Stable isotope signatures reveal small-scale spatial separation in populations of European sea
 bass

3

- 4 Giulia Cambiè¹, Michel J. Kaiser¹, Andrew L. Marriott², Jennifer Fox¹, Gwladys Lambert¹,
- 5 Jan G. Hiddink¹, Thomas Overy¹, Sarah A. Bennet³, Melanie J. Leng⁴ and Ian D. McCarthy¹
- 6
- ⁷ ¹ School of Ocean Sciences, Bangor University, Askew Street, Menai Bridge LL59 5AB, UK
- ² Inorganic Geochemistry Facility, Centre for Environmental Geochemistry, British
 Geological Survey, Keyworth, Nottingham NG12 5GG, UK
- ³School of Life Sciences, Gibbet Hill Campus, The University of Warwick, Coventry, CV4
 7AL, UK
- ⁴ NERC Isotope Geosciences Laboratory, British Geological Survey, Keyworth, Nottingham
 NG12 5GG, UK
- 14
- 15 Running head: Isotopic signature in sea bass populations
- 16

17 ABSTRACT

Scientific information about European sea bass (*Dicentrarchus labrax*) stocks in NE Atlantic 18 is limited and a more accurate definition of the stock boundaries in the area is required to 19 improve assessment and management advice. Here we study the connectivity and movement 20 patterns of European sea bass in Wales (UK) using the stable isotope (δ^{13} C and δ^{15} N) 21 composition of their scales. Analysis of fish scale δ^{13} C and δ^{15} N values in the last growing 22 season was performed on 189 adult sea bass caught at nine coastal feeding grounds. Fish >50 23 cm total length (TL) caught in estuaries had very low δ^{13} C and this is characteristic of fresh 24 water (organic/soil) input, indicating the primary use of estuaries as feeding areas. A random 25 forest classification model was used to test if there was a difference in $\delta^{15}N$ and $\delta^{13}C$ values 26 between north, mid and south Wales and whether it was possible to correctly assign the fish 27 to the area where it was caught. This analysis was restricted to fish of a similar size range 28 (40-50 cm TL) caught in open coastal areas (n=156). The random forest classification model 29 showed that about 75% of the fish could be correctly assigned to their collection region based 30 31 on their isotope composition. The majority of the misclassifications of fish were fish from north Wales classifying to mid Wales and vice versa, while the majority of fish from south 32 Wales were correctly assigned (80%). Our findings suggest that two sub-populations of sea 33

bass in Welsh waters use separate feeding grounds (south vs. mid/north Wales), and may
need separate management.

36

KEY WORDS: European sea bass, Stable isotopes, Random forest, Feeding ground, Stockboundaries.

- 39
- 40

INTRODUCTION

Most current fisheries controls are applied at large geographic scales that often encompass entire sea basins. Effort controls such as Minimum Landing Size (MLS) are only effective if the life history and growth of fish is uniform across the scale at which they are applied. However, for many species of fish, we have limited understanding of their home range and hence the interaction between local environmental parameters and life history traits such as growth rate potentially leading to a mismatch with the scale of management.

The European sea bass (Dicentrarchus labrax) is an economically important species 47 48 exploited by multiple fishing fleets across Europe (Pawson et al. 2007a, ICES 2013). In northern Europe, the recent combination of declining recruitment and increasing fishing 49 mortality has led to a rapid decline in stock biomass and has triggered management advice for 50 an 80% reduction in catches as an immediate conservation measure (ICES 2014). A package 51 of emergency management measures was implemented in 2015 and then strengthened for 52 2016. For commercial fishing, these measures included a temporal ban on pelagic trawling 53 (six months) and on hooks, lines and fixed gill nets (two months), a monthly catch limit, an 54 increase in the MLS from 36 to 42 cm for northern sea bass and an area closure around 55 Ireland. For recreational fishers the measures include a six month moratorium followed by 56 57 one fish bag limit and the same increase of the MLS of the commercial sector (http://ec.europa.eu/fisheries/cfp/fishing rules/sea-bass/index en.htm). 58

Although the stock structure of sea bass in the northeast Atlantic has not been clearly delineated, there is evidence that sea bass around Ireland and in the Bay of Biscay could be treated as two populations separate from the eastern Celtic Sea, English Channel, and North Sea (Fritsch et al. 2007, ICES 2012). In addition, previous proposals of stock boundaries, based on conventional tagging studies (ICES 2001, ICES 2002, ICES 2004, Fritsh et al. 2007, Pawson et al. 2007b), concluded that there were two separate stock units between the east and west UK (Pawson et al. 2007b). However, despite the inferences from tagging studies, ICES has concluded that current evidence supports the view that sea bass in the North Sea (ICES
Division IVb&c) and in the Irish Sea, the English Channel and Celtic Sea (Ices Divisions
VIIa,d,e,f,g&h) should be treated as a functional stock unit as there is no clear basis at present
to subdivide them into independent stock units (ICES 2012). Clearly, a more accurate
definition of the stock boundaries is required to improve assessment and management advice
to underpin sustainable exploitation.

Previous tagging studies around England and Wales have shown a tendency for adult sea bass 72 to migrate to the south and west in autumn (English Channel), during the pre-spawning 73 season, and to return north and eastwards in spring to feeding areas (Pawson et al. 2007b, 74 Pawson et al. 2008). These mark-recapture studies have provided evidence of philopatry in 75 relation to feeding and spawning areas (Pawson et al. 2007b, Pawson et al. 2008). Pawson et 76 al. (2008) reported that 55% of sea bass > 40 cm that were tagged and released during the 77 summer were subsequently recaptured within 16 km of their original tagging location on their 78 79 summer feeding grounds. A further 23% were recaptured during winter at least 80 km from 80 their release site. These data provided strong evidence that sea bass in England and Wales may share common migration routes to the same spawning grounds but may exhibit 81 segregation by returning to specific summer feeding grounds (Pawson et al. 2008). If this is 82 the case, then management of sea bass stocks may need to be applied at a much smaller 83 regional level than currently proposed by ICES. 84

Although the use of external tags can provide valuable information on stock structure and 85 movement patterns of fish (e.g. Dunn and Pawson 2002, Pawson et al. 2007b, Neuenfeldt et 86 87 al. 2013) there are multiple problems with their use that include, for example, transmission of 88 data to the researchers, and poor recapture rates (Block et al. 2011). Natural biogeochemical markers (trace elements and stable isotopes) located in the hard parts of fishes (e.g otoliths 89 and scales) have great potential as 'internal' tags to study stock structure, and to study 90 connectivity between fish populations in, and movements between, chemically distinctive 91 water bodies by fishes during their lifetime (Elsdon et al. 2008, Trueman et al. 2012). 92 Structures such as otoliths and scales deposit new material incrementally as the fish grows, 93 and can provide a record of the elemental and isotopic composition of the water in which the 94 fish has lived at each stage of its lifetime (e.g. Thorrold et al. 1998, Cadrin et al. 2013). 95

96 The use of stable isotopes remains a relatively underused tool for tracking migration and 97 general movements in marine animals (Trueman et al. 2012). This is because there can be

uncertainties in the use of stable isotopes, in particular in the spatial distribution of stable-98 isotope values across marine basins and in understanding patterns of isotope fractionation in 99 biological systems (Vander Zanden & Rasmussen 2001, Caut et al. 2009). In addition, 100 isotopic signatures vary temporally between body tissues that differ in their metabolic activity 101 102 and therefore represent an integration of feeding history over varying timescales from weeks to months (Vander Zanden et al. 2015). However, incrementally growing hard tissue 103 104 structures such as scales and otoliths will embed within their structure an isotopic signature laid down during a specific period of growth (specific timescale) (Rooker et al. 2008, 105 Sepulveda et al. 2009, Trueman et al. 2012). For fish scales in particular, the analysis of the 106 isotopic signature generally only refers to the most recent season of growth due to the 107 limitations imposed by scale architecture (Hutchinson & Trueman 2006). In fact, a typical sea 108 bass scale consists of two portions: a hard upper layer composed of calcium phosphate 109 overlying a poorly mineralised layer composed largely of collagen (Hutchinson & Trueman 110 2006). The collagenous layer grows by a process of underplating and for this reason only the 111 most recent season of growth is characterised by younger collagen (Hutchinson & Trueman 112 2006). 113

As our understanding of the spatial variation of isotope ratios in the aquatic environment has 114 115 developed, it has been possible to produce 'isoscapes' that map geographic changes in aquatic isotopic signatures (see Graham et al. 2010, West et al. 2010). These isoscape maps 116 can then provide information on the movement patterns and foraging behaviour of study 117 species (see Graham et al. 2010, Hobson et al. 2010). For example, recent studies of scale 118 δ^{13} C and otolith δ^{13} C and δ^{18} O chemistry has provided valuable insights into the large-scale 119 marine migrations of Atlantic salmon and identification of their feeding areas at sea 120 (Mackenzie et al. 2011, Hanson et al. 2013). 121

The aims of this paper were to (1) measure the δ^{13} C and δ^{15} N composition in the last season 122 of growth in scales from adult sea bass caught at several locations around Wales (UK) and (2) 123 to use these isotopic signatures to infer patterns of movement and connectivity between 124 feeding areas to determine the possible presence of different stock units within Welsh coastal 125 waters. Spatial variation in $\delta^{15}N$ of the base of the foodweb for the Irish Sea has been 126 established with a 3% difference in δ^{15} N between north and south Wales (Jennings & Warr 127 2003). Therefore, if sea bass show philopatry to regional feeding grounds around the UK, as 128 suggested by Pawson et al. (2008), we predicted that sea bass caught in north and south 129

Wales would exhibit distinct scale isotopic signatures that could be used to classify fish backto summer feeding region.

132

133

MATERIALS AND METHODS

Data collection and preparation of samples

135

134

Scales from 189 adult sea bass (38.6-60.7 cm total length TL) were collected from a geographically representative range of coastal sites in Wales (Figure 1), 101 scales were collected during the feeding season (July-December 2013) and 88 during the spawning season (March-May 2014).

140 Due to the scale architecture, using collagen from the most recent growth period is the only way to obtain isotope data from the last (most recent) feeding season. Although sea bass 141 scales are relatively large, the last season is often thin (depending on the month of capture, 142 with an average width of section of $0.44 \ (\pm 1.7 \text{ SD}) \text{ mm}$), and therefore several scales from 143 each individual fish were used to gather enough material (0.6 mg) for analysis. Individual 144 scales were briefly soaked in Millipore[™] ultra-pure water and manually cleansed using non-145 metallic forceps and a small nylon brush to remove any remaining adhering vestigial tissue. 146 The last season of growth was trimmed from the top edge of the scale, weighed and placed 147 into pre-weighed tin capsules. Decalcification of sea bass scales was not performed prior to 148 isotopic analysis since the removal of inorganic carbonates has no significant effect on scale 149 δ^{13} C and δ^{15} N values (Sinnatamby et al. 2007, Woodcock & Walther 2014). 150

Scale samples were analysed in a mass spectrometer. Analytical precision for δ^{13} C and δ^{15} N was based on both the long term reproducibility of calibrated in-house standards and repeat analysis of sample material. Standard deviation error for δ^{13} C was <0.1‰ and for δ^{15} N was <0.2‰.

155

Data analysis

156

157 Isotope ratios were expressed using a delta (δ) notation, representing parts per thousand (∞) 158 deviations from the international standards PeeDee Belemnite (PDB) for carbon and air for 159 nitrogen, according to the following equation:

160 $X = [(R_{sample} - R_{standard})/R_{standard}] \times 1000$

where X is δ^{13} C or δ^{15} N and R is the ratio of the heavy isotope to the light isotope (13 C: 12 C or 15 N: 14 N) in the sample and the reference material.

We first measured the intra fish variability to determine whether the use of a single sample per fish provided an accurate measurement of the isotopic signal. We tested seven fish (three from north, one from mid and three from south Wales). For each fish, we prepared and compared three replicates of scale material (last growing season) of 0.6 mg obtained from the same single mix of scales collected from the body area under the pectoral fin. A repeatedmeasures ANOVA was then applied to assess the presence of significant differences in the isotopic signal between samples of the same fish.

Of the 189 fish whose samples were analysed in the present study, 28 (38.6-60.7 cm TL) 170 were caught in estuaries (Aberdovey-mid Wales, n=13 and Burry Port-south Wales, n=15) 171 while the remainder were caught in open coastal waters. A comparison of the $\delta^{13}C$ values 172 between the two groups (estuarine fish vs. marine fish) was undertaken using Welch's t-test. 173 This test was not performed for δ^{15} N, due to the influence of the fish size on the accumulation 174 of the isotope ratio of this element (13 estuarine fish were > 50 cm TL) (Jennings et al. 2002). 175 To assess the possible differences in the isotope signal of sea bass collected during the 176 feeding and spawning season, sea bass from the same location (Hell's Mouth-Mid Wales, fish 177 provided by the same fisher and caught in the same spot, n=15 for both seasons) were 178 compared using a one-way ANOVA. This analysis explores how long sea bass stay in the 179 180 same feeding area. As the scale material is laid down during the feeding season, the scales collected during the spawning season represent the isotopic signature of the area occupied by 181 182 the fish during the whole preceding feeding season and not only during part of it.

To assess the presence of spatial differences in the isotope composition between areas a 183 random forest classification model (R package "randomForest", Liaw & Weiner 2002) was 184 used. Random forest analysis (Breiman 2001) is a nonparametric technique derived from 185 classification and regression trees (CART). The decision-tree modelling approach requires 186 fewer assumptions than traditional parametric methods (e.g. Linear Discriminant Analysis) 187 (Strobl et al. 2009). In particular random forest analysis allows correlated predictor variables 188 to be utilised without transformation or exclusion to obtain unbiased predictions and 189 estimates of variable importance (Strobl et al. 2009). In this context, random forest analysis 190 has been shown to be preferred for discrimination based on otolith microchemistry when the 191

192 assumptions of the traditional parametric methods cannot be reached (Mercier et al. 2011). 193 The random forest model produces many classification trees from which are derived an 194 ensemble of classifications to predict the dependent variable (in our case "geographic 195 location" of sea bass) as a result of average assignment across trees (Strobl et al. 2007, Strobl 196 et al. 2009). By default, the random forest model partitions the data into 'training' (generally 197 70% of data) and 'test' samples selected at random from the data set. Whilst the training 198 samples are used to build the model, the test set is used to validate its performance.

Estuarine fish were excluded from this analysis, to remove the effect of the different salinities 199 encountered between freshwater and marine habitats on δ^{13} C (Doucett et al. 1999). The 200 analysis was also restricted to fish of a similar size range (40-50 cm TL) to remove the effect 201 of the fish size on the variation of δ^{15} N between individuals (e.g. Jennings et al. 2002). The 202 δ^{13} C and δ^{15} N values of 156 sea bass between 40 and 50 cm TL caught in coastal areas (98 203 caught during the feeding season and 58 during the spawning season) were then compared to 204 assess if spatial differences in isotope composition existed. The nine different capture 205 locations were first aggregated into three main groups: north, mid and south Wales (Figure 206 1). The random forest classification model was then used to test if there was a difference in 207 scale δ^{13} C and δ^{15} N values between north, mid and south Wales and whether it was possible 208 to correctly assign the fish to the area where it was caught based on scale isotopic signature. 209 In addition this statistical technique allowed the importance of each predictor variable in the 210 211 classification process to be evaluated and ultimately to identify specific isotopic signatures by area. Based on the level of separation and/or overlap of the isotopic signatures between areas 212 we expected to derive insights on movement patterns related to feeding behaviour for adult 213 sea bass. 214

The association, or proximity, between each fish (characterized by the combination of δ^{13} C and δ^{15} N), is the number of times that they occur together in the same terminal node. The "randomForest" package normalized these counts to produce a proximity matrix that can be analysed using a metric scaling method. The resultant Multi-Dimensional Scaling plot represented the degree of differentiation in stable isotopes values between the three geographic locations.

221 Conditional variable importance was reported to show the relative contribution of each 222 predictor variable (δ^{13} C and δ^{15} N) to the classification performance. To evaluate the 223 conditional variable importance we measured the Mean Decrease Accuracy (MDA) of the forest when the values of each predictor (δ^{13} C and δ^{15} N) are randomly excluded (or permuted). The greater the decrease in the accuracy of the random forest resulting from the exclusion (or permutation) of a single variable, the more important that variable is for classification of the data.

To aid interpretation of the results of the random forest analysis, a conditional inference tree was used. In particular this single-tree method helped to trace the effects of δ^{13} C and δ^{15} N on the classification of the feeding locations of bass and ultimately to identify the range of δ^{13} C and δ^{15} N values associated with each location. To this end, the "ctree" function for conditional inference trees in the "party" R package (Hothorn et al. 2006) was used.

We finally assessed the degree of correlation between the average value of δ^{15} N for sea bass 233 scales for each location sampled in this study and the corresponding values of predicted 234 spatial variation in δ^{15} N baseline estimated from the isoscape map (Jennings & Warr 2003). 235 The model developed by Jennings & Warr (2003), which related the $\delta^{15}N$ of scallops of each 236 sampling site to day of sampling, shell height, depth, surface temperature, bottom 237 temperature and summer salinity was used to calculate the δ^{15} N baseline corresponding to our 238 capture locations. Since not all our sampling sites corresponded exactly to the scallop 239 samplings sites in Jennings & Warr (2003), we used the δ^{15} N baseline values corresponding 240 to the closest areas to our sampling sites (Colwyn Bay, site 28; Anglesey, site 27; Trefor, site 241 30; Hells Mouth, site 31; Aberdovey, site 33; Skokholm Island, site 34; Tenby, Burry Port 242 and Gower, site 37). Although this isoscape map was developed in 2002, the hydrodynamic 243 and biogeochemical processes controlling the distribution of carbon and nitrogen isotope 244 values are temporally stable (MacKenzie et al. 2014). All analyses were carried out using R 245 version 3.0.2 (R Core Team 2013). 246

247

248

RESULTS

The repeated measures ANOVA showed no significant difference between the three replicates of the scale material in δ^{15} N (F_{2,12}=0.33, p=0.73) and δ^{13} C (F_{2,12}=1.14, p=0.35) values (Table S1 in Supplement 1). This demonstrated that the use of a single sample per fish provided an accurate measurement of the isotopic signal in scale material laid down during the last feeding season.

No significant difference was found in the isotope data between the feeding and spawning season for the fish caught in Hell's Mouth (North Wales) (One-way ANOVA, $F_{1,28}=0.17$, p=0.68 for δ^{15} N; F_{1,28}=2.59, p=0.12 for δ^{13} C) (absolute values are reported in Table S2 in Supplement 1). This result suggested that adult sea bass caught in the same area have similar isotope composition even when caught at different times of the year.

The 28 sea bass caught in estuaries had a significantly lower δ^{13} C values (-18.22 ± 1.49‰) than those caught in coastal areas (-13.31 ± 0.99‰) (Welch's t-test, t = 16.8, P < 0.001; absolute values are reported in Table S3 in Supplement 1) which was expected given that freshwater ecosystems generally have lower δ^{13} C values relative to marine systems (Doucett et al. 1999). Out of these fish, 13 were adult sea bass > 50 cm TL.

Regional difference in the marine δ^{13} C and δ^{15} N values was detected (Table 1), such that values from south Wales were separated from those of mid and north Wales (Figure 2a). The two dimensional representation of the random forest analysis also showed the degree of overlap of the isotopic signature between north and mid Wales (Figure 2b).

The random forest classification model built on the training samples had a discrimination 268 capacity c. 75%, which means that most fish were correctly classified between north, mid and 269 south Wales. For both the training (n=116) and test (n=40) data sets, the majority of the 270 misclassifications were fish from north Wales that were classified as mid Wales and vice 271 versa, while the majority of fish from south Wales were correctly assigned (80%) (Table 2). 272 In this case, the misclassification corresponded to regions (Tenby and Skokholm Island) 273 geographically closer to Mid Wales than the rest of the samples (Burry Port and Oxwich-274 Gower). 275

The Mean Decrease Accuracy (MDA) of the forest showed that both variables (δ^{13} C and δ^{15} N) were important for the classification process. On average, δ^{13} C seemed slightly more important than δ^{15} N for classifying the fish. The importance of each isotope varied regionally such that δ^{15} N was more important for classifying fish in north Wales, δ^{13} C for mid Wales and both isotopes were important for classifying fish to south Wales (Table 3).

The most important primary split in the conditional inference tree was the δ^{13} C value; sea bass from north and mid Wales were allocated on the branch corresponding to δ^{13} C \leq -13.2‰ on the proportion of 82% (n=42) and 88% (n=30) respectively (Figure 3). Out of these proportions, most of the fish from north Wales (83%, n=35) had a δ^{13} C value in the range of -14.3‰ and -13.2‰ while fish from mid Wales equally split between the δ^{13} C value comprised between -14.3‰ and -13.2‰ (47%, n=14) and δ^{13} C \leq -14.3‰. Only 13% (n=9) of sea bass caught in south Wales was allocated on the branch corresponding to δ^{13} C \leq - 13.2‰. In fact most of the sea bass from south Wales (87%) were characterized by $\delta^{13}C > -$ 13.2‰. In particular 100% of fish with $\delta^{13}C > -13.2$ ‰ and $\delta^{15}N \le 15.47$ ‰ were from south Wales. This specific signature characterised 68% of the total samples from south Wales (Figure 3).

A strong linear relationship was found between the average values of scale δ^{15} N measured in this study by location (y) and the corresponding values of the δ^{15} N baseline in the vicinity of the sampling location (from Jennings & Warr 2003) (x) (y=0.674x + 9.83, P<0.001, adjusted R² = 0.81) (Figure 4). This demonstrates that the spatial differences in δ^{15} N detected in this study are consistent with the spatial patterns in δ^{15} N that occur at the base of the food web suggesting localised feeding in Welsh sea bass.

298

299

DISCUSSION

The use of isotope tags to study the movement of fish by linking the stable isotope 300 composition measured in fish tissue to that of the base of the food chain at the site of origin is 301 a powerful tool that can help to better define stock boundaries at a local level (Trueman et al. 302 2012). The δ^{13} C and δ^{15} N values of tissue protein are controlled by the trophic level of the 303 fish and δ^{13} C and δ^{15} N values for primary production at the location occupied by the fish 304 during periods of active growth (e.g. Deniro & Epstein 1978, Fry 1981). The highly 305 significant relationship between the average value of $\delta^{15}N$ found in fish scales and the 306 average value of $\delta^{15}N$ baseline by location (Jennings & Warr 2003) confirmed our 307 assumption that the observed spatial differences in sea bass δ^{15} N reflected spatial differences 308 in $\delta^{15}N$ of the base of the food web rather than differences in sea bass feeding strategies. In 309 310 this context the use of the last growing season of sea bass scales can represent an advantage in the analysis because the corresponding isotopic signature takes into account the temporal 311 variability in prey type and abundance across whole of the previous feeding season. Our 312 results thus suggest that much of the spatial variance in the $\delta^{15}N$ of sea bass along the Welsh 313 coast could be attributed to differences in base $\delta^{15}N$ and not in the diet composition, which 314 could be considered, to a first approximation, similar across Wales. Similar results have been 315 obtained for other predatory fishes such as dab Limanda limanda and whiting Merlangius 316 *merlangus*, for which the spatial variance in $\delta^{15}N$ was mostly attributed to differences in $\delta^{15}N$ 317 at the base of the food chain (Jennings & Warr 2003). 318

Our results also showed that the average $\delta^{15}N$ in fish scales tended to increase with latitude. 319 This south-north gradient of $\delta^{15}N$ seems to be confirmed from other studies, which 320 documented higher values of δ^{15} N associated with cooler temperatures (Jennings et al. 2008) 321 and higher levels of oxygen (Radabaugh et al. 2013). While the mechanisms underpinning the 322 spatial isotopic gradients are not fully understood and several factors may influence the 323 spatial variation in δ^{15} N values, the degree of resuspension of particulate organic nitrogen is 324 likely to be a major factor (e.g. Saino & Hattori 1987). Aberdovey (mid Wales) was the only 325 location where sea bass had lower δ^{15} N compared to two areas further south. It is possible 326 that the sea bass from Aberdovey (a site at the mouth of a large estuary) were caught along a 327 freshwater-marine gradient, a fact that could explain this apparent discrepancy, as terrestrial 328 inputs to freshwater bodies are more δ^{15} N-depleted than marine plankton (Schoeninger & 329 DeNiro 1984, Owens 1987). This interpretation would also be consistent with lower $\delta^{13}C$ 330 with respect to fish from the other coastal areas, which could confirm that sea bass caught in 331 Aberdovey originated from a feeding area with freshwater influence (Doucett et al. 1999). 332

Our study also demonstrates that the random forest analysis, although not yet widely used in 333 marine ecology for hierarchical classification (e.g Mercier et al. 2011), is a powerful 334 statistical tool, as it allows not only predictors to be used without transformation or exclusion 335 but also the relative importance of the variables in the classification process to be estimated 336 (Strobl et al. 2009). Therefore, while δ^{15} N appeared more important than δ^{13} C in classifying 337 adult sea bass in north Wales (c.f. fish from mid Wales), both variables were necessary to 338 correctly classify fish from south Wales. Moreover the conditional inference tree allowed us 339 to identify the range of $\delta^{13}C$ and $\delta^{15}N$ values associated with each location and thus to 340 establish a geographic range of the isotopic signatures. Additional variables (e.g. element 341 concentration, Sr, Ba, Mn and Mg) could be used in the future to improve the discriminatory 342 power of the random forest and thus the precision level of the biogeochemical tag (Wells et 343 al. 2000, Ramsay et al. 2011, Seeley et.al 2015). 344

Our results showed that the stable isotope composition of fish scales provides unique insights into fish movement at a regional scale (e.g. the Welsh coastline) without the need for expensive conventional tagging studies. While fine-scale ontogenetic changes in habitat use have been identified for other fish species (e.g. NE Atlantic orange roughly, Shephard et al. (2007)), the use of fish scales has been limited to few species to date (Ramsay et al. 2012; Woodcock & Walther 2014). The analysis of δ^{13} C and δ^{15} N in adult sea bass collected around Welsh waters showed a high degree of overlap in the isotope composition between north and

mid Wales. This finding could be explained by ecological patterns of sea bass population in 352 mid and north Wales and/or by abiotic factors such as temperature in these areas. While the 353 overlapping of the isotopic signature in sea bass scales could indicate the presence of 354 connectivity between north and mid Wales for feeding behaviour, it might also depend on a 355 similar temperature range in these areas, as the isotopic signature is related to productivity, 356 which is temperature-driven (e.g Fogel & Cifuentes 1993). Fish from south Wales appeared 357 more isolated and characterised by a very distinctive isotopic signature, especially fish from 358 the most southern areas (Burry Port and Oxwich-Gower). This finding suggests the possible 359 360 presence of (at least) two separated sub-populations of sea bass in Welsh waters with little mixing between sea bass in the south with those in mid and north Wales. 361

This separation is based on fish locations during the feeding season, and thus our study 362 363 confirms that adult sea bass do spend extended periods of time feeding in regional inshore areas as suggested by Pawson et al. (2007b and 2008). In this sense, the lack of difference in 364 365 the isotopic signature of sea bass caught in Hell's Mouth-mid Wales (the only area where sea bass were caught in the same exact location) between feeding and spawning season, support 366 the previous evidence that adult fish spend most of the feeding season in the same location. In 367 fact the scale material is laid down during the feeding season and therefore the scales 368 collected during the spawning season represent the isotopic signature of the area occupied by 369 the fish during the whole preceding feeding season. Similar fidelity has been demonstrated, 370 through tagging experiments, for other species in the region, such as plaice *Pleuronectes* 371 platessa, which remained on the same feeding grounds throughout the summer and autumn 372 (Dunn & Pawson 2002). In addition, stock boundaries were defined at a regional level with 373 restricted stock units in the north-east Irish Sea, the western Irish Sea, and a stock in the 374 south-east Irish Sea (Dunn & Pawson 2002). 375

No information on the presence of local separate spawning areas is currently available. Due 376 to this lack of information, three possible scenarios remain open: 1) the two sub-populations 377 mix during the spawning season and are part of the same stock, 2) they do not mix during the 378 spawning season and can be therefore considered as two separate sub-populations, 3) they 379 mix only partially. Future research should address this topic and the isotopic signatures 380 identified in this study could be used to track the origin of individuals from spawning 381 382 aggregations. However, whichever scenario is correct, it is already clear that different spatial scales of movement characterize the two sub-populations. Adult sea bass from south Wales 383 are likely to have a restricted range of movement, not only for feeding behaviour as 384

demonstrated by this study but also for spawning, considering the proximity of a well-known 385 spawning ground of the species outside the Bristol Channel (known as "Trevose Head", 386 Lancaster et al. 1998, Reynolds et al. 2003). In this sense, the south Wales sub-population 387 would have a "resident" behaviour, where feeding grounds and spawning grounds are all 388 within close geographical proximity to one another. This trait has been found in many 389 populations of the Atlantic cod Gadus morhua where the residence areas (nursery, feeding 390 391 and spawning grounds) were geographically close and had relatively stable environmental conditions (Robichaud & Rose 2004). In contrast, adult sea bass from mid and north Wales 392 393 either undertake a long-distance migration to the Trevose Head spawning area and/or other potential spawning grounds in south Wales/England or, alternatively, a small-scale migration 394 to possible spawning grounds located off the coast of mid and/or north Wales. Our results 395 provide for the first time insights into a regional segregation of the sea bass stock, which 396 implies that local/regional management may be more appropriate than the present single 397 stock approach. In this context, specific regulations based on regional variation in the ecology 398 and life history of sea bass could facilitate a more rapid achievement of the conservation 399 objectives. 400

Our results also suggest that a portion of the largest sea bass adopt estuaries as preferential 401 402 feeding areas. In fact all fish with a total length >50 cm caught in estuarine areas showed very low δ^{13} C, indicating the primary use of estuaries as feeding areas, as freshwater ecosystems 403 are generally δ^{13} C depleted relative to marine systems (Doucett et al. 1999). It is possible that 404 interspecific competition for resources has resulted in some big adult sea bass adopting an 405 estuarine feeding habit. Estuaries have been recognised to support the early phase of 406 development of the species (Kelley 1988, Lancaster et al. 1998) as well as part of the juvenile 407 stock (3-5 years old) (Pawson et al. 2005) and the present study stressed the importance of 408 409 this habitat for large adult sea bass. While other fish species, such as flathead mullet Mugil cephalus, have been recognised as using estuarine areas through the juvenile and/or adult life 410 stages (Elliott et al. 2007), few previous studies have mentioned the presence of adult sea 411 bass in estuarine habitats in NE Atlantic (Kelley 1987, Kennedy & Fitzmaurice 1972, Pickett 412 & Pawson 1994). Moreover, these studies focused on describing the opportunistic feeding 413 behaviour of the species and its different habitat use, without associating estuaries with a 414 specific ecological role in the adult phase. Our results indicated estuarine feeding of some 415 large sea bass during extended time periods, possibly during the whole feeding season. 416

417 Estuarine areas could therefore be characterised by higher protection levels, if protecting418 large sea bass (e.g. large spawners) is a management target.

In conclusion, the use of the natural variability of the stable isotope composition of fish scales 419 to describe movement patterns of a migratory fish species is an emerging area of research 420 (e.g. MacKenzie et al. 2011, Ramsay et al. 2012, Woodcock & Walther 2014). This technique 421 has useful applications to identify the use of different feeding grounds by individuals from 422 the same population, and also offers a non-lethal alternative to the use of otoliths to determine 423 geographic segregation and/or connectivity between foraging areas. Combined $\delta^{13}C$ and $\delta^{15}N$ 424 measurements of fish scales can clearly identify regional feeding areas providing insights on 425 the feeding ecology at local level and contributing to a better definition of the stock structure 426 and the related management units. 427

428

Acknowledgements. Special thanks to the fishers involved in data collection for investing 429 their time and for their essential feedback. Many thanks to all the staff of the fish processors 430 involved, particularly Mermaid Seafoods in Llandudno and Welsh Seafoods in Milford 431 Haven. Thanks to Hilmar Hinz for his help during the initial phase of this work and to Natalie 432 Hold and Lee G. Murray for managing the project. Thanks to the staff in Nuffield Lab and to 433 all the research assistants, in particular: Harriet Salomonsen, Julia Pantin and Thomas 434 Busbridge. Finally, we would like to thank three anonymous reviewers for the extremely 435 useful feedback. Funding for this work was provided by the European Fisheries Fund. 436

437

438

LITERATURE CITED

439

Block BA, Jonsen ID, Jorgensen SJ, Winship AJ, Shaffer SA, Bograd SJ, Hazen EL, Foley
DG, Breed GA, Harrison AL, Ganong JE, Swithenbank A, Castleton M, Dewar H, Mate BR,
Shillinger GL, Schaefer KM, Benson SR, Weise MJ, Henry RW, Costa DP (2011) Tracking
apex marine predator movements in a dynamic ocean. Nature 475:86–90

444 Breiman, L (2001) Random Forest. Mach Learn 45:5–32

Cadrin S, Kerr LA, Mariani S (2013) Stock Identification Methods 2nd edition. Academic
Press.

- 447 Caut S, Angulo E, Courchamp F (2009) Variation in discrimination factors (Delta N-15 and
- 448 Delta C-13): the effect of diet isotopic values and applications for diet reconstruction. J App
- 449 Ecol 46:443–453
- 450 Deniro MJ, Epstein S (1978) Influence of diet on distribution of carbon isotopes in animals.
- 451 Geochim Cosmochim Acta 42:495–506
- 452 Doucett RR, Hooper W, Power G (1999) Identification of anadromous and nonanadromous
- 453 adult brook trout and their progeny in the Tabusintac River, New Brunswick, by means of
- 454 multiple-stable-isotope analysis. Trans Am Fish Soc 128:278–288
- Dunn MR, Pawson MG (2002). The stock structure and migrations of plaice populations on
 the west coast of England and Wales. J Fish Biol 61:360–393
- 457 Elliott M, Whitfield AK, Potter IC, Blaber SJM, Cyrus DP, Nordlie FG, Harrison TD (2007)
- The guild approach to categorizing estuarine fish assemblages: a global review. Fish Fish8:241–268
- Elsdon TS, Wells BK, Campana SE, Gillanders BM, Jones CM, Limburg KE, Secor DH,
 Thorrold SR, Walther BD (2008) Otolith chemistry to describe movements and life-history
 parameters of fishes: Hypotheses, assumptions, limitations and inferences. Oceanogr Mar
 Biol 46:297–330
- Fogel ML, Cifuentes LA (1993) Isotope fractionation during primary production. In: Engel
 MH, Macko SA (eds) Organic Geochemistry: Principles and Applications. Plenum Press,
 New York; p 73–98
- 467 Fritsch M, Morizur Y, Lambert E, Bonhomme F, Guinand B (2007) Assessment of sea bass
 468 (*Dicentrarchus labrax*, L.) stock delimitation in the Bay. Fish Res 83:123–132
- 469 Fry B (1981) Natural stable carbon isotope tag traces Texas shrimp migrations. Fish Bull
 470 79:337–345
- 471 Graham BS, Koch PL, Newsome SD, McMahon KW, Aurioles D (2010) Using isoscapes to
- trace the movements and foraging behavior of top predators in oceanic ecosystems. In: (West
- 473 JB, Bowen GJ, Dawson TE, Tu KP (eds) Isoscapes: Understanding Movement, Pattern, And
- 474 Process On Earth Through Isotope Mapping. Springer, New York; p 299–318

- 475 Hanson NN, Wurster CM, Todd CD (2013) Reconstructing marine life-history strategies of
- 476 wild Atlantic salmon from the stable isotope composition of otoliths. Mar Ecol Prog Ser
- 477 475:249–266
- 478 Hobson KA, Barnett-Johnson R, Cerling T (2010) Using isoscapes to track animal migration.
- 479 In: West JB, Bowen GJ, Dawson TE, Tu KP (eds) Isoscapes: Understanding Movement,
- 480 Pattern, And Process On Earth Through Isotope Mapping. Springer, New York; p 273–298
- 481 Hothorn T, Hornik K, Zeileis A (2006) Unbiased Recursive Partitioning: A Conditional
- 482 Inference Framework. J Comput Graph Stat 15:651–674
- Hutchinson JJ, Trueman CN (2006) Stable isotope analyses of collagen in fish scales:
 limitations set by scale architecture. J Fish Biol 69:1874–1880
- .
- 485 ICES (2001) Report on the ICES Study Group on bass. CM 2001/ACFM:25
- 486 ICES (2002) Report on the ICES Study Group on bass. CM 2002/ACFM:11 ref.G
- ICES (2004) Report of the Study Group on Bass, Lowestoft, England, August 2003. ICES
 Document, CM 2004/ACFM: 04
- 489 ICES (2012) Report of the Inter-Benchmark Protocol on New Species (Turbot and Sea bass;
- 490 IBPNew 2012), 1–5 October 2012, Copenhagen, Denmark. ICES CM 2012/ACOM:45
- 491 ICES (2013) Report of the Working Group for Celtic Seas Ecoregion (WGCSE), 8–17 May
 492 2013, Copenhagen, Denmark. ICES CM 2013/ACOM:12
- 493 ICES (2014) European sea bass in Divisions IVbc, VIIa, and VIId-h (Irish Sea, Celtic Sea,
- 494 English Channel, and southern North Sea). ICES Advice 2014, Book 5
- Jennings S, Barnes C, Sweeting CJ, Polunin NVC (2008) Application of nitrogen stable
 isotope analysis in size-based marine food web and macroecological research. Rapid
 Commun Mass Spectrom 22:1673–1680
- Jennings S, Pinnegar JK, Polunin NVC, Warr KJ (2002) Linking size-based and trophic
 analyses of benthic community structure. Mar Ecol Prog Ser 226:77–85
- Jennings S, Warr KJ (2003) Environmental correlates of large-scale spatial variation in the δ^{15} N of marine animals. Mar Biol 142:1131–1140
- 502 Kelley DF (1987) Food of bass in U.K. waters. J Mar Biol Assoc UK 67:275–286

- Kelley DF (1988) The importance of estuaries for sea bass *Dicentrarchus Labrax* (L.). J Fish
 Biol 33 (Suppl. A):25–33
- Kennedy M, Fitzmaurice P (1972) The biology of the bass, *Dicentrarchus labrax*, in Irish
 Waters. J Mar Biol Assoc UK 52:557–597
- 507 Lancaster JE, Pawson MG, Pickett GD, Jennings S (1998) The Impact of the "Sea Empress"
- oil spill on seabass recruitment. Mar Pollut Bull 36:677–688
- 509 Liaw A, Wiener M (2002) Classification and regression by randomForest. R News 2/3:18–22
- MacKenzie K, Longmore C, Preece C, Lucas C, Trueman C (2014) Testing the long-term
 stability of marine isoscapes in shelf seas using jellyfish tissues. Biogeochemistry 121:441–
 454
- 513 MacKenzie KM, Palmer MR, Moore A, Ibbotson AT, Beaumont WRC, Poulter DJS,
- 514 Trueman CN (2011) Locations of marine animals revealed by carbon isotopes. Sci Rep 1:21
- 515 Mercier L, Darnaude AM, Bruguier O, Vasconcelos RP, Cabral HN, Costa MJ, Lara M,
- 516 Jones DL, Mouillot D (2011) Selecting statistical models and vari- able combinations for
- 517 optimal classification using otolith microchemistry. Ecol Appl 21:1352–1364
- 518 Neuenfeldt S, Righton D, Neat F, Wright PJ, Svedang H, Michalsen K, Subbey S, Steingrund
- 519 P, Thorsteinsson V, Pampoulie C, Andersen KH, Pedersen MW, Metcalfe J (2013) Analysing
- migrations of Atlantic cod Gadus morhua in the north-east Atlantic Ocean: then, now and the
 future. J Fish Biol 82:741–763
- 522 Owens NJP (1987) Natural variations in I5N in the marine environment. Adv Mar Biol 523 24:389–451
- Pawson MG, Brown M, Leballeur J, Pickett GD (2008) Will philopatry in sea bass,
 Dicentrarchus labrax, facilitate the use of catch-restricted areas for management of
 recreational fisheries? Fish Res 93:240–243
- Pawson MG, Kupschus S, Pickett GD (2007a) The status of sea bass (Dicentrarchus labrax)
 stocks around England and Wales, derived using a separable catch-at-age model, and
 implications for fisheries management. ICES J Mar Sci 64:346–356

- 530 Pawson MG, Pickett GD, Leballeur J, Brown M, Fritsch M (2007b) Migrations, fishery
- 531 interactions and management units of sea bass (*Dicentrarchus labrax*) in Northwest Europe.

532 ICES J Mar Sci 64:332–345

- Pawson MG, Pickett GD, Smith MT (2005) The role of technical measures in the recovery of
- the UK sea bass (*Dicentrarchus labrax*) fishery 1980–2002. Fish Res 76:91–105
- Pickett GD, Pawson MG, 1994. Sea bass. Biology, exploitation and conservation. Chapman
 & Hall, London
- R Core Team (2013) R: a language and environment for statistical computing. R Foundation
 for Statistical Computing, Vienna, Austria. <u>http://www.R-project.org/</u>
- 539 Radabaugh KR, Hollander DJ, Peebles EB (2013) Seasonal δ^{13} C and δ^{15} N isoscapes of fish
- populations along a continental shelf trophic gradient. Cont Shelf Res 68:112–122
- 541 Ramsay AL, Milner NJ, Hughes RN, McCarthy ID (2011) Comparison of the performance of
- scale and otolith microchemistry as fisheries research tools in a small upland catchment. Can
 J Fish Aquat Sci 68:1–11
- 544 Ramsay AL, Milner NJ, Hughes RN, McCarthy ID (2012) Fish scale δ^{15} N and δ^{13} C values
- provide biogeochemical tags of fish comparable in performance to element concentrations in
 scales and otoliths. Hydrobiologia 694:183–196
- 547 Reynolds WJ, Lancaster JE, Pawson MG (2003) Patterns of spawning and recruitment of
- bass to Bristol Channel nurseries in relation to the 1996 "Sea Empress" oil spill. J Mar Biol
 Assoc UK 83:1163–1170
- Robichaud D, Rose GA (2004) Migratory behaviour and range in Atlantic cod: inference
 from a century of tagging. Fish Fish 5:185–214
- 552 Rooker JR, Secor DH, DeMetrio G, Kaufman AJ, Rios AB, Ticina V (2008) Evidence of
- trans-Atlantic movement and natal homing of bluefin tuna from stable isotopes in otoliths.
 Mar Ecol Prog Ser 368:231–239
- 555 Saino T, Hattori A (1987) Geographical variation of the water column distribution of 556 suspended particulate organic nitrogen and its ¹⁵N natural abundance in the Pacific and its 557 marginal seas. Deep-Sea Res 34:807–827

- 558 Schoeninger MJ, DeNiro MJ (1984) Nitrogen and carbon isotopic composition of bone 559 collagen from marine and terrestrial animals Geochim Cosmochim Acta 48:625–639
- 560 Seeley M, Miller N, Walther B (2015) High resolution profiles of elements in Atlantic tarpon
- 561 (Megalops atlanticus) scales obtained via cross-sectioning and laser ablation ICP-MS: a
- 562 literature survey and novel approach for scale analyses. Environ Biol Fish 98:2223–2238
- 563 Sepulveda AJ, Colver WT, Lowe WH, Vinson MR (2009) Using nitrogen stable isotopes to
- 564 detect long-distance movement in threatened cutthroat trout (*Oncorhynchus clarkii* utah). Can
- 565 J Fish Aquat Sci 66:672–682
- 566 Shephard S, Trueman C, Rickaby R, Rogan E (2007) Juvenile life history of NE Atlantic
- orange roughy from otolith stable isotopes. Deep Sea Res Part 1 Oceanogr Res Pap 54:1221–
 1230
- 569 Strobl C, Boulesteix AL, Zeileis A, Hothorn T (2007) Bias in random forest variable 570 importance measures: illustrations, sources and a solution. BMC Bioinformatics 8:25
- 571 Strobl C, Malley J, Tutz G (2009) An introduction to recursive partitioning: rationale, 572 application, and characteristics of classification and regression trees, bagging, and random 573 forests. Psychol Methods 14:323–348
- 574 Thorrold SR, Jones CM, Swart PK, Targett TE (1998) Accurate classification of juvenile 575 weakfish *Cynoscion regalis* to estuarine nursery areas based on chemical signatures in 576 otoliths. Mar Ecol Prog Ser 173:253–265
- 577 Trueman CN, MacKenzie KM, Palmer MR (2012) Identifying migrations in marine fishes
 578 through stable-isotope analysis. J Fish Biol 81:826–847
- Vander Zanden MJ, Clayton MK, Moody EK, Solomon CT, Weidel BC (2015) Stable
 isotope turnover and half-life in animal tissues: a literature synthesis. PLoS ONE
 10(1):e0116182
- 582 Vander Zanden MJ, Rasmussen JB (2001) Variation in δ^{15} N and δ^{13} C trophic fractionation: 583 Implications for aquatic food web studies. Limnol Oceanogr 46:2061–2066
- 584 Wells BK, Thorrold SR, Jones CM (2000) Geographic variation in elemental signatures of
- weakfish scales. T Am Fish Soc 129:889–900.

- 586 West JB, Bowen GJ, Dawson TE, Tu KP (2010) Isoscapes: Understanding Movement,
- 587 Pattern, And Process On Earth Through Isotope Mapping. Springer, New York
- Woodcock SH, Walther BD (2014) Trace elements and stable isotopes in Atlantic tarpon
 scales reveal movements across estuarine gradients. Fish Res 153:9–17

Table 1. Values (mean \pm SD) of δ^{13} C and δ^{15} N in scale material formed during the last growth period of the adult sea bass *Dicentrarchus labrax* (40-50 cm TL, n=156) caught in coastal areas around Wales.

Area	Location	δ^{13} C (VPDB)‰	δ^{15} N (AIR)‰
North Wales	Colwyn Bay	-13.85 ± 0.61	16.76 ± 1.06
(n=51)	Anglesey	-13.35 ± 0.69	15.96 ± 0.50
	Trefor	-13.62 ± 0.61	16.06 ± 1.40
Mid Wales (n=34)	Hells Mouth	-14.02 ± 0.70	15.64 ± 1.09
	Aberdovey	-15.05 ± 0.61	14.84 ± 0.39
South Wales	Skokholm Island	-12.98 ± 1.20	15.43 ± 0.79
(n=71)	Tenby	-12.45 ± 0.50	14.82 ± 0.62
	Burry Port	-12.78 ± 0.68	15.24 ± 1.27
	Gower	-12.36 ± 0.48	14.52 ± 0.70

		Predicted							
		No	orth	Ν	/lid	So	uth	Erro	or (%)
		train	test	train	test	train	test	train	test
pa	North	28	8	3	4	8	0	28.2	33
serve	Mid	8	1	18	4	2	1	35.7	33
Obs	South	7	3	4	0	38	19	22.4	13.6

Table 2. Observed vs. predicted feeding locations of adult sea bass derived from the randomforest classification model for training and testing (cross-validation) data sets.

600	Table 3. Conditional variable importance from random forest analysis, expressed as the mean
601	decrease accuracy (MDA) of the random forest model associated with the removal of each
602	predictor (δ^{13} C and δ^{15} N) for each location.

Mean decrease accuracy					
	North	Mid	South	average	
$\delta^{15}N$	33.20	5.98	21.53	31.36	
$\delta^{13}C$	14.66	26.44	37.04	47.70	



Figure 1. Map of Wales indicating the locations where sea bass were caught for the analysisof scale carbon and nitrogen stable isotope ratios.



610

Figure 2. a) Isotopic plot showing the δ^{13} C and δ^{15} N in the scales of adult sea bass caught in nine different locations around the coast of Wales. b) Two-dimensional representation of the random forest analysis, showing the degree of separation between locations based on scale δ^{15} N and δ^{13} C. Open symbols, north Wales; grey symbols, mid Wales and black symbols, south Wales.



Figure 3. Conditional inference tree on the full dataset (n=156 adult bass, comprising the 116 training cases and the 40 testing cases used in the random forest analysis) using the variable δ^{13} C and δ^{15} N. Inner nodes (ovals) indicate which variables were used for splitting (threshold values on the line) and n is the number of adult sea bass falling in each terminal node. Bars express the proportion of sea bass in the node that are from north, mid or south Wales.



Figure 4. Relationship (regression line) between the average scale $\delta^{15}N$ in sea bass captured in 9 locations around the coast of Wales and the corresponding values of predicted spatial variation in $\delta^{15}N$ baseline estimated from the isoscape map in the Irish Sea (Jennings and Warr, 2003) (grey area: 95% confidence interval of the slope of the regression line).

-

- 640 Supplementary material
- 641 Supplement 1

Table S1. Absolute isotope values used in the repeated-measures ANOVA to assess thepresence of significant differences in the isotopic signature between samples of the same fish.

Location	Individual	$\delta^{15}N$	$\delta^{13}C$
Colwyn Bay	I1	17.9	-13.8
Colwyn Bay	I1	17.6	-14.1
Colwyn Bay	I1	17.5	-14.2
Trefor	I2	15.6	-13.2
Trefor	I2	16.1	-13.2
Trefor	I2	16.7	-13.3
Hell's Mouth	I3	15.7	-14.1
Hell's Mouth	I3	15.1	-13.8
Hell's Mouth	I3	15.4	-13.9
Aberdovey	I4	14.7	-15.2
Aberdovey	I4	14.8	-15.1
Aberdovey	I4	14.7	-15.0
Skokholm Island	15	16.4	-12.8
Skokholm Island	15	16.5	-12.8
Skokholm Island	15	16.6	-12.9
Tenby	I6	14.8	-12.6
Tenby	I6	14.8	-12.1
Tenby	I6	15.0	-12.8
Gower	I7	15.5	-12.1
Gower	I7	15.5	-12.3
Gower	I7	15.3	-12.3

Table S2. Absolute isotope values used in the one-way ANOVA to assess possible
differences between the feeding and spawning season for the fish caught in Hell's Mouth
(North Wales)

Season	Month	$\delta^{15}N$	$\delta^{13}C$
feeding	October	16.8	-14.0
feeding	October	17.8	-14.0
feeding	October	15.0	-14.6
feeding	October	14.4	-15.5
feeding	October	15.0	-14.8
feeding	November	16.5	-14.0
feeding	November	16.3	-14.4
feeding	November	15.4	-13.9
feeding	November	14.7	-14.5
feeding	November	17.6	-13.1
feeding	November	14.1	-14.4
feeding	November	15.3	-13.4
feeding	November	14.3	-14.0
feeding	November	14.5	-14.9
feeding	November	15.6	-13.8
spawning	April	15.7	-13.1
spawning	April	15.2	-14.5
spawning	April	14.2	-13.7
spawning	March	15.4	-14.2
spawning	March	14.7	-14.3
spawning	March	16.2	-12.8
spawning	March	15.7	-14.2
spawning	March	17.8	-13.2
spawning	March	14.8	-14.8
spawning	March	15.7	-14.0
spawning	March	16.7	-13.2
spawning	March	16.7	-12.2
spawning	March	14.9	-14.5
spawning	March	14.8	-14.2
spawning	March	17.3	-14.4

Area	Location	$\delta^{15}N$	δ ¹³ C
North	coast	14.9	-14.1
North	coast	16.8	-14.0
North	coast	19.0	-14.2
North	coast	17.0	-13.8
North	coast	16.0	-14.6
North	coast	16.1	-14.5
North	coast	17.0	-13.2
North	coast	15.8	-14.1
North	coast	17.6	-14.0
North	coast	18.6	-13.9
North	coast	16.0	-13.9
North	coast	16.3	-12.2
North	coast	17.0	-14.2
North	coast	16.4	-13.8
North	coast	16.7	-13.1
North	coast	14.7	-15.7
North	coast	16.0	-12.4
North	coast	15.8	-13.2
North	coast	16.3	-12.8
North	coast	15.0	-13.5
North	coast	16.3	-14.2
North	coast	16.3	-14.0
North	coast	16.1	-13.3
North	coast	15.9	-14.1
North	coast	16.1	-12.8
North	coast	17.1	-13.2
North	coast	17.3	-13.8

Table S3. Absolute isotope values of the 189 sea bass collected around Wales and indicationof the capture location (coast vs. estuary) used for the Welch's t-test.

North	coast	17.9	-13.7
North	coast	15.8	-14.2
North	coast	17.4	-13.3
North	coast	16.0	-12.7
North	coast	17.3	-13.4
North	coast	15.5	-12.9
North	coast	18.1	-13.3
North	coast	16.6	-12.5
North	coast	17.1	-13.7
North	coast	18.0	-13.4
North	coast	14.3	-14.1
North	coast	16.2	-13.2
North	coast	16.3	-13.6
North	coast	16.2	-13.2
North	coast	15.7	-14.6
North	coast	15.4	-14.6
North	coast	11.1	-14.4
North	coast	14.2	-14.6
North	coast	15.9	-13.7
North	coast	13.7	-14.2
North	coast	16.4	-13.3
North	coast	16.4	-14.8
North	coast	15.6	-13.0
North	coast	16.5	-13.3
North	coast	15.8	-13.8
Mid	coast	16.5	-14.0
Mid	coast	16.3	-14.4
Mid	coast	15.4	-13.9
Mid	coast	14.7	-14.5
Mid	coast	17.6	-13.1

Mid	coast	14.1	-14.4
Mid	coast	15.3	-13.4
Mid	coast	14.3	-14.0
Mid	coast	14.5	-14.9
Mid	coast	15.6	-13.8
Mid	coast	16.8	-14.0
Mid	coast	17.8	-14.0
Mid	coast	15.0	-14.6
Mid	coast	14.4	-15.5
Mid	coast	15.0	-14.8
Mid	coast	14.7	-15.2
Mid	coast	14.4	-14.4
Mid	coast	14.9	-15.8
Mid	coast	15.3	-14.8
Mid	coast	15.2	-14.3
Mid	coast	13.9	-15.3
Mid	coast	15.4	-14.2
Mid	coast	14.7	-14.3
Mid	coast	16.2	-12.8
Mid	coast	15.7	-14.2
Mid	coast	17.8	-13.2
Mid	coast	14.8	-14.8
Mid	coast	15.7	-14.0
Mid	coast	16.7	-13.2
Mid	coast	16.7	-12.2
Mid	coast	14.9	-14.5
Mid	coast	14.8	-14.2
Mid	coast	17.3	-14.4
Mid	coast	15.7	-13.1
Mid	coast	15.2	-14.5

Mid	coast	14.2	-13.7
Mid	estuary	15.9	-19.1
Mid	estuary	15.2	-20.5
Mid	estuary	16.2	-19.5
Mid	estuary	15.5	-19.1
Mid	estuary	15.3	-19.5
Mid	estuary	15.5	-19.2
Mid	estuary	16.1	-19.1
Mid	estuary	15.3	-18.5
Mid	estuary	16.1	-19.4
Mid	estuary	15.9	-19.4
Mid	estuary	15.5	-20.1
Mid	estuary	15.6	-19.5
Mid	estuary	15.8	-19.8
South	coast	16.0	-13.9
South	coast	15.3	-12.3
South	coast	14.8	-15.1
South	coast	14.1	-15.2
South	coast	14.7	-12.6
South	coast	14.8	-12.6
South	coast	14.6	-13.4
South	coast	16.8	-13.0
South	coast	14.9	-13.2
South	coast	16.4	-12.8
South	coast	16.1	-12.4
South	coast	16.2	-10.3
South	coast	15.8	-13.0
South	coast	15.4	-13.1
South	coast	15.7	-11.8
South	coast	14.6	-13.2

South	coast	14.4	-13.1
South	coast	15.1	-12.9
South	coast	15.7	-13.0
South	coast	14.4	-13.2
South	coast	15.6	-12.7
South	coast	16.0	-12.2
South	coast	14.9	-12.5
South	coast	14.8	-12.6
South	coast	14.6	-12.4
South	coast	14.0	-12.3
South	coast	14.8	-12.3
South	coast	14.9	-13.8
South	coast	15.2	-12.1
South	coast	15.0	-12.9
South	coast	14.3	-13.0
South	coast	14.1	-13.0
South	coast	14.5	-12.6
South	coast	14.7	-12.5
South	coast	15.4	-12.5
South	coast	14.9	-12.7
South	coast	14.6	-12.2
South	coast	15.5	-11.9
South	coast	15.5	-12.1
South	coast	13.8	-12.0
South	coast	13.4	-12.3
South	coast	14.3	-11.5
South	coast	13.4	-11.7
South	estuary	15.6	-17.5
South	estuary	14.9	-16.6
South	estuary	16.7	-16.1

South	estuary	16.1	-16.2
South	estuary	14.9	-17.2
South	estuary	14.3	-20.4
South	estuary	16.5	-16.4
South	estuary	14.0	-17.6
South	estuary	14.9	-16.5
South	estuary	16.2	-16.4
South	estuary	16.1	-17.1
South	estuary	16.7	-17.9
South	estuary	15.2	-16.6
South	estuary	17.0	-19.1
South	estuary	16.7	-15.9
South	coast	14.5	-12.1
South	coast	15.5	-12.9
South	coast	15.4	-12.4
South	coast	13.7	-12.2
South	coast	16.7	-12.9
South	coast	16.2	-14.1
South	coast	17.9	-13.2
South	coast	13.2	-14.2
South	coast	14.8	-13.0
South	coast	16.7	-12.1
South	coast	15.0	-12.2
South	coast	14.7	-12.5
South	coast	14.9	-12.6
South	coast	14.1	-12.6
South	coast	15.4	-11.7
South	coast	14.2	-12.4
South	coast	14.8	-12.7
South	coast	14.9	-11.8

South	coast	14.2	-12.7
South	coast	14.8	-12.7
South	coast	14.5	-11.8
South	coast	14.3	-12.0
South	coast	14.7	-12.4
South	coast	14.3	-11.9
South	coast	14.4	-11.8
South	coast	14.9	-12.1
South	coast	15.0	-11.7
South	coast	15.0	-12.4
South	coast	13.9	-12.5
South	coast	16.7	-12.5