# 1 Powering ocean giants: the energetics of shark and ray megafauna

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#### 28 Abstract

29 Shark and ray megafauna play crucial roles as top predators in many marine ecosystems, but 30 are currently among the most threatened vertebrates and, based on historical extinctions, may be highly susceptible to future environmental perturbations. However, our 31 32 understanding of their energetics lags behind that of other taxa. Such knowledge is required to answer important ecological questions and predict their responses to ocean warming, 33 34 which may be limited by expanding ocean deoxygenation and declining prey availability. To 35 develop bioenergetics models for shark and ray megafauna, incremental improvements in 36 respirometry systems are useful but unlikely to accommodate the largest species. Advances in biologging tools and modelling could help answer the most pressing ecological questions 37 about these iconic species. 38

#### 39 Glossary

Mesothermy: Some elasmobranchs are able to retain heat generated by metabolic
 processes to keep certain body parts warmer than the surrounding water, e.g.
 elevated temperature of the brain, eyes and body musculature of some sharks may
 maintain physiological performance in colder waters. Sometimes referred to as
 'regional endothermy'.

Metabolic scope: Also known as maximum factorial scope; the difference between
 maximum aerobic metabolism and standard aerobic metabolism. Metabolic scope
 indicates the capacity to perform energy-using processes, such as locomotion or
 digestion, beyond SMR, and is dictated by the rate of oxygen supply to organs. This
 supply is dependent on an animal's physiology and ambient environmental

50		conditions. Metabolic scope is distinguished from routine factorial scope, which is
51		the ratio of daily energy use to standard metabolic rate.
52	•	Obligate ram ventilation: Many large elasmobranchs lack the anatomical features
53		to pump water over their gills, and hence rely on moving their entire body to
54		oxygenate their gills. This requires them to constantly move forwards through the
55		water.
56	•	$\mathbf{Q}_{10}$ : The factorial increase in metabolic rate associated with a 10 °C increase in
57		temperature.
58	•	Respirometry: Indirect calorimetry via respirometry quantifies the aerobic
59		respiration of an animal by measuring its oxygen consumption. By estimating the
60		substrates being metabolised (commonly 70% carbohydrate, 20% lipid and 10%
61		protein), these measurements of oxygen consumption can be converted to energy
62		expenditure.
63	•	Standard Metabolic Rate (SMR): The minimum energy expenditure of an
64		ectotherm for body functioning and maintenance. As locomotor activity is essential
65		for respiration in obligate ram ventilators, measures of their SMR include some
66		locomotor costs.
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## 75 The value of measuring the energetics of large sharks and rays

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77 Understanding how megafauna are able to maintain energy balance has intrigued physiological ecologists for decades. Fundamental to this research is the measurement, or 78 79 at least estimation, of metabolic rates. The ability of megafauna to sustain energy balance has been explored in extinct groups of large vertebrates, such as dinosaurs [1], as well as 80 some of the largest extant vertebrates such as whales [2, 3], and may help inform our 81 82 understanding of the evolution of body size [4, 5]. Large sharks, rays and skates 83 (elasmobranchs) are a group of megafauna that include the largest fish and have a range of 84 diets spanning plankton, fish, reptiles and marine mammals. They also include many species threatened with over-fishing and climate change [6, 7]. Yet how energy balance is 85 maintained across this diverse group remains poorly understood. Given these long-standing, 86 87 unresolved questions and the advent of new technology and methods, it is timely to review current knowledge of the metabolic rates of large elasmobranchs and examine how our 88 89 understanding of this group is changing.

90 Logistical problems of working with large elasmobranchs means there are few 91 studies that quantify their energy use [8]. These species are too large for housing in the 92 laboratory, and, unlike marine mammals, do not surface to respire where they can be easily 93 accessed. Further, the few existing bioenergetics models (Box 1) for elasmobranch 94 megafauna extrapolate from species that are orders of magnitude smaller [9] (Figure 1B). In 95 fact, the heaviest elasmobranch for which metabolic rate (MR) – a key aspect of energetics - has been measured is only 47.7 kg, despite many elasmobranchs weighing >1,000 kg, with 96 97 the whale shark (*Rhincodon typus*) reaching 34,000 kg. Further, the thermal sensitivity of MR 98 (Q10; see Glossary) varies substantially across ectotherms [10] and is not well established for

elasmobranchs (Figure 1C). This enormous uncertainty in MR for large elasmobranchs 99 100 hinders our ability to answer important ecological questions concerning this group, which includes several unusual species: the largest ectotherm, the planktivorous whale shark; the 101 longest-lived vertebrate (392 years) - the Greenland shark (Somniosus microcephalus) [11]; 102 103 and some of the largest apex predators including the white shark (Carcharodon carcharias). 104 Large elasmobranchs are also disproportionately at risk of extinction within an 105 already vulnerable group [6]. The class Chondrichthyes (that contains about 1050 species, of 106 which 96% are elasmobranchs) has the lowest proportion of non-threatened species of any vertebrate group based on IUCN Red List criteria [6]. Ocean warming is also expected to raise 107 the MR of ectotherms [12], which will impact energy balance and reproductive performance 108 109 [13]. In previous mass extinctions, large ectotherms and top predators were among the animals most affected by ocean warming and the resultant water deoxygenation [14, 15]. 110 111 Ocean deoxygenation limits metabolic scope, prey availability and 'aerobic habitat' - zones 112 with sufficient oxygen to meet metabolic demands [16, 17].

Current climate change predictions mimic conditions of past extinctions, suggesting 113 114 the largest ectotherms are again likely to be heavily impacted [14, 16, 17]. Most large shark species went extinct during warming at the end-Cretaceous period, while smaller sharks 115 survived [14]. Nine of the 15 largest extant ectotherms are elasmobranchs. Unlike other 116 117 animals, most large elasmobranchs must move continuously, which requires from 34% to almost 100% of their metabolic scope, limiting growth, reproduction and foraging ability 118 when MR is elevated [18]. Prey availability could also be impacted by climate change, with 119 mean global zooplankton biomass predicted to fall by ~14% this century [19], and in some 120 121 areas by 50% [20]. Some large elasmobranchs are hypothesised to rely on efficient foraging 122 behaviours to overcome an apparent 'energy-budget paradox' caused by high feeding costs

and sparse prey orders of magnitude smaller than themselves [21, 22]. Large elasmobranch
species may therefore be sensitive to a range of environmental perturbations, and the
effects of such perturbations may be mediated by changes in their energy balance.

Although investigating the energetics of large elasmobranchs is challenging, it remains an important goal in light of their critical role in food webs and the need to understand their response to climate change [23]. Here, our aim is to promote energetics research in elasmobranch megafauna and to provide a roadmap for the most promising research methods and most pressing questions concerning the energetics of this group. This will enable an understanding of how metabolism, locomotion and feeding affect their growth, reproduction, ecology, and ultimately their fitness, which will provide the basis for population and ecosystem models [24]. 

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**148** Box 1. What is a bioenergetics model?

149 A bioenergetics model describes the energy requirements of an animal and how energy is 150 distributed to processes in the body (i.e. the animal's 'energetics') [25]. It has four main component 151 processes: standard metabolic rate (SMR); active metabolic rate (cost of activity); food assimilation 152 (specific dynamic action); and growth and reproduction (Figure I). If the consumption rate is known, 153 bioenergetics models are commonly used to predict and assess rates of growth and reproduction 154 [26], both of which are crucial for survival at the individual and population level. However, growth 155 and reproduction can be depressed in times of elevated energy demand, a situation predicted under 156 future ocean warming [25].

157 An animal's energy requirements can increase in a number of ways, but is most commonly 158 due to elevated SMR or activity levels. In ectotherms, SMR increases approximately exponentially 159 with temperature [12]. Activity levels may increase with migration, changed prey availability, 160 predator evasion, mating behaviours, weather patterns, tidal state, temperature, or human 161 interference [27-31]. Consequently, an individual must either increase its food consumption to meet 162 these new energy requirements, or energy must be diverted away from growth or reproduction and 163 towards SMR or activity costs [13, 25]. Animals may be able to increase their total energy intake by 164 consuming more energy-dense food, rather than consuming greater biomass [32], however energy-165 dense diets are often lacking in sufficient nitrogen for synthesising proteins [33]. 166

# 168 What can we learn from the energetics of megafauna?

### 169 Terrestrial megafauna and hard-to-study species

Although megafauna are difficult to study because they are often highly mobile, too 170 171 large for captivity, dangerous to handle, or live in extreme environments, energetics studies can reveal how these animals survive, especially in unusual environments or with unusual 172 life histories. For example, energetically-economical long-distance travel and foraging are 173 174 achieved through morphological adaptation in large sea-birds that stay airborne for months [34], and through migratory and feeding behaviour strategies in African elephants 175 (Loxodonta africana) [35] and blue whales (Balaenoptera musculus) [2]. Likewise, MR 176 177 measurements have shown how energetic costs of prey capture are linked to hunting 178 success and hence population viability at small population sizes in endangered species 179 including cheetah (Acinonyx jubatus) and puma (Puma concolor) [36, 37]. Similarly, MR measurements can help identify challenges and threats that animals face with climate 180 181 change. The measured MR of free-living polar bears (Ursus maritimus) has revealed unusually high energy requirements associated with carnivory, and hence how increased 182 183 search costs for prey due to sea ice loss linked to warming threatens their survival [38].

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#### 185 Elasmobranchs

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187 Although energetics studies of large elasmobranchs are challenging, they have
 188 increased our fundamental understanding of how these species survive. Megafauna,
 189 particularly ectotherms, may have advantages for feeding on widely dispersed prey, since
 190 energy reserves of larger individuals sustain them for longer because of their lower mass-

specific SMR [39]. Conversely, filter-feeding and **mesothermy** appear as the two key evolutionary pathways for the largest elasmobranchs, but also puts them at greater extinction risk due to potentially unsustainable metabolic demands if prey availability declines in changed environmental conditions [40]. Human activities are probably already having direct consequences on the energetics of elasmobranchs; for example tourism operations cause reef sharks (*Triaenodon obesus*) to be active during times they normally rest, increasing energy expenditure [29].

198 Many energetics studies of elasmobranchs have focused on how they minimise transport costs. Locomotion is energetically costly, yet many megafauna regularly migrate 199 200 thousands of kilometres. Understanding long-distance migrations, and how climate change 201 impacts them, is a key research topic for marine megafauna [41]. Studying energetics of 202 animals has informed why they move to areas that yield the greatest net energy gain. For 203 example, movement patterns of many terrestrial and marine animals are partially driven 204 by the energetic demands of traversing different environments [35, 42]. It is proposed that ectotherms choose habitats with high food availability regardless of temperature to 205 206 maximise net energy gains when feeding, but preference colder areas to minimise energy 207 expenditure when not feeding [30, 43, 44]. For example, basking shark (Cetorhinus 208 maximus) movements are both thermally driven and dependent on prey (zooplankton) biomass [45, 46]. 209

Energetics has provided insights into the underlying reasons for long-distance migrations in elasmobranchs. When the purpose of migration is primarily to improve foraging, then marine megafauna ensure that the energy gained through better feeding conditions exceeds the energetic cost of migration, in a number of ways. For example, whale sharks may reduce locomotor costs by 32% by adjusting swimming patterns while

215	foraging [22]. Most sharks are negatively buoyant and are able to use a passive, downward
216	glide to cover considerable horizontal distances without expending energy on swimming
217	(e.g. [47, 48]) and reduce the cost of vertical movements, in which they may dive 2000 m in
218	search of prey [49, 50]. Blue sharks ( <i>Prionace glauca</i> ) remain at ~400 m when migrating to
219	reduce their cost of transport [51]. Their MR in this cool water is estimated to be 40% of
220	that in warmer surface waters, though such estimates are based on the measured
221	temperature dependence of metabolism in other ectotherms [51]; no data are available
222	for blue sharks. Although energetics studies have improved our understanding of the
223	behaviour, morphology and ecology of elasmobranch megafauna, several methods must
224	be developed further to fill the information void regarding fundamental energy use.
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227	Future Research
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229	Building on recent methodological advances in measuring the energetics of
230	marine megafauna
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232	Mega-flumes and respirometry: measuring the metabolism of large animals
233	<b>Respirometry</b> remains the primary method for measuring the MR of water-breathers,
234	as other methods used for terrestrial and aquatic air-breathing species are not applicable
235	(Box 2). Advances in laboratory infrastructure, animal husbandry, and capture and transport
236	techniques have allowed researchers to perform respirometry on increasingly large marine
237	animals [52]. The MR of freely-swimming elasmobranchs up to 47.7 kg has been measured

using large (up to ~3,800 L) respirometers [9]. The main issue with this method remains the
size-limitation of laboratory respirometry systems. To understand allometric scaling of MR
in elasmobranchs (Figure 1B), the MR of incrementally larger individuals may be measured
with larger systems or opportunistic methods. For example, the MR of white sharks up to
36.2 kg was measured in an 11,360-L transport tank en-route to an aquarium [53].

243 Besides SMR, another key component of bioenergetics is the energy cost of activity. 244 Total activity cost (e.g. daily) can be determined if the cost of transport (COT) per unit 245 distance or swimming effort is known. Total activity cost is often estimated as a constant value proportional to SMR, although this is usually a spurious assumption [54]. Total activity 246 cost can be quantified using a combination of laboratory and field techniques [55]. In water-247 248 breathers, this is usually achieved using a swim-tunnel ('flume'), which comprises a closed system where water is circulated at specific speeds, and the subject animal swims to hold its 249 250 position while its MR or COT is monitored [9]. Until recently, the largest elasmobranch for 251 which COT was measured was a 9.5-kg mako shark (Isurus oxyrinchus) in a 3,000-L flume 252 [56]. The 'mega-flume' is a 26,000-L sea-deployable flume that was originally tested by 253 measuring the COT of a 36-kg zebra shark (Stegostoma fasciatum) [9], and this remains the largest animal that it has been used to measure. Flumes are yet to overcome the logistical 254 challenges required to measure COT in the largest sharks, including the potential of stress to 255 256 inflate MR and that animals may require training to swim in a flume appropriately [57]. 257 Although flumes can be used to calibrate a measure of activity to then quantify total activity cost in free-ranging animals (see below), similar results may be achieved in simple 258 respirometry setups relying on volitional activity by the animal [58]. Flumes may overcome 259 260 their logistical challenges and continue to increase in size, allowing COT measurements of

larger animals, but are unlikely to accommodate an animal >10 m in length such as a whale

262 shark.

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264 Box 2. What can we learn about elasmobranch energetics from other megafauna?

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#### 266 What can we learn from other megafauna?

267 One approach to gain insights into the energetics of elasmobranchs is to compare them with other

268 species exhibiting similar traits (Table I) and for which energetics is more easily measured. For

- 269 example, feeding, reproductive costs and costs of activity can be more easily measured in air-
- 270 breathers such as cetaceans (whales, dolphins and porpoises; see below).
- 271 Feeding: A shared zooplankton diet between some elasmobranchs and cetaceans implies similar
- 272 problems in energy acquisition, including patchy prey, high feeding costs, minimum prey density
- 273 requirements, and declining zooplankton biomass in the future [19, 59]. The assumed energy cost of
- whale lunge-feeding is used to estimate minimum prey intake from feeding events [60]. A similar
- approach could be used for sharks hunting large vertebrate prey, whereby the estimated cost of
- 276 hunting attempts could define the required success rate.

277 **Reproductive costs**: Although elasmobranchs exhibit diverse reproductive methods, many large

- 278 species produce a few, well-developed offspring, similarly to cetaceans. Therefore both may have
- similar gestation costs, although parental energy input differs substantially due to lactation costs incetaceans [61].
- Cost of activity: Although SMR cannot be extrapolated from mammals to fish [62], activity cost may
   be estimated using kinematic modelling [63], and these groups may have similar costs of transport
   per unit body weight.
- 284

- 285 Table I: Similarities and differences in traits of elasmobranch and marine mammal megafauna. Marine
- 286 mammals represent the largest group of non-elasmobranch marine megafauna, and of these, the cetaceans
- are the most morphologically and ecologically similar to large elasmobranchs.

Trait	Large elasmobranchs	Marine mammals
Oxygen exchange	Water (gills), most must swim	Air (lungs), no swimming
	continuously	required
Energy storage	Primarily in liver as lipids	Subcutaneous fat (blubber)
Thermoregulation	Mostly ectothermic	Endothermic
Fecundity	Mostly low	Low
Diet	Largest are planktivores, some	Largest are planktivores, some
	are highly active hunters	are highly active hunters
Buoyancy	Changes with body size and	Generally positive at surface,
	habitat. Large or deep-sea	may decrease with depth as is
	sharks may be neutral or	influenced by lung volume
	slightly positive, others are	
	slightly negative	
Parental care	None (but offspring may be	Substantial in most species,
	large at birth)	and offspring large at birth
Migrations	Some long distance (1000s of	Some long distance (1000s of
	km)	km)

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#### 290 Currently non-transferable methods

- 291 Unfortunately, many insights into the energetics of non-elasmobranch megafauna have been
- 292 derived using methods that are inapplicable to elasmobranchs.

293 Surfacing rates to estimate MR: Air-breathing marine megafauna surface for respiration, offering an

- 294 opportunity to estimate their MR that is not possible with water-breathers. Specifically, MR for
- 295 cetaceans can be estimated from respiration timing, pre-existing oxygen stores, tidal lung volume
- and oxygen extracted from inspired air (measured) [3].
- 297 **Doubly-labelled water (DLW)**: DLW is used to quantify energy use in wild terrestrial megafauna (e.g.
- polar bears [38] and cheetahs [37]), whereby stable isotopes of oxygen and hydrogen trace the flow
- of CO<sub>2</sub> and water through the body to quantify MR. Unfortunately, the DLW method is unsuitable for
- 300 fish due to high water fluxes between the body and environment.

301 **MR estimates from fish otoliths:** A new method of determining energy use in teleosts has been

developed using isotopic composition of carbon found in ear bones (otoliths) [64]. This method has

provided total MR estimates to a resolution of approximately 15 d [64]. Elasmobranchs lack the

- 304 calcified structures necessary for this analysis.
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# 306 Using captive elasmobranchs and back-calculating metabolic costs

307 Commercial aquaria may be valuable for determining fundamental relationships such 308 as allometric and thermal scaling of MR in the largest elasmobranchs. Some aquaria house many large elasmobranchs, including the largest ectotherm, the whale shark. Harnessing 309 these commercial enterprises can advance our understanding of the charismatic megafauna 310 311 they display, without the need for capturing wild animals. Existing infrastructure may allow large animals to be isolated and their MR measured via respirometry, providing a solution to 312 313 the size-limitation of traditional laboratory respirometry systems. However, issues remain in 314 obtaining MR measurements that reflect a normal existence in the wild [58].

315 Captive elasmobranchs can provide insight into energy requirements without the need for directly measuring MR. If the food intake, growth, and assimilation costs (dependent on 316 diet composition) of an individual are monitored, the remaining energy devoted to MR can 317 318 be calculated (Box 1). This has been partially achieved in captive white sharks; in a simple 319 bioenergetics model, assimilation costs were assumed constant (27%) and the food 320 consumption and mass of individuals were tracked [65]. The model estimated that metabolism (SMR + activity cost) and growth accounted for 45% and 28% of energy use, 321 respectively [65]. However, white sharks in captivity grew at twice the rate of wild sharks, 322 suggesting energy allocation to growth is probably higher in captive sharks due to reduced 323 324 activity and/or, as is commonly the case, energy ingestion may typically be a limiting factor

325 for growth [65]. Nevertheless, this 'back-calculation' method can estimate feeding

326 requirements to maintain growth rates, indicate metabolic sensitivity to temperature, and

327 provide insight into the interplay between reproductive rate and body size [66].

#### 328 Using new biologgers in the field to estimate the energy expenditure of locomotion

329 Advances in biologging technology have allowed researchers to progress from 330 describing animal movement or behaviour patterns to explaining their underlying causes. 331 Growing demand and commercial viability has seen a surge in development and use of new animal-borne sensors, along with attachment and retrieval methods, to explore the activity, 332 behaviour, movement and physiology of wild animals [73] (Figure 2, Box 2). For example, 333 334 combined with respirometry, animal activity measures can provide proxies of energy 335 expenditure [67]. In the laboratory, data loggers can calibrate an activity measure, often 336 body acceleration or tail-beat frequency, against MR by simultaneously measuring both at 337 multiple swim speeds [68]. The same activity measure can then be recorded in free-ranging animals via animal-borne loggers, enabling activity energy cost to be quantified [55, 68]. 338 While activity measures enable an understanding of total energy use (e.g. [69]), they can 339 340 also elucidate costs of fine-scale behaviours including differences with swimming patterns [28], time of day [55], weather [31], tides [27] or tourism operations [29]. The rigid fins of 341 342 large sharks represent ideal surfaces for sensor attachment, allowing a range of data to be 343 measured. However, the reliance on laboratory calibration of such devices inherits the same problems as traditional respirometry for megafauna, in that the required laboratory systems 344 are size limiting. Calibration with larger elasmobranchs will require advances such as the 345 346 mega-flume or other large respirometry setups [9]. However, the MR-activity relationship

may shift reliably with body mass, which would allow extrapolations from smaller animals
with similar morphologies [70].

Heart rate is commonly included in terrestrial and aquatic biologging studies to 349 estimate activity cost and total energy expenditure. For example, heart rate measurements 350 351 have shown unexpected efficiencies in birds on long-distance migrations [34], a low daily 352 activity cost in great cormorants (Phalacrocorax carbo) allowing them to survive Arctic 353 winters [71], and unusual energy usage in narwhals (Monodon monoceros) following net 354 entanglement [72]. However, the use of heart rate sensors in free-ranging fish has lagged behind other taxa [73]. Traditionally, the variation among individuals in the MR-heart rate 355 356 relationship was thought to be greater in fish (including elasmobranchs) than other taxa 357 [74], which may have slowed the development of commercially-available heart rate sensors for fish [73]. However, more recent work suggests there is a reliable relationship between 358 359 heart rate and MR (e.g. [75]). As with activity, heart rate must first be calibrated in the 360 laboratory before being employed as a quantitative measure in the field, thus making it problematic for megafauna for the reasons described above. Additionally, loggers must be 361 physically retrieved by re-capture; a difficult task for large elasmobranchs that traverse 362 oceans. 363

Swim speed has occasionally been used as a proxy for activity energy cost in bioenergetics models (e.g. [76]) and has provided useful qualitative insights into elasmobranch energetics (e.g. [77]). Swim speed is an essential parameter of kinematic modelling (KM), a promising approach that uses hydrodynamics to estimate transport cost. KM estimates forces such as lift and drag produced as an animal of known morphology moves though water at a certain speed. In combination with estimates of muscle efficiency, KM can be used to infer activity cost [63]. This modelling can outline differences in activity

cost of different movements within individuals and environments, and can be incorporated 371 372 into bioenergetics models if data on SMR, prey capture and prey energy content are available [63]. KM is also useful in understanding the evolution of morphology associated 373 with different lifestyles. For example, similarly to seabirds with high aspect-ratio wings, 374 375 pelagic sharks in oligotrophic environments have disproportionately large pectoral fins that 376 reduce the cost of a high cruising speed needed to search vast areas for prey [63]. 377 Additionally, the kinematics of breaching events may provide insights into maximum 378 movement speeds, power outputs and energy expenditure [78]. However, KM remains vulnerable to uncertainty of influential parameters that are difficult to measure, such as 379 hydrodynamic efficiency (see Outstanding Questions), for which more work is required 380 381 before reliable comparisons can be made between species and morphologies. Nevertheless, if swim speed measurements are obtained for more large elasmobranchs, KM can provide 382 383 insights into activity cost without laboratory calibration, such as relative cost of activity or 384 environmental influence on activity levels [63]. Finally, machine-learning and artificial intelligence are already being used to identify 385 386 elasmobranch behaviour based on an acceleration signature [79]. Future use of machinelearning, especially in conjunction with improved biologging sensors and an understanding 387

of activity cost, will allow for an array of pattern recognition in energy use of wild animals;

389 comparable to machine-learning that out-performs human specialists in diagnosing disease

390 [80]. This may elucidate fine-scale differences in energy use linked to particular behaviours

391 or prey availability, or prediction of metabolic responses to complex environmental

392 conditions associated with future warming and deoxygenation.

393

#### 394 Energetics of large elasmobranchs in the Anthropocene

By building on recent methodological advances in measuring the energetics of marine megafauna, a suite of important questions can be answered related to the long-term survival of elasmobranch megafauna in the Anthropocene.

#### 398 Warming and elasmobranch movement

399 Unlike most other ectotherms, the relative increase of SMR with warming in 400 elasmobranchs is not well established (Figure 1C), making it difficult to predict how growth, 401 reproduction or activity may change as oceans warm. Further, activity cost increases with 402 warming, but not necessarily in line with SMR [54]. Warming raises activity cost in multiple 403 ways: energy cost of transport per unit distance increases [63], the speed at which minimum cost of transport occurs increases (therefore increasing absolute energy use)[63], and 404 activity rates may increase [54, 81]. Animals may be able to buffer against these effects by 405 406 increasing rest periods or tracking optimal temperatures [81], e.g. some elasmobranchs 407 avoid warm surface waters once a temperature threshold is met [82]. However, obligate 408 ram ventilating elasmobranchs may have limited ability to reduce activity, and optimal 409 temperatures may no longer coincide with suitable prey habitats, aggregation sites, spawning grounds, or migration patterns [83]. Any unavoidable increases in activity cost 410 may strain the already limited metabolic scope of obligate ram ventilators, reducing their 411 412 ability for growth and reproduction [18]. Understanding plasticity in thermal responses will be important for predicting resilience of ectotherms to climate change and the sustainability 413 of long-distance migrations [10]. 414

#### 415 Ocean deoxygenation

Accelerating water deoxygenation, now seen in all oceans, is considered one of the 416 417 most significant ecological consequences of climate change [84]. Future deoxygenation is predicted to mimic conditions during the end-Permian period, when a collapse of suitably 418 419 aerobic habitat caused the largest marine extinction in history [16]. Although similarities in 420 modern climate change to conditions seen in multiple previous extinctions of large sharks suggest elasmobranch megafauna are particularly vulnerable to the effects of ocean 421 422 deoxygenation and warming [14], further work is needed to understand their metabolic 423 responses to these changing conditions and their ability to find suitably oxygenated habitats. 424

Lower levels of dissolved oxygen have far-reaching effects on animal physiology and 425 426 behaviour, including the ability to maintain energy balance [85]. Reduced oxygen partial 427 pressure and increased oxygen demand in warmer water (due to elevated SMR) reduces the 'metabolic index' of ectotherms (the ratio between O<sub>2</sub> partial pressure and SMR [86]), 428 meaning a limited capacity to increase foraging time when needed [16]. Additionally, 429 430 elevated environmental CO<sub>2</sub> associated with deoxygenation can increase metabolic demands of maintaining pH homeostasis, and activity may increase with low dissolved 431 432 oxygen as animals move to find oxygenated water [85]. Deep anoxic zones can limit the 433 diving of water-breathers, but do not directly affect air-breathers [87]. Therefore, deepdiving water-breathers may miss opportunities to hunt hypoxia-tolerant prey [85, 87]. 434 Historically, as the ratio between dissolved oxygen supply and demand declines, animals 435 that have been most susceptible include those with high energy requirements (e.g. highly 436 437 active sharks), a reduced available metabolic scope (e.g. obligate ram ventilating

elasmobranchs) [18], and the largest ectotherms [16]. The five largest extant ectotherm
species are elasmobranchs, and future research needs to examine if their size, energy use
and lifestyle are sustainable with declining ocean oxygen levels. For example, it would be
valuable to determine the reliance of elasmobranchs on deep cooler waters for energy
acquisition or reducing MR (Figure 2), that may become hypoxic and uninhabitable.

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#### 444 Concluding remarks

445 Elasmobranchs play important roles as top predators in ecosystems, but are threatened 446 by fishing and climate change. More work is needed to understand fundamental physiological traits and the vulnerability of elasmobranch megafauna compared to other 447 448 marine life. Of particular importance is their metabolic response to elevated temperatures 449 and body size, reliance on deep waters that may become hypoxic, and ability to increase 450 energy acquisition under climate change. By obtaining fundamental metabolic relationships (e.g. Q<sub>10</sub>) for large elasmobranchs, biologging and modelling can enable a better 451 452 understanding of their energetics without the need for size-limiting laboratory systems. MR 453 data for large elasmobranchs is difficult to obtain, but similar data in other hard-to-study groups has provided crucial insights into their ecology [38]. Collaboration between 454 455 researchers in different fields may be key to predicting future impacts of climate change on 456 the largest ectotherms. Already, work between ecologists and physicists has used kinematic 457 modelling to understand energy use [63, 78], and machine-learning may be incorporated 458 into existing work between physiologists and oceanographers to understand future ocean 459 warming and deoxygenation scenarios, and how this will affect animals with extreme traits.

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## 461 References

462 1. Grady, J.M. *et al.* (2014) Evidence for mesothermy in dinosaurs. *Science* 344, 1268-1272

463 2. Hazen, E.L. et al. (2015) Blue whales (Balaenoptera musculus) optimize foraging efficiency by

balancing oxygen use and energy gain as a function of prey density. *Science Adv.* 1, e1500469

465 3. Roos, M.M. *et al.* (2016) The significance of respiration timing in the energetics estimates of free-466 ranging killer whales (*Orcinus orca*). *J. Exp. Biol.* 219, 2066-77

- 467 4. Goldbogen, J. (2018) Physiological constraints on marine mammal body size. *PNAS* 115, 3995-3997
- 468 5. Gearty, W. *et al.* (2018) Energetic tradeoffs control the size distribution of aquatic mammals. *Proc*469 *Natl Acad Sci U S A* 115, 4194-4199
- 470 6. Dulvy, N.K. *et al.* (2014) Extinction risk and conservation of the world's sharks and rays. *eLife* 3,
  471 e00590
- 472 7. Queiroz, N. *et al.* (2016) Ocean-wide tracking of pelagic sharks reveals extent of overlap with
  473 longline fishing hotspots. *PNAS* 113, 1582-1587
- 474 8. Lauder, G.V. and Di Santo, V. (2015) Swimming Mechanics and Energetics of Elasmobranch Fishes.
- 475 In Physiology of Elasmobranch Fishes: Structure and Interaction with Environment (Shadwick, R.E. et
- 476 al. eds), pp. 219-253, Academic Press.
- 477 9. Payne, N.L. *et al.* (2015) A new method for resolving uncertainty of energy requirements in large
- water breathers: The 'mega-flume' seagoing swim-tunnel respirometer. *Methods Ecol. Evol.* 6, 668-677
- 480 10. Seebacher, F. *et al.* (2015) Physiological plasticity increases resilience of ectothermic animals to
  481 climate change. *Nat. Clim. Change* 5, 61-66
- 482 11. Nielsen, J. *et al.* (2016) Eye lens radiocarbon reveals centuries of longevity in the greenland shark
  483 (*Somniosus microcephalus*). *Science* 353, 702-704
- 484 12. Gillooly, J.F. *et al.* (2001) Effects of size and temperature on metabolic rate. *Science* 293, 2248-485 2251
- 486 13. Brown, J.H. *et al.* (2004) Toward a metabolic theory of ecology. *Ecology* 85, 1771-1789
- 487 14. Belben, R.A. *et al.* (2017) Ecological impact of the end-cretaceous extinction on lamniform
  488 sharks. *PLoS One* 12, e0178294
- 489 15. Daufresne, M. *et al.* (2009) Global warming benefits the small in aquatic ecosystems. *PNAS* 106,
  490 12788-12793
- 491 16. Penn, J.L. *et al.* (2018) Temperature-dependent hypoxia explains biogeography and severity of
  492 end-permian marine mass extinction. *Science* 362, eaat1327
- 493 17. Sheridan, J.A. and Bickford, D. (2011) Shrinking body size as an ecological response to climate 494 change. *Nat. Clim. Change* 1, 401-406
- 495 18. Dowd, W. *et al.* (2006) Standard and routine metabolic rates of juvenile sandbar sharks
- 496 (*Carcharhinus plumbeus*), including the effects of body mass and acute temperature change. *Fish.* 497 *Bull.* 104, 323-331
- 498 19. Kwiatkowski, L. *et al.* (2018) Consistent trophic amplification of marine biomass declines under
  499 climate change. *Glob. Chang. Biol.* 25, 218-229
- 500 20. Woodworth-Jefcoats, P.A. et al. (2017) Climate change is projected to reduce carrying capacity
- and redistribute species richness in north pacific pelagic marine ecosystems. *Glob. Chang. Biol.* 23,
   1000-1008
- 503 21. Sims, D.W. (1999) Threshold foraging behaviour of basking sharks on zooplankton: Life on an 504 energetic knife-edge? *Proc. R. Soc. London, B* 266, 1437-1443
- 505 22. Meekan, M. *et al.* (2015) Swimming strategy and body plan of the world's largest fish:
- 506 Implications for foraging efficiency and thermoregulation. *Front. Mar. Sci.* 2, 64
- 507 23. Estes, J.A. *et al.* (2011) Trophic downgrading of planet Earth. *Science* 333, 301-306
- 508 24. Christensen, V. and Walters, C.J. (2004) Ecopath with Ecosim: methods, capabilities and
- 509 limitations. Ecol. Model. 172, 109-139

- 510 25. Kitchell, J.F. *et al.* (1977) Applications of a bioenergetics model to yellow perch (*Perca flavescens*)
- 511 and walleye (Stizostedion vitreum vitreum). J. Fish. Board Canada 34, 1922-1935
- 512 26. Deslauriers, D. *et al.* (2017) Fish bioenergetics 4.0: An R-based modeling application. *Fisheries* 42, 513 586-596
- 514 27. Bouyoucos, I.A. *et al.* (2017) Swimming speeds and metabolic rates of semi-captive juvenile
- lemon sharks (*Negaprion brevirostris*, Poey) estimated with acceleration biologgers. *J. Exp. Mar. Biol. Ecol.* 486, 245-254
- 517 28. Bouyoucos, I.A. *et al.* (2018) In situ swimming behaviors and oxygen consumption rates of
- 518 juvenile lemon sharks (*Negaprion brevirostris*). Environ. Biol. Fish. 101, 761-773
- 519 29. Barnett, A. *et al.* (2016) Ecotourism increases the field metabolic rate of whitetip reef sharks.
- 520 Biol. Conserv. 199, 132-136
- 521 30. Gleiss, A.C. *et al.* (2017) Are vertical migrations driven by circadian behaviour? Decoupling of
- activity and depth use in a large riverine elasmobranch, the freshwater sawfish (*Pristis pristis*). *Hydrobiologia* 787, 181-191
- 524 31. Payne, N.L. *et al.* (2013) Rain reverses diel activity rhythms in an estuarine teleost. *Proc. R. Soc.* 525 *London, B* 280, 20122363
- 526 32. Lawson, C.L. et al. (2018) The influence of ontogenetic diet variation on consumption rate
- 527 estimates: A marine example. Sci. Rep. 8, 10725
- 528 33. Elser, J.J. *et al.* (1996) Organism size, life history, and N: P stoichiometry: toward a unified view of 529 cellular and ecosystem processes. *BioScience* 46, 674-684
- 530 34. Weimerskirch, H. *et al.* (2016) Frigate birds track atmospheric conditions over months-long
- transoceanic flights. *Science* 353, 74-78
- 532 35. Wall, J. et al. (2006) Elephants avoid costly mountaineering. Curr. Biol. 16, R527-R529
- 533 36. Williams, T.M. *et al.* (2014) Instantaneous energetics of puma kills reveal advantage of felid
- sneak attacks. *Science* 346, 81-85
- 535 37. Scantlebury, D.M. *et al.* (2014) Flexible energetics of cheetah hunting strategies provide
- 536 resistance against kleptoparasitism. *Science* 346, 79-81
- 38. Pagano, A. *et al.* (2018) High-energy, high-fat lifestyle challenges an arctic apex predator, the
  polar bear. *Science* 359, 568-572
- 39. Hays, G.C. *et al.* (2018) A paradigm shift in the trophic importance of jellyfish? *Trends Ecol. Evol.*33, 874-884
- 40. Pimiento, C. *et al.* (2019) Evolutionary pathways toward gigantism in sharks and rays. *Evolution*73, 588-599
- 41. Hays, G.C. *et al.* (2016) Key questions in marine megafauna movement ecology. *Trends Ecol. Evol.*31, 463-475
- 42. McElroy, B. *et al.* (2012) Optimum swimming pathways of fish spawning migrations in rivers. *Ecology* 93, 29-34
- 547 43. McLaren, I.A. (1963) Effects of temperature on growth of zooplankton, and the adaptive value of 548 vertical migration. *J. Fish. Board Canada* 20, 685-727
- 44. Papastamatiou, Y.P. *et al.* (2015) Drivers of Daily Routines in an Ectothermic Marine Predator:
  Hunt Warm, Rest Warmer? *PLoS One* 10, e0127807
- 45. Sims, D.W. *et al.* (2006) Encounter success of free-ranging marine predator movements across a dynamic prey landscape. *Proc. R. Soc. London, B* 273, 1195-1201
- 553 46. Cotton, P.A. *et al.* (2005) The effects of climate variability on zooplankton and basking shark
- 554 (*Cetorhinus maximus*) relative abundance off southwest britain. *Fish. Oceanogr.* 14, 151-155
- 47. Gleiss, A.C. *et al.* (2011) Convergent evolution in locomotory patterns of flying and swimming
- animals. Nat. Commun. 2, 352
- 48. Gleiss, A.C. *et al.* (2011) Moved by that sinking feeling: variable diving geometry underlies
- 558 movement strategies in whale sharks. *Funct. Ecol.* 25, 595-607
- 49. Thorrold, S.R. et al. (2014) Extreme diving behaviour in devil rays links surface waters and the
- 560 deep ocean. Nat. Commun. 5, 4274

- 561 50. Tyminski, J.P. *et al.* (2015) Vertical movements and patterns in diving behavior of whale sharks as
- revealed by pop-up satellite tags in the eastern gulf of mexico. *PloS One* 10, e0142156
- 563 51. Campana, S.E. *et al.* (2011) Migration pathways, behavioural thermoregulation and
- 564 overwintering grounds of blue sharks in the northwest atlantic. *PloS One* 6, e16854
- 565 52. Smith, M. *et al.* eds (2017) *The Elasmobranch Husbandry Manual II Recent Advances in the Care* 566 *of Sharks, Rays and their Relatives,* Ohio Biological Survey.
- 567 53. Ezcurra, J. et al. (2012) Oxygen consumption rate of young-of-the-year white sharks,
- 568 Carcharodon carcharias, during transport to the Monterey Bay aquarium. In Global Perspectives on
- 569 the Biology and Life History of the White Shark (1st edn) (Domeier, M.L. ed), pp. 17-26, CRC Press.
- 570 54. Halsey, L.G. *et al.* (2015) The interactions between temperature and activity levels in driving
- 571 metabolic rate: Theory, with empirical validation from contrasting ectotherms. *Oecologia* 177, 1117 572 1129
- 573 55. Brodie, S. *et al.* (2016) Improving consumption rate estimates by incorporating wild activity into a 574 bioenergetics model. *Ecol. Evol.* 6, 2262-2274
- 575 56. Sepulveda, C. *et al.* (2007) Aerobic metabolic rates of swimming juvenile mako sharks, *Isurus* 576 *oxyrinchus. Mar. Biol.* 152, 1087-1094
- 577 57. Lear, K.O. *et al.* (2019) Treading water: respirometer choice may hamper comparative studies of
- 578 energetics in fishes. Mar. Freshwater Res. 70, 437-448
- 579 58. Lear, K.O. *et al.* (2017) Correlations of metabolic rate and body acceleration in three species of 580 coastal sharks under contrasting temperature regimes. *J. Exp. Biol.* 220, 397-407
- 581 59. Acevedo-Gutiérrez, A. *et al.* (2002) High feeding costs limit dive time in the largest whales. *J. Exp.*582 *Biol.* 205, 1747-1753
- 583 60. Goldbogen, J. et al. (2011) Mechanics, hydrodynamics and energetics of blue whale lunge
- 584 feeding: Efficiency dependence on krill density. J. Exp. Biol. 214, 131-146
- 585 61. Braithwaite, J.E. *et al.* (2015) Optimal migration energetics of humpback whales and the 586 implications of disturbance. *Conserv. Physiol.* 3, cov001
- 586 Implications of disturbance. *Conserv. Physiol.* 3, covoul
- 587 62. White, C.R. *et al.* (2006) The scaling and temperature dependence of vertebrate metabolism.
  588 *Biol. Lett.* 2, 125-127
- 589 63. losilevskii, G. and Papastamatiou, Y.P. (2016) Relations between morphology, buoyancy and 590 energetics of requiem sharks. *R Soc Open Sci* 3, 160406
- 591 64. Chung, M.-T. *et al.* (2019) Field metabolic rates of teleost fishes are recorded in otolith
- 592 carbonate. Comm. Biol. 2, 24
- 593 65. Ezcurra, J.M. *et al.* (2012) Captive feeding and growth of young-of-the-year white sharks,
- 594 *Carcharodon carcharias*, at the monterey bay aquarium. In *Global Perspectives on the Biology and* 595 *Life History of the White Shark* (Domeier, M.L. ed), pp. 3-16, CRC Press.
- 596 66. Marshall, D.J. and White, C.R. (2018) Have we outgrown the existing models of growth? *Trends* 597 *Ecol. Evol.* 34, 102-111
- 598 67. Metcalfe, J.D. *et al.* (2016) Recent advances in telemetry for estimating the energy metabolism of 599 wild fishes. *J Fish Biol* 88, 284-97
- 600 68. Gleiss, A.C. et al. (2011) Making overall dynamic body acceleration work: On the theory of
- acceleration as a proxy for energy expenditure. *Methods Ecol. Evol.* 2, 23-33
- 602 69. Semmens, J. *et al.* (2013) Feeding requirements of white sharks may be higher than originally
- 603 thought. Sci. Rep. 3, 1471
- 604 70. Halsey, L.G. et al. (2011) Assessing the development and application of the accelerometry
- technique for estimating energy expenditure. *Comp. Biochem. Physiol., Part A: Mol. Integr. Physiol.*158, 305-314
- 71. White, C.R. *et al.* (2011) Metabolic rate throughout the annual cycle reveals the demands of an
- arctic existence in great cormorants. *Ecology* 92, 475-486
- 609 72. Williams, T.M. *et al.* (2017) Paradoxical escape responses by narwhals (*Monodon monoceros*).
- 610 Science 358, 1328-1331

- 611 73. Cooke, S.J. et al. (2016) Remote bioenergetics measurements in wild fish: Opportunities and
- 612 challenges. Comp. Biochem. Physiol., Part A: Mol. Integr. Physiol. 202, 23-37
- 74. Thorarensen, H. *et al.* (1996) The limitations of heart rate as a predictor of metabolic rate in fish. *J. Fish Biol.* 49, 226-236
- 615 75. Clark, T.D. et al. (2010) Simultaneous biologging of heart rate and acceleration, and their
- 616 relationships with energy expenditure in free-swimming sockeye salmon (Oncorhynchus nerka). J.
- 617 Comp. Physiol. B 180, 673-684
- 618 76. Sundström, L.F. and Gruber, S.H. (1998) Using speed-sensing transmitters to construct a
- bioenergetics model for subadult lemon sharks, *Negaprion brevirostris*, in the field. *Hydrobiologia*371, 241-247
- 621 77. Watanabe, Y.Y. *et al.* (2012) The slowest fish: Swim speed and tail-beat frequency of greenland 622 sharks. *J. Exp. Mar. Biol. Ecol.* 426, 5-11
- 78. Johnston, E.M. *et al.* (2018) Latent power of basking sharks revealed by exceptional breaching
  events. *Biol. Lett.* 14, 20180537
- 79. Brewster, L.R. *et al.* (2018) Development and application of a machine learning algorithm for
   classification of elasmobranch behaviour from accelerometry data. *Mar. Biol.* 165, 62
- 627 80. Rajpurkar, P. *et al.* (2017) Chexnet: Radiologist-level pneumonia detection on chest x-rays with 628 deep learning. *arXiv preprint* 1711, 05225
- 629 81. Payne, N.L. *et al.* (2018) Combining abundance and performance data reveals how temperature
- regulates coastal occurrences and activity of a roaming apex predator. *Glob. Change Biol.* 24, 1884 1893
- 632 82. Andrzejaczek, S. et al. (2018) Temperature and the vertical movements of oceanic whitetip
- 633 sharks, Carcharhinus longimanus. Sci Rep 8, 8351
- 83. Pecl, G.T. *et al.* (2017) Biodiversity redistribution under climate change: Impacts on ecosystems
  and human well-being. *Science* 355, eaai9214
- 84. Oschlies, A. *et al.* (2018) Drivers and mechanisms of ocean deoxygenation. *Nat. Geosci.* 11, 467–
  473
- 85. Breitburg, D. *et al.* (2018) Declining oxygen in the global ocean and coastal waters. *Science* 359,
  eaam7240
- 640 86. Deutsch, C. *et al.* (2015) Climate change tightens a metabolic constraint on marine habitats.
- 641 Science 348, 1132-1135
- 642 87. Stramma, L. *et al.* (2012) Expansion of oxygen minimum zones may reduce available habitat for 643 tropical pelagic fishes. *Nat. Clim. Change* 2, 33-37
- 644 88. Norin, T. and Gamperl, A.K. (2018) Metabolic scaling of individuals vs. populations: Evidence for 645 variation in scaling exponents at different hierarchical levels. *Funct. Ecol.* 32, 379-388
- 646 89. Chapman, C.A. *et al.* (2011) The physiological tolerance of the grey carpet shark (*Chiloscyllium*
- 647 *punctatum*) and the epaulette shark (*Hemiscyllium ocellatum*) to anoxic exposure at three seasonal
- 648 temperatures. Fish Physiol. & Bioc. 37, 387-399
- 649 90. Clarke, A. and Johnston, N.M. (1999) Scaling of metabolic rate with body mass and temperature 650 in teleost fish. *J. Anim. Ecol.* 68, 893-905
- 651 91. Clarke, A. ed (2017) *Principles of thermal ecology: Temperature, energy and life*, Oxford

652 University Press.

- 653 92. Nakamura, I. *et al.* (2015) Ocean sunfish rewarm at the surface after deep excursions to forage 654 for siphonophores. *J. Anim. Ecol.* 84, 590-603
- 655 93. Hight, B.V. and Lowe, C.G. (2007) Elevated body temperatures of adult female leopard sharks,
- 656 *Triakis semifasciata*, while aggregating in shallow nearshore embayments: Evidence for behavioral 657 thermoregulation? *J. Exp. Mar. Biol. Ecol.* 352, 114-128
- 658 94. Whitney, N.M. *et al.* (2019) Advances in the application of high-resolution biologgers to
- 659 Elasmobranch fishes. In Shark Research: Emerging Technologies and Applications for the Field and
- 660 Laboratory (Carrier, J. et al. eds), CRC Press.

- 661 95. Bernal, D. and Lowe, C.G. (2015) Field studies of elasmobranch physiology. In *Fish Physiology*, pp.
- 662 311-377, Elsevier.
- 663 96. Downs, C.J. et al. (2008) Scaling metabolic rate with body mass and inverse body temperature: a
- test of the Arrhenius fractal supply model. *Funct. Ecol.* 22, 239-244
- 665 97. Del Raye, G. et al. (2013) Travelling light: White sharks (Carcharodon carcharias) rely on body
- 666 lipid stores to power ocean-basin scale migration. Proc. R. Soc. B 280, 20130836
- 667 98. Lawson, G.L. *et al.* (2015) Development of an animal-borne "sonar tag" for quantifying prey
- availability: test deployments on northern elephant seals. Animal Biotelemetry 3,
- 669 99. Melvin, G.D. (2016) Observations of in situ Atlantic bluefin tuna (*Thunnus thynnus*) with 500-kHz
- 670 multibeam sonar. ICES J. Mar. Sci. 73, 1975-1986
- 671 100. Grady, J.M. *et al.* (2019) Metabolic asymmetry and the global diversity of marine predators.
- 672 *Science* 363, eaat4220
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Figure 1. Extrapolation to obtain metabolic rate estimates for large elasmobranchs. To 679 calculate the MR of large animals, smaller species or juveniles of the same species are 680 681 commonly used to calculate the allometric scaling of MR, and the trend extrapolated (e.g. [18, 69]). Choice of proxy species is important, as metabolism is affected by physiology, 682 683 morphology, activity levels, swimming style, thermal acclimation, and endothermy [9, 54, 62]. 684 How MR scales with body mass is an on-going debate (e.g. [62, 88]), and small errors can result in large differences in MR estimates when extrapolating to megafauna [9]. Similarly, 685 686 the sensitivity of MR to temperature  $(Q_{10})$  for elasmobranchs is not well established 687 compared with other taxa [18, 26, 58]. Pictured (A) a whale shark (Rhincodon typus); the world's largest ectotherm. (B) The range of SMR estimates for large sharks based on the 688 common range of allometric scaling exponents (b = 0.67-0.89), with all other parameters held 689 690 constant (temperature=20°C, Q<sub>10</sub>=2.1). For example, the estimated SMR of a 10,000 kg whale shark is likely to be between 69 and 523 kJ  $h^{-1}$  (>7-fold range). (C) The range of inferred SMR 691 for a 10,000 kg whale shark within its natural temperature range based on Q<sub>10</sub> values 692 693 described in the literature for sharks  $(Q_{10}=1.3-2.9)$  [18, 89], teleosts  $(Q_{10}=1.83)$  [90] and the 694 mean across ectotherm taxa ( $Q_{10}=2.19$ ) [91], with other parameters kept constant (b=0.70). 695 At 30°C, estimated SMR ranges from 45 to 502 kJ h<sup>-1</sup>. Photo and silhouettes reproduced from 696 Simon Pierce (simonjpierce.com; (A)), and Natasha Sinegina, T. Michael Keesey, Steven Traver 697 (phylopic.org; (B, C)). 698

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Figure 2. Estimating metabolic rate for free-living sharks using data from animal-borne 701 702 tags. It is now possible to track sharks routinely (e.g. via satellite or by using acoustic tags) while obtaining continuous data that enable changes in MR over time to be estimated. (A) A 703 704 blue shark (Prionace glauca). (B) Tracks of seven individuals that moved in and out of the Gulf Stream [51]. When in warm waters of the Gulf Stream, the sharks tend to dive and occupy 705 706 deeper, relatively cool waters (~400 m). In surface waters of the Gulf Stream, the MR of 707 sharks is estimated to be 2.5x higher than at depth, once body temperature equilibrates with water temperature [51]. (C) Simulated biologging data on the depth of blue sharks and 708 709 associated water and body temperatures. Water temperature is taken from dive profiles of 710 blue sharks [51], with resulting body temperature based on the measured thermal inertia of leopard sharks (Triakis semifasciata) and ocean sunfish (Mola mola) during short dives [92, 711 93]. The rate of change in body temperature (thermal inertia) will depend on animal body 712 713 size and gill surface area (where most heat-loss occurs in fish), the difference in temperature between the body and surrounding water, and heat generated by metabolism. Some sharks 714 715 demonstrate mesothermy, and so their heat loss may differ. Thermal inertia has not been measured in large, fully ectothermic elasmobranchs, but likely causes substantial differences 716 717 between water and body temperature of large elasmobranchs that traverse the water column. (D) Simulated data to illustrate how instantaneous estimates of MR could be 718 719 achieved by integrating animal-borne sensor data such as body temperature and 720 acceleration. For example, metabolic costs of blue sharks diving to cooler waters can be more 721 accurately determined if their body temperature and swimming effort are considered; the 722 sharks likely have a low cost of activity as they passively glide to descend, but increase activity 723 costs at depth as they forage and then return to the warmer surface (see reviews on

724	elasmobranch biologging [94, 95]). Reproduced from Mark Conlin/NMFS (A) and [51] under
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753 Box 1 Figure I. (A) A bioenergetics model calculates total daily energy expenditure by summing the 754 energy costs of four broad body processes. These components may be simple (a single number for 755 each) or more complex (multiple interdependent equations). Generally, four components are 756 measured (SMR, active metabolic rate, assimilation, and growth and reproduction) and summed to 757 calculate total energy requirements. Food consumption is then assumed to be equal to those total 758 energy requirements. Alternatively, consumption is measured directly and then any other single 759 unknown component can be estimated. (B) The mass-balance equation of a bioenergetics model. 760 Processes on the left sum to the daily energy requirements, but are also dependent on consumption 761 level. For example, if warmer temperatures cause an increase in SMR, the animal must either 762 consume more energy or compensate (keeping consumption constant) by devoting less energy to 763 another process (usually growth or reproduction). Silhouette reproduced from Dmitry Bogdanov 764 (vectorized by T. Michael Keesey).