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MINI-REVIEW

Fungal bioremediation of soil co-contaminated with petroleum hydrocarbons and toxic metals



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Abstract

Much research has been carried out on the bacterial bioremediation of soil contaminated with petroleum hydrocarbons and toxic metals but much less is known about the potential of fungi in sites that are co-contaminated with both classes of pollutants. This article documents the roles of fungi in soil polluted with both petroleum hydrocarbons and toxic metals as well as the mechanisms involved in the biotransformation of such substances. Soil characteristics (e.g., structural components, pH, and temperature) and intracellular or excreted extracellular enzymes and metabolites are crucial factors which affect the efficiency of combined pollutant transformations. At present, bioremediation of soil co-contaminated with petroleum hydrocarbons and toxic metals is mostly focused on the removal, detoxification, or degradation efficiency of single or composite pollutants of each type. Little research has been carried out on the metabolism of fungi in response to complex pollutant stress. To overcome current bottle-necks in understanding fungal bioremediation, the potential of new approaches, e.g., gradient diffusion film technology (DGT) and metabolomics, is also discussed.

Key points

- Fungi play important roles in soil co-contaminated with TPH and toxic metals.
- Soil characteristics, enzymes, and metabolites are major factors in bioremediation.
- DGT and metabolomics can be applied to overcome current bottlenecks.

Keywords Fungi · Bioremediation · Petroleum hydrocarbons · Toxic metals

Co-contamination in the soil environment

With the accelerating pace of industrialization and urbanization, soil contamination has become a critical worldwide concern because of the threat to natural ecosystems and human health and much research has been carried out on innovative and cost-effective remediation technologies (Dong et al. 2013; Khan et al. 2018; Song et al. 2017). Soils co-contaminated with petroleum hydrocarbons and toxic metals are one of the major challenging problems in petroleum-producing countries, such as Qatar (Freije 2015), China (Cheng et al. 2019; Dong et al. 2013), and Russia (Kuvukina et al. 2018). Petroleum hydrocarbons and associated by-products found in soil are usually generated from accidental spills of crude oil, fuel contamination, refining processes, and subsequent problems associated with distribution and utilization. Spilled contaminants penetrate into soil pores and adsorb onto soil particles, moving vertically with capillary and gravitational forces which alters chemical, physical and biological properties, and composition (Czarny et al. 2020; dos Santos and Maranho 2018). Alkanes, aromatic compounds, nitrogensulfur-oxygen-containing compounds, and asphaltene are the major constituents of total petroleum hydrocarbons (TPH). The aromatic fraction refers to those compounds with benzene rings including polycyclic aromatic hydrocarbons (PAHs), which contain multiple fused aromatic rings, and are listed as priority pollutants due to their carcinogenic, mutagenic, and toxic properties as well as environmental recalcitrance (Czarny et al. 2020; Khan et al. 2018).

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Toxic metals found in petroleum-contaminated soils include As, Ba, Cd, Cr, Pb, Hg, Ni, V, and Zn, and these are mainly associated with petroleum extraction and refining, and combustion of fuel for heat and transport (Adeniyi and Afolabi 2002; Klimek et al. 2016; Kuyukina et al. 2018; Muniz et al. 2004). The heavy oils in Russia are enriched with V and Ni, and trace amounts of Cd, Pb, and Zn (Kuyukina et al. 2018). More than 20 soil samples collected from petroleum-producing sites in China contained Cd, Ni, Cr, and Zn at concentrations ranging from 0.08–8.18, 21.6–40.7, 25.9–71.5, and 36.7–226.0 mg/kg dry weight, respectively (Cheng et al. 2019).

Remediation of soil co-contaminated with organic and inorganic pollutants is a complex problem as these two pollutant classes need to be treated differently (Gadd 2004). The presence of toxic metals in co-contaminated soil can inhibit petroleum biodegrading microorganisms, affecting growth and metabolism, nitrogen and sulfur conversions, and dehalogenation (Biswas et al. 2015; El-Azeem et al. 2013; Sandrin and Maier 2003). Toxic metal species may exert a plethora of toxic effects depending on metal concentration and speciation, physicochemical factors, and the organism's ability to respond to metal stress through intrinsic or induced mechanisms (Gadd 1993; Gadd 2007; Rangel et al. 2018). Metals can exert toxicity in many ways, e.g., inhibition of enzymes, displacement or substitution of essential metals, disruption of cell and organellar membranes, and interaction with normal cellular homeostatic and stress response systems (Gadd 1993; Gadd 2007; Sullivan and Gadd 2019). For example, toxic metal cations may substitute for essential metal co-factors within an enzyme (e.g., Cd²⁺ may substitute for Zn²⁺) resulting in enzyme dysfunction (Sandrin and Maier 2003). Petroleum hydrocarbons in toxic metal-contaminated soils are hydrophobic materials with low water solubility and preferentially attach to the soil matrix which reduces the bioavailability of toxic metals to potential bioremediating microorganisms (Lai et al. 2009). Although research has been carried out on the bioremediation of co-contaminated soil with bacterial systems, much less attention has been paid to the potential roles of fungi in soils contaminated with petroleum hydrocarbons and toxic metals.

Fungi in soil co-contaminated with petroleum hydrocarbons and toxic metals

Fungi are ubiquitous chemoorganotrophic (heterotrophic) organisms (Gadd 2008; Gadd 2017), and are one of the three major clades of eukaryotic life that independently evolved multicellular organization (Stajich et al. 2010). The colonization of soil by fungal mycelium results in enmeshment and aggregation of soil particles and improvement of soil structure, sometimes facilitating contaminant bioavailability (Harms et al. 2011). Compared with bacteria, filamentous fungi show some advantages in the transport or translocation of essential substances, including nutrients and water, and the pollutant itself, over significant distances (Boswell et al. 2003; Furuno et al. 2012; Boswell et al. 2002; Harms et al. 2011; Jacobs et al. 2002; Worrich et al. 2018). It is also significant that fungal mycelia can act as "highways" in facilitating the transport of pollutant-degrading bacteria over distance in soil which can enhance bioremediation (Banitz et al. 2013; Kohlmeier et al. 2005; Wick et al. 2007).

Many fungi can survive and grow in the presence of toxic metals and this depends on intrinsic biochemical and structural properties, physiological and/or genetic adaptation, including morphological changes, and environmental modification of metal speciation, bioavailability, and toxicity (Gadd 1993; Gadd 2010; Glasauer et al. 2004; Sullivan and Gadd 2019). Filamentous fungi, e.g., Aspergillus and Penicillium spp., have been investigated for the degradation of aliphatic hydrocarbons, chlorophenols, and polycyclic aromatic hydrocarbons, with the organic pollutants serving as carbon and energy sources (Harms et al. 2011; Hofrichter et al. 1994; Pinedo-Rivilla et al. 2009) (Table 1). The ability of ureolytic fungi, such as Neurospora crassa, to immobilize metals has been investigated because, when incubated in urea-supplemented media, toxic metals are precipitated as carbonates and/or oxides (Li et al. 2015; Li and Gadd 2017a; Li and Gadd 2017b; Li et al. 2019; Li et al. 2016; Li et al. 2014). When grown in urea-containing media supplemented with heavy oil and Ca²⁺, such mineral precipitation tended to aggregate along the edge of the heavy oil which may provide an additional carbon or energy source during the biomineralization process (Fig. 1). Moreover, fungi are primary decomposers of organic matter and plant biomass in soil with the chemical structure of lignin polymers of wood being comparable with the aromatic structure of PAHs (Haritash and Kaushik 2009; Vanholme et al. 2010) (Fig. 2). Because of this, many lignin-degrading fungi, e.g., Phanerochaete chrysosporium, have been investigated for degradation of PAHs and other aromatic compounds because of the wide range of substances that can be attacked by such organisms (Gadd 2004; Gadd 2001). Some fungi convert high-molecular-mass PAHs such as the highly carcinogenic benzo $\left[\alpha\right]$ pyrene into water-soluble products using nonspecific detoxification mechanisms (Harms et al. 2011). Fusarium solani and Hypocrea lixii isolated from petrol station soil were investigated for the degradation of pyrene and tolerance to copper and zinc. These organisms degraded more than 60% of the supplied pyrene and could also accumulate Cu and Zn (Hong et al. 2010). In co-contaminated soil, Fe(III) coordinating fungal siderophores could play an important role not only by binding metals other than Fe(III), e.g., Cd, Cu, Ni, Pb, Zn, Th (IV), U(IV), and Pu (IV) (Ahmed and Holmström 2014) but also by facilitating the biodegradation of petroleum hydrocarbons by satisfying the Fe requirement for the

Species	Hydrocarbons	Formula	Structure	Removal efficiency (%)	Treatment length (d)	Reference
Penicillium sp.	Decane	$C_{10}H_{22}$	~~~~~	49.0	28	Govarthanan et al. (2017)
Aspergillus sp.	N-hexadecane	$C_{16}H_{34}$	~~~~~~	86.3	10	Al-Hawash et al. (2018)
Fusarium sp.	N-octadecane	C ₁₈ H ₃₈	~~~~~~	89	60	Hidayat and Tachibana (2013)
Phomopsis liquidambari	Phenanthrene	$C_{14}H_{10}$	\bigcirc	77.4	10	Fu et al. (2018)
Irpex lacteus	Anthracene	$C_{14}H_{10}$		60	25	Drevinskas et al. (2016)
Pleurotus ostreatus	Anthracene	$C_{14}H_{10}$		56	23	Drevinskas et al. (2016)
Ganoderma lucidum	Pyrene	C ₁₆ H ₁₀		99.6	30	Agrawal et al. (2018)
Polyporus sp.	Chrysene	$C_{18}H_{12}$		65	30	Hadibarata et al. (2009)

Table 1 Some examples of degradation of petroleum hydrocarbons by different fungal species

degrading microorganisms in Fe-limited habitats. In cocontaminated soil, PAHs may interact with lipophilic components of the fungal cytoplasmic membrane, changing the permeability, which can result in penetration of toxic metals into cells and resulting effects/on cellular functions. Shen et al. (2005) investigated the effects of Cd and phenanthrene (Phe) on the growth of certain soil fungi and showed that growth was strongly inhibited in soil containing Cd and Phe compared with soil containing only Cd. Examples of selected fungal species interacting with PAHs and toxic metals are presented in Table 2.

Factors affecting the efficiency of fungal bioremediation

Soil characteristics

Soil components, pH, and temperature are key factors in fungal bioremediation and metal speciation and affect the transportation and bioavailability of contaminants (Liu et al. 2017; Rangel et al. 2018). Organic matter and clay minerals significantly reduce the solution-phase concentration of metal ions. It was reported that in mineral-dominated soil, 0.01-mg L⁻¹ Cd²⁺ inhibited the dechlorination of trichloroaniline (TCA) while 0.2-mg L⁻¹ Cd²⁺ was necessary in an organicdominated soil, which correlated with the metal-binding capacity of the organic material (Zhang et al. 2016). Clay minerals, e.g., montmorillonite, possess high cation exchange capacities (CECs), and can efficiently reduce metal bioavailability and toxicity (Sandrin and Maier 2003). Moreover, metals in soil may react with the organic pollutants to affect the speciation, bioavailability, and toxicity of the metal and the organic pollutant (Ceci et al. 2019).

The pH is another crucial factor in determining the biodegradation of petroleum hydrocarbons and biotransformation of toxic metals. Changes in pH can alter fungal, and bacterial, community structure and enzyme activities as well as affecting metal speciation. Such effects of pH on the speciation of metal ions can be simulated using geochemical modeling software, e.g., Geochemists' workbench (GWB) (Carrillo-Chávez et al. 2014; Li et al. 2019), MINEQL+ (Cloutier-Hurteau et al. 2007; Kocaoba 2020), and PHREEQC (Ceci et al. 2015; Liang et al. 2016b). For example, the speciation and solubility of Zn²⁺ in a simulated fungal system for metal remediation calculated using GWB showed that smithsonite (ZnCO₃) (pH = 1.7–3.2) and Zn₃(PO₄)₂·4H₂O (pH = 3.2–14) were the main mineral phases over different pH ranges (Fig. 3).

Temperature can influence the bioremediation of cocontaminated soil by affecting the chemistry of pollutants and fungal biodiversity (Rangel et al. 2018). The viscosity of petroleum increases at low temperatures and volatility is reduced which results in retardation of biodegradation. The highest degradation rates for hydrocarbon pollutants generally occur around 30–40 °C in the soil environment (Das and Chandran 2011). At higher temperatures, the solubility of PAHs and toxic metal ions increases which improves their bioavailability, although such high temperatures will also affect microbial community structure and activity. Compared with effects at 20 °C and 40 °C, 30 °C was found to be the optimum temperature for metal removal by *Beauveria bassiana* due to increased biomass production which provided more metal-binding sites (Gola et al. 2016).

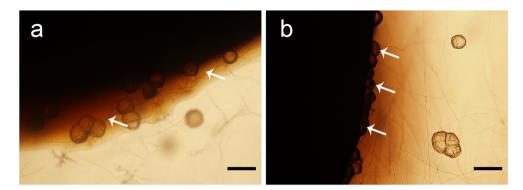
Fungus	PAHs	Initial content Removal	Removal	Toxic	Initial content	Bioremediation Data source Reference	Data source	Reference
			efficiency	metals		efficiency		
Acremonium sp.	Naphthalene, fluorine, phenanthrene,	25 mg L^{-1}	64.9–96.9%	64.9-96.9% Mn, Fe, Zn, 50 mg L ⁻¹	$50 \text{ mg } \mathrm{L}^{-1}$. 1	Lab	Ma et al. (2014)
Pleurotus ostreatus	Pyrene, benzo[a]anthracene, chrysene, benzo[k]fluoranthene, benzo[a]pyrene,	10 ppm	0.5-52.2%	Cd, Hg	10-500 ppm	I	Lab	Baldrian et al. (2000)
Fusarium flocciferum, Trichoderma sp., Trametes versicolor, Pleurotus sp.	dibenzo[a,h]anthracene, benzo[ghi]perylene Benzo[a]anthracene, benzo[a]fluoranthene, benzo[a]pyrene, chrysene, phenanthrene	150– 250 mg kg ⁻¹	21–93%	Cd, Ni	50–500 mg kg ⁻¹	1	Lab	Al-Turki (2009)
Pleurotus ostreatus	Crude oil	Ι	I	Pb, Cu, Mn, Cd, Ni	Pb, Cu, Mn, 0.79–52.07 mg kg ⁻¹ 28.2–75.9% Cd, Ni	28.2–75.9%	Lab	Anacletus et al. (2017)
-, data unavailable								

Importance of metabolites and enzymes

Many transforming interactions between fungi and different pollutants depend on a variety of extracellular excreted substances and metabolites (Gadd 2004; Kirtzel et al. 2020). Fungi are capable of degrading petroleum hydrocarbons by secreting enzymes (e.g., laccases, tyrosinases, manganese peroxidases, cytochrome P450 monooxygenases, reductive dehalogenases), and affecting metal speciation by excretion of a variety of other metabolites (e.g., organic acids, amino acids, siderophores, extracellular proteins, etc.) (Fig. 4).

Petroleum hydrocarbons can be used by several fungal species as a carbon and energy source and assimilated into fungal biomass. Fungal taxa including Amorphoteca, Neosartorya, Talaromyces, Aspergillus, Fusarium, Paecilomyces, Sporobolomyces, Cephalosporium, Penicillium, and Graphium have all been reported to include potential degraders of petroleum hydrocarbons (Chulalaksananukul et al. 2006; Das and Chandran 2011; Varjani 2017). Some species can oxidize pollutants (e.g., phenols and aromatic amines) extracellularly by the production of laccases (Martínková et al. 2016), manganese peroxidases (Zhang et al. 2016), or lignin peroxidases (Falade et al. 2017; Grossart and Rojas-Jimenez 2016). Moreover, fungal cell membranes are permeable to many organic pollutants and these can be degraded by intracellular enzymes, e.g., cytochrome P450 (Ostrem Loss and Yu 2018), reductive dehalogenases (Stella et al. 2017), and nitroreductases (Tripathi et al. 2017; Xu and Zhou 2016), to simpler organic compounds, followed by further metabolism such as β oxidation and entry into the tricarboxylic acid (TCA) cycle (Varjani 2017). Degradation of petroleum hydrocarbons consists of several different enzymatic steps with biodegradability depending on the chemical structure and other factors that affect fungal growth and metabolism, and pollutant chemistry and speciation. In general, biodegradation efficiency can be ranked as linear alkanes > branched alkanes > small aromatics > cyclic alkanes (Das and Chandran 2011; Guermouche M'rassi et al. 2015; Varjani 2017).

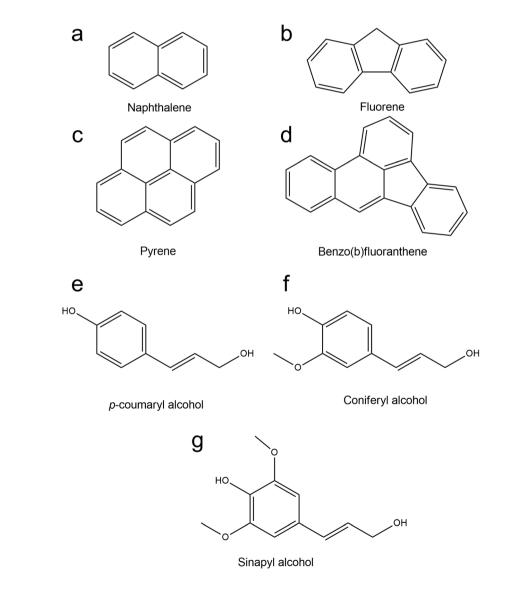
In co-contaminated soil, petroleum hydrocarbons may provide a carbon and energy source for certain fungal species while toxic metals can also exert significant effects on fungal populations and activity. Despite the potential toxicity of many metal species, many fungi can flourish in contaminated conditions although there may be shifts in species composition (Fomina et al. 2005; Gadd 2005; Gadd 2007). The major survival mechanisms involved can be explained as changes in toxic metal mobility resulting from various tolerance and resistance mechanisms (Gadd 2007; Gadd 2010). Fungi possess many mechanisms or properties that influence metal toxicity and mobility, including the production of metal-binding proteins, organic and inorganic precipitation, active transport, and intracellular compartmentalization, while cell walls and associated pigments and **Fig. 1** Fungal biomineralization of carbonates in media amended with heavy oil. *Neurospora crassa* was incubated on solid agar medium, supplemented with 40-mM urea and 50-mM CaCl₂, at 25 °C in the dark for 5 days. Four wells (5-mm diameter) were made in the agar medium using a sterile cork borer and filled with heavy oil prior to fungal inoculation. Scale bars = 200 µm (Li et al., unpublished data)



polysaccharides have significant metal-binding abilities (Gadd 1993; Gadd 2007). The secretion of fungal metabolites (e.g., metal-binding peptides, polysaccharides, amino acids, organic acids) is particularly important for metal and mineral transformations playing roles in both mobilization and immobilization

of metal species (Gadd 2007; Gadd et al. 2014). Moreover, fungal phenolic polymers and melanin possess many potential metal-binding sites with oxygen-containing groups, such as carboxyl, phenolic and alcoholic hydroxyl, carbonyl, and methoxyl groups (Fomina and Gadd 2014). Fungal surface

Fig. 2 Chemical structure of some commonly studied PAHs and three constitutive monomers of lignin. PAHs are aromatic hydrocarbons with two or more fused benzene rings. **a** Naphthalene. **b** Fluorene. **c** Pyrene. **d** Benzo(b)fluoranthene. Lignin results from the enzymatic copolymerization of three phenolic monomers (monolignols): **e** *p*-coumaryl alcohol, **f** coniferyl alcohol, and **g** sinapyl alcohol



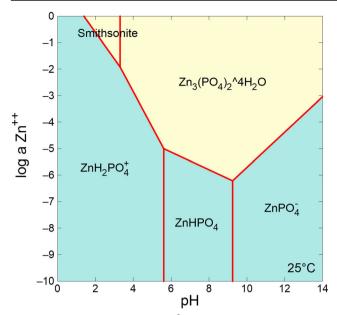


Fig. 3 Geochemical simulation of Zn^{2+} versus pH in a simulated fungal system at 25 °C. The chemical parameters were set at 0.33-M CO₃²⁻, 6.1- mM Cl⁻, 0.83-mM SO₄²⁻, 0.66-M NH₄⁺, 4-mM K⁺, 0.8-mM Mg²⁺, 1.7- mM Na⁺, 0.2-mM Ca²⁺, 0.02-mM Mn²⁺, and 9-µm Fe³⁺. The letter *a* on the *y*-axis represents the effective concentration of a given chemical species in the mixture (Li et al., unpublished data)

complex formation may be related to the coordination of metal ions with oxygen donor atoms and proton release (Gadd 2009):

$$S-OH + Cu^{2+} \rightleftharpoons S - OCu^{+} + H^{-}$$

Bidentate surface complexation may also result:

$$-S-OH + Cu^{2+} \implies -S-O \\ -S-OH + Cu^{2+} \implies -S-O \\ -S-O + 2H^{+}$$

Metal immobilization appears particularly relevant to bioremediation approaches and fungi are capable of mediating precipitation of metals as insoluble oxalates, oxides, carbonates, and phosphates (Fomina et al. 2008; Gadd et al. 2014; Liang and Gadd 2017; Suyamud et al. 2020). For example, the liberation of phosphate from organic or inorganic phosphate hydrolysis proved to be an efficient method for metal immobilization, including Zn, Pb, La, and U, which were precipitated on and around hyphal surfaces (Ezawa and Saito 2017; Fomina et al. 2008; Liang et al. 2016a; Liang and Gadd 2017; Liang et al. 2015; Suyamud et al. 2020). Urease-positive fungi (e.g., *Neurospora crassa*, *Pestalotiopsis* sp., and *Myrothecium gramineum*) are promising candidates for the immobilization of toxic metals because the mechanism involved is associated with urea degradation. Such fungi grown

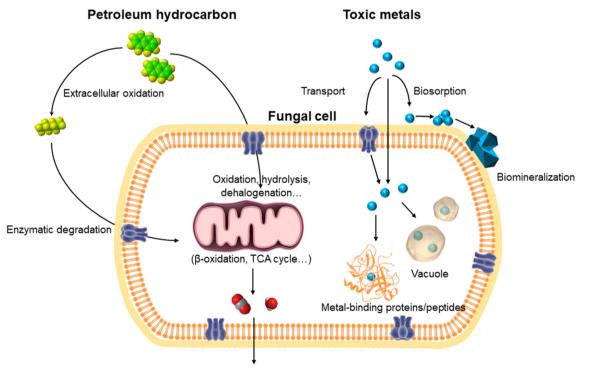


Fig. 4 Simplified diagram of fungal interactions with petroleum hydrocarbons and toxic metals. Fungal cell membranes are permeable to petroleum hydrocarbons or simpler organic compounds oxidized by extracellular enzymes, which can undergo further metabolism including hydrolysis, dehalogenation, β -oxidation, and entry into the tricarboxylic acid cycle. Toxic metals can accumulate on fungal cell surfaces through

biosorption, which can result in nucleation and subsequent precipitation of biominerals. Some metals can be intracellularly accumulated by active transport or diffusion through the cell membrane, and localized within vacuolar or other organellar compartments and/or be sequestered by sulfide, metal-binding proteins/peptides, and other macromolecules in urea-containing medium hydrolyze urea producing ammonia and free carbonate which results in the precipitation of metals as carbonates, e.g., BaCO₃, CdCO₃, CoCO₃, Cu₂(OH)₂CO₃, La₂(CO₃)₃, and NiCO₃ (Li and Gadd 2017b; Li et al. 2019; Liang and Gadd 2017; Liu et al. 2019; Rautaray et al. 2004). Fungi can produce a variety of metal oxalates on interacting with metals and metal-bearing minerals including those of Ca, Cd, Co, Cu, Mg, Mn, Sr, Zn, Ni, and Pb (Gadd et al. 2014). Extracellular proteins, amino acids, and polysaccharides also play an important role in toxic metal immobilization. Extracellular nickel precipitation was associated with the removal of extracellular protein (Li et al. 2019), and it has been demonstrated that extracellular protein may act as a template for mineral formation, influencing the size of the resultant biominerals (Li and Gadd 2017a; Li et al. 2019; Liu et al. 2019).

Future perspectives and conclusions

At present, bioremediation of soil co-contaminated with petroleum hydrocarbons and toxic metals is mostly focused on the removal, detoxification, or degradation efficiency of single or composite pollutants of each type. Little research has been carried out on the metabolism of fungi in response to complex pollutant stress. Fungal responses to petroleum hydrocarbons can be reflected by differences in metabolic responses, enzyme induction and synthesis, and extracellular metabolite production, which will also affect the migration and transformation of toxic metals. This is clearly a complex problem, affected by many variables, and sometimes limited by the availability of appropriate analytical techniques. For example, understanding the spatial distribution of toxic metals in the soil and/or the migration and transformation processes mediated by different fungal species depends on the sampling and analysis methods used. Sampling technology can be the main bottleneck that limits understanding due to heterogeneity of the soil in vertical and horizontal dimensions. In fact, toxic metals may show different gradient distributions over very small interfaces. Conventional techniques are also based on sampling and subsequent transport to the laboratory for analysis, but there may be subsequent changes during collection and storage due to, e.g., contamination and changes in environmental conditions (e.g., metabolic activity, pH, dissolved oxygen, Eh), which conceal the dynamic changes in biodiversity or chemical speciation that may occur in contaminated soil. To overcome these difficulties, new technology, including microbial metabolomics (Dombrowski et al. 2016; Tian et al. 2018) and proteomics, needs to be applied to the study of fungal bioremediation in co-contaminated soil, which could inform about metabolic responses under multiple pollutant conditions. This could provide scientific explanations for fungal responses to multiple contaminants at the molecular level (Aydin et al. 2017; Wang et al. 2017). Further, to obtain highresolution spatial distribution characteristics of toxic metals and reveal interface reaction processes of soil-metals-fungal interactions, new methods such as gradient diffusion film technology (DGT) combined with laser ablation-inductively coupled plasma mass spectrometry could be applied to analyze dynamic changes in toxic metal speciation at soilmicrobe interfaces (Challis et al. 2018; Feng et al. 2016; Koppel et al. 2020; Wang et al. 2018). Such approaches will further clarify fungal transformation mechanisms in soil contaminated with petroleum hydrocarbons and toxic metals and may contribute to more effective strategies for fungal bioremediation.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

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References

- Adeniyi AA, Afolabi JA (2002) Determination of total petroleum hydrocarbons and heavy metals in soils within the vicinity of facilities handling refined petroleum products in Lagos metropolis. Environ Int 28(1):79–82. https://doi.org/10.1016/S0160-4120(02)00007-7
- Agrawal N, Verma P, Shahi SK (2018) Degradation of polycyclic aromatic hydrocarbons (phenanthrene and pyrene) by the ligninolytic fungi *Ganoderma lucidum* isolated from the hardwood stump. Bioresources Bioprocess 5(1):11–20. https://doi.org/10.1186/ s40643-018-0197-5
- Ahmed E, Holmström SJM (2014) Siderophores in environmental research: roles and applications. Microb Biotechnol 7(3):196–208. https://doi.org/10.1111/1751-7915.12117
- Al-Hawash AB, Zhang J, Li S, Liu J, Ghalib HB, Zhang X, Ma F (2018) Biodegradation of n-hexadecane by Aspergillus sp. RFC-1 and its mechanism. Ecotoxicol Environ Saf 164:398–408. https://doi.org/ 10.1016/j.ecoenv.2018.08.049

- Al-Turki AI (2009) Microbial polycyclic aromatic hydrocarbons degradation in soil. R J Environ Toxicol 3(1):1–8. https://doi.org/10.3923/ rjet.2009.1.8
- Anacletus F, Nwauche K, Ighorodje-Monago C (2017) Mineral and heavy metal composition of crude oil polluted soil amended with non-ionic surfactant (Triton X-100) and white rot fungus (*Pleurotus* ostratus). J Environ Anal Toxicol 7:449–451. https://doi.org/10. 4172/2161-0525.1000449
- Aydin S, Karaçay HA, Shahi A, Gökçe S, Ince B, Ince O (2017) Aerobic and anaerobic fungal metabolism and omics insights for increasing polycyclic aromatic hydrocarbons biodegradation. Fungal Biol Rev 31(2):61–72. https://doi.org/10.1016/j.fbr.2016.12.001
- Baldrian P, in der Wiesche C, Gabriel J, Nerud F, Zadražil F (2000) Influence of cadmium and mercury on activities of ligninolytic enzymes and degradation of polycyclic aromatic hydrocarbons by *Pleurotus ostreatus* in soil. Appl Environ Microbiol 66(6):2471– 2478. https://doi.org/10.1128/aem.66.6.2471-2478.2000
- Banitz T, Johst K, Wick LY, Schamfuß S, Harms H, Frank K (2013) Highways versus pipelines: contributions of two fungal transport mechanisms to efficient bioremediation. Env Microbiol Rep 5(2): 211–218. https://doi.org/10.1111/1758-2229.12002
- Biswas B, Sarkar B, Mandal A, Naidu R (2015) Heavy metalimmobilizing organoclay facilitates polycyclic aromatic hydrocarbon biodegradation in mixed-contaminated soil. J Hazard Mater 298:129–137. https://doi.org/10.1016/j.jhazmat.2015.05.009
- Boswell GP, Jacobs H, Davidson FA, Gadd GM, Ritz K (2002) Functional consequences of nutrient translocation in mycelial fungi. J Theor Biol 217:459–477. https://doi.org/10.1006/jtbi.2002.3048
- Boswell GP, Jacobs H, Davidson FA, Gadd GM, Ritz K (2003) Growth and function of fungal mycelia in heterogeneous environments. Bull Math Biol 65(3):447–477. https://doi.org/10.1016/S0092-8240(03) 00003-X
- Carrillo-Chávez A, Salas-Megchún E, Levresse G, Muñoz-Torres C, Pérez-Arvizu O, Gerke T (2014) Geochemistry and mineralogy of mine-waste material from a "skarm-type" deposit in Central Mexico: modeling geochemical controls of metals in the surface environment. J Geochem Explor 144:28–36. https://doi.org/10.1016/j. gexplo.2014.03.017
- Ceci A, Rhee YJ, Kierans M, Hillier S, Pendlowski H, Gray N, Persiani AM, Gadd GM (2015) Transformation of vanadinite [Pb₅(VO₄)₃Cl] by fungi. Environ Microbiol 17(6):2018–2034. https://doi.org/10. 1111/1462-2920.12612
- Ceci A, Pinzari F, Russo F, Persiani AM, Gadd GM (2019) Roles of saprotrophic fungi in biodegradation or transformation of organic and inorganic pollutants in co-contaminated sites. Appl Microbiol Biotechnol 103(1):53–68. https://doi.org/10.1007/s00253-018-9451-1
- Challis JK, Stroski KM, Luong KH, Hanson ML, Wong CS (2018) Field evaluation and in situ stress testing of the organic-diffusive gradients in thin-films passive sampler. Environ Sci Technol 52(21):12573– 12582. https://doi.org/10.1021/acs.est.8b03622
- Cheng J, Sun Z, Yu Y, Li X, Li T (2019) Effects of modified carbon black nanoparticles on plant-microbe remediation of petroleum and heavy metal co-contaminated soils. Int J Phytoremediat 21(7):634–642. https://doi.org/10.1080/15226514.2018.1556581
- Chulalaksananukul S, Gadd GM, Sangvanich P, Sihanonth P, Piapukiew J, Vangnai AS (2006) Biodegradation of benzo(a)pyrene by a newly isolated *Fusarium* sp. FEMS Microbiol Lett 262(1):99–106. https://doi.org/10.1111/j.1574-6968.2006.00375.x
- Cloutier-Hurteau B, Sauvé S, Courchesne F (2007) Comparing WHAM 6 and MINEQL⁺ 4.5 for the chemical speciation of Cu²⁺ in the *Rhizosphere* of forest soils. Environ Sci Technol 41(23):8104– 8110. https://doi.org/10.1021/es0708464
- Czarny J, Staninska-Pięta J, Piotrowska-Cyplik A, Juzwa W, Wolniewicz A, Marecik R (2020) *Acinetobacter* sp. as the key player in diesel oil

degrading community exposed to PAHs and heavy metals. J Hazard Mater 383:121168. https://doi.org/10.1016/j.jhazmat.2019.121168

- Das N, Chandran P (2011) Microbial degradation of petroleum hydrocarbon contaminants: an overview. Biotechnol Res Int 2011:1–13. https://doi.org/10.4061/2011/941810
- Dombrowski N, Donaho JA, Gutierrez T, Seitz KW, Teske AP, Baker BJ (2016) Reconstructing metabolic pathways of hydrocarbondegrading bacteria from the Deepwater Horizon oil spill. Nat Microbiol 1(7):16057. https://doi.org/10.1038/nmicrobiol.2016.57
- Dong Z-Y, Huang W-H, Xing D-F, Zhang H-F (2013) Remediation of soil co-contaminated with petroleum and heavy metals by the integration of electrokinetics and biostimulation. J Hazard Mater 260: 399–408. https://doi.org/10.1016/j.jhazmat.2013.05.003
- dos Santos JJ, Maranho LT (2018) Rhizospheric microorganisms as a solution for the recovery of soils contaminated by petroleum: a review. J Environ Manag 210:104–113. https://doi.org/10.1016/j. jenvman.2018.01.015
- Drevinskas T, Mickienė R, Maruška A, Stankevičius M, Tiso N, Mikašauskaitė J, Ragažinskienė O, Levišauskas D, Bartkuvienė V, Snieškienė V, Stankevičienė A, Polcaro C, Galli E, Donati E, Tekorius T, Kornyšova O, Kaškonienė V (2016) Downscaling the in vitro test of fungal bioremediation of polycyclic aromatic hydrocarbons: methodological approach. Anal Bioanal Chem 408(4): 1043–1053. https://doi.org/10.1007/s00216-015-9191-3
- El-Azeem SAA, Ahmad M, Usman AR, Kim K-R, Oh S-E, Lee SS, Ok YS (2013) Changes of biochemical properties and heavy metal bioavailability in soil treated with natural liming materials. Environ Earth Sc 70(7):3411–3420. https://doi.org/10.1007/s12665-013-2410-3
- Ezawa T, Saito K (2017) How do arbuscular mycorrhizal fungi handle phosphate? New insight into fine-tuning of phosphate metabolism. New Phytol 220(4):1116–1120. https://doi.org/10.1111/nph.15187
- Falade AO, Nwodo UU, Iweriebor BC, Green E, Mabinya LV, Okoh AI (2017) Lignin peroxidase functionalities and prospective applications. Microbiol Open 6(1):e00394. https://doi.org/10.1002/mbo3. 394
- Feng Z, Zhu P, Fan H, Piao S, Xu L, Sun T (2016) Effect of biofilm on passive sampling of dissolved orthophosphate using the diffusive gradients in thin films technique. Anal Chem 88(13):6836–6843. https://doi.org/10.1021/acs.analchem.6b01392
- Fomina M, Gadd GM (2014) Biosorption: current perspectives on concept, definition and application. Bioresour Technol 160:3–14. https://doi.org/10.1016/j.biortech.2013.12.102
- Fomina M, Gadd GM, Burford E (2005) Toxic metals and fungal communities. In: Dighton J, White JF, Oudemans P (eds) The fungal community its organization and role in the ecosystem. CRC Press, Boca Raton, pp 733–758
- Fomina M, Charnock JM, Hillier S, Alvarez R, Livens F, Gadd GM (2008) Role of fungi in the biogeochemical fate of depleted uranium. Curr Biol 18(9):375–377. https://doi.org/10.1016/j.cub.2008.03.011
- Freije AM (2015) Heavy metal, trace element and petroleum hydrocarbon pollution in the Arabian Gulf: review. J Associa Arab Uni Basic Appl Sci 17(1):90–100. https://doi.org/10.1016/j.jaubas.2014.02. 001
- Fu W, Xu M, Sun K, Hu L, Cao W, Dai C, Jia Y (2018) Biodegradation of phenanthrene by endophytic fungus *Phomopsis liquidambari* in vitro and in vivo. Chemosphere 203:160–169. https://doi.org/ 10.1016/j.chemosphere.2018.03.164
- Furuno S, Foss S, Wild E, Jones KC, Semple KT, Harms H, Wick LY (2012) Mycelia promote active transport and spatial dispersion of polycyclic aromatic hydrocarbons. Environ Sci Technol 46(10): 5463–5470. https://doi.org/10.1021/es300810b
- Gadd GM (1993) Interactions of fungi with toxic metals. New Phytol 124:25–60
- Gadd GM (ed) (2001) Fungi in Bioremediation. Cambridge University Press, Cambridge

- Gadd GM (2004) Mycotransformation of organic and inorganic substrates. Mycologist 18(2):60–70. https://doi.org/10.1017/ S0269915X04002022
- Gadd GM (2005) Microorganisms in toxic metal polluted soils. In: Buscot F, Varma A (eds) Microorganisms in soils: roles in genesis and functions. Springer-Verlag, Berlin, pp 325–356
- Gadd GM (2007) Geomycology: biogeochemical transformations of rocks, minerals, metals and radionuclides by fungi, bioweathering and bioremediation. Mycol Res 111(1):3–49. https://doi.org/10. 1016/j.mycres.2006.12.001
- Gadd GM (2008) Fungi and their role in the biosphere. In: Encyclopedia of ecology, Ed: S.E. Jorgensen and B. Fath. Elsevier, Amsterdam, pp 1709–1717
- Gadd GM (2009) Biosorption: critical review of scientific rationale, environmental importance and significance for pollution treatment. J Chem Technol Biotechnol 84(1):13–28. https://doi.org/10.1002/ jctb.1999
- Gadd GM (2010) Metals, minerals and microbes: geomicrobiology and bioremediation. Microbiol 156(3):609–643. https://doi.org/10.1099/ mic.0.037143-0
- Gadd GM (2017) Geomicrobiology of the built environment. Nat Microbiol 2(4):1–9. https://doi.org/10.1038/nmicrobiol.2016.275
- Gadd GM, Bahri-Esfahani J, Li Q, Rhee YJ, Wei Z, Fomina M, Liang X (2014) Oxalate production by fungi: significance in geomycology, biodeterioration and bioremediation. Fungal Biol Rev 28(2–3):36– 55. https://doi.org/10.1016/j.fbr.2014.05.001
- Glasauer S, Beveridge TJ, Burford EP, Harper FA, Gadd GM (2004) Metals and metalloids, transformations by microorganisms. In: Hillel D, Rosenzweig C, Powlson DS, Scow KM, Singer MJ, Sparks DL, Hatfield J (eds) Encyclopedia of soils in the environment. Elsevier, Amsterdam, pp 438–447
- Gola D, Dey P, Bhattacharya A, Mishra A, Malik A, Namburath M, Ahammad SZ (2016) Multiple heavy metal removal using an entomopathogenic fungi *Beauveria bassiana*. Bioresour Technol 218: 388–396. https://doi.org/10.1016/j.biortech.2016.06.096
- Govarthanan M, Fuzisawa S, Hosogai T, Chang Y-C (2017) Biodegradation of aliphatic and aromatic hydrocarbons using the filamentous fungus *Penicillium* sp. CHY-2 and characterization of its manganese peroxidase activity. RSC Adv 7(34):20716–20723. https://doi.org/10.1039/C6RA28687A
- Grossart H-P, Rojas-Jimenez K (2016) Aquatic fungi: targeting the forgotten in microbial ecology. Curr Opin Microbiol 31:140–145. https://doi.org/10.1016/j.mib.2016.03.016
- Guermouche M'rassi A, Bensalah F, Gury J, Duran R (2015) Isolation and characterization of different bacterial strains for bioremediation of n-alkanes and polycyclic aromatic hydrocarbons. Environ Sci Pollut Res 22(20):15332–15346. https://doi.org/10.1007/s11356-015-4343-8
- Hadibarata T, Tachibana S, Itoh K (2009) Biodegradation of chrysene, an aromatic hydrocarbon, by *Polyporus* sp. S133 in liquid medium. J Hazard Mater 164(2):911–917. https://doi.org/10.1016/j.jhazmat. 2008.08.081
- Haritash AK, Kaushik CP (2009) Biodegradation aspects of polycyclic aromatic hydrocarbons (PAHs): a review. J Hazard Mater 169(1):1– 15. https://doi.org/10.1016/j.jhazmat.2009.03.137
- Harms H, Schlosser D, Wick LY (2011) Untapped potential: exploiting fungi in bioremediation of hazardous chemicals. Nat Rev Microbiol 9(3):177–192. https://doi.org/10.1038/nrmicro2519
- Hidayat A, Tachibana S (2013) Crude oil and n-octadecane degradation under saline conditions by *Fusarium* sp., F092. J Environ Sci Technol 6(1):29–40
- Hofrichter M, Bublitz F, Fritsche W (1994) Unspecific degradation of halogenated phenols by the soil fungus *Penicillium frequentans* Bi 7/2. J Basic Microbiol 34(3):163–172. https://doi.org/10.1002/ jobm.3620340306

- Hong JW, Park JY, Gadd GM (2010) Pyrene degradation and copper and zinc uptake by *Fusarium solani* and *Hypocrea lixii* isolated from petrol station soil. J Appl Microbiol 108(6):2030–2040. https://doi. org/10.1111/j.1365-2672.2009.04613.x
- Jacobs H, Boswell GP, Ritz K, Davidson FA, Gadd GM (2002) Solubilization of calcium phosphate as a consequence of carbon translocation by *Rhizoctonia solani*. FEMS Microbiol Ecol 40(1): 65–71. https://doi.org/10.1111/j.1574-6941.2002.tb00937.x
- Khan MAI, Biswas B, Smith E, Naidu R, Megharaj M (2018) Toxicity assessment of fresh and weathered petroleum hydrocarbons in contaminated soil - a review. Chemosphere 212:755–767. https://doi. org/10.1016/j.chemosphere.2018.08.094
- Kirtzel J, Ueberschaar N, Deckert-Gaudig T, Krause K, Deckert V, Gadd GM, Kothe E (2020) Organic acids, siderophores, enzymes and mechanical pressure for black slate bioweathering with the basidiomycete *Schizophyllum commune*. Environ Microbiol 22(4):1535– 1546. https://doi.org/10.1111/1462-2920.14749
- Klimek B, Sitarz A, Choczyński M, Niklińska M (2016) The effects of heavy metals and total petroleum hydrocarbons on soil bacterial activity and functional diversity in the Upper Silesia industrial region (Poland). Water Air Soil Pollut 227(8):265–274. https://doi. org/10.1007/s11270-016-2966-0
- Kocaoba S (2020) Adsorption of Fe(II) and Fe(III) from aqueous solution by using sepiolite: speciation studies with MINEQL+ computer program. Sep Sci Technol 55(5):896–906. https://doi.org/10.1080/ 01496395.2019.1579841
- Kohlmeier S, Smits TH, Ford RM, Keel C, Harms H, Wick LY (2005) Taking the fungal highway: mobilization of pollutant-degrading bacteria by fungi. Environ Sci Technol 39(12):4640–4646. https:// doi.org/10.1021/es047979z
- Koppel DJ, King CK, Brown KE, Price GAV, Adams MS, Jolley DF (2020) Assessing the risk of metals and their mixtures in the Antarctic nearshore marine environment with diffusive gradients in thin-films. Environ Sci Technol 54(1):306–315. https://doi.org/ 10.1021/acs.est.9b04497
- Kuyukina M, Krivoruchko A, Ivshina I (2018) Hydrocarbon- and metalpolluted soil bioremediation: progress and challenges. Microbiology Australia 39(3):133–136. https://doi.org/10.1071/MA18041
- Lai C-C, Huang Y-C, Wei Y-H, Chang J-S (2009) Biosurfactantenhanced removal of total petroleum hydrocarbons from contaminated soil. J Hazard Mater 167(1):609–614. https://doi.org/10.1016/ j.jhazmat.2009.01.017
- Li Q, Gadd GM (2017a) Biosynthesis of copper carbonate nanoparticles by ureolytic fungi. Appl Microbiol Biotechnol 101(19):7397–7407. https://doi.org/10.1007/s00253-017-8451-x
- Li Q, Gadd GM (2017b) Fungal nanoscale metal carbonates and production of electrochemical materials. Microb Biotechnol 10(5):1131– 1136. https://doi.org/10.1111/1751-7915.12765
- Li QW, Csetenyi L, Gadd GM (2014) Biomineralization of metal carbonates by *Neurospora crassa*. Environ Sci Technol 48(24):14409– 14416. https://doi.org/10.1021/es5042546
- Li Q, Csetenyi L, Paton GI, Gadd GM (2015) CaCO₃ and SrCO₃ bioprecipitation by fungi isolated from calcareous soil. Environ Microbiol 17(8):3082–3097. https://doi.org/10.1111/1462-2920. 12954
- Li Q, Liu D, Jia Z, Csetenyi L, Gadd GM (2016) Fungal biomineralization of manganese as a novel source of electrochemical materials. Curr Biol 26(7):950–955. https://doi.org/10.1016/j.cub.2016.01.068
- Li Q, Liu D, Chen C, Shao Z, Wang H, Liu J, Zhang Q, Gadd GM (2019) Experimental and geochemical simulation of nickel carbonate mineral precipitation by carbonate-laden ureolytic fungal culture supernatants. Environ Sci Nano 6(6):1866–1875. https://doi.org/10.1039/ C9EN00385A
- Liang X, Gadd GM (2017) Metal and metalloid biorecovery using fungi. Microb Biotechnol 10(5):1199–1205. https://doi.org/10.1111/1751-7915.12767

- Liang X, Hillier S, Pendlowski H, Gray N, Ceci A, Gadd GM (2015) Uranium phosphate biomineralization by fungi. Environ Microbiol 17(6):2064–2075. https://doi.org/10.1111/1462-2920.12771
- Liang X, Csetenyi L, Gadd GM (2016a) Uranium bioprecipitation mediated by yeasts utilizing organic phosphorus substrates. Appl Microbiol Biotechnol 100(11):5141–5151. https://doi.org/10.1007/ s00253-016-7327-9
- Liang X, Kierans M, Ceci A, Hillier S, Gadd GM (2016b) Phosphatasemediated bioprecipitation of lead by soil fungi. Environ Microbiol 18(1):219–231. https://doi.org/10.1111/1462-2920.13003
- Liu S-H, Zeng G-M, Niu Q-Y, Liu Y, Zhou L, Jiang L-H, Tan X-f XP, Zhang C, Cheng M (2017) Bioremediation mechanisms of combined pollution of PAHs and heavy metals by bacteria and fungi: a mini review. Bioresour Technol 224:25–33. https://doi.org/10.1016/ j.biortech.2016.11.095
- Liu F, Csetenyi L, Gadd GM (2019) Amino acid secretion influences the size and composition of copper carbonate nanoparticles synthesized by ureolytic fungi. Appl Microbiol Biotechnol 103(1):7217–7230. https://doi.org/10.1007/s00253-019-09961-2
- Ma X-k, Wu LL, Fam H (2014) Heavy metal ions affecting the removal of polycyclic aromatic hydrocarbons by fungi with heavy-metal resistance. Appl Microbiol Biotechnol 98(23):9817–9827. https:// doi.org/10.1007/s00253-014-5905-2
- Martínková L, Kotik M, Marková E, Homolka L (2016) Biodegradation of phenolic compounds by Basidiomycota and its phenol oxidases: a review. Chemosphere 149:373–382. https://doi.org/10.1016/j. chemosphere.2016.01.022
- Muniz P, Danulat E, Yannicelli B, Garcia-Alonso J, Medina G, Bicego MC (2004) Assessment of contamination by heavy metals and petroleum hydrocarbons in sediments of Montevideo Harbour (Uruguay). Environ Int 29(8):1019–1028. https://doi.org/10.1016/ S0160-4120(03)00096-5
- Ostrem Loss EM, Yu J-H (2018) Bioremediation and microbial metabolism of benzo(a)pyrene. Mol Microbiol 109(4):433–444. https://doi. org/10.1111/mmi.14062
- Pinedo-Rivilla C, Aleu J, Collado I (2009) Pollutants biodegradation by fungi. Curr Org Chem 13(12):1194–1214. https://doi.org/10.2174/ 138527209788921774
- Rangel DE, Finlay RD, Hallsworth JE, Dadachova E, Gadd GM (2018) Fungal strategies for dealing with environment-and agriculture-induced stresses. Fungal Biol 122(6):602–612. https://doi.org/10. 1016/j.funbio.2018.02.002
- Rautaray D, Ahmad A, Sastry M (2004) Biological synthesis of metal carbonate minerals using fungi and actinomycetes. J Mater Chem 14(14):2333–2340. https://doi.org/10.1039/B401431F
- Sandrin TR, Maier RM (2003) Impact of metals on the biodegradation of organic pollutants. Environ Health Perspect 111(8):1093–1101. https://doi.org/10.1289/ehp.5840
- Shen G, Cao L, Lu Y, Hong J (2005) Influence of phenanthrene on cadmium toxicity to soil enzymes and microbial growth (5 pp). Environ Sci Pollut Res 12(5):259–263. https://doi.org/10.1065/ espr2005.06.266
- Song B, Zeng G, Gong J, Liang J, Xu P, Liu Z, Zhang Y, Zhang C, Cheng M, Liu Y, Ye S, Yi H, Ren X (2017) Evaluation methods for assessing effectiveness of in situ remediation of soil and sediment contaminated with organic pollutants and heavy metals. Environ Int 105:43–55. https://doi.org/10.1016/j.envint.2017.05.001
- Stajich JE, Wilke SK, Ahrén D, Au CH, Birren BW, Borodovsky M, Burns C, Canbäck B, Casselton LA, Cheng CK, Deng J, Dietrich FS, Fargo DC, Farman ML, Gathman AC, Goldberg J, Guigó R, Hoegger PJ, Hooker JB, Huggins A, James TY, Kamada T, Kilaru S, Kodira C, Kües U, Kupfer D, Kwan HS, Lomsadze A, Li W, Lilly

WW, Ma L-J, Mackey AJ, Manning G, Martin F, Muraguchi H, Natvig DO, Palmerini H, Ramesh MA, Rehmeyer CJ, Roe BA, Shenoy N, Stanke M, Ter-Hovhannisyan V, Tunlid A, Velagapudi R, Vision TJ, Zeng Q, Zolan ME, Pukkila PJ (2010) Insights into evolution of multicellular fungi from the assembled chromosomes of the mushroom *Coprinopsis cinerea (Coprinus cinereus)*. Proc Natl Acad Sci U S A 107(26):11889–11894. https://doi.org/10. 1073/pnas.1003391107

- Stella T, Covino S, Čvančarová M, Filipová A, Petruccioli M, D'Annibale A, Cajthaml T (2017) Bioremediation of long-term PCB-contaminated soil by white-rot fungi. J Hazard Mater 324: 701–710. https://doi.org/10.1016/j.jhazmat.2016.11.044
- Sullivan TS, Gadd GM (2019) Metal bioavailability and the soil microbiome. Adv Agronomy 155:79–120
- Suyamud B, Ferrier J, Csetenyi L, Inthorn D, Gadd GM (2020) Biotransformation of struvite by *Aspergillus niger*: phosphate release and magnesium biomineralization as glushinskite. Environ Microbiol 22(4):1588–1602. https://doi.org/10.1111/1462-2920. 14949
- Tian Z, Vila J, Yu M, Bodnar W, Aitken MD (2018) Tracing the biotransformation of polycyclic aromatic hydrocarbons in contaminated soil using stable isotope-assisted metabolomics. Environ Sci Technol Lett 5(2):103–109. https://doi.org/10.1021/acs.estlett. 7b00554
- Tripathi V, Edrisi SA, Chen B, Gupta VK, Vilu R, Gathergood N, Abhilash PC (2017) Biotechnological advances for restoring degraded land for sustainable development. Trends Biotechnol 35(9): 847–859. https://doi.org/10.1016/j.tibtech.2017.05.001
- Vanholme R, Demedts B, Morreel K, Ralph J, Boerjan W (2010) Lignin biosynthesis and structure. Plant Physiol 153(3):895–905. https:// doi.org/10.1104/pp.110.155119
- Varjani SJ (2017) Microbial degradation of petroleum hydrocarbons. Bioresour Technol 223:277–286. https://doi.org/10.1016/j. biortech.2016.10.037
- Wang S, Wang Z, Zhou L, Shi X, Xu G (2017) Comprehensive analysis of short-, medium-, and long-chain acyl-coenzyme a by online twodimensional liquid chromatography/mass spectrometry. Anal Chem 89(23):12902–12908. https://doi.org/10.1021/acs.analchem. 7b03659
- Wang S, Wu Z, Luo J (2018) Transfer mechanism, uptake kinetic process, and bioavailability of P, Cu, Cd, Pb, and Zn in macrophyte rhizosphere using diffusive gradients in thin films. Environ Sci Technol 52(3):1096–1108. https://doi.org/10.1021/acs.est.7b01578
- Wick LY, Remer R, Würz B, Reichenbach J, Braun S, Schäfer F, Harms H (2007) Effect of fungal hyphae on the access of bacteria to phenanthrene in soil. Environ Sci Technol 41(2):500–505. https://doi. org/10.1021/es061407s
- Worrich A, Wick LY, Banitz T (2018) Ecology of contaminant biotransformation in the mycosphere: role of transport processes. Adv Appl Microbiol 104:93–133
- Xu Y, Zhou N-Y (2016) Microbial remediation of aromaticscontaminated soil. Front Env Sci Eng 11(2):1–9. https://doi.org/10. 1007/s11783-017-0894-x
- Zhang H, Zhang S, He F, Qin X, Zhang X, Yang Y (2016) Characterization of a manganese peroxidase from white-rot fungus *Trametes* sp.48424 with strong ability of degrading different types of dyes and polycyclic aromatic hydrocarbons. J Hazard Mater 320: 265–277. https://doi.org/10.1016/j.jhazmat.2016.07.065

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