A-I gene response to virus infection in grouper. *Fish Shellfish Immun* (2015) 43:396–404. doi: 10.1016/j.fsi.2015.01.006
84. Pridgeon JW, Klesius PH. Apolipoprotein A1 in channel catfish: transcriptional analysis, antimicrobial activity, and efficacy as plasmid DNA immunostimulant against *Aeromonas hydrophila* infection. *Fish Shellfish Immun* (2013) 35:1129–37. doi: 10.1016/j.fsi.2013.07.028
85. Hyka N, Dayer J-M, Modoux C, Kohno T, Edwards III CK, Roux-Lombard P, et al. Apolipoprotein A-I inhibits the production of interleukin-1 $\beta$  and tumor necrosis factor- $\alpha$  by blocking contact-mediated activation of monocytes by T lymphocytes. *Blood* (2001) 97:2381–9. doi: 10.1182/blood.V97.8.2381
86. Wilhelm AJ, Zabalawi M, Owen JS, Shah D, Grayson JM, Major AS, et al. Apolipoprotein A-I modulates regulatory T Cells in autoimmune LDLr<sup>-/-</sup>, ApoA-I<sup>-/-</sup> Mice. *J Bio Chem* (2010) 285:36158–69. doi: 10.1074/jbc.M110.134130
87. Bonacina F, Coe D, Wang G, Longhi MP, Baragetti A, Moregola A, et al. Myeloid apolipoprotein E controls dendritic cell antigen presentation and T cell activation. *Nat Commun* (2018) 9:1–15. doi: 10.1038/s41467-018-05322-1
88. Brada N, Gordon MM, Wen J, Alpers DH. Transfer of cobalamin from intrinsic factor to transcobalamin II. *J Nutr Biochem* (2001) 12:200–6. doi: 10.1016/S0955-2863(00)00129-7
89. Benoit CR, Stanton AE, Tartanian AC, Motzer AR, McGaughey DM, Bond SR, et al. Functional and phylogenetic characterization of noncanonical vitamin B12-binding proteins in zebrafish suggests involvement in cobalamin transport. *J Biol Chem* (2018) 293:17606–21. doi: 10.1074/jbc.RA118.005323
90. Wong MK-S, Takei Y. Lack of plasma kallikrein-kinin system in teleosts. *PLoS One* (2013) 8:e81057. doi: 10.1371/journal.pone.0081057
91. Chang C, Hu M, Zhu Z, Lo LJ, Chen J, Peng J. Liver-enriched gene 1a and 1b encode novel secretory proteins essential for normal liver development in zebrafish. *PLoS One* (2011) 6:e22910. doi: 10.1371/journal.pone.0022910
92. Kaattari SL, Zhang HL, Khor IW, Kaattari IM, Shapiro DA. Affinity maturation in trout: Clonal dominance of high affinity antibodies late in the immune response. *Dev Comp Immunol* (2002) 26:191–200. doi: 10.1016/S0145-305X(01)00064-7
93. Cain KD, Jones DR, Raison RL. Antibody-antigen kinetics following immunization of rainbow trout (*Oncorhynchus mykiss*) with a T-cell dependent antigen. *Dev Comp Immunol* (2002) 26:181–90. doi: 10.1016/S0145-305X(01)00063-5
94. Costa G, Danz H, Kataria P, Bromage E. A holistic view of the dynamics of teleost IgM: A case study of *Streptococcus iniae* vaccinated rainbow trout (*Oncorhynchus mykiss*). *Dev Comp Immunol* (2012) 36:298–305. doi: 10.1016/J.DCI.2011.04.011



95. Huang Q, Yang L, Luo J, Guo L, Wang Z, Yang XJ, et al. SWATH enables precise label-free quantification on proteome scale. *Proteomics* (2015) 15:1215–23. doi: 10.1002/pmic.201400270

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

*Copyright © 2020 Bakke, Monte, Stead, Causey, Douglas, Macqueen and Dooley. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.*