

- 7 Mueller BU, Pabst T, Osato M, Asou N, Johansen LM, Minden MD *et al.* Heterozygous PU.1 mutations are associated with acute myeloid leukemia. *Blood* 2002; **100**: 998–1007.
- 8 Rosenbauer F, Wagner K, Kutok JL, Iwasaki H, Le Beau MM, Okuno Y *et al.* Acute myeloid leukemia induced by graded reduction of a lineage-specific transcription factor, PU.1. *Nat Genet* 2004; **36**: 624–630.
- 9 Metcalf D, Dakic A, Mifsud S, Di Rago L, Wu L, Nutt S. Inactivation of PU.1 in adult mice leads to the development of myeloid leukemia. *Proc Natl Acad Sci USA* 2006; **103**: 1486–1491.
- 10 Stoletov K, Klemke R. Catch of the day: zebrafish as a human cancer model. *Oncogene* 2008; **27**: 4509–4520.
- 11 Orkin SH, Zon LI. Hematopoiesis: an evolving paradigm for stem cell biology. *Cell* 2008; **132**: 631–644.

- 12 Dakic A, Metcalf D, Di Rago L, Mifsud S, Wu L, Nutt SL. PU.1 regulates the commitment of adult hematopoietic progenitors and restricts granulopoiesis. *J Exp Med* 2005; **201**: 1487–1502.
- 13 Pigneux A, Perreau V, Jourdan E, Vey N, Dastugue N, Huguet F *et al.* Adding lomustine to idarubicin and cytarabine for induction chemotherapy in older patients with acute myeloid leukemia: the BGMT 95 trial results. *Haematologica* 2007; **92**: 1327–1334.
- 14 Rabbani A, Finn RM, Ausio J. The anthracycline antibiotics: antitumor drugs that alter chromatin structure. *Bioessays* 2005; **27**: 50–56.
- 15 Anderson KL, Smith KA, Pio F, Torbett BE, Maki RA. Neutrophils deficient in PU.1 do not terminally differentiate or become functionally competent. *Blood* 1998; **92**: 1576–1585.

Supplementary Information accompanies this paper on the Leukemia website (<http://www.nature.com/leu>)

OPEN

A powerful molecular synergy between mutant Nucleophosmin and Flt3-ITD drives acute myeloid leukemia in mice

Leukemia (2013) **27**, 1917–1920; doi:10.1038/leu.2013.77

Acute myeloid leukemia (AML) is the commonest myeloid malignancy, yet there has been little therapeutic progress for this disease in decades, and only 25–30% of patients survive long term.¹ This reflects its pathogenetic complexity and the fact that the molecular basis of its largest cytogenetic subgroup, AML with a normal karyotype (AML-NK), was obscure until recently. Recent advances in DNA sequencing have revealed that AML-NK is molecularly heterogeneous with >30 genes recurrently targeted by somatic mutations in this disease.² What is also evident is that each individual case of AML-NK appears to harbor only a small number of coding driver mutations, often as few as three and usually no more than five.^{2,3} Furthermore, it is manifest that the precise combination of driver mutations in the genome of each AML impacts on its salient features, including responsiveness to treatments and prognosis.³

These observations provide a sound starting point for systematic mechanistic studies to understand the pathogenesis and improve the treatment of AML-NK. Carefully designed mouse models are the gold standard in the study of normal and malignant hemopoiesis, and are already instructing our understanding of AML-NK.^{4,5} Here, we report that the two most commonly co-occurring somatic mutations in AML, namely Nucleophosmin (*NPM1*) exon 12 mutations (*NPM1c*) and internal tandem duplications of *FLT3* (*FLT3-ITD*), cooperate explosively to induce AML in knock-in mice. In revealing this striking molecular synergy, our work offers a basis for the frequent co-occurrence of these two mutations and provides a valuable model for in-depth studies of the pathogenesis and treatment of this large subgroup of AML.

NPM1 is a nucleolar phosphoprotein involved in many cellular processes. For many of its roles, it relies on its ability to shuttle between the nucleolus, nucleus and cytoplasm using subcellular localization signals.⁶ This ability is impaired in 30% of AMLs as a result of *NPM1c* mutations, which disrupt the nucleolar localization signal of *NPM1* and generate a nuclear export signal in its place.⁷ Mutant *NPM1* is known to bind to and alter the subcellular distribution of several proteins, including HEXIM1, p19Arf and

nuclear factor- κ B;⁸ however, the relevance of these interactions to AML is unclear. *FLT3-ITD* mutations occur in 20–25% of AML⁹ and result in ligand-independent receptor dimerization and constitutive *FLT3* signaling,¹⁰ and are associated with an increased risk of relapse. Moreover, patients with low or absent levels of wild-type (WT) *FLT3*, consistent with loss-of-heterozygosity (LOH) for this locus, have a particularly poor outcome.⁹

Recently, we described a conditional knock-in mouse model of *NPM1c* mutations and demonstrated that one-third of mice developed delayed-onset AML, suggesting a requirement for cooperating mutations. We went on to show that insertional mutagenesis with transposons led rapidly to AML in 80% of *Npm1c* mice, in association with specific recurrent mutations including activating insertions in *Csf2* and *Flt3*.⁴ *Flt3-ITD* homozygous mutant mice exhibit enhanced proliferation and survival properties in hemopoietic progenitors and develop a late-onset disease akin to chronic myelomonocytic leukemia.¹¹

To study the combined effects of *NPM1c* with *FLT3-ITD* we crossed conditional *Npm1^{fllox-cA/+}* with constitutive *Flt3^{ITD/+}* to generate *Npm1^{fllox-cA/+}; Flt3^{ITD/+}* double heterozygous mice, then crossed into *Mx1-Cre* transgenic mice to induce recombination of *Npm1^{fllox-cA}* in hematopoietic stem cells.⁴ The *Mx1-Cre* allele requires induction by interferon, usually achieved by intraperitoneal injection of polyinosinic-polycytidylic acid (plpC). However, we observed universal and rapid emergence of AML (myeloid leukemia with maturation) in uninjected *Npm1^{fllox-cA}; Flt3^{ITD/+}; Mx1-Cre+* mice (hereafter referred to as '*Npm1c/Flt3-ITD* mice'). *Mx1-Cre* is known to 'leak' in 2–4% of hemopoietic stem/progenitor cells,¹² and this was sufficient to rapidly generate AML from double mutant cells signifying a striking cooperativity between *Npm1c* and *Flt3-ITD*. The presence of the cytoplasmic *NPM1* was confirmed on protein blots (Figure 1a).

All *Npm1c/Flt3-ITD* mice developed AML and became moribund aged 31–68 days (median 49 days; $n = 29$). By contrast, no case of AML was observed in *Npm1^{fllox-cA}; Mx1-Cre+* mice (hereafter referred to as '*Npm1c* mice'; $n = 30$, of which 15 received plpC aged 6–8 weeks), *Flt3^{ITD/+}* mice (hereafter referred to as '*Flt3-ITD* mice'; $n = 34$) or WT mice (WT, $n = 29$) aged to at least 8 months (Figure 1b). Weekly blood counts from 19 mice with each genotype showed a progressive increase in blood leukocyte counts in *Npm1c/Flt3-ITD* mice, to more than 25-fold that of

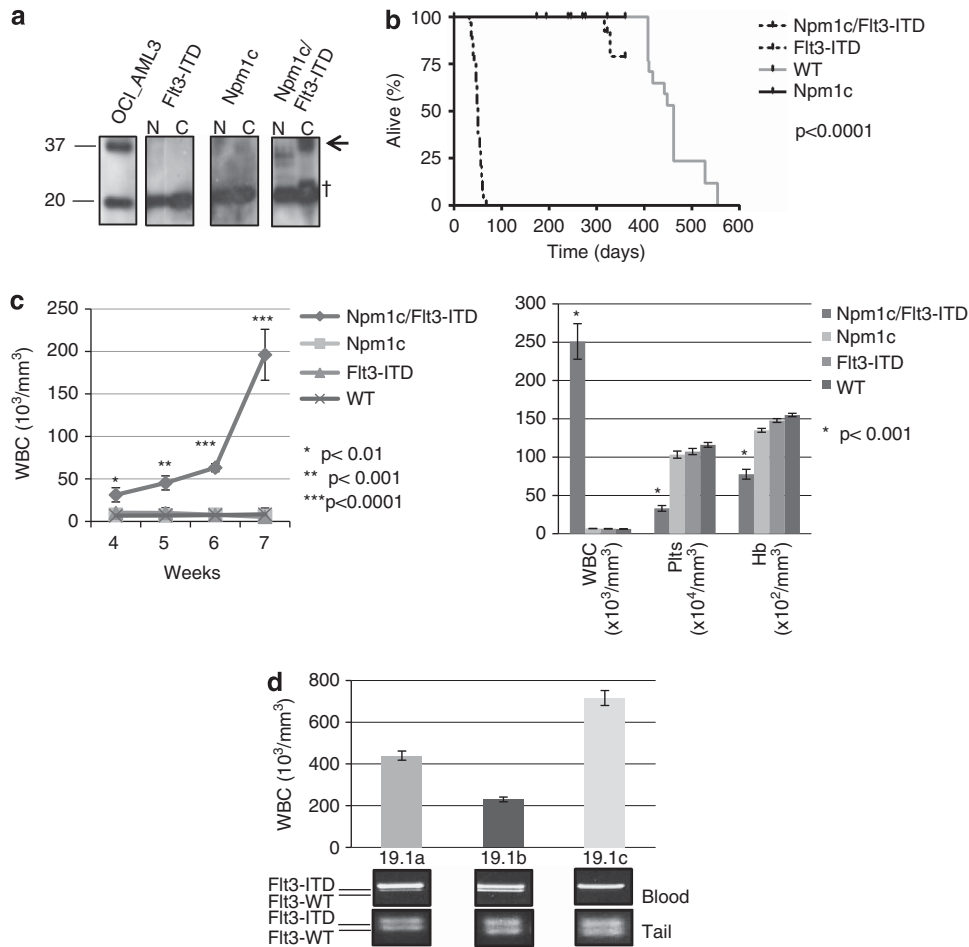


Figure 1. *Npm1c* and *Flt3-ITD* collaborate to drive rapid-onset leukemogenesis with frequent occurrence of *Flt3* LOH. **(a)** *Npm1* mutant protein (arrow) accumulates in the cytoplasm of spleen cells collected from 3-week-old *Npm1c/Flt3-ITD*, but not *Npm1c* or *Flt3-ITD* single-mutant mice. **(b)** Kaplan-Meier survival plots showing the rapid demise of *Npm1c/Flt3-ITD* mice compared with all other genotypes. **(c)** Serial blood counts highlight a consistent explosive increase in blood leukocytes counts (WBC) between 4 and 7 weeks in *Npm1c/Flt3-ITD* mice (left) and the markedly abnormal WBC, platelet count (Plts) and hemoglobin concentration (Hb) of sick leukemic *Npm1c/Flt3-ITD* mice compared with age-matched control mice. **(d)** Loss of the *Flt3* WT allele in blood DNA from *Npm1c/Flt3-ITD* AMLs is demonstrated as loss of intensity of the *Flt3*-WT PCR band. By contrast, constitutional tail DNA shows no LOH. In these three littermates (19.1a–c), the extent of *Flt3*-LOH associates with the degree of leukocytosis (N = nuclear lysate, C = cytoplasmic lysate, OCL-AML3 lysate as positive control, † nonspecific band).

age-matched control littermates, whereas the hemoglobin and platelet counts were significantly reduced (Figure 1c).

Interestingly, *Npm1c/Flt3-ITD* siblings/littermates often progressed to AML at different rates or developed more/less aggressive disease. To explain this observation we hypothesized that, as seen in human AML, LOH for *Flt3-ITD* may be responsible. We found evidence for significant spontaneous loss of the WT *Flt3* allele in blood samples from *Npm1c/Flt3-ITD* mice and a tendency for higher blood leukocyte counts (Figure 1d) when LOH was present. LOH was also seen in bone marrow and spleen but not tail DNA, in keeping with somatic loss of the WT allele in leukemic cells (Figure 1d). At the time mice became sick with AML, LOH was detected in 12 of 15 spleen samples tested.

Flow cytometric analysis of blood samples demonstrated, in *Npm1c/Flt3-ITD* mice, a population of blasts/immature cells with low side scatter (SSC) and CD45^{dim} (Figure 2a) and a large number of single Mac1⁺ precursors (Figure 2b). In addition, we also observed an increased number of mature myeloid (Gr1⁺/Mac1⁺) cells in *Npm1c/Flt3-ITD* mice, indicating that any maturation block was incomplete (Figure 2b). The relative numbers of circulating B (B220⁺) and T (CD3⁺) lymphocytes were reduced (data not shown). To assay their self-renewal potential, bone marrow cells from *Npm1c* ($n=4$), *Flt3-ITD* ($n=4$),

WT ($n=4$) and *Npm1c/Flt3-ITD* ($n=4$) were studied in serial replating assays. *Npm1c/Flt3-ITD* cells gave rise to significantly more colonies at first and subsequent platings than any other genotype (Figure 2c), demonstrating a significantly increased self-renewal potential.

Blood smears from sick mice confirmed the presence of blasts, and histological sections demonstrated widespread infiltration of solid organs by abnormal myeloid cells (Supplementary Figure S1). Cells infiltrating the bone marrow and spleen were Gr1⁺/Mac1⁺ or Gr1⁻/Mac1⁺, and there were increased numbers of Mac1⁺/cKit⁺ cells compared with other genotypes (Supplementary Figure S2). Compared with single mutant and WT mice, sick *Npm1c/Flt3-ITD* mice had marked splenomegaly (0.95 ± 0.27 g vs 0.13 ± 0.02 g; $P < 0.0001$) and hepatomegaly (2.33 ± 0.26 g vs 1.6 ± 0.17 g, $P < 0.0001$) at the time of death. *Npm1c/Flt3-ITD* leukemias were transplantable into both syngeneic and NOD SCID γ mice demonstrating their true neoplastic nature (data not shown).

AML is a molecularly and clinically heterogeneous disease and recent studies have revealed that this heterogeneity is derived, to a large extent, from the specific combinations of somatic driver mutations present in individual cases. Here, we show that the combination of *Npm1c* and *Flt3-ITD*, the two most commonly co-occurring AML mutations, is rapidly and universally

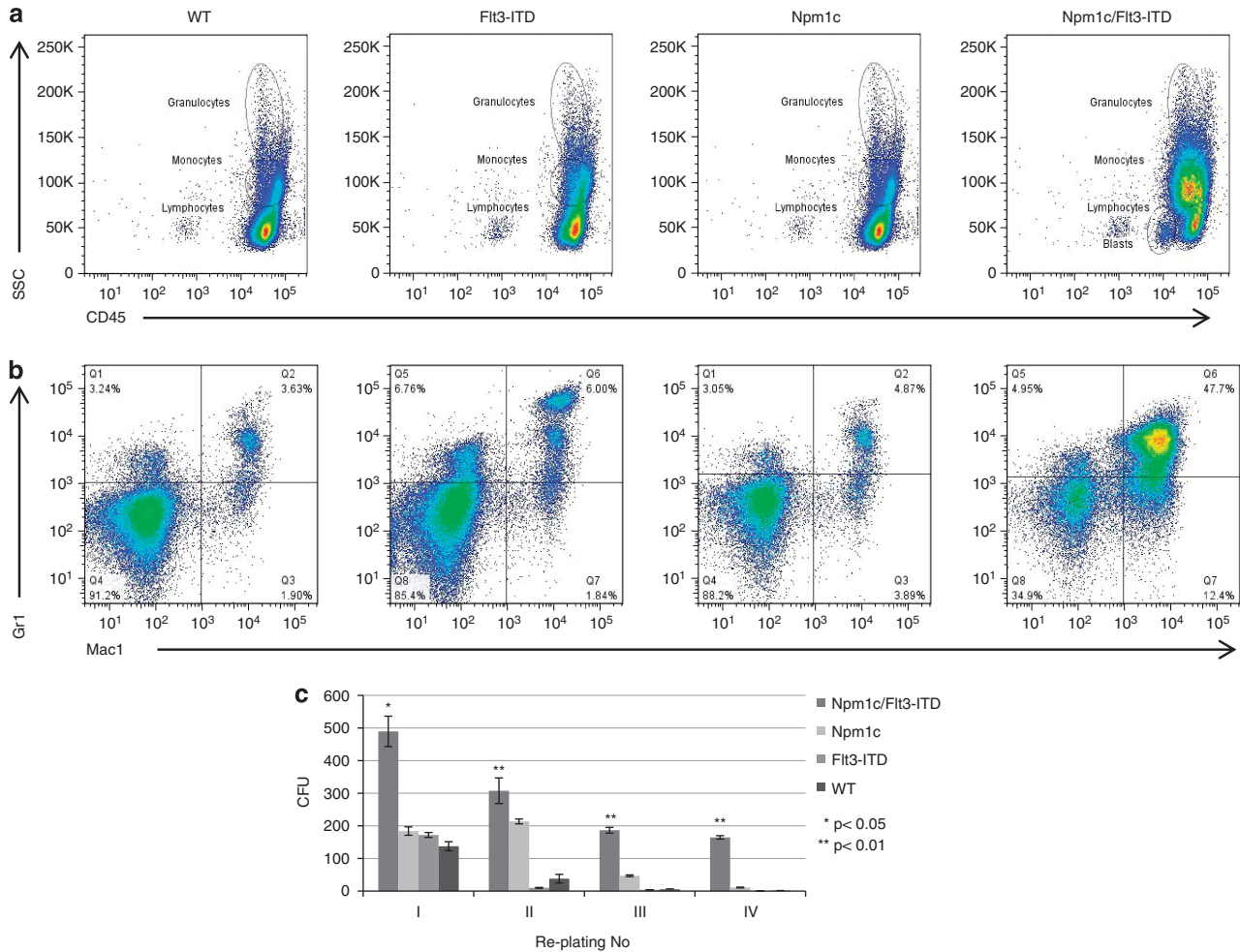


Figure 2. Expansion of circulating myeloid cells in *Npm1c/Flt3-ITD* mice culminating to aggressive AML. (a) Representative flow cytometric analysis of peripheral blood from 7-week-old WT, *Npm1c*, *Flt3-ITD* and *Npm1c/Flt3-ITD* mice shows the presence of a low SSC; CD45^{dim} population of immature/blast cells in double mutant mice and (b) an increase in both mature granulocytic (Gr1⁺/Mac1⁺) and monocytic (Gr1^{low}/Mac1⁺) populations. (c) Colony-forming assays of BM cells derived from WT and mutant mice showing a markedly increased replating ability of *Npm1c/Flt3-ITD* cells compared with other genotypes, indicative of an increased self-renewal potential. As we described before, a lesser increase in replating ability is observed with *Npm1c* cells.

leukemogenic in knock-in mice. These findings are particularly striking in light of the fact that, in isolation, both *Npm1c*⁴ and *Flt3-ITD*¹¹ mutations have relatively subtle effects on mouse hemopoiesis and lead to leukemia or a myeloproliferative disorder only after prolonged latencies and in a minority of mice.

What is most remarkable about our findings is the very short latency of AML in *Npm1c/Flt3-ITD* mice, which suggests either: (i) that the two mutations are sufficient to promote AML in this strain of mice (C57BL6/N) or (ii) that additional mutations are acquired very rapidly in the pool of cells susceptible to leukemic transformation. The later possibility is supported by the fact that at least one type of somatic mutation, namely LOH for *Flt3-ITD*, was frequently observed in our mouse AMLs over this short time span. We recently reported that *Npm1c* can generate AML in collaboration with, amongst others, activating insertions of the *GrOnc* transposon in intron 9 of mouse *Flt3*. These insertions led to aberrant expression of a *Flt3* messenger RNA predicted to code for an amino-terminal truncated version of Flt3⁴ which, like Flt3-ITD, was thought to be constitutively active. Most of these murine AMLs harbored additional transposon insertions thought to be important in leukemogenesis. Thus, at this stage it appears more likely that additional mutations may be required for leukemogenesis in our *Npm1c/Flt3-ITD* mice, but this cannot be stated unequivocally.

In interesting contrast to our present work, a recent report demonstrated that the combination of *Flt3-ITD* with *NUP98-HOXD13* in mice led to AML after a much longer latency (median 95 days),¹⁴ despite the fact that, unlike *Npm1c*, *NUP98-HOXD13* alone leads to a highly penetrant myelodysplastic syndrome with a high risk of leukemic transformation. This relative delay is particularly intriguing as *NUP98-HOXD13* can promote leukemic transformation in association with simple overexpression of WT *FLT3*.¹⁵ By contrast, in two large transposon-mediated insertional mutagenesis screens, one published⁴ and one ongoing, we never observed transposon insertions leading to simple *Flt3* overexpression amongst > 100 mouse *Npm1c* + ve AMLs.

Notwithstanding the above, our observations emphasize the remarkable complementarity between *Npm1c* and *Flt3-ITD*. In the context of a stochastic model for AML pathogenesis,² this potent molecular synergy goes some way toward explaining why *NPM1c* and *FLT3-ITD* co-occur so frequently and make the model described here a valuable tool for the study of the pathogenesis and treatment of one of the largest molecularly defined subgroups of AML.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

ACKNOWLEDGEMENTS

We are grateful to Dr Gary Gilliland for his kind donation of the Flt3-ITD mice. We thank James Bussell and the members of the Wellcome Trust Sanger Institute's Research Support Facility for help with mouse colony management, controlled procedures and daily welfare checks. We also thank the Haematology Diagnostic service and the Pathology Tissue Bank of Cambridge University NHS Trust for their help with sample processing for microscopy. We are grateful to the Wellcome Trust and the Kay Kendall Leukaemia Fund for funding this work and also to Leukaemia Lymphoma Research for funding related work in GV's laboratory.

All material is original research, and has neither been previously published or submitted for publication elsewhere.

A Mupo¹, L Celani^{1,2}, O Dovey¹, JL Cooper¹, C Grove¹, R Rad¹,
P Sportoletti², B Falini², A Bradley¹ and GS Vassiliou¹

¹The Wellcome Trust Sanger Institute, Wellcome Trust Genome
Campus, Hinxton, Cambridge, UK and

²Hematology and Clinical Immunology Section, Department of
Clinical and Experimental Medicine, University of Perugia,
Perugia, Italy

E-mail: gsv20@sanger.ac.uk

REFERENCES

- Grimwade D, Hills RK. Independent prognostic factors for AML outcome. *Hematology Am Soc Hematol Educ Program* 2009; **1**: 385–395.
- Welch JS, Ley TJ, Link DC, Miller CA, Larson DE, Koboldt DC *et al.* The origin and evolution of mutations in acute myeloid leukemia. *Cell* 2012; **150**: 264–278.
- Patel JP, Gonen M, Figueroa ME, Fernandez H, Sun Z, Racevskis J *et al.* Prognostic relevance of integrated genetic profiling in acute myeloid leukemia. *N Engl J Med* 2012; **366**: 1079–1089.
- Vassiliou GS, Cooper JL, Rad R, Li J, Rice S, Uren A *et al.* Mutant nucleophosmin and cooperating pathways drive leukemia initiation and progression in mice. *Nat Genet* 2011; **43**: 470–475.
- Sasaki M, Knobbe CB, Munger JC, Lind EF, Brenner D, Brustle A *et al.* IDH1(R132H) mutation increases murine haematopoietic progenitors and alters epigenetics. *Nature* 2012; **488**: 656–659.
- Hingorani K, Szebeni A, Olson MO. Mapping the functional domains of nucleolar protein B23. *J Biol Chem* 2000; **275**: 24451–24457.
- Falini B, Mecucci C, Tiacci E, Alcalay M, Rosati R, Pasqualucci L *et al.* Cytoplasmic nucleophosmin in acute myelogenous leukemia with a normal karyotype. *N Engl J Med* 2005; **352**: 254–266.
- Falini B, Bolli N, Liso A, Martelli MP, Mannucci R, Pileri S *et al.* Altered nucleophosmin transport in acute myeloid leukaemia with mutated NPM1: molecular basis and clinical implications. *Leukemia* 2009; **23**: 1731–1743.
- Whitman SP, Archer KJ, Feng L, Baldus C, Becknell B, Carlson BD *et al.* Absence of the wild-type allele predicts poor prognosis in adult de novo acute myeloid leukemia with normal cytogenetics and the internal tandem duplication of FLT3: a cancer and leukemia group B study. *Cancer Res* 2001; **61**: 7233–7239.
- Kiyoi H, Towatari M, Yokota S, Hamaguchi M, Ohno R, Saito H *et al.* Internal tandem duplication of the FLT3 gene is a novel modality of elongation mutation which causes constitutive activation of the product. *Leukemia* 1998; **12**: 1333–1337.
- Lee BH, Tothova Z, Levine RL, Anderson K, Buza-Vidas N, Cullen DE *et al.* FLT3 mutations confer enhanced proliferation and survival properties to multipotent progenitors in a murine model of chronic myelomonocytic leukemia. *Cancer Cell* 2007; **12**: 367–380.
- Kemp R, Ireland H, Clayton E, Houghton C, Howard L, Winton DJ. Elimination of background recombination: somatic induction of Cre by combined transcriptional regulation and hormone binding affinity. *Nucleic Acids Res* 2004; **32**: e92.
- Kiyoi H, Ohno R, Ueda R, Saito H, Naoe T. Mechanism of constitutive activation of FLT3 with internal tandem duplication in the juxtamembrane domain. *Oncogene* 2002; **21**: 2555–2563.
- Greenblatt S, Li L, Slape C, Nguyen B, Novak R, Duffield A *et al.* Knock-in of a FLT3/ITD mutation cooperates with a NUP98-HOXD13 fusion to generate acute myeloid leukemia in a mouse model. *Blood* 2012; **119**: 2883–2894.
- Palmqvist L, Argiropoulos B, Pineault N, Abramovich C, Sly LM, Krystal G *et al.* The Flt3 receptor tyrosine kinase collaborates with NUP98-HOX fusions in acute myeloid leukemia. *Blood* 2006; **108**: 1030–1036.



This work is licensed under a Creative Commons Attribution 3.0 Unported License. To view a copy of this license, visit <http://creativecommons.org/licenses/by/3.0/>

Supplementary Information accompanies this paper on the Leukemia website (<http://www.nature.com/leu>)

Phase I study of ON 01910.Na (Rigosertib), a multikinase PI3K inhibitor in relapsed/refractory B-cell malignancies

Leukemia (2013) **27**, 1920–1923; doi:10.1038/leu.2013.79

Chronic lymphocytic leukemia (CLL) and mantle cell lymphoma (MCL) are incurable B-cell malignancies usually responsive to initial immunochemotherapy, but virtually all patients experience relapse. Salvage therapy choices for relapsed/refractory disease are often limited by resultant cytopenias and acquired drug resistance. The current priority in these B-cell malignancies, therefore, is to develop agents with novel mechanisms of action that are selective for tumor cells, overcome shared patterns of acquired drug resistance and exhibit limited toxicities.

Styrylbenzylsulfones are a new family of non-ATP-competitive anticancer agents that induce apoptosis in a variety of tumor cell lines, including those resistant to many chemotherapy agents.^{1,2} As a class, styrylbenzylsulfones inhibit cell cycle progression and induce mitotic arrest of tumor cells with less toxicity to normal human cells.^{3,4} ON 01910.Na (rigosertib) is a styryl sulfonol compound that demonstrated inhibition of

phosphatidylinositol-3-kinase (PI3K), preferentially targeting the PI3K α and PI3K β isoforms, and triggered apoptosis via the release of cytochrome c from mitochondria in MCL cell lines.³ Rigosertib's mechanism of action was initially considered to include inhibition of polo-like 1 kinase,⁴ but evidence for direct inhibition was not confirmed in subsequent studies⁵ and its antimitotic activity may rely on the phosphorylation of mitosis coordinator RanGAP1 SUMO1.⁶ First-in-man studies of rigosertib in solid tumors demonstrated excellent tolerability with limited hematologic toxicity.⁷ Rigosertib has also demonstrated preclinical and early clinical activities in myelodysplastic syndromes (MDS),^{8,9} and it is currently being tested in a randomized phase III trial in patients with relapsed/refractory MDS (NCT01241500).

We have previously reported that rigosertib induces rapid apoptosis in CLL cells with the relative sparing of normal B-cells and T-cells.¹⁰ We demonstrated that the *in vitro* activity of rigosertib involved a dual mechanism of inhibition of PI3K pathway signaling coupled with the induction of an oxidative stress response. Importantly, activity of rigosertib was equally observed against