1 2	Title:	Global REACH 2018: Renal oxygen delivery is maintained during early acclimatization to 4330 m	
3	Running Hea	ad: Renal oxygen delivery at high-altitude (4330 m)	
4	Journal:	American Journal of Physiology – Renal Physiology	
5 6 7	Authors:	Andrew R Steele <sup>1</sup> , Michael M Tymko <sup>1</sup> , Victoria L Meah <sup>1,7,8</sup> , Lydia L Simpson <sup>2</sup> , Christopher Gasho <sup>3</sup> , Tony G Dawkins <sup>4</sup> , Francisco C Villafuerte <sup>5</sup> , Philip N Ainslie <sup>6</sup> , Michael Stembridge <sup>4</sup> , Jonathan P Moore <sup>2</sup> , Craig D Steinback <sup>1,7,8,9</sup>	
8 9	Affiliations:	<sup>1</sup> Neurovascular Health Lab, Faculty of Kinesiology, Sport, & Recreation, University of Alberta, Canada;	
10 11		<sup>2</sup> Extremes Research Group, School of Sport, Health and Exercise Sciences, Bangor University, Bangor, UK;	
12 13		<sup>3</sup> Division of Pulmonary and Critical Care, School of Medicine, Loma Linda University, Loma Linda, CA, USA;	
14 15		<sup>4</sup> Cardiff School of Sport and Health Sciences, Cardiff Metropolitan University, Cardiff, UK;	
16 17		<sup>5</sup> Department of Biological and Physiological Sciences, Universidad Peruana Cayetano Heredia, Lima, Peru;	
18 19		<sup>6</sup> Centre for Heart, Lung, and Vascular Health, University of British Columbia Okanagan, Kelowna, Canada;	
20 21		<sup>7</sup> Women and Children's Health Research Institute, University of Alberta, Canada; <sup>8</sup> Alberta Diabetes Institute, University of Alberta, Canada;	
22		<sup>9</sup> Neuroscience and Mental Health Institute, University of Alberta, Canada.	
23			
24	Corresponde	ence:	
25 26 27 28 29 30 31 32 33 34	Craig D. Steinback, PhD Associate Professor Faculty of Kinesiology, Sport, and Recreation University of Alberta 1-059D Li Ka Shing Centre for Health Research Innovation Edmonton, Alberta, Canada T6G 2E1 Tel: (780)492-5553 Fax: (780) 492-4249		
36	Number of F	igures: 2	
37	Number of Table: 4		

#### Word Count: 6564

#### Abstract:

38

39

Early acclimatization to high-altitude is characterized by various respiratory, hematological, and 40 41 cardiovascular adaptations that serve to restore oxygen delivery to tissue. However, less is understood about renal function and the role of renal oxygen delivery (RDO<sub>2</sub>) during high-42 altitude acclimatization. We hypothesized that: 1) RDO2 would be reduced after 12-hours of 43 high-altitude exposure (high-altitude day1) but restored to sea-level values after one-week (high-44 altitude day7); and 2) RDO<sub>2</sub> would be associated with renal reactivity (RR), an index of acid-45 base compensation at high-altitude. Twenty-four healthy lowlander participants were tested at 46 sea-level (344m; Kelowna, Canada), on day1 and day7 at high-altitude (4330m; Cerro de Pasco, 47 Peru). Cardiac output, renal blood flow, arterial and venous blood sampling for renin-48 angiotensin-aldosterone-system hormones and NT pro-B type natriuretic peptides were collected 49 50 at each time point. RR was calculated as: ( $\Delta$  arterial bicarbonate)/( $\Delta$  partial pressure of arterial carbon dioxide) between sea-level and high-altitude day1, and sea-level and high-altitude day7. 51 The main findings were: 1) RDO<sub>2</sub> was initially decreased at high-altitude compared to sea-level 52 (ΔRDO<sub>2</sub>: -22±17%, P<0.001), but was restored to sea-level values on high-altitude day7 53  $(\Delta RDO_2: -6\pm14\%, P=0.36)$ . The observed improvements in RDO<sub>2</sub> resulted from both changes in 54 renal blood flow ( $\Delta$  from high-altitude day1: +12±11%; P=0.008), and arterial oxygen content ( $\Delta$ 55 from high-altitude day1 +44.8 $\pm$ 17.7%; P=0.006); and 2) RR was positively correlated with 56 RDO<sub>2</sub> on high-altitude day7 (r=0.70; P<0.001), but not high-altitude day1 (r=0.26; P=0.29). 57 These findings characterize the temporal responses of renal function during early high-altitude 58 acclimatization, and the influence of RDO<sub>2</sub> in the regulation of acid-base. 59

#### **Introduction:**

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

75

76

77

78

79

80

81

82

High-altitude acclimatization is characterized by varying elevations in ventilation, hemoglobin concentration, heart rate, and redistribution of blood flow, which serves to restore arterial oxygen content (CaO<sub>2</sub>) and preserve oxygen delivery to vital organs (10, 13, 44, 53, 56). Alterations in renal function are also critical during high-altitude acclimatization; however, there are few studies exploring renal acclimatization in comparison to ventilatory and hematological factors (4, 10, 22, 43). This is noteworthy since there are unique characteristics of renal oxygenation that renders the kidney susceptible to hypoxia. For example, the partial pressure of oxygen (PO<sub>2</sub>) in renal tissue is typically tightly controlled through a coupling between renal blood flow (i.e. oxygen delivery) and sodium reabsorption load (i.e. oxygen utilization) (31). Hence, unlike other tissues, greater renal blood flow does not necessarily influence renal oxygenation since renal oxygen consumption (i.e. metabolic rate) can rapidly adapt to maintain constant oxygen delivery (31). Furthermore, portions of the medulla have a tissue PO<sub>2</sub> of ~10-15 mmHg, which is near the "critical PO<sub>2</sub>", which the enzyme mitochondrial cytochrome oxidase becomes reduced, in turn limiting adenosine triphosphate production (25, 30). This, coupled with the fact that 95-99% of renal energy is via oxidative phosphorylation (30), highlights the importance of controlled renal oxygen delivery (RDO<sub>2</sub>) for normal kidney function (7). Despite the kidney's precise maintenance of RDO2 in normoxia, RDO2 has not been quantified during early acclimatization to severe hypoxia (e.g. >4000 m).

While data sets are limited, renal blood flow (as indexed via the effective renal plasma flow) appears to decrease following acute (48-hours) exposure to 4350 m (37) and following a 60 day stay at 3500 m (43). Only two studies to our knowledge (37, 38) have investigated the mechanism(s) regulating renal blood flow at high-altitude. Olsen and colleagues (37) reported a

reduction in effective renal blood flow indicating a pre-existing increase in renal vascular tone. The authors from this study speculated that elevated catecholamines (e.g. noradrenaline), were responsible for the observed renal vasoconstriction and consequential reduction in renal blood flow. However, systemic hypoxia stimulates numerous factors that independently influence renal blood flow control such as natriuretic peptides (5) and RAAS hormones (6, 13). Specifically, the influence of renin during hypoxia has been unclear: some studies have reported that renin is elevated (12, 35, 40) while others have documented no change (3, 45) and others still have reported a decrease (6, 37). To our knowledge, no study has investigated the integrative mechanisms controlling renal blood flow at high-altitude following 12-hours and a week of acclimatization.

Under normal oxygen conditions (e.g. sea-level), arterial oxygen content (CaO<sub>2</sub>) is relatively stable and RDO<sub>2</sub> is directly related to renal blood flow (31). However, since early exposure to high-altitude decreases both renal blood flow (37), and CaO<sub>2</sub> (43), this may hence effectively decrease RDO<sub>2</sub>. Yet, RDO<sub>2</sub> may return to sea-level values as CaO<sub>2</sub> becomes restored with acclimatization (19, 43), which may offset the high-altitude related reductions in renal blood flow, and mediates the reduction in RDO<sub>2</sub> as demonstrated in animal studies (48). To our knowledge no study has characterized RDO<sub>2</sub> during acclimatization to high-altitude in humans. The contributions of renal blood flow and CaO<sub>2</sub> on RDO<sub>2</sub> has yet to be characterized at high-altitude, and may have functional consequences on acid-base acclimatization (18, 49).

The tight regulation of blood pH is critical for homeostasis and regular cellular function. High-altitude driven hyperventilation decreases the partial pressure of arterial carbon dioxide (PaCO<sub>2</sub>), resulting in respiratory alkalosis (10, 47). Renal acid (H<sup>+</sup>) retention and bicarbonate (HCO<sub>3</sub>)<sup>-</sup> excretion aims to normalize arterial pH towards standard sea-level values (pH  $\sim$  7.4) (9,

13, 28, 43, 45). A recent study by Zouboules and colleagues (58) proposed a novel renal reactivity index ( $\Delta[HCO_3^-]/\Delta$  [PaCO<sub>2</sub>]), to quantify the relationship between HCO<sub>3</sub><sup>-</sup> and PaCO<sub>2</sub> at high-altitude (58). While bicarbonate excretion occurs to normalize pH as a function of PaCO<sub>2</sub>, this response might be linked to RDO<sub>2</sub>, since bicarbonate reabsorption is dependent on renal cortex tissue PO<sub>2</sub> in hypoxic animals (49).

The purpose of the current investigation was to assess the mechanism(s) that govern renal oxygen delivery during early high-altitude acclimatization. We hypothesized the following: 1) after rapid ascent from sea-level to high-altitude (4330 m), RDO<sub>2</sub> would decrease 12-hours after arrival, but thereafter following a week of acclimatization, RDO<sub>2</sub> would increase through an increase in arterial oxygen delivery rather than renal blood flow; and 2) an association between RDO<sub>2</sub> and renal reactivity would be present after 12-hours and one week of high-altitude acclimatization.

#### **Methods:**

# Ethical Approval

This *a priori* study was conducted as part of the Global Research Expedition on Altitude-Related Chronic Health (REACH) expedition to Instituto de Investigacions de Altura at Cerro de Pasco, Peru (4330 m). Participants were researchers involved in the expedition and as such were in numerous studies; however, care was taken to ensure adequate washout between studies to avoid cross-over or contamination between investigations. An overview of our research team's expedition has been previously published (50).

This study abided by the Canadian Government Tri-council Policy on Research Ethics Policy Statement (TCPS2) and the Declaration of Helsinki, apart from registration in a publicly accessible database. Ethical approval was obtained in advanced through the Clinical Research Ethics Board of the University of British Columbia (H17-02687 and H18-01404), the University of Alberta Biomedical Ethics 100 Board (Pro00077330) and the Universidad Peruana Cayetano Heredia Comité de Ética (no. 101686). Participants were given in-depth study information and provided written consent.

# **Participants**

Participants (20 males, 4 females) were recruited from the research expedition team and had no history of pre-existing neurological, cardiovascular or renal dysfunction prior to testing. Participants were born and lived at or near sea-level and had not traveled to high-altitude within 6 months prior to experimentation (50).

# Experimental overview

Sea-level testing occurred at the University of British Columbia – Okanagan Campus, BC (altitude = ~344 m) ~three months prior to departure to high-altitude. The research team travelled to Lima, Peru (altitude = ~150m) in June 2018, spent three days in Lima before the expedition preparing to depart and then traveled via automobile directly to Cerro de Pasco, Peru (4330 m) over 6-8 hours. Participants were tested the morning immediately following ascent having spent ~12-hours at high-altitude (high-altitude day1), and again following seven days of acclimatization (high-altitude day7).

At both sea-level and high-altitude participants arrived at the laboratory between 0600 and 1030 following a 12-hour fast and having avoided caffeine, alcohol and exercise. Throughout the week of acclimatization, participants were asked to avoid exercise to not contaminate results. Participants were asked to complete a nine-hour urinary collection from the previous night, which was used to calculate glomerular filtration rate. Participants were asked to complete an acute mountain sickness questionnaire at high-altitude prior to testing on both high-altitude day1 and high-altitude day7 (i.e. Lake Louise Questionnaire) (41). Experimentation commenced with participants laying supine and resting quietly for ~ten-minutes prior to collecting measurements of renal blood flow via duplex ultrasound, venous blood samples, echocardiography, and radial artery blood samples were taken. These methodologies are discussed in further detail below.

Acute mountain sickness was identified using the standard 2018 Lake Louis acute mountain sickness scoring system. As per the recommendations, the scoring system was not used until at least six hours prior of ascent. Acute mountain sickness is identified via four categories: headache, gastrointestinal symptoms, fatigue and/or weakness and dizziness/light-headedness with each category with a score between 0-3. Acute mountain sickness is diagnosed as a score of three with an associated headache. As per the guidelines, participants with mild symptoms of acute mountain sickness had scores between 3-5; moderate between 6-9 and severe 10-12 points [refer to (41) for more details].

#### Heart rate and blood pressure

Continuous heart rate (electrocardiogram Lead II) was recorded and integrated with a data acquisition system (Powerlab 16/30; ADInstruments, Australia) and stored for subsequent analysis using associated software (Labchart 8.0 Pro; ADInstruments, Australia). Systolic and diastolic blood pressures were measured using an automated cuff (Omron M2 Classic; Japan). Mean arterial pressure was subsequently calculated as: (1/3 x systolic blood pressure) + (2/3 x diastolic blood pressure). Arterial oxygen saturation was estimated by pulse oximetry (N-595; Nellcor Oximax, Boulder, USA) using an index finger sensor.

#### **Blood** measures

Venous blood samples were taken from the antecubital vein immediately centrifuged, aliquoted and frozen until analysis in Edmonton, Alberta, Canada. Frozen samples were transported by a commercial company (Marken, New York, USA). Plasma aldosterone

concentration (LDN REF: MS E-5200) and active renin (LDN REF: MS E-5300) were measured using a solid phase enzyme-linked immunosorbent assay (ELISA), based on the principle of competitive binding. N-terminal pro-B-type natriuretic peptide (NT pro-BNP) (R&D systems REF: DY3604-05) was quantified using a sandwich solid phase ELISA.

Radial artery blood samples were collected using a lithium heparin-coated auto fill syringe and analyzed using point-of-care device i-STAT (Abbott Laboratories, Chicago, USA) for blood gases using the CG4+ (lactate, pH, PaCO<sub>2</sub>, arterial partial pressure of oxygen (PaO<sub>2</sub>), HCO<sub>3</sub> and oxygen saturation (SaO<sub>2</sub>)), and CHEM8+ (glucose, urea nitrogen, creatinine, sodium, potassium, chloride, ionized calcium, TCO<sub>2</sub>, anion gap, hematocrit and hemoglobin) test cartridges. The point of care device, i-STAT, has been validated on altitude up to 5043 meters (32).

CaO<sub>2</sub> was calculated with measures of oxygen saturation (SaO<sub>2</sub>), [Hb] and arterial partial pressure of oxygen (mmHg) using the following formula:

Equation 1:

$$CaO_2 \text{ (ml dl}^{-1}) = \left( [Hb] \times 1.36 \times \frac{SaO_2}{100} \right) + \left( 0.003 \times PaO_2 \right)$$

where [Hb] is the concentration of hemoglobin (g dL<sup>-1</sup>), 1.36 is the affinity of oxygen to hemoglobin, SaO<sub>2</sub>, is the percentage of hemoglobin saturated with oxygen, 0.003 is the fraction of free oxygen dissolved in the blood.

Renal reactivity on high-altitude day1 and high-altitude day7 was calculated using relative changes with respect to sea-level values as previously described (58).

*Equation 2:* 

Renal Reactivity = 
$$\left(\frac{\Delta HCO_3}{\Delta PaCO_2}\right) = \left(\frac{\left(HCO_3\right)_{altitude} - \left(HCO_3\right)_{sea-level}}{\left(PaCO_2\right)_{altitude} - \left(PaCO_2\right)_{sea-level}}\right)$$

Where HCO<sub>3</sub><sup>-</sup> is arterial bicarbonate (mmol L<sup>-1</sup>) and PaCO<sub>2</sub> (mmHg) is partial pressure of arterial carbon dioxide

#### *Transthoracic echocardiography*

Echocardiography was performed using an ultrasound system (as above) and a phased-array transducer (1.5 – 3.6 MHz M4S-RS, GE Healthcare, Piscataway, NJ, USA) by the same sonographer (V.L.M.). A three-lead electrocardiograph was attached to the participant and connected to the ultrasound system to allow cardiac cycle gating. Images were acquired at end expiration over five cardiac cycles and data was stored for later offline analysis (EchoPAC, GE Medical, Horton, Norway). Measurements were made in triplicate from different cardiac cycles and averaged for use in statistical analyses. With the participant lying supine, subcostal images were acquired for assessment of inferior vena cava diameter. With the participant in the left lateral decubitus position, images were acquired for assessment of cardiac function according to current guidelines (29). Left ventricular stroke volume using end-diastolic and end-systolic volume that, were derived using the Simpson's biplane method from apical 4- and 2-chamber views. Cardiac output was calculated as stroke volume x heart rate. Total peripheral resistance was calculated as: mean arterial pressure (mmHg) / mean cardiac output (ml/min).

#### Renal function

#### Duplex ultrasound

Renal artery diameter and blood flow were measured with a convex-array transducer (2.0 – 5.5 MHz 4C-RS Probe, GE Healthcare, Piscataway, NJ, USA) on a commercially available ultrasound system (Vivid Q, GE Healthcare, Piscataway, NJ, USA) by a single trained sonographer (V.L.M). The probe was placed at the midpoint between the xiphoid process and the umbilicus where the aorta was identified in a transverse section and the origin of the renal arteries was obtained using B-mode. Images were collected for measurement of renal artery diameter and allowing subsequent calculation of cross-sectional area. Renal artery blood flow was calculated as the product of the cross-sectional area and the velocity-time integral (pulse-wave Doppler). Absolute renal blood flow and normalized renal blood flow ([renal blood flow / cardiac output] \*100) are reported. Renal vascular resistance was calculated as: mean arterial pressure (mmHg) / mean renal blood flow (ml/min). All measurements were made in triplicate from different cardiac cycles and averaged for use in statistical analyses.

The product of renal blood flow (ml min<sup>-1</sup>) and CaO<sub>2</sub> (ml dl<sup>-1</sup>) was used to calculate convective RDO<sub>2</sub>:

Equation 3:

238 RDO<sub>2</sub> (ml O<sub>2</sub> min<sup>-1</sup>)= 
$$\frac{\text{(mean renal blood flow} \times CaO2)}{\text{(100)}}$$

Urine collection and analysis

Participants were asked to complete a 9-hour urinary collection to calculate glomerular filtration rate. Due to limitations associated with conducting field research, were unable to control for salt and fluid intake. Participants were asked to maintain normal drinking habitats

throughout the week of high-altitude and we specifically requested participants to drink a standardized 200 mL of water forty-five minutes before testing. Urine was refrigerated until analysis (4 °C). Urine pots were shaken vigorously before analysis to ensure a homogenous mixture. Volumes were measured using graduated cylinders. Urine analysis was performed using a DCA Vantage Analyzer (Siemens Healthineers Global; Germany) for creatinine and microalbumin. Creatinine clearance was used to calculate glomerular filtration rate using the standard formula:

Equation 4:

Glomerular filtration rate (ml/min/1.73m<sup>2</sup>)=
$$\frac{(Ux)\times(\dot{V})}{(Px)}$$

Where Ux is urine creatinine concentration (mol  $L^{-1}$ ),  $\dot{V}$  is urine production rate (ml min<sup>-1</sup>) and Px is serum creatinine concentration (mol  $L^{-1}$ ). Glomerular filtration rate was then scaled to body surface area as determined through the Dubois and Dubois formula (8).

Filtration fraction was calculated using the following:

*Equation 5:* 

Filtraction Fraction (%)= 
$$\frac{\text{(glomerular filtration rate)}}{\text{(1-hematocrit)} \times \text{(mean renal blood flow)}}$$

Where the ratio between glomerular filtration (ml/min/1.73m²) and renal plasma blood flow (renal blood flow [ml min⁻¹] × 1- hematocrit [%]) is expressed as a percent.

### Muscle sympathetic nerve activity

261

262

263

264

265

266

267

268

269

270

271

272

273

274

275

276

277

278

279

Muscle sympathetic nerve activity was measured in subset of individuals who participated in an associated study on high-altitude day7. As such basal muscle sympathetic nerve activity data and characteristics have been previously reported in nine participants (n=9; 7 males; age  $25 \pm 3$  years and BMI  $23 \pm 2$ ) (42); however, these data are presented in the current manuscript to provide novel insight related to sympathetic-mediated mechanisms governing renal blood flow at high-altitude. Muscle sympathetic nerve activity was recorded from the peroneal nerve using microneurography as previously described (42). The raw sympathetic signal was amplified (preamplifier 1000× and variable-gain, isolated amplifier (10000×), bandpass filtered (700–2000 Hz), rectified and integrated (Bio amp 16/30; ADInstruments, Australia). A suitable signal was confirmed by the presence of pulse-synchronous bursts of activity that increased in response to apnea, but not loud noise. Muscle sympathetic nerve activity raw and integrated signals were analysed using semi-automated peak detection algorithms (Labchart 8.0 Pro; ADInstruments, Australia) and bursts were confirmed by a trained observer (L.L.S. and C.D.S.) based on physiological principles. To account for differences in microelectrode position, burst amplitude data were normalized by assigning a value of 100 to the largest burst observed. Mean muscle sympathetic nerve activity was expressed as integrated burst frequency (burst min 1), incidence (burst 100 cardiac cycles<sup>-1</sup>) and total activity [mean burst amplitude × burst frequency (a.u.·min<sup>-1</sup>)].

280

281

#### Data and statistical analyses

Data was assessed for normality and variance using the Sharpiro-Wilk and the Bron-Forsythe test. A linear mixed-effect model analysis was performed to test for significance between sea-level vs. high-altitude day1 vs. high-altitude day7. Tukey post-hoc analyses were used if main effects existed. Acute mountain sickness scores were assessed using paired nonparametric tests (Wilcoxon signed-rank test). Pearson product moment correlations were used to assess associations between:  $\Delta$  RDO<sub>2</sub> and glomerular filtration rate / renal reactivity; and renal blood flow and muscle sympathetic nerve activity. Statistical analyses were performed using Graph Pad, Prism 8.3.0. All reported data is presented as the mean  $\pm$  SD with statistical significance set at p < 0.05

#### 291 Results:

# **Participants**

Participant demographics are presented in *table 1*. Twenty-four participants were recruited but only twenty-two full data sets across all three assessments were obtained because two participants (both male) did not complete measurements on high-altitude day7 due to unexpected departure back to Lima, Peru. The values for these two participants at sea-level and high-altitude day1 are included in the group analysis. Thirteen of the twenty-four participants had mild acute mountain sickness (Lake Louise scores between 3-5) (41) on high-altitude day1. All participants refrained from taking acetazolamide (i.e. diamox), and other medications for altitude (e.g. dexamethasone) or travel-related illness (anti-biotics). No participants experienced acute mountain sickness on high-altitude day7.

#### Blood gas changes with high-altitude

High-altitude caused an initial decrease in both  $PaO_2$  and  $SaO_2$  that improved on high-altitude day7 (*table 2*).  $PaCO_2$  decreased longitudinally with high-altitude, while  $HCO_3^-$  was progressively decreased (*table 2*). Respiratory alkalosis developed on high-altitude day1 (P<0.001); there was partial correction to pH via renal compensation on high-altitude day7 (*table 2*).  $CaO_2$  decreased initially with high-altitude (P<0.001) but improved to pre-altitude values on high-altitude day7 (P=0.31) through increases in  $PaO_2$ ,  $SaO_2$  and hemoglobin concentration (*figure 1B*).

high-altitude day1 (P=0.018), but not high-altitude day7 (P=0.62; table 3).

but systolic and mean arterial pressure remained unchanged (P=0.30 P=0.098, respectfully table

3) Cardiac output increased on high-altitude day1 compared to sea-level (P<0.001), but fell to

sea-level values on high-altitude day7 (P=0.67; table 3). Total peripheral resistance decreased on

returned to sea-level values on high-altitude day7 (P=0.54; figure 1A). Accordingly, renal

vascular resistance was increased on high-altitude day1 (P=0.016), but not high-altitude day7

(P=0.76; table 4). RDO<sub>2</sub> was decreased by -22±17% on high-altitude day1 (P<0.001), due to a

simultaneous reduction in both renal blood flow and CaO<sub>2</sub> but was normalized back to sea-level

values on high-altitude day? ( $-6\pm14\%$ ) (P=0.36; figure 1C). Total normalized sympathetic nerve

activity was calculated in a subset of participants on high-altitude day7 and was negatively

correlated with renal blood flow normalized to cardiac output (r=-0.69; P=0.039; See figure

supplemental 1) (https://doi.org/10.6084/m9.figshare.12860744.v1). RAAS hormones: active

renin, and plasma aldosterone concentration, both decreased at high-altitude (P=0.025 and

Renal blood flow was decreased at high-altitude on high-altitude day1 by 17±15% but

313

312

Diastolic pressure was elevated at high-altitude compared to sea-level (P=0.0092 table 3),

314

315

316

317

318

319

320

321

322

323

324

325

326

327

328

329

330

331

332

333

level and high-altitude day7 (r=0.70; P<0.001; figure 2B) and between high-altitude day1 and

Association between renal oxygen delivery and renal reactivity

P=0.018, respectively), while NT pro-BNP did not change (P=0.15; table 4).

(P=0.0016). A positive correlation was found between  $\Delta$  RDO<sub>2</sub> and renal reactivity between sea-

Renal reactivity was increased between high-altitude day1 and high-altitude day7

high-altitude day7 (r=0.49; P=0.022; figure~2C), but not between sea-level and high-altitude day1 (r=0.26; P=0.29 figure~2A). No relationships were found between  $\Delta$  renal blood flow (r=0.10; P=0.67),  $\Delta$  CaO<sub>2</sub> (r=0.25; P=0.23), or  $\Delta$  glomerular filtration rate (r=0.15; P=0.63), and renal reactivity on high-altitude day7.

#### **Discussion:**

To our knowledge, this study is the first to assess RDO<sub>2</sub> after rapid ascent to high-altitude in a large cohort of lowlander participants whom have refrained from taking high-altitude medications (e.g. acetazolamide). The main findings were: 1) there was a reduction in RDO<sub>2</sub> on high-altitude day1; however, RDO<sub>2</sub> was restored to sea-level values on high-altitude day7 through an increase in both CaO<sub>2</sub> and renal blood flow; and 2) the relative change in RDO<sub>2</sub> at high-altitude compared to sea-level was associated with renal reactivity on high-altitude day7, indicating that acid-base regulation is linked to renal oxygenation after exposure to severe hypobaric hypoxia. Together, these data demonstrate that RDO<sub>2</sub> is normalized after a week of high-altitude acclimatization and provides novel insight on the critical role of renal adaptation and acid-base balance under hypoxic conditions.

# Renal blood flow control at high-altitude

Compared to ventilatory and hematological acclimatization responses (10, 13, 43, 55, 57), less is known on the impact of renal blood flow on high-altitude acclimatization. While short exposure to hypoxia (e.g. 20 minutes) augments renal blood flow (5, 51), this is not apparent during chronic hypoxia (1, 37, 38, 43). Renal blood flow has been reported as unchanged (38), and decreased (37), after 48-hours at 4350 m, but longer duration studies (weeks) have shown a decrease in renal blood flow (1, 43). Together, these findings indicate that the renal blood flow response to hypoxia is highly dependent on exposure time. We saw an early high-altitude renal vasoconstriction with a decreased renal blood flow, which normalized to sealevel values following a week of acclimatization. Numerous factors can influence renal blood

flow such as reactive oxygen species, RAAS, phosphodiesterase type 5 upregulation, renal sympathetic nerve activity, circulating catecholamines, natriuretic peptides and ET during hypoxia (11, 16, 34, 36, 43, 52). In this investigation, NT pro-BNP was unchanged during acclimatization. However, analyzed venous blood samples for RAAS hormones both renin activity and plasma aldosterone concentrations were decreased occurring on high-altitude day7, but not high-altitude day1. Prolonged hypoxia may depress RAAS to increase excretory function. This depression would counter the effects of increased renal vascular resistance and may explain the observed +12% increase in renal blood flow seen between high-altitude day1 and high-altitude day7 (37). The renal system may decrease renin secretion to preserve excretory function via decreased renal vascular resistance following a week of acclimatization (6, 38).

Sympathetic nerve activity may also influence renal blood flow at high-altitude (10, 37, 42). A previous study demonstrated that renal vascular vasodilation to dopamine at high-altitude (~48 hours at 4350 m) was attenuated, and plasma circulating norepinephrine concentrations were increased, indicating greater renal arteriole vasoconstriction potentially through increased adrenergic activity (37). Furthermore, a study conducted in dogs demonstrated an augmented renal blood flow response to hypoxia after kidney denervation (27), while another study conducted in conscious rabbits subjected to 0.14 and 0.10 fraction inspired oxygen content, had a 14% and 38% increase in renal sympathetic nerve activity, respectively, and congruent decreases in renal blood flow that were abolished following renal denervation (34). In the current study, we observed a significant negative relationship between total normalized muscle sympathetic nerve activity and normalized renal blood flow on high-altitude day? (See figure supplemental 1) (https://doi.org/10.6084/m9.figshare.12860744.v1). In other words, participants with greater total normalized muscle sympathetic nerve activity had lower normalized renal blood flows.

Collectively, this latter observation and previous findings (34, 37) would suggest the level of sympathetic nerve activity is an important determinant of renal blood flow during hypoxia. We acknowledge the requirement of sea-level and high-altitude day1 muscle sympathetic nerve activity data, as well as acute manipulation of sympathetic nerve activity, to draw further conclusions.

#### *RDO*<sup>2</sup> *at high-altitude*

To date, no previous studies have calculated RDO<sub>2</sub> at high-altitude in humans (48). The data from the current investigation demonstrated that only 12 hours of high-altitude exposure resulted in a concomitant decrease in renal blood flow and CaO<sub>2</sub>, resulting in a reduction in RDO<sub>2</sub> by 22%. The acute reduction in RDO<sub>2</sub> was offset by elevated CaO<sub>2</sub> and renal blood flow after 7 days of high-altitude acclimatization (*see figure 1*). We report similar findings as a previous animal study (48). Since sodium tubular load accounts for 99.5% of renal metabolic activity (14, 25, 31), renal blood flow may decrease in order to limit renal oxygen consumption, effectively preserving oxygen for other organs (20, 33). This is supported by reciprocal changes in cardiac output and renal blood flow observed where renal blood flow decreased was by 17%, while cardiac output was augmented by 20%. The limited oxygen supply is being directed away from the metabolic demanding kidneys conserving systemic oxygen (20, 33).

#### *RDO*<sup>2</sup> *and acid-base acclimatization*

There have been several previous studies that have characterized renal acid-base acclimatization at high-altitude. Renal alterations are initiated within two hours after the onset of hypocapnia,

and current data indicates incomplete pH compensation is present (metabolic alkalosis) at altitudes above 2800 m (17, 20, 28, 58). Renal reactivity, an index of acid-base compensation between  $HCO_3^-$  and  $PaCO_2$  ( $\Delta[HCO_3^-]/\Delta$  [PaCO<sub>2</sub>]), (58), has been shown to increase at altitudes up to 3800 m, and then decreases with further increases in altitude (58). In the current study, renal reactivity was greater on high-altitude day7 compared to high-altitude day1, indicating renal reactivity has a temporal component that is influenced by early acclimatization. Compared to Zouboules and colleagues (58) expedition, which was conducted at 4240 m after incremental ascent over seven days, we observed similar renal reactivity response to high-altitude. It is important to note, however, that the ascent profile used in this current study and Zouboules and colleagues (58) expedition were very different. For example, Zouboules and colleagues (58) trekked most days towards Everest basecamp where acclimatization was obviously influenced by the daily changes in altitude. In our study, we ascended via automobile to 4330 m where we resided for the duration of the study. Hence, the current study enabled the question of acclimatization to be addressed over time at the same altitude. Therefore, to address the question and to extend the data presented by Zouboules and colleagues (58), we assessed both renal reactivity and RDO2 at high-altitude, and found an association between these two physiological parameters on high-altitude day7 (see figure 2B and 2C). Interestingly, a relationship was not observed between renal reactivity and renal blood flow, CaO<sub>2</sub> or glomerular filtration rate. One interpretation of these findings is that the reduction in renal blood flow or glomerular filtration rate seen at high-altitude (37, 39, 43) does not influence the kidneys capacity to filtrate and excrete HCO<sub>3</sub><sup>-</sup> in the urine as previously hypothesized (39, 58). Conversely, this may imply RDO<sub>2</sub> influences the tubular handling of HCO<sub>3</sub> and H<sup>+</sup> (18, 49). RDO<sub>2</sub> at high-altitude may impact the activity of intracellular carbonic anhydrase (23), proton secretion via the Na<sup>+</sup>-H<sup>+</sup>

405

406

407

408

409

410

411

412

413

414

415

416

417

418

419

420

421

422

423

424

425

426

427

exchanger (NHE3) (2) and/or activity of intercalated cells on the collecting ducts (15). However, considering the known linkage between sodium and HCO<sub>3</sub><sup>-</sup> reabsorption in the proximal (18, 54), we must acknowledge that the independent influence of sodium on acid-base regulation. That is, renal reactivity and arterial HCO<sub>3</sub><sup>-</sup> may actually correlate with sodium excretion rather than changes in RDO<sub>2</sub>. We recommend that these findings be interpreted cautiously. Future endeavours should determine the influence of sodium (and other electrolyte) excretion on acid-base regulation during acclimatization.

#### Experimental limitations and considerations

The current investigation was the first to assess RDO<sub>2</sub> at high-altitude; however, there are some experimental considerations that warrant discussion. First, para-aminohippurate would provide a more specific measure of renal perfusion. However, renal ultrasound is strongly correlated to effective renal blood flow when flows are above 280 ml min<sup>-1</sup> as seen this study (46). Second, muscle sympathetic nerve activity was recorded in a subset of individuals and used as a surrogate for renal sympathetic nerve activity. We acknowledge that sympathetic vasomotor outflow to skeletal muscle vasculature may not reflect renal sympathetic nerve activity and may exhibit differential reflex responses (42). While renal sympathetic nerve activity and muscle sympathetic nerve activity are strongly correlated in animals (26), these findings should be interpreted cautiously and used to inform future research. Third, salt and fluid intake was not controlled for during testing. We acknowledge that changes in fluid and salt may have contributed to the change in renal function and renal oxygen delivery (21). However, we feel this has limited influence on our findings. Previous findings have demonstrated that high-altitude changes renal blood flow and Sprague-Dawley rats during hypobaric hypoxia have a temporal

RDO<sub>2</sub> response to our findings (48). Future endeavours should investigate this physiological phenomenon while controlling salt and fluid intake. Fourth, we did not calculate metabolic efficacy of sodium reabsorption across the proximal tubule or renal oxygen consumption. High-altitude may change both of these to maintain normoxic filtration (48). However, this should be addressed in future studies specifically looking at renal metabolic function during hypobaric hypoxia. Lastly, no comparisons were made between sexes despite knowing there is a difference in renal blood flow and RAAS regulation between men and women (24). However, since this was a repeated measures assessment comparing within individuals and females were only a small subset this should not greatly impact our findings. Future endeavours should examine the impact of sex on RDO<sub>2</sub> at high-altitude.

#### Significance and perspective

Our data characterizes renal acclimatization following 12-hours and one-week exposure to 4300 m. Renal oxygen delivery fell immediately with initial high-altitude exposure but was restored on high-altitude day7 by increases in both CaO<sub>2</sub> and renal blood flow. In addition, relative changes to RDO<sub>2</sub> from sea-level were positively correlated with renal reactivity on high-altitude day7, indicating a potential link between RDO<sub>2</sub> and acid-base compensation during high-altitude acclimatization. Together, these data demonstrate that RDO<sub>2</sub> is normalized following a week of acclimatization and may contribute to pH normalization.

- 470 1. Anand IS, Chandrashekhar Y, Rao SK, Malhotra RM, Ferrari R, Chandana J, Ramesh B,
- Shetty KJ, Boparai MS. Body fluid compartments, renal blood flow, and hormones at 6,000 m in
- 472 normal subjects. J Appl Physiol (1985). 1993;74(3):1234-9.
- 473 2. Aronson PS, Nee J, Suhm MA. Modifier role of internal H+ in activating the Na+-H+
- exchanger in renal microvillus membrane vesicles. Nature. 1982;299(5879):161-3.
- 475 3. Ashack R, Farber MO, Weinberger MH, Robertson GL, Fineberg NS, Manfredi F. Renal
- and hormonal responses to acute hypoxia in normal individuals. J Lab Clin Med.
- 477 1985;106(1):12-6.
- 478 4. Bartsch P, Pfluger N, Audetat M, Shaw S, Weidmann P, Vock P, Vetter W, Rennie D,
- Oelz O. Effects of slow ascent to 4559 M on fluid homeostasis. Aviat Space Environ Med.
- 480 1991;62(2):105-10.
- 481 5. Berger EY, Galdston M, et al. The effect of anoxic anoxia on the human kidney. J Clin
- 482 Invest. 1949;28(4):648-52.
- 483 6. Bestle MH, Olsen NV, Poulsen TD, Roach R, Fogh-Andersen N, Bie P. Prolonged
- 484 hypobaric hypoxemia attenuates vasopressin secretion and renal response to osmostimulation in
- 485 men. J Appl Physiol (1985). 2002;92(5):1911-22.
- 486 7. Bullen A, Liu ZZ, Hepokoski M, Li Y, Singh P. Renal Oxygenation and Hemodynamics
- 487 in Kidney Injury. Nephron. 2017;137(4):260-3.
- 488 8. Daugirdas JT, Meyer K, Greene T, Butler RS, Poggio ED. Scaling of measured
- 489 glomerular filtration rate in kidney donor candidates by anthropometric estimates of body surface
- area, body water, metabolic rate, or liver size. Clin J Am Soc Nephrol. 2009;4(10):1575-83.
- 491 9. de Seigneux S, Malte H, Dimke H, Frøkiaer J, Nielsen S, Frische S. Renal compensation
- 492 to chronic hypoxic hypercapnia: downregulation of pendrin and adaptation of the proximal
- 493 tubule. Am J Physiol Renal Physiol. 2007;292(4):F1256-66.
- 494 10. Dempsey JA, Powell FL, Bisgard GE, Blain GM, Poulin MJ, Smith CA. Role of
- chemoreception in cardiorespiratory acclimatization to, and deacclimatization from, hypoxia. J
- 496 Appl Physiol (1985). 2014;116(7):858-66.
- 497 11. Edwards RM. Segmental effects of norepinephrine and angiotensin II on isolated renal
- 498 microvessels. Am J Physiol. 1983;244(5):F526-34.
- 499 12. Epstein M, Saruta T. Effects of simulated high altitude on renin-aldosterone and Na
- 500 homeostasis in normal man. J Appl Physiol. 1972;33(2):204-10.
- 501 13. Forster HV, Dempsey JA, Chosy LW. Incomplete compensation of CSF [H+] in man
- during acclimatization to high altitude (48300 M). J Appl Physiol. 1975;38(6):1067-72.
- 503 14. Friederich-Persson M, Thörn E, Hansell P, Nangaku M, Levin M, Palm F. Kidney
- 504 hypoxia, attributable to increased oxygen consumption, induces nephropathy independently of
- 505 hyperglycemia and oxidative stress. Hypertension. 2013;62(5):914-9.
- 506 15. Galla JH, Rome L, Luke RG. Bicarbonate transport in collecting duct segments during
- 507 chloride-depletion alkalosis. Kidney Int. 1995;48(1):52-5.
- 508 16. Gao M, Wang R, Jiayong Z, Liu Y, Sun G. NT-ProBNP levels are moderately increased
- in acute high-altitude pulmonary edema. Exp Ther Med. 2013;5(5):1434-8.
- 510 17. Ge RL, Babb TG, Sivieri M, Resaland GK, Karlsen T, Stray-Gundersen J, Levine BD.
- 511 Urine acid-base compensation at simulated moderate altitude. High Alt Med Biol. 2006;7(1):64-
- 512 71.
- 513 18. Gibson KJ, McMullen JR, Lumbers ER, Renal acid-base and sodium handling in hypoxia
- and subsequent mild metabolic acidosis in foetal sheep. Clin Exp Pharmacol Physiol. 2000;27(1-
- 515 2):67-73.

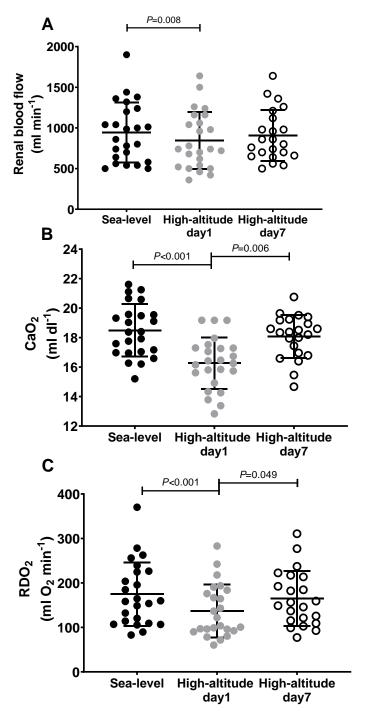
- 516 19. Goldberg S, Buhbut E, Mimouni FB, Joseph L, Picard E. Effect of moderate elevation
- above sea level on blood oxygen saturation in healthy young adults. Respiration.
- 518 2012;84(3):207-11.
- 519 20. Goldfarb-Rumyantzev AS, Alper SL. Short-term responses of the kidney to high altitude
- in mountain climbers. Nephrol Dial Transplant. 2014;29(3):497-506.
- 521 21. Graudal NA, Hubeck-Graudal T, Jurgens G. Effects of low sodium diet versus high
- sodium diet on blood pressure, renin, aldosterone, catecholamines, cholesterol, and triglyceride.
- 523 Cochrane Database Syst Rev. 2011(11):Cd004022.
- 524 22. Haditsch B, Roessler A, Krisper P, Frisch H, Hinghofer-Szalkay HG, Goswami N.
- Volume regulation and renal function at high altitude across gender. PLoS One.
- 526 2015;10(3):e0118730.
- 527 23. Hamm LL, Nakhoul N, Hering-Smith KS. Acid-Base Homeostasis. Clin J Am Soc
- 528 Nephrol. 2015;10(12):2232-42.
- 529 24. Hilliard LM, Nematbakhsh M, Kett MM, Teichman E, Sampson AK, Widdop RE, Evans
- RG, Denton KM. Gender differences in pressure-natriuresis and renal autoregulation: role of the
- Angiotensin type 2 receptor. Hypertension. 2011;57(2):275-82.
- 532 25. Johannes T, Mik EG, Nohé B, Unertl KE, Ince C. Acute decrease in renal microvascular
- PO2 during acute normovolemic hemodilution. Am J Physiol Renal Physiol. 2007;292(2):F796-
- 534 803
- 535 26. Kamiya A, Kawada T, Yamamoto K, Michikami D, Ariumi H, Miyamoto T, Uemura K,
- Sugimachi M, Sunagawa K. Muscle sympathetic nerve activity averaged over 1 minute parallels
- renal and cardiac sympathetic nerve activity in response to a forced baroreceptor pressure
- 538 change. Circulation. 2005;112(3):384-6.
- 539 27. Karim F, Poucher SM, Summerill RA. The effects of stimulating carotid chemoreceptors
- on renal haemodynamics and function in dogs. The Journal of physiology. 1987;392:451-62.
- 541 28. Krapf R, Beeler I, Hertner D, Hulter HN. Chronic respiratory alkalosis. The effect of
- sustained hyperventilation on renal regulation of acid-base equilibrium. N Engl J Med.
- 543 1991;324(20):1394-401.
- Lang RM, Badano LP, Mor-Avi V, Afilalo J, Armstrong A, Ernande L, Flachskampf FA,
- Foster E, Goldstein SA, Kuznetsova T, Lancellotti P, Muraru D, Picard MH, Rietzschel ER,
- Rudski L, Spencer KT, Tsang W, Voigt JU. Recommendations for cardiac chamber
- 547 quantification by echocardiography in adults: an update from the American Society of
- 548 Echocardiography and the European Association of Cardiovascular Imaging. J Am Soc
- 549 Echocardiogr. 2015;28(1):1-39.e14.
- 550 30. Lee CJ, Gardiner BS, Evans RG, Smith DW. Analysis of the critical determinants of
- renal medullary oxygenation. Am J Physiol Renal Physiol. 2019;317(6):F1483-f502.
- 552 31. Levy MN. Effect of variations of blood flow on renal oxygen extraction. Am J Physiol.
- 553 1960;199:13-8.
- 554 32. Lewis CT, Malein WL, Chesner I, Clarke S. High altitude arterialised capillary earlobe
- blood gas measurement using the Abbott i-STAT. J R Army Med Corps. 2018;164(5):335-7.
- 556 33. Luks AM, Johnson RJ, Swenson ER. Chronic kidney disease at high altitude. J Am Soc
- 557 Nephrol. 2008;19(12):2262-71.
- 558 34. Malpas SC, Shweta A, Anderson WP, Head GA. Functional response to graded increases
- in renal nerve activity during hypoxia in conscious rabbits. Am J Physiol. 1996;271(6 Pt
- 560 2):R1489-99.

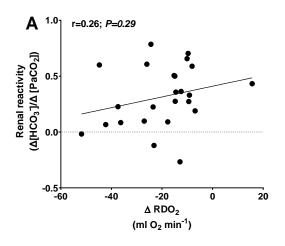
- 35. Milledge JS, Catley DM, Williams ES, Withey WR, Minty BD. Effect of prolonged
- exercise at altitude on the renin-aldosterone system. J Appl Physiol Respir Environ Exerc
- 563 Physiol. 1983;55(2):413-8.
- 36. Modesti PA, Vanni S, Morabito M, Modesti A, Marchetta M, Gamberi T, Sofi F, Savia
- 565 G, Mancia G, Gensini GF, Parati G. Role of endothelin-1 in exposure to high altitude: Acute
- Mountain Sickness and Endothelin-1 (ACME-1) study. Circulation. 2006;114(13):1410-6.
- 567 37. Olsen NV, Hansen JM, Kanstrup IL, Richalet JP, Leyssac PP. Renal hemodynamics,
- tubular function, and response to low-dose dopamine during acute hypoxia in humans. J Appl
- 569 Physiol (1985). 1993;74(5):2166-73.
- 570 38. Olsen NV, Kanstrup IL, Richalet JP, Hansen JM, Plazen G, Galen FX. Effects of acute
- 571 hypoxia on renal and endocrine function at rest and during graded exercise in hydrated subjects.
- 572 J Appl Physiol (1985). 1992;73(5):2036-43.
- 573 39. Pichler J, Risch L, Hefti U, Merz TM, Turk AJ, Bloch KE, Maggiorini M, Hess T,
- Barthelmes D, Schoch OD, Risch G, Huber AR. Glomerular filtration rate estimates decrease
- during high altitude expedition but increase with Lake Louise acute mountain sickness scores.
- 576 Acta Physiol (Oxf). 2008;192(3):443-50.
- 577 40. Ritthaler T, Schricker K, Kees F, Krämer B, Kurtz A. Acute hypoxia stimulates renin
- secretion and renin gene expression in vivo but not in vitro. Am J Physiol. 1997;272(4 Pt
- 579 2):R1105-11.
- 580 41. Roach RC, Hackett PH, Oelz O, Bärtsch P, Luks AM, MacInnis MJ, Baillie JK, Lake
- Louise AMSSCC. The 2018 Lake Louise Acute Mountain Sickness Score. High Alt Med Biol.
- 582 2018;19(1):4-6.
- 583 42. Simpson LL, Meah VL, Steele A, Thapamagar S, Gasho C, Anholm JD, Drane AL,
- Dawkins TG, Busch SA, Oliver SJ, Lawley JS, Tymko MM, Ainslie PN, Steinback CD,
- 585 Stembridge M, Moore JP. Evidence for a physiological role of pulmonary arterial baroreceptors
- in sympathetic neural activation in healthy humans. J Physiol. 2020;598(5):955-65.
- 587 43. Singh MV, Salhan AK, Rawal SB, Tyagi AK, Kumar N, Verma SS, Selvamurthy W.
- Blood gases, hematology, and renal blood flow during prolonged mountain sojourns at 3500 and
- 589 5800 m. Aviat Space Environ Med. 2003;74(5):533-6.
- 590 44. Stembridge M, Ainslie PN, Hughes MG, Stöhr EJ, Cotter JD, Nio AQ, Shave R.
- Ventricular structure, function, and mechanics at high altitude: chronic remodeling in Sherpa vs.
- short-term lowlander adaptation. J Appl Physiol (1985). 2014;117(3):334-43.
- 593 45. Swenson ER, Duncan TB, Goldberg SV, Ramirez G, Ahmad S, Schoene RB. Diuretic
- effect of acute hypoxia in humans: relationship to hypoxic ventilatory responsiveness and renal
- 595 hormones. J Appl Physiol (1985). 1995;78(2):377-83.
- 596 46. Takano R, Taniguchi N, Itoh K, Kusano E. Measurement of renal blood flow in human
- subjects using the ultrasound velocity profiling technique. J Med Ultrason (2001).
- 598 2006;33(2):91-7.
- 599 47. Teppema LJ, Dahan A. The ventilatory response to hypoxia in mammals: mechanisms,
- measurement, and analysis. Physiol Rev. 2010;90(2):675-754.
- Thron CD, Chen J, Leiter JC, Ou LC. Renovascular adaptive changes in chronic hypoxic
- 602 polycythemia. Kidney Int. 1998;54(6):2014-20.
- 603 49. Torrance SM, Wittnich C. Neonatal hemodynamic responses to extreme ranges of
- controlled graded hypoxia. Crit Care Med. 1996;24(11):1886-92.
- 50. Tymko MM, Hoiland RL, Tremblay JC, Stembridge M, Dawkins TG, Coombs GB,
- Patrician A, Howe CA, Gibbons TD, Moore JP, Simpson LL, Steinback CD, Meah VL, Stacey

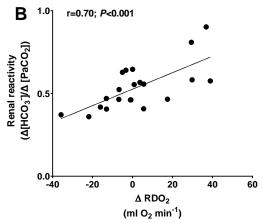
- BS, Bailey DM, MacLeod DB, Gasho C, Anholm JD, Bain AR, Lawley JS, Villafuerte FC,
- Vizcardo-Galindo G, Ainslie PN. The 2018 Global Research Expedition on Altitude Related
- 609 Chronic Health (Global REACH) to Cerro de Pasco, Peru: an Experimental Overview. Exp
- 610 Physiol. 2020.
- 51. Vidiendal Olsen N, Christensen H, Klausen T, Fogh-Andersen N, Plum I, Kanstrup IL,
- Hansen JM. Effects of hyperventilation and hypocapnic/normocapnic hypoxemia on renal
- function and lithium clearance in humans. Anesthesiology. 1998;89(6):1389-400.
- 614 52. Wallin BG, Thompson JM, Jennings GL, Esler MD. Renal noradrenaline spillover
- correlates with muscle sympathetic activity in humans. J Physiol. 1996;491 ( Pt 3)(Pt 3):881-7.
- 616 53. Wang ZY, Olson EB, Jr., Bjorling DE, Mitchell GS, Bisgard GE. Sustained hypoxia-
- 617 induced proliferation of carotid body type I cells in rats. J Appl Physiol (1985). 2008;104(3):803-
- 618 8.
- 619 54. Weinstein SW, Klose R, Szyjewicz J. Proximal tubular Na, Cl, and HCO3 reabsorption
- and renal oxygen consumption. Am J Physiol. 1984;247(1 Pt 2):F151-7.
- 621 55. Woods D, Hooper T, Hodkinson P, Ball S, Wakeford R, Peaston B, Bairsto C, Green N,
- Mellor A. Effects of altitude exposure on brain natriuretic peptide in humans. Eur J Appl
- 623 Physiol. 2011;111(11):2687-93.
- 624 56. Woods DR, Mellor A, Begley J, Stacey M, O'Hara J, Hawkins A, Yarker J, Foxen S,
- 625 Smith C, Boos C. Brain natriuretic peptide and NT-proBNP levels reflect pulmonary artery
- 626 systolic pressure in trekkers at high altitude. Physiol Res. 2013;62(6):597-603.
- 57. Young AJ, Karl JP, Berryman CE, Montain SJ, Beidleman BA, Pasiakos SM. Variability
- in human plasma volume responses during high-altitude sojourn. Physiol Rep. 2019;7(6):e14051.
- 58. Zouboules SM, Lafave HC, O'Halloran KD, Brutsaert TD, Nysten HE, Nysten CE,
- 630 Steinback CD, Sherpa MT, Day TA. Renal reactivity: acid-base compensation during
- incremental ascent to high altitude. J Physiol. 2018;596(24):6191-203.

<u>Figure 1</u>: RDO<sub>2</sub> and determinants. RDO<sub>2</sub> is acutely decreased during initial exposure to high-altitude (4330 m), however increases to sea-level thereafter at high-altitude day7 from restored renal blood flow and CaO<sub>2</sub>. Participants were tested at sea-level (Kelowna, BC 344 m), the morning immediately following ascent having spent ~12-hours at high-altitude (high-altitude day1) (Cerro de Pasco, Peru 4330m) and again following seven days of acclimatization (high-altitude day7) (Cerro de Pasco, Peru 4330m).

Figure 2: Renal reactivity and Δ RDO<sub>2</sub> at high-altitude day1, high-altitude day7 and between high-altitude day1 and high-altitude day7. While the change in renal reactivity between sea-level and high-altitude day1 was not associated with the concurrent change Δ RDO2 (A), there was a strong correlation between the changes in renal reactivity and RDO2 when considering the differences between sea-level and high-altitude day7 (B). There was also a correlation between changes renal reactivity and RDO2 during acclimatization (between high-altitude days 1 and 7) (C). Renal reactivity is higher in participants with greater RDO2 suggesting acid-base compensation is dictated by RDO2 at high-altitude. Participants were tested at sea-level (Kelowna, BC 344 m), the morning immediately following ascent having spent ~12-hours at high-altitude (high-altitude day1) (Cerro de Pasco, Peru 4330m) and again following seven days of acclimatization (high-altitude day7) (Cerro de Pasco, Peru 4330m).







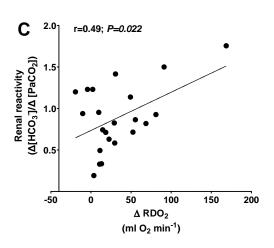


Table 1: Participant demographics and acute mountain sickness scores.

	Low altitude	High-altitude			
	Sea-level (n=24)	High-altitude day1 (n=24)	High-altitude day7 (n=22)	P-Value	
Age	$28 \pm 6.4$	-	-	-	
Weight (kg)	74 ± 8	73 ± 10	$72 \pm 10$	0.57	
Height (cm)	$176 \pm 10$	-	-	-	
BMI (kg m <sup>-1</sup> )	$24.3 \pm 2.4$	$23.6 \pm 9.6$	$22.8 \pm 3.5$	0.19	
AMS scores	-	$3.0 \pm 1.9$	0.4 ± 0.9 #	0.046	

List of Abbreviations: BMI, body mass index; and AMS; acute mountain sickness.

Participants were tested at sea-level (Kelowna, BC 344 m), the morning immediately following ascent having spent ~12-hours at high-altitude (high-altitude day1) (Cerro de Pasco, Peru 4330m) and again following seven days of acclimatization (high-altitude day7) (Cerro de Pasco, Peru 4330m).

P-value for a linear mixed-effect model analysis (effect of time) indicated for each variable. Symbols indicate significant post-hoc comparisons,

# Represents a significant difference between Sea-level vs High-altitude day7 (p<0.05),

	Low altitude	High-altitude		
	Sea-level (n=24)	High-altitude day1 (n=24)	High-altitude day7 (n=22)	P-Value
рН	$7.43 \pm 0.033$	7.48 ± 0.034 *	7.45 ± 0.031 #	< 0.001
Bicarbonate (mmol L <sup>-1</sup> )	$25.8 \pm 1.7$	24.6 ± 1.9	19.9 ± 2.0 #†	< 0.001
PaCO <sub>2</sub> (mmHg)	$38.4 \pm 3.2$	33.1 ± 3.3 *	28.2 ± 2.6 #†	< 0.001
Renal reactivity (Δ[HCO <sub>3</sub> <sup>-</sup> ]/Δ [PaCO <sub>2</sub> ])	-	0.098±0.75	0.54±0.14†	0.0016
PaO <sub>2</sub> (mmHg)	$100.6 \pm 18.4$	41.5 ± 7.3 *	50.7 ± 3.9 #†	< 0.001
SaO <sub>2</sub> (%)	$97.6 \pm 1.2$	78.9 ± 8.4 *	$87.6 \pm 2.1$	< 0.001
Hemoglobin (g dl <sup>-1</sup> )	$14.2 \pm 1.3$	$15.2 \pm 1.1$	15.6 ± 1.2 #	< 0.001
Hematocrit (%)	$42.3 \pm 4.4$	$44.3 \pm 2.7$	46.5 ± 2.4 #	< 0.001
CaO <sub>2</sub> (ml dl <sup>-1</sup> )	$15.2 \pm 1.8$	12.8 ± 1.7*	$18.1 \pm 1.4  \dagger$	< 0.001

List of Abbreviations: PaO<sub>2</sub>, arterial partial pressure of oxygen and PaCO<sub>2</sub>, arterial partial pressure of carbon

Participants were tested at sea-level (Kelowna, BC 344 m), the morning immediately following ascent having spent ~12-hours at high-altitude (high-altitude day1) (Cerro de Pasco, Peru 4330m) and again following seven days of acclimatization (high-altitude day7) (Cerro de Pasco, Peru 4330m).

P-value for a linear mixed-effect model analysis indicated for each variable. Symbols indicate significant post-hoc comparisons,

- \* Represents a significant difference between Sea-level vs High-altitude day1 (p<0.05),
- # Represents a significant difference between Sea-level vs High-altitude day7 (p<0.05),
- † Represents a significant difference between High-altitude day1 vs High-altitude day7 (p<0.05).

Table 2: Arterial blood data

Table 3: Cardiovascular hemodynamics and muscle sympathetic nerve activity						
	Low altitude High-altitude					
	Sea-level (n=24)	High-altitude day1 (n=24)	High-altitude day7 (n=22)	P-Value		
Cardiovascular hemodynamic	S					
Heart rate (beats min <sup>-1</sup> )	56 ± 12	77 ± 13 *	66 ± 13	<0.001		
Cardiac output (L min <sup>-1</sup> )	$4.0\pm0.8$	5.0 ± 1.1 *	4.1 ± 0.9 †	< 0.001		
Mean arterial pressure (mmHg)	88 ± 6	89 ± 7	90 ± 8	0.098		
Systolic pressure (mmHg)	117 ± 9	118 ± 8	$119 \pm 10$	0.30		
Diastolic pressure (mmHg)	$70 \pm 7$	78 ± 7 *	76.± 7	0.001		
Total peripheral resistance (mmHg L <sup>-1</sup> min <sup>-1</sup> )	$21.9 \pm 3.9$	18.9 ± 4.1 *	22.7 ± 4.6 †	0.001		
Muscle sympathetic nerve acti	vity (n = 9)					
Burst frequency (bursts min <sup>-1</sup> )	-	-	32 ± 15	-		
Burst incidence (bursts 100HB <sup>-1</sup> )	-	-	42 ± 15	-		
Mean burst amplitude (a.u.)	-	-	39 ± 9	-		
Total activity (a.u. min <sup>-1</sup> )	-	-	1284 ± 411	-		

List of Abbreviations: HB, heartbeat and a.u, arbitrary units

Participants were tested at sea-level (Kelowna, BC 344 m), the morning immediately following ascent having spent ~12-hours at high-altitude (high-altitude day1) (Cerro de Pasco, Peru 4330m) and again following seven days of acclimatization (high-altitude day7) (Cerro de Pasco, Peru 4330m).

P-value for a linear mixed-effect model analysis indicated for each variable. Symbols indicate significant post-hoc comparisons,

- \* Represents a significant difference between Sea-level vs High-altitude day1 (p<0.05),
- # Represents a significant difference between Sea-level vs High-altitude day7 (p<0.05),
- $\dagger$  Represents a significant difference between High-altitude day1 vs High-altitude day7 (p<0.05).

Table 4: Renal function and volume regulatory hormones					
	Low altitude	High-altitude			
	Sea-level (n=24)	High-altitude day1	High-altitude day7	P-Value	
	(n 24)	(n=24)	(n=22)		
Renal function	I	1	1	I	
RDO <sub>2</sub> (ml O <sub>2</sub> min <sup>-1</sup> )	$174.8 \pm 71.7$	137.9 ± 59.2*	164.9 ± 61.9†	<0.001	
Renal blood flow (ml min <sup>-1</sup> )	924 ± 366	795 ± 351*	907 ± 312	0.019	
Normalized renal blood flow (%)	23 ± 3	16 ± 3*	22 ± 4†	<0.001	
Renal vascular resistance (mmHg ml <sup>-1</sup> min <sup>-1</sup> )	110 ± 50	129 ± 64 *	116 ± 47	0.046	
Glomerular filtration rate (ml/min/1.73 <sup>2</sup> )	102 ± 20	91 ± 31 *	86 ± 17 #	0.005	
Filtration fraction (%)	$21 \pm 10$	28 ± 9 *	24 ± 9	0.005	
Volume regulatory hormones					
Active renin (pg ml <sup>-1</sup> )	$59.2 \pm 23.1$	$49.4 \pm 38.9$	$37.2 \pm 24.1$	0.025	
Plasma aldosterone concentration (pg ml <sup>-1</sup> )	212.7 ± 104.9	$175.1 \pm 162.4$	111.7 ± 92.5 #	0.018	
NT-pro-BNP (pg ml <sup>-1</sup> )	$1753.1 \pm 600.2$	$1909 \pm 970.6$	$1460 \pm 764.6$	0.15	
Urinary volume (ml) (9 hours)	$510 \pm 198.5$	$680.1 \pm 405.8$	754.6 ± 255.8 #	0.022	
Urinary microalbumin (mg L <sup>-1</sup> )	5.9 ± 1.5	10.2 ± 3.5*	$6.4 \pm 2.0$	0.012	

List of Abbreviations: NT pro-BNP, N-terminal pro-B-type natriuretic peptide and pg, picogram

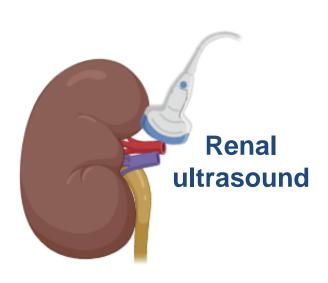
Participants were tested at sea-level (Kelowna, BC 344 m), the morning immediately following ascent having spent ~12-hours at high-altitude (high-altitude day1) (Cerro de Pasco, Peru 4330m) and again following seven days of acclimatization (high-altitude day7) (Cerro de Pasco, Peru 4330m).

P-value for a linear mixed-effect model analysis indicated for each variable. Symbols indicate significant post-hoc comparisons,

- \* Represents a significant difference between Sea-level vs High-altitude day1 (p<0.05),
- # Represents a significant difference between Sea-level vs High-altitude day7 (p<0.05),
- † Represents a significant difference between High-altitude day1 vs High-altitude day7 (p<0.05).

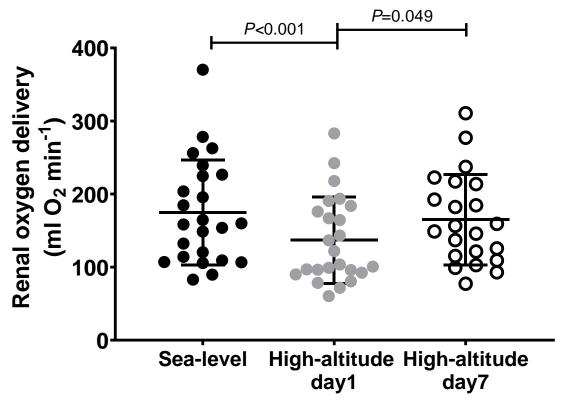
# Global REACH 2018: Renal oxygen delivery is maintained during early acclimatization to 4330 m

# **METHODS**



Radial arterial blood samples

Renal oxygen delivery at sea-level (344 m), after 12 hours and 7 days at high-altitude (4330 m)



CONCLUSION: Renal oxygen delivery is maintained at 4330 meters after a week of acclimatization.