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Williamson, Jennifer L.; Mills, Gina; Hayes, Felicity; Jones, Timothy; Freeman, Chris. 2016. How do increasing background concentrations of tropospheric ozone affect peatland plant growth and carbon gas exchange?

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1	How do increasing background concentrations of tropospheric ozone affect peatland
2	plant growth and carbon gas exchange?
3	
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6	
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11	Abstract
12	
13	In this study we have demonstrated that plants originating from upland peat bogs are
14	sensitive to increasing background concentrations of ozone. Peatland mesocosms
15	from an upland peat bog in North Wales, UK were exposed to eight levels of elevated
16	background ozone in solardomes for 4 months from May to August, with 24 hour
17	mean ozone concentrations ranging from 16 to 94 ppb and cumulative AOT024hr
18	ranging from 45.98 ppmh to 259.63 ppmh. Our results show that plant senescence
19	increased with increasing exposure to ozone, although there was no significant effect
20	of increasing ozone on plant biomass. Assessments of carbon dioxide and methane
21	fluxes from the mesocosms suggests that there was no change in carbon dioxide
22	fluxes over the 4 month exposure period but that methane fluxes increased as
23	cumulative ozone exposure increased to a maximum AOT 0_{24hr} of approximately 120
24	ppm h and then decreased as cumulative ozone exposure increased further.
25	
26	Key words: tropospheric ozone; methane; peatlands; wetlands; senescence

28 Highlights

29	•	Peatland plant senescence is increased by season-long exposure to elevated
30		ozone but above and below ground plant biomass is not significantly affected.
31	•	Methane emissions increase at low to moderate cumulative ozone exposure
32		but decrease as cumulative ozone exposure increases further.
33	٠	Dissolved organic carbon in the peat pore water and carbon dioxide exchange
34		are not significantly affected by increasing background ozone concentrations.

35

36 Introduction

37 Peat-forming wetlands are an important carbon storage ecosystem with global estimates of carbon sequestration in the region of 20-30 gCm⁻²yr⁻¹ (Wieder, 2001). Carbon 38 dioxide is taken up by vascular plants and mosses during photosynthesis and, although 39 40 some is released back to the environment during plant respiration, the remainder is stored in plant tissue or transported through the plant and released as exudates of 41 dissolved organic material (Schutz et al., 1991). Once the plants die, the carbon in their 42 tissues is not broken down and released to the atmosphere as peatland decomposition 43 rates are low so the plant material is laid down as peat. The low molecular weight 44 45 exudates such as sugars and amino acids form an energy source for bacteria and archaea living in the peatland and their metabolism contributes to the carbon dioxide release 46 and to the release of methane. The dissolved carbon in the pore water can be exported 47 48 out of the peatland in streams and this can be an important loss point for carbon in the 49 peatland carbon cycle. The majority of peat-forming wetlands in northern Europe are in upland areas where ozone concentrations are higher than adjacent low-lying areas 50 51 (Royal Society, 2008), and thus any changes that affect plant growth and carbon gas exchange have the potential to affect carbon storage within peatlands. 52

Annual mean tropospheric ozone concentrations in Northern Europe are currently in the region of 30-35 ppb, having increased from less than 20 ppb in the mid-20th century. Although there has been little change in mean concentrations in the past decade (Hartmann *et al.*, 2014), climate change and hemispheric transport of pollutants may affect future ozone levels. One future scenario predicts that background ozone concentrations in Northern Europe will continue to increase during the 21st century due to hemispheric transport of ozone precursor molecules (Royal Society 2008).

Tropospheric ozone is a phytotoxic pollutant and wetland vascular plants have been 60 61 found to be relatively sensitive to elevated ozone concentrations (Franzaring et al., 2000, Power & Ashmore, 2002, Williamson et al., 2010) with symptoms including 62 premature senescence, reductions in photosynthesis and reduced biomass. However, 63 64 Sphagnum mosses have been shown to be relatively tolerant to elevated ozone concentrations (Rinnan 2003) during both short term acute ozone fumigation (Potter et 65 al., 1996a) and during longer term exposure with only Sphagnum recurvum showing a 66 67 reduction in shoot growth under elevated ozone (Potter et al., 1996b).

68 Carbon dioxide uptake in wetland mesocosms takes place during photosynthesis and,

69 if plant growth is reduced by increasing background ozone, it may be expected that

70 carbon dioxide uptake would be reduced. Increasing tropospheric ozone

concentrations during short-term exposure was found to transiently increase the rate

of dark respiration (Niemi et al., 2002, Rinnan et al., 2003), possibly as a result of the

73 plants repairing ozone damaged tissues. Under a doubling in ambient ozone

concentrations Haapala et al., (2011) showed that photosynthesis was reduced during

the first year of a four year exposure period but during the fourth year both

76 photosynthesis and total respiration showed a tendency to be higher than under

77 ambient conditions.

As wetland plants play a major role in methane emission from wetlands it is possible 78 79 that any damage to wetland plant functioning by ozone could have a secondary effect on these. Two possible routes via which methane emissions are affected by plant 80 81 growth are: through the provision of a conduit for gas exchange via the aerenchyma (Chanton et al., 1997; Ding et al., 2005; Greenup et al., 2000; Thomas et al., 1996) 82 and the exudation of low molecular weight compounds to provide an energy source 83 for microbes (Schutz et al., 1991). A range of wetland adapted species demonstrate 84 active pressurised gas flow from the leaves to the roots to allowed continued oxygen 85 86 supply in waterlogged soils, whereby air is forced downwards through the aerenchyma as a result of the pressure generated through the gradient in temperature 87 and water vapour pressure. The return flow of gas is through the older leaves of the 88 89 plants as they are unable to support the pressure gradients required to force air down into the roots (Mitsch and Gosselink 2000). Ozone-induced increased senescence 90 could increase the available pathways for gas release and hence the flow of methane 91 92 between the substrate and the atmosphere. Previous studies have shown that recently fixed photosynthate is preferentially retained in leaves rather than transported through 93 the plant (Andersen, 2003; Grantz and Farrar, 1999, 2000), which could lead to a 94 reduction in root exudates meaning that there would be less energy available for 95 methanogens. 96

97 Previous published research on the effect of elevated ozone on methane emissions 98 from peatlands ranged from showing an increase when mesocosms were exposed to 99 100ppb ozone (Niemi *et al.*, 2002), a non-significant increase after 50 days exposure 100 to 200ppb ozone (Rinnan *et al.*, 2003), a transient decrease at ozone concentrations 101 double the current ambient (Morsky *et al.*, 2008) to a significant decrease in methane 102 emissions during the growing seasons of a two year exposure period (Toet *et al.*, 2011). A previous study investigating the impacts of elevated background ozone
using open top chambers showed that seasonal exposure of meadow mesocosms to
elevated ozone over a three year period did not change methane fluxes (Kanerva *et al.*, 2007).

Here, we have studied the effects of increasing background ozone concentrations on 107 plant senescence, plant growth and carbon gas exchange in peatlands, an ecosystem 108 recognised as having the potential to exert profound changes to the planet's climate 109 through the storage of carbon and the emission of methane (Bridgham et al., 2013, 110 111 Freeman et al., 2001). By using a wide range of ozone treatments, we hoped to increase our understanding of tropospheric ozone effects on plant growth and carbon 112 gas exchange from peatlands and shed some light on the conflicting results found in 113 114 other studies.

115

116 Materials and Methods

117 Ozone exposure:

Forty-eight mesocosms (diameter 16cm, depth 40cm) were collected from the 118 Migneint, a large area of oligotrophic, blanket bog in North Wales, UK (3°48.8' W, 119 52°59.6' N) dominated by the NVC vegetation type M6 (Carex echinata-Sphagnum 120 121 recurvum/auriculatum mire) (Buckton & Ormerod, 1997), following the method of 122 Freeman et al., (1993) and exposed to ozone in specially constructed greenhouses (solardomes), with 6 replicate mesocosms per ozone treatment. Mesocosms were 123 selected to ensure as uniform as possible vegetation cover, with the vascular plants 124 125 being Juncus effusus and Carex echinata. Sphagum mosses were not identified to species level but the majority of the Sphagnum present in the area sampled was 126 Sphagnum fallax. The water table was maintained within 2cm of the surface of the 127

128 mesocosms throughout the exposure period.

The solardome facility consists of eight hemispherical glass domes, 3 m in diameter 129 and 2 m tall, situated on an East-West line to minimise differences in shading as used 130 in previous experiments including: Hayes et al (2015), Hayes et al (2011), and Mills et 131 al (2009). Ozone was generated by passing oxygen (from a Workhorse 8 oxygen 132 generator, Ozone Industries Ltd.) through a G11 ozone generator (Ozone Industries 133 Ltd.). A computer-controlled (Lab-VIEW version 7) mass-flow controller system was 134 used to deliver ozone to the solardomes, where it was mixed with charcoal filtered air 135 136 and the fan system ensured two complete air changes per minute. The ozone concentration in the centre of each of the solardomes was measured on a 30 minute 137 cycle by two API400 ozone analysers (Envirotech) with matched calibrations. Ozone 138 139 concentrations in one solardome were continually sampled to provide a feedback system using a Model 49C ozone analyser (Thermo Electron) and the ozone supply to 140 all domes was adjusted accordingly. 141 142 The ozone profile used in the solardomes was based on concentrations measured at the Snowdonia ozone monitoring site at Marchlyn Mawr, Wales, UK(4°03.4' W, 143 53°08.2' N) during a typical week with no marked ozone episodes but relatively high 144 background ozone: 31st May - 6th June 2006 (AA treatment) and with incremental 145 starting points. The target treatments consisted of a sub-ambient treatment (AA-20 146 147 ppb), a simulated ambient treatment (AA) and six treatments with increasing background ozone (AA+12 ppb, AA+24 ppb, AA+36 ppb, AA+48 ppb, AA+60 ppb 148 and AA+72 ppb). These were applied as a continuous, repeated, weekly regime 149 designed to simulate increased background ozone concentrations. The exposure 150 period within the solardomes was from 9th May 2008 -2nd September 2008. 151

153 Gas and water sampling and analysis:

Gas exchange samples were taken fortnightly by placing a two litre transparent, 154 plastic chamber over the mesocosms and attaching with a rubber seal between the 155 156 headspace and the outer casing of the mesocosm to ensure that the soil structure was not disturbed by the attachment of the chamber. A 30ml sample of the background 157 gas was taken at the moment of capping and a second sample of the gas within the 158 chamber was taken after one hour. The accumulation of methane and carbon dioxide 159 within the chamber was found to be linear over this time period when measured in a 160 161 preliminary experiment prior to the mesocosms being placed in the solardomes. Gas samples were stored under positive pressure in airtight glass vials (Perkin Elmer) that 162 were evacuated prior to use and analysed within 24 hours of sample collection. Gas 163 164 samples were analysed for the concentration of methane and carbon dioxide using a Perkin Elmer Gas Chromatograph (GC) fitted with a flame ionisation detector (FID) 165 to detect methane and a methaniser to convert carbon dioxide to methane. Gas 166 167 samples were pressurised with a known amount of nitrogen in the headspace autosampler (Turbo-Matrix) and samples were injected into the GC at 23.2psi with 168 nitrogen carrier gas. Samples were passed through a Poropak QS ceramic column, 169 hydrogen flow was set at 45ml min⁻¹ and airflow was set to 450 ml min⁻¹. The FID 170 (flame ionisation detector) temperature was 375°C. 171 172 Pore water samples were taken at three weekly intervals from the wetland mesocosms. Samples were filtered through a 0.45µm cellulose acetate filter immediately following 173

174 collection. Total dissolved carbon was measured using a ThermaloxTM elemental

analyser. Samples were injected over a platinum-coated, mesh catalyst. Oxygen was

176 used as the carrier gas and thermal catalytic oxidation was used to oxidise carbon

177 compounds in the sample to carbon dioxide, which was detected and measured using

a non-dispersive infra-red detector.

179

180 Plant Growth:

During the growing season visible senescence on vascular plants growing in the mesocosms was assessed at two week intervals. Vascular plant leaves were counted as senesced if more than 25% of an individual leaf had died back and the percentage of the entire plant that was senesced was calculated.

Above and below ground vascular plant and moss biomass present in the mesocosms was measured following 16 weeks of ozone exposure. Plant biomass was harvested and dried to constant mass at 65°C.

188

189 Statistical analysis:

190 Relationships between ozone exposure, plant senescence, plant biomass and methane emissions were analysed using regression analysis in R v 2.14.2. Ozone exposure is 191 192 reported as accumulated hourly mean ozone concentration over 24 hours without a threshold ozone concentration (AOT024hr). This measure incorporated the effects of 193 elevated ozone throughout the night, rather than daylight hours as is more usually 194 used. It also included the potential effects of ozone in treatments that were below 40 195 ppb, which would be omitted if the more commonly used parameter AOT40 had been 196 197 calculated. Use of AOT0_{24hr} also allowed the cumulative effect of ozone to be assessed, irrespective of the time scale of ozone exposure. Gas exchange and 198 senescence measurements are plotted grouped by cumulative ozone exposure; dome 199 200 mean values for senescence, carbon dioxide and methane fluxes measured through the 4 month ozone exposure were ordered by cumulative ozone dose and averaged by 201 202 each 20 ppm h increase in ozone exposure.

204 Results

Seasonal mean ozone concentrations ranged from 16 ppb in the lowest ozone
treatment to 94 ppb in the highest treatment, while AOT0_{24hr} and daylight AOT40

ranged from 45 to 260 ppm h and 0 to 73 ppm h respectively (Table 1).

208

209 Vascular plant species emerging or germinating in the mesocosms consisted of *Juncus* effusus, Carex echinata and small quantities of Poa triviata. Although each 210 211 mesocosm did not have the same number of plants per species, there was no significant difference in the species present across the eight ozone treatments. Using 212 combined data from all ozone treatments and assessments, vascular plant senescence 213 214 on the species growing in the mesocosms showed a positive relationship (P < 0.05) 215 with increasing AOT0_{24hr} (Figure 1a), indicating senescence increased to a greater extent in the mesocosms exposed to higher doses of background ozone. Elevated 216 217 background ozone over the 16 week period caused an increase in the percentage of senesced vascular plant material from 5% in the lowest exposure to 25% in the 218 219 highest accumulated ozone exposure. There were no significant differences in senescence seen in the individual vascular plant species present in the mesocosms so 220 221 senescence data was pooled across all vascular plant species for analysis and 222 presentation. After 16 weeks of ozone exposure there was a significant relationship between ozone exposure and senescence, with the mesocosms exposed to higher 223 ozone showing higher vascular plant senescence (Figure 1b). In contrast, there was 224 225 no significant effect of ozone on vascular plant cover, above or below ground vascular plant biomass (for all biomass combined and for individual species present in each 226 mesocosm) or Sphagnum spp. moss biomass (Table 2). 227

228	Methane fluxes showed an inverse polynomial relationship with accumulated ozone
229	exposure. At low to moderate AOT 0_{24hr} values (ranging from $0 - 120$ ppm h)
230	methane fluxes increased as accumulated ozone exposure increased, whereas from
231	AOT $0_{24 hr}$ values of 120 ppm h to 220 ppm h methane fluxes decreased as AOT $0_{24 hr}$
232	values increased (Figure 2). When methane fluxes after 16 weeks of ozone exposure
233	were correlated with vascular plant senescence, vascular plant biomass and moss
234	biomass there was no correlation between methane fluxes and above or below
235	vascular plant biomass, vascular plant cover or moss biomass (Table 3). However,
236	there was a trend towards a significant correlation ($P = 0.08$) between vascular plant
237	senescence and methane fluxes (Table 3). Carbon dioxide fluxes did not show a
238	statistically significant change with increasing exposure to ozone and showed high
239	levels of variability within mesocosms exposed to similar levels of ozone, though
240	mesocosms showed a net uptake of carbon dioxide throughout the exposure period
241	(data not presented).
242	Dissolved organic carbon within the pore waters of the wetland mesocosms showed
243	no relationship with increasing exposure to elevated background ozone, remaining
244	unchanged over time and as ozone exposure increased (data not presented).
245	
246	Discussion
247	This experiment has shown that elevating the background ozone throughout the
248	growing season increases vascular plant senescence and changes methane fluxes from
249	wetlands, although plant biomass, carbon dioxide fluxes and dissolved organic carbon

250 concentrations were unchanged.

The increase in vascular plant senescence caused by exposure to elevated ozone 252 agrees with published results showing that wetland plants were sensitive to mean 253 daily peak concentrations of ozone of 77 ppb, 80 ppb and 150 ppb respectively 254 255 (Franzaring et al., 2000, Power & Ashmore, 2002, Williamson et al., 2010). In this study the linear relationship between AOT 024 hr indicates that vascular plants exposed 256 to lower concentrations of ozone for a longer time period showed similar levels of 257 258 senescence to those exposed to higher ozone concentrations for shorter time periods at any given value of AOT 0_{24 hr}. This suggests that a growing season with moderately 259 260 high background tropospheric ozone could be as detrimental to plant health as one where there are a small number of high peaks in tropospheric ozone. 261 Above and below ground vascular plant biomass was unaffected by increasing 262 263 exposure to ozone, a finding that agrees with published results from Toet et al (2011) who found that vascular plant biomass in wetland mesocosms was unaffected by two 264 years exposure to elevated ozone. However, this is in contrast to studies on other 265 266 semi-natural vegetation types where increasing ozone exposure reduced vascular plant biomass with examples from grasslands (Barbo et al., 1998; Hayes et al., 2006; Ramo 267 et al., 2006; Ramo et al., 2007) and trees (Paakkonen et al., 1996; Saleem et al., 2001). 268 Similarly to the vascular plants in this study Sphagnum moss biomass was unaffected 269 by elevated ozone exposure, which agrees with previous studies showing that the 270 271 majority of *Sphagnum* species are relatively tolerant to ozone (Morsky et al., 2011, Toet et al., 2011). In addition, as carbon dioxide fluxes were unchanged by increasing 272 ozone exposure, this suggests that ozone exposure does not have a significant impact 273 274 on wetland plant photosynthesis and respiration. Methane emissions increased with increasing AOT024hr to a maximum at 275 approximately 120 ppm h and then decreased as AOT024hr continued to increase. It is 276

possible that effects of ozone on methane fluxes seen in published papers, ranging 277 from large increases through to decreases in methane emissions, are due to differences 278 in cumulative ozone exposure between experiments taking into account the ambient 279 ozone concentrations at the experimental sites used. The results of Lloyd (2004) 280 showed large increases in methane emission but the cumulative ozone exposure of the 281 highest treatment was an AOT024hr of 120 ppm h which coincides with the peak in 282 methane emissions shown in this experiment. The positive effects of ozone on 283 methane emissions shown by Niemi et al., (2002) occurred at an estimated cumulative 284 285 ozone exposure of up to 48.8 ppm h in their highest ozone treatment, which, when compared with an AOT0_{24hr} of 9.7 ppm h in their control exposure would be in the 286 range of the results in this experiment that show an increase in methane. The results 287 288 of Morsky et al., (2008) and Toet et al., (2011), showing a decrease in methane 289 emissions after exposing peatland mesocosms to increases in ozone above current ambient concentrations of tropospheric ozone, potentially coincided with the 290 291 decreasing methane phase of the relationship found in our experiment. Using ozone concentrations measured at the Snowdonia ozone monitoring site at Marchlyn Mawr 292 over a four month summer growing season the average ozone exposure ($AOT0_{24hr}$) 293 between 2006 and 2010 was 101 ppm h, a value that is close to the ozone exposure 294 corresponding to the highest methane emissions seen in our study. This could be one 295 296 possible explanation for the decrease in methane emissions measured here and in other studies under ozone concentrations above ambient levels, although it should be 297 remembered that the different studies took place in very different locations with 298 299 different ambient ozone characteristics. The increase in methane fluxes from peatlands seen following exposure to low to 300

301 moderate accumulated levels of ozone in this study could either be due to an increase

in methanogenic activity, a decrease in methanotropic activity or an increase in the 302 release of methane through the aerenchyma of the vascular plants. Toet et al., (2009) 303 showed that ozone does not diffuse more than a few millimetres into the substrate, 304 particularly in waterlogged soils, suggesting that it is unlikely that the change in 305 methane fluxes seen is a result of the direct impact of ozone on methanogenic or 306 methanotrophic bacteria. This is corroborated by results from Morsky et al., (2008) 307 308 and Rinnan et al., (2003) showing that exposure to elevated ozone had no impact on potential methane production or consumption in peat taken from mesocosms. 309 310 Wetland vascular plants show two main types of gas exchange between the atmosphere and their roots: passive diffusion and active pressurised flow (Brix et al., 311 1992, Whiting & Chanton, 1993) and the major flow mechanism differs between 312 313 species (Roura-Carol & Freeman, 1999, Thomas et al., 1996, Van der Nat et al., 1998). Passive molecular diffusion occurs as a result of a concentration gradient 314 between the methane within the substrate and the atmosphere. If this were the 315 316 dominant gas exchange mechanism then an increase in methane production would have to occur for methane emissions to increase. DOC concentrations were 317 unchanged by exposure to elevated ozone, which suggests that substrate availability to 318 methanogens is not increasing. However, active pressurised flow occurs because of a 319 pressure differential developing between green, photosynthesising leaves and older, 320 senescing leaves (Chanton & Whiting, 1996, Chanton et al., 1997, Chanton et al., 321 1993, Shannon et al., 1996, Yavitt & Knapp, 1998) forcing the flow of methane into 322 the roots, through the aerenchyma and out through inter-cellular pore spaces in 323 324 senesced leaves. As the senesced leaf area was increased by elevated ozone, then the lower pressure "leaky" leaf area is increased, thus there may be more gas flow 325 through plants resulting in more methane being transported from the peat to the 326

atmosphere. Senesced leaves are also unlikely to show any stomatal control and it has 327 been shown that methane emissions from *Carex* species are partially under stomatal 328 control (Morrissey et al., 1993), which suggests that this could also be a factor behind 329 330 the increasing methane emissions seen in this experiment as many of the mesocosms were dominated by Carex echinata. A further indication that the influence of elevated 331 ozone on methane fluxes may be under stomatal control comes from Mills et al., 332 (2009) and Wagg et al., (2012) who showed that grassland plants exposed to elevated 333 ozone lost their usual response to ABA and no longer had stomatal control when 334 335 exposed to extreme drought. The natural variation in species cover and composition of the mesocosms means that further work would be required to further explore this 336 relationship and test the hypothesis that the change in methane emissions seen 337 338 following exposure to elevated background ozone is because of increased senescence 339 present on aerenchymatous vascular plants. As a recent review by Rinnan et al., (2013) concludes; methane and carbon dioxide 340 341 fluxes from peatland systems are under the control of many different factors including temperature, water table height and fluctuation and light availability meaning that 342 there are many ways the impact of elevated tropospheric ozone on these carbon gas 343 fluxes may be masked. This study has provided a potential explanation for the 344 345 seemingly contradictory methane emissions from previous studies but further 346 investigation of the interactions between the factors that affect methane emission would increase our understanding of the effect of ozone exposure on methane 347 emissions. We have shown that peatland ecosystems have the potential to be changed 348 349 by relatively low accumulations of tropospheric ozone, when considered over the period of a growing season. Increases in plant senescence may affect the long-term 350 viability of sensitive peatland plants, and, although biomass was unaffected by 351

elevated background ozone over one growth season, it is possible that the increased
resources needed by the plants to replenish damaged tissue may have longer-term
implications for plant health and carbon sequestration by wetlands.

355

356 Conclusions:

Our results have shown that peatland plants are sensitive to increasing background 357 ozone concentrations, which adds new knowledge to previous published work 358 showing that peaks in tropospheric ozone also damage wetland plants. The gas 359 360 exchange measurements made during this study suggest that methane fluxes from wetlands can be very sensitive to relatively small changes in background ozone 361 concentrations, and that the results from five separate studies on the impacts of 362 363 elevated ozone on methane fluxes fit within the pattern found in our study. We hypothesize that there is a relationship between plant senescence resulting in changes 364 in methane fluxes (P = 0.08 in our study). Further work carrying out more intensive 365 gas exchange sampling from peatlands would indicate whether the effects we have 366 seen occur on a wider scale. Further understanding of the mechanism for how 367 changes in plant growth following ozone exposure are resulting in changes in methane 368 fluxes and the interactions between the factors that affect methane emission and 369 tropospheric ozone exposure are needed to fully assess the implications of 370 371 tropospheric ozone increases for global greenhouse gas budgets. 372 Acknowledgements: 373 374 Thanks to Steve Hughes and Aled Williams for their help and support. This work was

funded by a NERC PhD studentship (project number C03028).

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Figure 1a: Vascular plant senescence against AOT0 ppm h through the 16 week experimental period. P < 0.05, $R^2 = 0.509$, F stat = 8.30. Error bars show the standard error of the mean for each data point with AOT0 meaned per 20 ppm h intervals.



Figure 1b: Vascular plant senescence at the end of the 16 weeks of ozone exposure period. See Table 2 for statistical relationships.



Figure 2: Methane flux plotted against AOT0 ppm h accumulated throughout the 16 week experimental period. P < 0.01, $R^2 = 0.719$, F stat = 8.93. Values are shown as the mean \pm SE and where SE bars are not present variation was within the size of the points on the plot.