## What Are the Roles for Dehydroascorbate Reductases and Glutathione in Sustaining Ascorbate Accumulation?

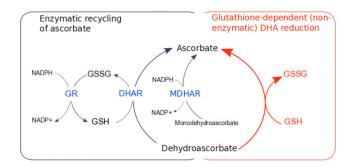
Ascorbate (vitamin C) is one of the main redox buffers required for metabolizing reactive oxygen species. Its action is essential to protect cells against free oxygen radicals, especially under stress conditions, such as high light. Since ascorbate biosynthesis is slow and energy consuming, various strategies are in place to ensure its recycling. Ascorbate can be regenerated from its oxidized forms, monodehydroascorbate (MDHA) and dehydroascorbate (DHA), by MDHA and DHA reductases (MDHARs and DHARs), respectively (Smirnoff, 2018) The activity of DHARs is coupled to oxidation of glutathione, which is also required for spontaneous, nonenzymatic reduction of DHA to ascorbate (Fig. 1).

The role of DHAR proteins in ascorbate regeneration has been demonstrated in tobacco (Nicotiana *benthamiana*), since DHAR-down-regulating tobacco lines show reduced total ascorbate levels, lower dry weight, and diminished photosynthetic efficiency (Chen and Gallie, 2006, 2008). Likewise, DHAR overexpression in tobacco and maize (Zea mays) led to an increase in ascorbate and glutathione concentration, as well as a shift toward the reduced state for glutathione (Chen et al., 2003). However, the physiological importance of DHARs in ascorbate regeneration has been recently called into question. An Arabidopsis (Arabidopsis thaliana) mutant lacking all three *DHARs* ( $\Delta$ *dhar*), with negligible DHAR activity, was shown to be equivalent to wild-type plants in terms of growth and development, as well as ascorbate levels (Rahantaniaina et al., 2017). Analysis of the  $\Delta dhar$  mutant showed that DHARs are required to couple hydrogen peroxide metabolism to glutathione oxidation and that this is functionally important for downstream activation of the salicylic acid pathway. Thus, the role of DHARs in ascorbate recycling remains controversial.

In this issue of *Plant Physiology*, Terai et al. (2020) dissect the functions of DHARs in a context of highlight stress, a condition that increases ascorbate oxidation as a protective mechanism and increases the ascorbate pool (Dowdle et al., 2007). Analysis of the Arabidopsis  $\Delta dhar$  mutant confirmed indeed that DHAR activity is dispensable for growth and ascorbate homeostasis under low light. When subjected to high-light stress, both the wild-type plants and  $\Delta dhar$  mutants accumulated ascorbate to high levels, but minor differences were observed after a prolonged stress. The lower ascorbate accumulation of  $\Delta dhar$  relative to the wild type was associated with a slight overaccumulation of threonate, an ascorbate degradation

product. The authors hypothesized that the physiological significance of DHAR activity depends on the ascorbate pool size and that DHARs are required only at high levels of ascorbate. In agreement with this hypothesis, both the ascorbate-deficient mutant *vtc2-4* (which contains only 30% of the wild-type ascorbate) and the *vtc2-4*  $\Delta dhar$  quadruple mutant showed comparable ascorbate accumulation in response to high light. In other words, when the Arabidopsis pool of ascorbate is small, DHAR appears unnecessary for its regeneration.

Terai et al. (2020) examined if nonenzymatic reduction of ascorbate by glutathione could function as a backup for DHAR activity. Consistent with earlier reports (Han et al., 2013), analysis of the glutathionedeficient pad2-1 plants demonstrated that 30% of the wild-type glutathione level is enough to sustain ascorbate recycling during high-light exposure. However, ascorbate accumulation was completely blocked in the  $\Delta dhar pad2-1$  quadruple mutants, which consistently showed bleaching, lower photochemical activity, and a higher accumulation of threonate in response to high light. Collectively, these data suggest that under high-light stress, glutathione can compensate for DHAR loss of function. A blockage of ascorbate accumulation in response to high light was also observed when glutathione deficiency was induced pharmacologically by buthionine sulfoximine treatment, providing extra evidence that, in high-light conditions, glutathione acts as a substitute for ascorbate reduction (Fig. 1).



**Figure 1.** Ascorbate recycling pathways. Since ascorbate biosynthesis is slow and energy consuming, plants have evolved efficient ascorbate recycling mechanisms. MDHA and DHA can be enzymatically reconverted to ascorbate via MDHA or DHA reductases, respectively. Alternatively, glutathione reacts chemically with DHA to regenerate ascorbate—a reaction long thought to be negligible. Terai et al. (2020) demonstrate that DHAR function is dispensable in optimal growth conditions, while, under conditions of high-light stress, DHAR and glutathione nonenzymatic reduction cooperate to ensure appropriate accumulation of ascorbate. Figure prepared by A.M.

<sup>&</sup>lt;sup>1</sup>Author for contact: elisa.dell-aglio@insa-lyon.fr.

<sup>&</sup>lt;sup>2</sup>Senior author.

www.plantphysiol.org/cgi/doi/10.1104/pp.20.00388

This report constitutes a meaningful advancement on the physiological role of DHARs. It now seems that, in Arabidopsis, DHARs are not normally required for ascorbate recycling or to maintain ascorbate redox state even under conditions that are known to stimulate glutathione oxidation without affecting ascorbate levels (Rahantaniaina et al., 2017), contrary to previously reported findings in tobacco (Chen and Gallie, 2006, 2008). Rather, Terai et al. (2020) raise the possibility that, in Arabidopsis, DHAR activity is required for ascorbate recycling only when glutathione availability is low and in conditions of high ascorbate accumulation (such as high light). Studies on a larger number of plant models, crops, and growth conditions, especially those such as cadmium stress that affect glutathione availability, might help to further clarify whether and when DHARs fulfill a direct role in ascorbate recycling.

Elisa Dell'Aglio<sup>1</sup> ORCID ID: 0000-0001-6363-9109 Institut National des Sciences Appliquées-Lyon, 69100 Villeurbanne, France

> Amna Mhamdi<sup>2</sup> ORCID ID: 0000-0001-9959-1362 Department of Plant Biotechnology and

## Bioinformatics and VIB Center for Plant Systems Biology, Ghent University, 9052 Ghent, Belgium

## LITERATURE CITED

- Chen Z, Gallie DR (2006) Dehydroascorbate reductase affects leaf growth, development, and function. Plant Physiol 142: 775–787
- Chen Z, Gallie DR (2008) Dehydroascorbate reductase affects nonphotochemical quenching and photosynthetic performance. J Biol Chem 283: 21347–21361
- Chen Z, Young TE, Ling J, Chang SC, Gallie DR (2003) Increasing vitamin C content of plants through enhanced ascorbate recycling. Proc Natl Acad Sci USA 100: 3525–3530
- Dowdle J, Ishikawa T, Gatzek S, Rolinski S, Smirnoff N (2007) Two genes in Arabidopsis thaliana encoding GDP-L-galactose phosphorylase are required for ascorbate biosynthesis and seedling viability. Plant J 52: 673–689
- Han Y, Chaouch S, Mhamdi A, Queval G, Zechmann B, Noctor G (2013) Functional analysis of *Arabidopsis* mutants points to novel roles for glutathione in coupling H<sub>2</sub>O<sub>2</sub> to activation of salicylic acid accumulation and signaling. Antioxid Redox Signal **18**: 2106–2121
- Rahantaniaina MS, Li S, Chatel-Innocenti G, Tuzet A, Issakidis-Bourguet E, Mhamdi A, Noctor G (2017) Cytosolic and chloroplastic DHARs cooperate in oxidative stress-driven activation of the salicylic acid pathway. Plant Physiol 174: 956–971
- Smirnoff N (2018) Ascorbic acid metabolism and functions: A comparison of plants and mammals. Free Radic Biol Med 122: 116–129
- Terai Y, Ueno H, Ogawa T, Sawa Y, Miyagi A, Kawai-Yamada M, Ishikawa T, Maruta T (2020) Dehydroascorbate reductases and glutathione set a threshold for high light-induced ascorbate accumulation. Plant Physiol 183: 112–122