CORE

# LAMA2 gene mutation update: Toward a more comprehensive picture of the laminin- $\alpha 2$ variome and its related phenotypes 

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#### Abstract

Congenital muscular dystrophy type 1A (MDC1A) is one of the main subtypes of early-onset muscle disease, caused by disease-associated variants in the laminin- $\alpha 2$ (LAMA2) gene. MDC1A usually presents as a severe neonatal hypotonia and failure to thrive. Muscle weakness compromises normal motor development, leading to the inability to sit unsupported or to walk independently. The phenotype associated with LAMA2 defects has been expanded to include milder and atypical cases, being now collectively known as LAMA2-related muscular dystrophies (LAMA2-MD). Through an international multicenter collaborative effort, 61 new LAMA2 disease-associated variants were identified in 86 patients, representing the largest number of patients and new diseasecausing variants in a single report. The collaborative variant collection was supported by the LOVD-powered LAMA2 gene variant database (https://www.LOVD.nI/LAMA2), updated as part of this work. As of December 2017, the database contains 486 unique LAMA2 variants ( 309 diseaseassociated), obtained from direct submissions and literature reports. Database content was systematically reviewed and further insights concerning LAMA2-MD are presented. We focus on the impact of missense changes, especially the c.2461A > C (p.Thr821Pro) variant and its association with late-onset LAMA2-MD. Finally, we report diagnostically challenging cases, highlighting the


relevance of modern genetic analysis in the characterization of clinically heterogeneous muscle diseases.

## KEYWORDS

congenital, LAMA2, laminin- $\alpha 2$, locus-specific database, muscular dystrophy, mutation update

## 1 | BACKGROUND

Laminin-211 is a heterotrimetric cruciform-shaped complex that establishes a stable link between the sarcolemma of muscle fibers and the extracellular matrix, being a major component of the extrasynaptic skeletal muscle basement membrane (BM; Durbeej, 2015). The -211 classification derives from the three specific chains ( $\alpha 2, \beta 1$, and $\gamma 1$ ), which compose this specific laminin form (Aumailley et al., 2005). Laminin-211 binds to the glycosylated residues of $\alpha$-dystroglycan ( $\alpha$ DG) and also self-assembles (polymerizes) into networks through its N -terminal domain (Yurchenco, 2015). This supramolecular network connects to collagen IV and to perlecan (heparan sulfate proteoglycan) through nidogens cross-linking (Jones, Dehart, Gonzales, \& Goldfinger, 2000). Laminin-211 expression is not confined to skeletal muscle but has also been shown to be expressed in a variety of other tissues, more importantly in peripheral nerve (Schwann cells) and in brain (Yurchenco, 2015). Posttranslational changes have been reported in laminin-211 components. More specifically, laminin- $\alpha 2$ chain was found to undergo cleavage at residue 2580 under specific conditions to generate an N -terminal 300 kDa peptide and a Cterminal 80 kDa peptide, which are subsequently connected through a noncovalent process (Durbeej, 2015).

Disease-associated (pathogenic) variants located in the gene that codes for the $\alpha 2$ chain (LAMA2; MIM\# 156225) of laminin-211, give rise to a group of diseases collectively designated as LAMA2-related muscular dystrophy (LAMA2-MD). LAMA2 maps to chromosome $6 q 22.33$ and spans over 260 kb . It comprises 65 exons and codes for a protein with a molecular mass of approximately 390 kDa (Zhang, Vuolteenaho, \& Tryggvason, 1996). The majority of patients with LAMA2 mutations have a congenital muscular dystrophy (CMD) phenotype classified as type 1A (MDC1A; MIM\# 607855). The classical phenotype manifests as neonatal hypotonia or muscle weakness during the first months of life and reduced spontaneous movements (Helbling-Leclerc et al., 1995). As muscle weakness persists during development, it compromises the achievement of normal motor milestones (no cephalic control or inability to sit unsupported) and frequently gives rise to failure to thrive. Other manifestations such as gastroesophageal reflux, aspiration, recurrent chest infections, and even respiratory failure were reported in MDC1A (Jones et al., 2001). Facial muscle weakness, ophthalmoparesis, and macroglossia are also features present in these patients but are often beyond early childhood (Quijano-Roy, Sparks, \& Rutkowski, 2012). Other relevant clinical hallmarks of MDC1A include elevated creatine kinase (CK) levels and dystrophic changes (necrosis and regeneration of fibers, chronic inflammation, and fibrosis) recognizable in muscle biopsies of these patients (Tomé et al., 1994). Diagnostically important features are the complete absence of laminin- $\alpha 2$ staining evaluated by
immunohistochemistry (IHC) performed in muscle or in skin biopsies (Sewry et al., 1996) using specific antibodies, and typical white matter changes (WMC) in brain detectable by magnetic resonance imaging (MRI; Lamer et al., 1998). WMC are related with alterations in the brain's water content, due to modifications in the maturation and/or function of the blood-brain barrier, and are detectable after the first six months to one year of life (Menezes et al., 2014). Besides WMC, brain structural defects have been reported in patients with laminin deficiency, in an estimated $\sim 4 \%$ of LAMA2-MD cases (Jones et al., 2001). In some initial studies, performed before LAMA2 genotyping was available, this association was based solely on laminin staining by IHC (Brett et al., 1998; Martinello, Angelini, \& Trevisan, 1998; Philpot et al., 1999; Pini, Merlini, Tomé, Chevallay, \& Gobbi, 1996; Sunada, Edgar, Lotz, Rust, \& Campbell, 1995; Tsao, Mendell, Rusin, \& Luquette, 1998). It is plausible that any dystroglycanopathy could account for the partial laminin deficiency observed in some patients, explaining the diversity of structural brain defects reported. It is nonetheless consensual that primary laminin- $\alpha 2$ deficiency can contribute to structural abnormalities in the cerebral cortex during fetal development. Malformations found in patients with LAMA2 disease-causing variants includes: (a) cortical dysplasia (Mercuri et al., 1999), (b) changes within the lissencephaly spectrum, namely agyria or pachygyria (Geranmayeh et al., 2010), and (c) polymicrogyria (Vigliano, Dassi, Di Blasi, Mora, \& Jarre, 2009).

In a subset of MDC1A cases there is partial laminin- $\alpha 2$ deficiency (reduced/irregular laminin- $\alpha 2$ staining in IHC ), which translates into a CMD with a slower disease progression (Geranmayeh et al., 2010; Oliveira et al., 2008). There is some degree of correlation between independent ambulation and IHC status of laminin- $\alpha 2$. The majority of MDC1A patients that do not acquire independent locomotion have complete laminin- $\alpha 2$ deficiency on muscle biopsy, whereas in the majority of cases that are able to walk independently a partial laminin- $\alpha 2$ deficiency has been documented (Geranmayeh et al., 2010).

Further to MDC1A, "milder" LAMA2-related phenotypes have been increasingly reported over the past few years. These late-onset LAMA2-MD patients are mainly characterized by proximal muscle weakness with onset during childhood, delayed motor milestones, achievement of independent ambulation, and persistently elevated CK levels (Gavassini et al., 2011). Some reports classified these patients as a subtype of limb-girdle muscular dystrophy (LGMD). Patients included in this group may also show muscle hypertrophy, rigid spine syndrome, and pronounced joint contractures which are often more evident in the elbows. In addition to cardiac involvement in a limited number of cases, these clinical features are evocative of Emery-Dreifuss muscular dystrophy (EDMD; Nelson et al., 2015). It should be emphasized that patients with late-onset LAMA2-MD still manifest typical brain

WMC, but IHC labeling of laminin- $\alpha 2$ in muscle biopsy may show only very subtle changes

As laminin- $\alpha 2$ is also expressed in Schwann cells, there is a range of clinical features related with peripheral nerve involvement in LAMA2MD patients. In a particular series of MDC1A patients, the majority had decreased motor nerve conduction, suggesting that peripheral demyelinating neuropathy is a disease feature (Shorer, Philpot, Muntoni, Sewry, \& Dubowitz, 1995). Later it was also shown that laminin- $\alpha 2$ related neuropathic abnormalities also included sensory nerves (Quijano-Roy et al., 2004). More importantly, in a milder case of LAMA2-MD there was evidence of a myelinogenesis disorder, leading to the assumption that the neuropathy in laminin- $\alpha 2$ deficient cases is actually dysmyelinating (Di Muzio et al., 2003). These changes are more evident in milder LAMA2-MD patients (Chan et al., 2014; Deodato et al., 2002; Mora et al., 1996), whereas as in MDC1A presentations the more severe muscle involvement probably masks the subtle neuropathic features of the disease.

In terms of the mutation spectrum of the LAMA2 gene, four independent studies described cohorts with more than twenty patients (Geranmayeh et al., 2010; Oliveira et al., 2008; Pegoraro et al., 1998; Xiong et al., 2015). The most frequent reported genotypes include variants that create premature termination codons (PTC) in both disease alleles, and are associated with complete deficiency of laminin- $\alpha 2$ in muscle biopsy as well as an MDC1A phenotype. In contrast, missense variants are present in a smaller number of cases and usually correlate with partial laminin- $\alpha 2$ deficiency giving rise to milder phenotypes. The asymmetrical proportion between truncating and non-truncating variants, explains the higher prevalence of MDC1A as compared with other emerging LAMA2-related phenotypes.

A relatively high frequency (18.4\% of disease-causing variants) of large deletions and duplications in LAMA2 was also reported. Variants of this sort are detectable by multiplex ligation-dependent probe amplification (MLPA) or array comparative genomic hybridization (array-CGH; Oliveira et al., 2014).

The LAMA2 locus-specific database (LSDB), which we initiated in 2002, was continuously updated and used to assist the collection of new variants as reported here. Of the 486 unique variants registered to date (December 2017), a total of 61 novel disease-associated variants detected in 86 patients are reported for the first time. Database content is systematically presented and further insights into the genotypes and phenotypes of LAMA2-MD are presented.

## 2 | DEVELOPMENT AND UPDATE OF LAMA2 LSDB

As part of the work we report the development of a comprehensive database for LAMA2 variants, an important resource made available for the scientific community since 2002. The LOVD software (Fokkema et al., 2011) was used to store genetic and clinical data, allowing an off-the-shelf LSDB deployment in accordance with international guidelines for the curation and creation of these databases (Celli, Dalgleish, Vihinen, Taschner, \& den Dunnen, 2012; Vihinen, den Dunnen, Dalgleish, \& Cotton, 2012). The LSDB content was updated and migrated
to LOVD version 3.0, being completely redesigned in terms of its database architecture.

Variant data was collected from publications accessed by the curators (64\%) or through direct database submissions (36\%). Currently (by December 2017), the LAMA2-LOVD contains a total of 1,186 of entries (486 unique) identified in a total of 748 individuals. Based on disease impact, these entries comprise: 816 disease-associated variants (309 unique), 317 benign (141 unique), and 53 variants of unknown clinical significance (VUS, 38 unique).

## 3 | DESCRIPTION OF NOVEL LAMA2 VARIANTS

A total of 61 novel disease-associated or likely associated variants were identified in the LAMA2 gene (Table 1), representing more than 20\% (61/309) of the total disease variants currently listed in the LAMA2 LSDB. Variant interpretation followed the standards and guidelines for the classification of sequence variants, proposed by the American College of Medical Genetics and Genomics (ACMG; Richards et al., 2015). The LOVD LAMA2 database gives two classifications, a Functional classification (column Effect) and a Clinical classification (column ClassClinical). The functional classification indicates the consequences of the variant for the function of the gene/protein (e.g., affects function), the clinical classification the consequences for the individual carrying the variant (e.g., ACMG:5, disease-associated, autosomal recessive [pathogenic]). The summary conclusion of the curators for specific variants, based on all individual observations of the variants, is given in a SUMMARY record. All unpublished variants collected and/or classified in the course of this project can be retrieved from the database using the following link: https://databases.lovd.nl/ shared/references/DOI:10.1002/humu. 23599.

Variants were identified by different international groups (material and methods in Supporting Information I), which reflects by the diversity of the patients' geographical origins (11 distinct nationalities). Most variants are predicted to be truncating, 20 nonsense type and 23 small frame-shift variants ( 16 deletions and seven duplications). In addition, this list includes a significant number of variants affecting canonical splice-sites $(n=13$ ), the majority located in donor sites (+1 and +2 positions). Due to the inability to obtain proper biological samples or study limitations it was mostly not possible to evaluate their impact at the mRNA level. Thus, the impact of these splice-site variants was evaluated with bioinformatic tools (see Section 3.1), which for all variants indicated unequivocal deleterious effects. One fully characterized was $c .819+2 T>C$, located in the donor splice-site of intron 5. Analysis by RT-PCR followed by sequencing, showed the presence of aberrant transcripts (details in Section 6 and Supporting Information II Figures S1 and S2). In addition to the most prevalent type of variants already stated, the remainder include: (a) two missense variants (one of which might also have an effect on splicing), (b) one in-frame (IF) codon deletion, (c) one deletion-insertion variant, and (d) a large deletion encompassing exons 57 to 65 . This large deletion was detected in a homozygous patient with an MDC1A phenotype by array-CGH technique (Supporting Information Figure S3). The 61 new variants were
TABLE 1 Novel pathogenic variants identified in LAMA2 gene listed in the locus-specific database

| Exon/ Intron | DNA variant (NM_ <br> 000426.3) | Interpretation [a] | DNA variant (NC 000006.11) hg19 | RNA variant | Predicted effect on protein | External variant databases | Number of entries in LSDB | PatientID in LSBD | Gender | Geographic origin | Phenotype | IHC for laminin$\alpha 2$ in muscle/ fibroblasts | Zygosity/ 2nd variant/ orientation (cis, trans, or unknown) | Interpretation of the second variant |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | c.47del | Pathogenic | g.129204437delG | r.(?) | $\begin{aligned} & \text { p.(Gly16 } \\ & \text { Alafs*29) } \end{aligned}$ | - | 1 | 102376 | M | United States | MDC1A | - | $\begin{aligned} & \text { Het. } / c .2 T>C / \\ & \text { trans } \end{aligned}$ | Pathogenic |
| 1 | c. $94 \mathrm{C}>$ T | Likely pathogenic | g.129204484C > T | r.(?) | p.(Gln32*) | - | 1 | 102361 | F | Canada | MDC1A | Deficiency | Het. / c.82452A > G/ unknown | Likely pathogenic |
| 2 | c.164del | Likely pathogenic | g.129371114delA | r.(?) | p. (Asn55 Metfs*16) | - | 1 | 102378 | M | Canada | CMD | Deficiency | Het./?/ unknown | No second pathogenic variant found |
| 2 i | c. $283+2$ del | Likely pathogenic | g.129371235delT | $\mathrm{r} .(\mathrm{spl}$ ? $)$ | p.(?) | - | 1 | 102463 | F | United States | MDC1A | Deficiency | Het. / c. 1609-41_16097inv / unknown | VUS [1] |
| 3 i | c. $396+1 \mathrm{G}>\mathrm{T}$ | Pathogenic | g. $129381042 \mathrm{G}>\mathrm{T}$ | r.(spl?) | p.(?) | ClinVar (RCV000 316746.1); dbSNP (rs77061 7208); gnomAD (0.0024\%) | 6 | 102366 | F | United States | MDC1A | - | Het./ c. $498 \mathrm{G}>\mathrm{A}$ / unknown | Pathogenic |
|  |  |  |  |  |  |  |  | 102732 | F | Mexico | MDC1A | - | Het./ c. $5116 \mathrm{C}>\mathrm{T}$ /unknown | Pathogenic |
|  |  |  |  |  |  |  |  | 102386 | M | United States | MDC1A | - | Hom./ n.a./ n.a. | n.a. |
|  |  |  |  |  |  |  |  | 131976 | M | United States | MDC1A | - | Het./ c. $6501 \mathrm{C}>\mathrm{A}$ /unknown | Likely pathogenic |
|  |  |  |  |  |  |  |  | 102478 | F | United States | MDC1A | - | Het./ c. $6690 \mathrm{C}>\mathrm{A}$ /unknown | Pathogenic |
|  |  |  |  |  |  |  |  | 36041 | M | Lybia | Unknown | - | Het./ $\text { c. } 8586 \mathrm{~T}>\mathrm{G}$ <br> /trans | Pathogenic |
| $4 i$ | c. $639+2 \mathrm{~T}>\mathrm{A}$ | Likely pathogenic | g. $129419562 \mathrm{~T}>\mathrm{A}$ | r.spl? | p.(?) | - | 1 | 102476 | F | United States | MDC1A | - | Het./ c. 2049 _2050 del/trans | Pathogenic |
| $5 i$ | c. $819+2 \mathrm{~T}>\mathrm{C}$ | Pathogenic | g.129465227T > C | $\begin{aligned} & \text { r.[640_819 } \\ & \text { del; } 640 \\ & -1027 \mathrm{del}] \end{aligned}$ | p.[Ile214] Arg273̄ del; Ile214 Hisfs*22] | - | 3 | $\begin{gathered} 102735 \\ {[2]} \end{gathered}$ | M | Portugal | Late-onset LAMA2related MD | Partial deficiency | Het./ c. $3976 \mathrm{C}>\mathrm{T}$ / unknown | Pathogenic |

TABLE 1 (Continued)

| Exon/ Intron | DNA variant (NM_ 000426.3) | Interpretation [a] | DNA variant (NC 000006.11) hg19 | RNA variant | Predicted effect on protein | External variant databases | Number of entries in LSDB | PatientID in LSBD | Gender | Geographic origin | Phenotype | IHC for laminin$\alpha 2$ in muscle/ fibroblasts | Zygosity/ 2nd variant/ orientation (cis, trans, or unknown) | Interpretation of the second variant |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  |  |  |  |  | ${ }_{[2]}^{102736}$ | F | Portugal | Late-onset LAMA2related MD | - | Het./ c.3976C > T / unknown | Pathogenic |
|  |  |  |  |  |  |  |  | $\begin{gathered} 103207 \\ {[3]} \end{gathered}$ | F | Portugal | Late-onset LAMA2related MD | - | Het./ <br> c.1854_1861 dup/trans | Pathogenic |
| 7 | c.939_940del | Pathogenic | $\begin{aligned} & \text { g. } 129470153 \\ & 12947015 \overline{d e l} \end{aligned}$ | r.(?) | $\begin{gathered} \text { p.(Cys314 } \\ \text { Trpfs*3) } \end{gathered}$ | gnomAD (0.0028\%) | 7 | 102373 | F | United States | MDC1A | - | Het./ c. $7732 \mathrm{C}>\mathrm{T}$ / unknown | Pathogenic |
|  |  |  |  |  |  |  |  | $\underset{[2]}{102382}$ | F | United States | MDC1A | Partial deficiency | Het./ c. $5562+5 \mathrm{G}>$ C/unknown | Pathogenic |
|  |  |  |  |  |  |  |  | $\begin{gathered} 132007 \\ {[2]} \end{gathered}$ | M | United States | MDC1A |  | Het./ c. $5562+5 \mathrm{G}>\mathrm{C}$ /unknown | Pathogenic |
|  |  |  |  |  |  |  |  | 102396 | F | United States | MDC1A | - | Hom./ n.a./n.a. | n.a. |
|  |  |  |  |  |  |  |  | 132008 | F | United States | MDC1A | - | Het./ c.7658delC / unknown | Pathogenic |
|  |  |  |  |  |  |  |  | 132009 | M | United States | MDC1A | - | Hom./ n.a./ n.a. | n.a. |
|  |  |  |  |  |  |  |  | 102655 | M | United States | MDC1A | Deficiency | Het./ <br> c. $7732 \mathrm{C}>\mathrm{T}$ <br> /unknown | Pathogenic |
| 7 | c. $991 \mathrm{~A}>\mathrm{T}$ | Likely pathogenic | g. $129470205 \mathrm{~A}>\mathrm{T}$ | r.(?) | p.(Arg331*) | $\begin{aligned} & \text { gnomAD } \\ & (0.00041 \%) \end{aligned}$ | 1 | 102467 | M | United States | MDC1A | - | Het./ <br> c.5325dupA <br> / unknown | Pathogenic |
| 12 | c. 1762 del | Pathogenic | g.129513978delG | r.(?) | p.(Ala588 Leufs*11) | Clinvar (RCV000 171527.1); dbSNP (rs78620 5654) | 7 | 102328 | F | Saudi Arabia | MDC1A | - | Hom./ n.a./n.a. | n.a. |
|  |  |  |  |  |  |  |  | 102349 | F | Unknown | MDC1A | Deficiency | Hom./ n.a./n.a. | n.a. |
|  |  |  |  |  |  |  |  | 132010 | M | Saudi Arabia | MDC1A | - | Hom./ n.a./n.a. | n.a. |
|  |  |  |  |  |  |  |  | 132011 | M | Saudi Arabia | MDC1A | - | $\begin{aligned} & \text { Het./ } \\ & \text { c. } 1303 \mathrm{C}>\mathrm{T} / \\ & \text { trans } \end{aligned}$ | Pathogenic |

TABLE 1 (Continued)

| Exon/ Intron | DNA variant (NM 000426.3) | Interpretation [a] | DNA variant (NC 000006.11) hg19 | RNA variant | Predicted effect on protein | External variant databases | Number of entries in LSDB | PatientID in LSBD | Gender | Geographic origin | Phenotype | IHC for laminin$\alpha 2$ in muscle/ fibroblasts | Zygosity/ 2nd variant/ orientation (cis, trans, or unknown) | Interpretation of the second variant |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  |  |  |  |  | $\begin{gathered} 132012 \\ {[4]} \end{gathered}$ | M | Saudi Arabia | MDC1A | - | Hom./n.a./ n.a. | n.a. |
|  |  |  |  |  |  |  |  | $\begin{gathered} 132013 \\ {[4]} \end{gathered}$ | M | Saudi Arabia | MDC1A | - | Hom./n.a./ n.a. | n.a. |
|  |  |  |  |  |  |  |  | 102363 | F | Saudi Arabia | MDC1A | - | Hom./n.a. / n.a. | n.a. |
| 13 | $\begin{aligned} & \text { c. } 18233_{-} \\ & 1824 \mathrm{del} \end{aligned}$ | Likely pathogenic | $\begin{aligned} & \text { g. } 129571297 \\ & 129571298 \text { del } \end{aligned}$ | r.(?) | p.(Tyr608*) | dbSNP (rs754600 708); gnomAD (0.00041\%) | 1 | 102471 | F | United States | MDC1A | Deficiency | Hom./n.a. / n.a. | n.a. |
| 14 | c. 2017 G > T | Likely pathogenic | g. $129573361 G>T$ | r.(?) | p.(Glu673*) | - | 1 | 102460 | F | United States | MDC1A | Deficiency | Het. / c.2023 2024del/ unknown | Likely pathogenic |
| 14 | $\begin{aligned} & \text { c. } 20233_{-} \\ & 2024 \mathrm{del} \end{aligned}$ | Likely pathogenic | $\begin{aligned} & \text { g. } 129573367 \text { _- } \\ & 129573368 \text { del } \end{aligned}$ | r.(?) | p.(Met675 Aspfs*29) | $\begin{aligned} & \text { gnomAD } \\ & (0.00041 \%) \end{aligned}$ | 1 |  |  |  |  |  | Het. / c. $2017 \mathrm{G}>\mathrm{T}$ / unknown | Likely pathogenic |
| 17 | c.2350dup | Pathogenic | g. 129591796 dup | r.(?) | $\begin{aligned} & \text { p.(Tyr784 } \\ & \text { Leufs*3) } \end{aligned}$ | ClinVar (RCV000 486406.1 | 1 | 103206 | F | Spain | MDC1A | - | Het. /c. 4692 4695dup / unknown | Pathogenic |
| 17 | c. $2383 \mathrm{G}>\mathrm{T}$ | Likely pathogenic | g.129591829G > T | r.(?) | p.(Glu795*) | dbSNP (rs149896 793); ESP (0.01\%); gnomAD (0.00041\%) | 1 | 102397 | F | United States | MDC1A | - | Het./ c.4761dupT / unknown | Likely pathogenic |
| 17i | $\begin{gathered} \text { c. } 2450+4 \mathrm{~A} \\ >\mathrm{G} \end{gathered}$ | Likely pathogenic | g. 129591900 A > G | r.(spl?) | p.(?) | - | 2 | $103191$ | F | Portugal | MDC1A | Partial deficiency | Het./ c. $8244+1 G>f$ /unknown | Pathogenic |
|  |  |  |  |  |  |  |  | 103972 | M | Portugal | Late-onset LAMA2related MD | - | Het. / c.7750-1713_78992154del/ unknown | Pathogenic |
| 18i | c.2538-1G>A | Likely pathogenic | g. $129608991 \mathrm{G}>\mathrm{A}$ | r.spl? | p.(?) | - | 2 | 102547 | F | United States | MDC1A | Deficiency | Het./ $\text { c. } 3735+2 \mathrm{~T}>\mathrm{A}$ <br> / unknown | Likely pathogenic |
|  |  |  |  |  |  |  |  | 132014 | F | United States | MD | - | Het./?/ unknown | No second pathogenic variant found |
| 21 | c. $2875 \mathrm{C}>\mathrm{T}$ | Pathogenic | g.129618848C > T | r.(?) | p.(Gln959*) | - | 1 | 102661 | F | Saudi Arabia | MDC1A | - | Hom./n.a. / n.a. | n.a. |

TABLE 1 (Continued)

| Exon/ Intron | DNA variant (NM 000426.3) | Interpretation [a] | DNA variant (NC 000006.11) hg19 | RNA variant | Predicted effect on protein | External variant databases | Number of entries in LSDB | PatientID in LSBD | Gender | Geographic origin | Phenotype | IHC for laminin$\alpha 2$ in muscle/ fibroblasts | Zygosity/ 2nd variant/ orientation (cis, trans, or unknown) | Interpretation of the second variant |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 23 | $\begin{aligned} & \text { c. } 33388_{-} \\ & 3345 \text { dup } \end{aligned}$ | Likely pathogenic | $\begin{aligned} & \text { g. } 129634169- \\ & 12963417 \text { dup } \end{aligned}$ | r.(?) | $\begin{gathered} \text { p.(Thr1116 } \\ \text { Glnfs*26) } \end{gathered}$ | - | 1 | 102385 | M | United States | MDC1A | - | Het./ $\text { c. } 6207 \mathrm{C}>\mathrm{A}$ <br> /unknown | Likely pathogenic |
| 23 | c.3372dup | Likely pathogenic | g. 129634203 dup | r.(?) | $\begin{aligned} & \text { p.(Cys1125 } \\ & \text { Metfs*4) } \end{aligned}$ | - | 1 | 103970 | U [6] | Portugal | $\begin{aligned} & \text { LGMD/ } \\ & \text { EDMD } \\ & \text { [6] } \end{aligned}$ | - | Het./ $\text { c. } 2461 \mathrm{~A}>\mathrm{C}$ <br> /unknown | Pathogenic |
| 24 | c. $3472 \mathrm{~A}>\mathrm{T}$ | Likely pathogenic | g. $129635860 \mathrm{~A}>\mathrm{T}$ | r.(?) | p.(Lys1158*) | - | 1 | 102324 | F | United States | Unknown | - | Het. /?/ unknown | No second pathogenic variant found |
| 24 | c.3520C > T | Pathogenic | g. $129635908 \mathrm{C}>\mathrm{T}$ | r.(?) | p.(Gln1174*) | - | 1 | 103127 | F | Spain | MDC1A | - | Het./ $\begin{aligned} & \text { c. } 3976 \mathrm{C}>\mathrm{T} \\ & \text { / trans } \end{aligned}$ | Pathogenic |
| 25 | $\begin{aligned} & \text { c. } 3560-356 \text { del } \end{aligned}$ | Pathogenic | $\begin{aligned} & \mathrm{g} .129636625 \\ & 129636634 \text { del } \end{aligned}$ | r.(?) | p.(Thr1187 <br> Metfs*9) | - | 1 | 102486 | F | Canada | MDC1A | Deficiency | Hom./ n.a./n.a. | n.a. |
| $25 i$ | $\begin{aligned} & \text { c. } 3735 \\ & \quad+2 T>A \end{aligned}$ | Likely pathogenic | g. $129636802 \mathrm{~T}>\mathrm{A}$ | r.spl? | p.(?) | - | 1 | 102547 | F | United States | MDC1A | Deficiency | $\begin{aligned} & \text { Het. / c.2538- } \\ & \text { 1G > A/ } \\ & \text { unknown } \end{aligned}$ | Likely pathogenic |
| 26 | c.3829C > T | Likely pathogenic | g. $129637000 \mathrm{C}>\mathrm{T}$ | r.(?) | p.(Arg1277*) | - | 1 | 102383 | M | Canada | MDC1A | Deficiency | Het./ c. $4654 \mathrm{G}>\mathrm{A}$ /trans | VUS [1] |
| 27 | c. $4002 \mathrm{~T}>$ G | Pathogenic | g. $129637260 \mathrm{~T}>\mathrm{G}$ | r.(?) | p.(Tyr1334*) | - | 1 | 102535 | F | United States | MDC1A | - | Het. / c.7658delC/ unknown | Pathogenic |
| 27 | c.4049del | Pathogenic | g.129637307delG | r.(?) | $\begin{array}{r} \text { p.(Arg1350 } \\ \text { Hisfs*12) } \end{array}$ | - | 2 | $102663$ | M | United States | MDC1A | Deficiency | Het./ c.2049_2050 <br> del / <br> trans | Pathogenic |
|  |  |  |  |  |  |  |  | 102401 | F | United States | MDC1A | - | Het./ c. $5562+5 \mathrm{G}>\mathrm{C}$ <br> / trans | Pathogenic |
| 29 | c. $4261 \mathrm{C}>\mathrm{T}$ | Pathogenic | g. $129649507 \mathrm{C}>\mathrm{T}$ | r.(?) | p.(Gln1421*) | - | 1 | $\begin{gathered} 102662 \\ {[4]} \end{gathered}$ | M | United States | MDC1A | - | Het./ c. $283+1 \mathrm{G}>\mathrm{A}$ <br> /trans | Pathogenic |
| 30 | c.4348C > T | Pathogenic | g. $129663524 \mathrm{C}>\mathrm{T}$ | r.(?) | p.(Arg1450*) | ClinVar (RCV000 dbSNP (rs20092 gnomAD (0.0012\%) | $\begin{aligned} & 1 \\ & \text { 401.1); } \\ & \text { 3); } \end{aligned}$ | 102364 | M | United States | MDC1A | Deficiency | Het./ c.2049_2050 del / trans | Pathogenic |
| 33 | c.4761dup | Likely pathogenic | g.129687407dupT | r.(?) | $\begin{aligned} & \text { p.(Arg1588 } \\ & \text { Serfs*20) } \end{aligned}$ | - | 1 | 102397 | F | United States | MDC1A | - | Het./ c. $2383 \mathrm{G}>\mathrm{T}$ /unknown | Likely Pathogenic |

TABLE 1 (Continued)

| Exon/ Intron | DNA variant (NM <br> 000426.3) | Interpretation [a] | DNA variant (NC 000006.11) hg19 | RNA variant | Predicted effect on protein | External variant databases | Number of entries in LSDB | PatientID in LSBD | Gender | Geographic origin | Phenotype | IHC for laminin$\alpha 2$ in muscle/ fibroblasts | Zygosity/ 2nd variant/ orientation (cis, trans, or unknown) | Interpretation of the second variant |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 34 | c.4941del | Likely pathogenic | g.129691117delG | r.(?) | $\begin{aligned} & \text { p.(Met1647 } \\ & \text { Ilefs*5) } \end{aligned}$ | - | 1 | 102353 | M | United States | Father of affected child (carrier study) | - | Het./n.a./ n.a. | n.a. |
| 35 | c. $5050 \mathrm{G}>\mathrm{T}$ | Pathogenic | g. $129704357 \mathrm{G}>\mathrm{T}$ | r.(?) | p.(Glu1684*) | $\begin{aligned} & \text { ClinVar } \\ & \text { (RCV0000 } \\ & \text { 78775.3; } \\ & \text { RCV000 } \\ & \text { 177827.2): } \\ & \text { dbSNP } \\ & \text { (rs2016 } \\ & 32009 \text { ) } \end{aligned}$ | 1 | 131883 | M | Italy | MDC1A | Deficiency | Het. / $\text { c. } 2901 \mathrm{C}>\mathrm{A}$ <br> / trans | Pathogenic |
| $35 i$ | $\begin{aligned} & \text { c.5072- } \\ & 1454 \_5154 \\ & \text { delinsAGA } \\ & \text { TTGCC } \end{aligned}$ | Likely pathogenic | $\begin{aligned} & \text { g. } 129711182 \\ & 129712718 \\ & \text { delins } \\ & \text { AGATTGCC } \end{aligned}$ | r.spl? | p.(?) | - | 1 | 102381 | M | United States | MDC1A | - | Hom./n.a./n.a. | n.a. |
| 36 | c.5132del | Pathogenic | g.129712696delA | r.(?) | $\begin{gathered} \text { p.(Glu1711 } \\ \text { Glyfs*14) } \end{gathered}$ | - | 1 | 102658 | M | United States | MDC1A | - | Het. / $\begin{aligned} & \text { c. } 363 \mathrm{C}>\mathrm{A} / \\ & \text { trans } \end{aligned}$ | Pathogenic |
| 36 | $\begin{aligned} & \text { c. } 5134- \\ & 5153 \mathrm{del} \end{aligned}$ | Pathogenic | $\begin{aligned} & \text { g. } 129712698 \\ & 129712717 \mathrm{del} \end{aligned}$ | r.(?) | $\begin{gathered} \text { p.(Arg1712 } \\ \text { Glufs } 4 \text { ) } \end{gathered}$ | - | 1 | 111376 | M | Turkey | MDC1A | Deficiency | Hom./n.a./n.a. | n.a. |
| 36 | c.5182del | Likely pathogenic | g.129712746delC | r.(?) | p.(Leu1728*) | - | 1 | 102358 | F | United States | MDC1A | Deficiency | Hom./ n.a. / n.a. | n.a. |
| 37 | c.5259del | Pathogenic | g.129714214delA | r.(?) | p.(Val1754*) | - | 1 | 102469 | F | Israel | MDC1A | Deficiency | Het. / c. $7147 \mathrm{C}>\mathrm{T}$ / trans | Pathogenic |
| 37 | c. $5263 \mathrm{~A}>\mathrm{T}$ | Pathogenic | g. $129714218 \mathrm{~A}>\mathrm{T}$ | r.(?) | p.(Lys1755*) | - | 1 | 103189 | M | Iran | MDC1A | Deficiency | Het./ c. $6501 \mathrm{C}>\mathrm{G} /$ unknown | Pathogenic |
| 41 | c. $5914 \mathrm{C}>\mathrm{T}$ | Pathogenic | g. $129748945 \mathrm{C}>\mathrm{T}$ | r.(?) | p.(Gln1972*) |  | 4 | 102365 | M | United States | Unknown | - | Hom./n.a./n.a. | n.a. |
|  |  |  |  |  |  |  |  | 102384 | M | United States | MDC1A | Deficiency | Hom./n.a./n.a. | n.a. |
|  |  |  |  |  |  |  |  | $\begin{gathered} 132015 \\ {[4]} \end{gathered}$ | F | United States | Unknown | - | Het./?/ unknown | No second pathogenic variant found |

tABLE 1 (Continued)

| Exon/ Intron | DNA variant (NM <br> 000426.3) | Interpretation [a] | DNA variant (NC 000006.11) hg19 | RNA variant | Predicted effect on protein | External variant databases | Number <br> of <br> entries <br> in LSDB | Patient- <br> ID in LSBD | Gender | Geographic origin | Phenotype | IHC for laminin$\alpha 2$ in muscle/ fibroblasts | Zygosity/ 2nd variant/ orientation (cis, trans, or unknown) | Interpretation of the second variant |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  |  |  |  |  |  | 102461 | F | United States | MDC1A | - | Hom./n.a./n.a. | n.a. |
| 42 | c.5998del | Likely pathogenic | g.129759820delA | r.(?) | $\begin{aligned} & \text { p.(Thr2000 } \\ & \text { Profs*3) } \end{aligned}$ | - | 1 | 102362 | M | United States | MDC1A | Deficiency | Het. / c. $7147 \mathrm{C}>\mathrm{T}$ /unknown | Pathogenic |
| 43 | c. $6207 \mathrm{C}>\mathrm{A}$ | Likely pathogenic | g. $129762082 \mathrm{C}>\mathrm{A}$ | r.(?) | p.(Tyr2069*) | $\begin{aligned} & \text { dbSNP } \\ & \text { (rs1433 } \\ & \text { 43647); } \\ & \text { ESP } \\ & \text { (0.02\%) } \end{aligned}$ | 1 | 102385 | M | United States | MDC1A | - | $\begin{gathered} \text { Het. / c. } 3338 \\ \text { 3345dup } \\ \text { / unknown } \end{gathered}$ | Likely pathogenic |
| 43 | c.6266del | Pathogenic | g.129762141delA | r.(?) | $\begin{array}{r} \text { p.(Asn2089 } \\ \text { Thrfs*14) } \end{array}$ | - | 1 | 102371 | F | Mexico | MDC1A | Deficiency | Het. / $\text { c. } 2962 \mathrm{C}>\mathrm{T}$ <br> / trans | Pathogenic |
| $45 i$ | $\begin{gathered} c .6429+ \\ 1 G>T \end{gathered}$ | Likely pathogenic | g.129766967G > T | r.spl? | p.(?) | $\begin{aligned} & \text { gnomAD } \\ & \text { (0.0032\%) } \end{aligned}$ | 2 | 102400 | M | United States | MDC1A | Deficiency | Het./ c. 2901C > A / unknown | Pathogenic |
|  |  |  |  |  |  |  |  | 132016 | M | United States | Unknown | Partial deficiency | Hom./n.a./n.a. | n.a. |
| 46 | c. $6501 \mathrm{C}>\mathrm{G}$ | Pathogenic | g. $129774204 \mathrm{C}>\mathrm{G}$ | r.(?) | p.(Tyr2167*) | - | 1 | 103189 | M | Iran | MDC1A | Deficiency | Het./ c. $5263 \mathrm{~A}>\mathrm{T} /$ unknown | Pathogenic |
| 47 | c.6588dup | Likely pathogenic | g.129775314dupT | r.(?) | $\begin{array}{r} \text { p.(Ile2197 } \\ \text { Tyrfs*5) } \end{array}$ | $\begin{aligned} & \text { gnomAD } \\ & (0.00041 \%) \end{aligned}$ | 1 | 102379 | F | United States | MDC1A | Partial deficiency | Het./ c.7571A > T/ unknown | VUS [1] |
| 49 | c.6979G > T | Pathogenic | g. $129781456 \mathrm{G}>\mathrm{T}$ | r.(?) | p.(Gly2327*) | - | 1 | 103192 | M | Iran | MDC1A | - | Hom./n.a. / $\mathrm{n} . \mathrm{a}$. | n.a. |
| 51 | c. $7297 \mathrm{C}>$ T | Likely pathogenic | $\mathrm{g} .129786431 \mathrm{C}>\mathrm{T}$ | r.(?) | p.(Gln2433*) | - | 1 | 102334 | F | United States | MDC1A | Deficiency | Het. / c.35T > G /unknown | Pathogenic |
| 54 | c.7491del | Likely pathogenic | g.129799877delA | r.(?) | $\begin{aligned} & \text { p.(Asp2498 } \\ & \text { Ilefs*49) } \end{aligned}$ | - | 1 | 102480 | M | United States | MDC1A | Deficiency | ```Het. / c. }824 +3_8244+6 del/ unknown``` | Likely pathogenic |
| 54 i | $\begin{gathered} \text { c. } 7572+ \\ 1 \mathrm{G}>\mathrm{A} \end{gathered}$ | Pathogenic | g.129799959G $>$ A | r.spl? | p.(?) | - | 2 | 111379 | F | Germany | Late-onset LAMA2related MD | - | Het. / c. $245 \mathrm{~A}>\mathrm{T} /$ unknown | VUS [1] |
|  |  |  |  |  |  |  |  | 102548 | F | United States | MDC1A | Deficiency | Het. / c.2T > C / trans | Pathogenic |
| $56 i$ | $\begin{gathered} \mathrm{c} .7898+ \\ 2 \mathrm{~T}>\mathrm{G} \end{gathered}$ | Likely pathogenic | g. $129807769 \mathrm{~T}>\mathrm{G}$ | r.spl? | p.(?) | - | 1 | 111374 | M | Turkey | MDC1A | Deficiency | Hom./ n.a. / n.a. | n.a. |
| $\stackrel{56 i_{-}}{65}$ | $\begin{aligned} & \text { c. } 7898+732 \\ & \quad-* 39282 \mathrm{del} \end{aligned}$ | Likely pathogenic | $\begin{aligned} & \text { g. } 129808499 \\ & 129876774 \text { del } \end{aligned}$ | r.(?) | p.(?) | - | 1 | $\begin{gathered} 102475 \\ {[5]} \end{gathered}$ | M | Saudi Arabia | MDC1A | - | Hom./ n.a./n.a. | n.a. |

TABLE 1 (Continued)

| Exon/ Intron | DNA variant (NM 000426.3) | Interpretation [a] | DNA variant (NC 000006.11) hg19 | RNA variant | Predicted effect on protein | External variant databases | Number of entries in LSDB | PatientID in LSBD | Gender | Geographic origin | Phenotype | IHC for laminin$\alpha 2$ in muscle/ fibroblasts | Zygosity/ <br> 2nd variant/ orientation (cis, trans, or unknown) | Interpretation of the second variant |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 58 i | $\begin{gathered} \mathrm{c} .8244+ \\ 1 \mathrm{G}>\mathrm{C} \end{gathered}$ | Likely pathogenic | g.129813629G > C | r.spl? | p.(?) | - | 1 | 102380 | M | United States | MDC1A | - | Hom./ n.a. / n.a. | n.a. |
| $58 i$ | c.8244+2dup | Likely pathogenic | g.129813630dup | r.spl? | p.(?) | - | 1 | 111380 | F | Saudi Arabia | MDC1A | - | Hom./ n.a. / n.a. | n.a. |
| $59 i$ | $\begin{gathered} \text { c. } 8357+ \\ 1 G>A \end{gathered}$ | Pathogenic | g. $129823917 \mathrm{G}>\mathrm{A}$ | r.spl? | p.(?) | - | 1 | 102391 | M | United States | MDC1A | Deficiency | $\begin{aligned} & \text { Het. / c. } 2049 \\ & \text { 2050del } \\ & \text { /trans } \end{aligned}$ | Pathogenic |
| 61 | $\begin{aligned} & \text { c. } 8556 \text { - } \\ & 8558 \text { del } \end{aligned}$ | Likely pathogenic | $\begin{aligned} & \text { g. } 129826353 \text { _ } \\ & 129826355 \text { del } \end{aligned}$ | r.(?) | $\begin{aligned} & \text { p.(Ile2852 } \\ & \text { del) } \end{aligned}$ | ClinVar (RCV0000 78805.4); dbSNP rs39 8123389 | 1 | 102737 | M | Portugal | MDC1A | Partial deficiency | $\begin{aligned} & \text { Het. / c.5234+ } \\ & \text { 1G > A/ } \\ & \text { unknown } \end{aligned}$ | Pathogenic |
| 61 | c. $8586 \mathrm{~T}>$ G | Pathogenic | g. $129826383 \mathrm{~T}>$ G | r.(?) | p.(Tyr2862*) | - | 1 | 36041 | M | Lybia | Unknown | - | Het. / $c .396+1 G>A$ <br> / trans | Likely pathogenic |
| 61 | c.8669dup | Pathogenic | g.129826466dupT | r.(?) | $\begin{aligned} & \text { p.(Leu2890 } \\ & \text { Phefs*16) } \end{aligned}$ | - | 3 | 102395 | M | United States | MDC1A | Deficiency | Het./ c. $2370 \mathrm{~T}>\mathrm{A}$ / trans | VUS [1] |
|  |  |  |  |  |  |  |  | 102472 | M | United States | MDC1A | Deficiency | Het. /c.2049_ 2050del/ trans | Pathogenic |
|  |  |  |  |  |  |  |  | 102369 | M | United States | MDC1A | Partial deficiency | Het./?/ unknown | No second pathogenic variant found |
| 63 | c.8947C $>$ T | Pathogenic | g.129833597C > T | r.(?) | p.(Gln2983*) | - | 1 | 111373 | M | Turkey | MDC1A | Deficiency | Het./ $\text { c. } 6955 \mathrm{C}>\mathrm{T}$ <br> / trans | Pathogenic |
| 64 | c.9095dup | Likely pathogenic | g.129835624dupA | r.(?) | $\begin{gathered} \text { p.(Ile3033 } \\ \text { Aspfs*6) } \end{gathered}$ | - | 2 | 102474 | M | United States | MDC1A | Partial deficiency | Het./ c. $5562+5 \mathrm{G}>\mathrm{C}$ <br> /unknown | Pathogenic |
|  |  |  |  |  |  |  |  | $\begin{gathered} 132025 \\ {[4]} \end{gathered}$ | M | United States | Unknown | - | Het./ <br> c. $4860 \mathrm{G}>\mathrm{A}$ <br> / unknown | VUS |

[^0]identified in 87 patients ( 85 families) and in one obligate carrier. In terms of genotypes, a total of 25 patients had homozygous variants. In the remaining 57 cases compound heterozygous variants were found: 52 classified as pathogenic and five VUS. From this cohort, only five cases (5.7\%) had incomplete genotyping as only one mutated allele was identified. Four variants were detected in more than three unrelated patients: c.939_940del $(n=7), c .1762 d e l(n=7), c .396+1 G>T(n=6)$, and $c .5914 \mathrm{C}>\mathrm{T}(n=4)$. This is explainable by study inclusion criteria and the higher frequency of variants identified in patients of specific populations or ethnic groups still underrepresented in the literature (c.1762del in patients from Saudi Arabia, and the others were found in patients with Hispanic ancestry). Finally, concerning the clinical presentation, the majority of patients were classified as MDC1A, whereas only seven had onset beyond infancy, and achieved independent locomotion. This particular phenotype ("late-onset" LAMA2-MD) was seen in patients with splicing variants or missense substitutions, presumably non-truncating alleles, and with partial LAMA2 deficiency documented in some of the cases.

### 3.1 Bioinformatic analysis of novel LAMA2 variants

The novel LAMA2 variants, especially those of the missense type and/or predicted to affect splicing, were further assessed resorting to bioinformatic prediction tools. A total of 14 variants fitting into these categories are shown in Table 2. With the exception of homozygotes, all variants are heterozygous and found in combination with a second change known to be disease-associated or likely disease-associated. Since experimental evidence could not be obtained, bioinformatic analysis was pivotal to attempt their classification in terms of pathogenicity.

Listed in Supporting Information III are the in silico tools used to evaluate variants, more specifically tolerance predictors and splicing predictors. Considering the extensive list of tools available to evaluate missense variants, we sought to determine which could be most efficient in the case of LAMA2 variants. The performance measures for binary classifiers, as described by Niroula and Vihinen (2016), were calculated for nine tolerance predictors algorithms. Two control sets of LAMA2 variants consistently classified in LOVD database as either pathogenic $(n=29)$ or benign $(n=22)$ were used to perform these calculations (Supporting Table). The tools with best performance for our purpose, based on the accuracy and Matthews correlation coefficient, were MutPred2 (Pejaver, Mooney, \& Radivojac, 2017), PolyPhen-2 (HumVar; Adzhubei et al., 2010), SIFT (Kumar, Henikoff, \& Ng, 2009), and UMD-Predictor (Salgado et al., 2016; Supporting Information Table S1). These algorithms were subsequently applied to evaluate the new missense variants reported in this work (Table 2).

The majority of variants listed in Table 2 were inferred as being of the missense type ( $n=8$ ). Five variants were consistently classified as deleterious by all four tolerance predictors used, and two variants were classified as deleterious by three out of the four algorithms. The other missense variant was considered deleterious by half of the tolerance predictors.

In four variants, also inferred to be of the missense type, a dual effect was predicted as they could also influence the splicing mecha-
nism. In these, the majority of algorithms tested to evaluate missense variants consistently pointed toward intolerance (all except MutPred2 in three and SIFT in one of the variants) and also indicated an effect on splicing by all tools used, except GeneSplicer in two of the variants.

From the two variants remaining in Table 2, one is an apparently synonymous substitution predicted to create a new acceptor splice-site by three distinct algorithms (all except GeneSplicer) and the other is a large intronic inversion that was predicted to disrupt the canonical acceptor splice-site.

## 4 | BIOLOGICAL RELEVANCE: CONTENT ANALYSIS OF THE LAMA2 LSDB

An overview of disease-associated variants found in the LAMA2 gene is shown in Figures 1 and 2. These may be subdivided as: $59.6 \%$ single nucleotide variants (SNV) ( $n=184$ unique; 496 in total), 24.9\% small deletions ( $n=77 ; 214$ ), $8.7 \%$ small insertions ( $n=27 ; 62$ ), 6.2\% large deletions or duplications ( $n=19 ; 42$ ), and two deletion/insertions (0.6\%). In terms of their foreseeable impact, the most frequent group is that of variants that cause a PTC. These include nonsense $(n=79)$ and out-of-frame changes ( 65 deletions, 23 duplications). A total of 79 variants were predicted or experimentally demonstrated to affect splicing. The first and last two conserved nucleotides of introns concentrate the vast majority of splicing variants. It should be highlighted that both PTC-inducing and splice-site variants are widespread throughout the gene with no clear "mutational" hotspots. In terms of distribution throughout the gene, missense variants ( $n=40,13 \%$ of total diseaseassociated variants) do not follow a similar pattern; they seem to cluster in specific regions of laminin- $\alpha 2$ (Figure 1). The first group ( $n=11$, $27.5 \%$ of missense variants) affect residues located in domain VI corresponding to the N -terminal part of the laminin- $\alpha 2$. A possible explanation is that missense variants located in this region have a detrimental effect on laminin-211 function through the disruption of protein folding and loss of the ability for polymerization into supramolecular networks, that occurs through a cooperative self-assembly process of laminin-211 (Durbeej, 2015; Yurchenco, 2015). A subset of these missense substitutions (namely p.Tyr138His, p.Gln167Pro, p.Leu243Pro, and p.Gly284Arg) are located on the presumed polymerization face near a patch containing the sequence P-L-E-N-G-E, corresponding to residues 208-213 of laminin- $\alpha 2$ (Yurchenco, 2015). These changes were identified in patients with late-onset LAMA2-MD with moderately reduced protein levels.

The next cluster consists of missense variants ( $n=10,25 \%$ of total) that specifically alter cysteine residues, located in one of the three EGF-like repeats (domains V, IIIb, and IIIa), known to establish disulfide bridges. Here, the solenoid-like structure conveyed by these rigid rod-like structures is probably modified in a way that alters the integrity of the connection between the sarcolemma and extracellular matrix mediated by laminin- $\alpha 2$. The last group of missense variants affects residues located in the C-terminal region of the protein that contains a tandem of five laminin G-like (LG) domains-LG1-5. A total of seven disease-associated missense variants (17.5\% of all missense) are in LG2, LG3, or LG4 domains. LG4 and LG5 domains mediate
TABLE 2 New LAMA2 missense changes and other variants possibly affecting splicing based on bioinformatic prediction tools

| Exon/ Intron | DNA variant (NM <br> 000426.3) | Interpretation [a] | $\begin{aligned} & \text { DNA variant (NC } \\ & \text { 000006.11) } \mathrm{hg} 19 \end{aligned}$ | RNA variant | Predicted effect on protein | External variant databases | Bioinformatic assessment [b] | Patient- <br> ID in <br> LSDB | Gender | Geographic origin | Phenotype | IHC for LAMA2 in muscle/skin | Zygosity/ second variant/ orientation (cis, trans, or unknown) | Interpretation of the second variant |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 1 | c. $112 \mathrm{G}>\mathrm{A}$ | VUS | g. $129204502 \mathrm{G}>\mathrm{A}$ | $\begin{aligned} & \text { r.(?)^ } \\ & \text { r.(spl?) } \end{aligned}$ | $\begin{aligned} & \text { p.(Gly38Ser)^ } \\ & \text { p.(?) } \end{aligned}$ | - | Possibly affects function: -Missense variant not tolerated in 3/4 predictors used (all except MutPred2) <br> -Effect on splicing predicted in 4/4 of tools tested | 102536 | M | Saudi Arabia | MDC1A | Deficiency | $\begin{aligned} & \text { Hom. / n.a. / } \\ & \text { n.a. } \end{aligned}$ | n.a. |
| 2 | c. $245 \mathrm{~A}>\mathrm{T}$ | VUS | g. $129371195 \mathrm{~A}>\mathrm{T}$ | r.(?) | p.(Gln82Leu) | - | Possibly affects function: -Missense variant not tolerated in 3/4 predictors used (all except MutPred2) | 111379 | F | Germany | Late-onset LAMA2related MD | - | Het./ c. $7572+1 \mathrm{G}$ /unknown | Pathogenic |
| 3 | c. $437 \mathrm{C}>\mathrm{T}$ | VUS | g. $129419358 \mathrm{C}>\mathrm{T}$ | r.(?) | p.(Ser146Phe) | ```ClinVar (RCV000505761.1); dbSNP (rs143680577)``` | Possibly affects function: -Missense variant not tolerated in 4/4 predictors used | 102470 | M | United States | MDC1A | - | Het./ c.6617del / unknown | Pathogenic |
| 5 | c.745C > T | VUS | g. $129465151 \mathrm{C}>\mathrm{T}$ | r.(?) | p.(Arg249Cys) | ```dbSNP (rs376437110); ESP (0.01%); gnomAD (0.0014 %)``` | Possibly affects function: -Missense variant not tolerated in 4/4 predictors used | 102368 | M | United States | MDC1A | - | Het./ c.9101_910. unknown | Pathogenic |
| 5 | c. $818 \mathrm{G}>\mathrm{A}$ | VUS | g. $129465224 \mathrm{G}>\mathrm{A}$ | r.(?) | p.(Arg273Lys) | ClinVar (RCV000394734.1) | Possibly affects function: -Missense variant not tolerated in 3/4 predictors used (all except MutPred2) | 102398 | M | United States | MDC1A | - | Het. / c. $3976 \mathrm{C}>\mathrm{T}$ /unknown | Pathogenic |
| 10 | c. $1326 \mathrm{~T}>\mathrm{G}$ | VUS | $\mathrm{g} .129498870 \mathrm{~T}>\mathrm{G}$ | r.(?) | p.(Cys 442 Trp) | - | Possibly affects function: -Missense variant not tolerated in 4/4 predictors used | 102360 | M | United States | MDC1A | Deficiency | Het./ $c .3976 \mathrm{C}>\mathrm{T}$ <br> /unknown | Pathogenic |
|  |  |  |  |  |  |  |  | 102549 | M | United States | MDC1A | - | Het./ c.7658delC /unknown | Pathogenic |

TABLE 2 (Continued)
TABLE 2 (Continued)

| Exon/ Intron | DNA variant (NM <br> 000426.3) | Interpretation [a] | DNA variant (NC 000006.11) hg19 | RNA variant | Predicted effect on protein | External variant databases | Bioinformatic assessment [b] | PatientID in LSDB | Gender | Geographic origin | Phenotype | IHC for LAMA2 in muscle/skin | Zygosity/ <br> second <br> variant/ <br> orientation <br> (cis, trans, or unknown) | Interpretation of the second variant |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 46 | c. $6548 \mathrm{~T}>$ G | VUS | g. $129774251 \mathrm{~T}>\mathrm{G}$ | r.(?) | $\begin{gathered} \text { p.(Leu2183 } \\ \text { Arg) } \end{gathered}$ | - | Possibly affects function: <br> -Missense variant not tolerated in 4/4 predictors used | 102399 | M | Iran | CMD | - | Hom. / n.a./ n.a. | n.a. |
| 47 | c. $6707 \mathrm{G}>\mathrm{A}$ | VUS | g. $129775433 \mathrm{G}>\mathrm{A}$ | $\begin{aligned} & \text { r.(?)^ } \\ & \text { r.(spl?) } \end{aligned}$ | $\begin{aligned} & \text { p. (Arg2236 } \\ & \text { Lys) })^{\wedge} \text { p.(?) } \end{aligned}$ | - | Possibly affects function: <br> -Missense variant not tolerated in 4/4 predictors used <br> -Effect on splicing predicted in 3/4 of tools tested (all except GeneSplicer) | 103971 | U [1] | Portugal | LGMD/ EDMD [2] | - | Het./ c.2461A>C /unknown | Pathogenic |
| 54 | c. $7571 \mathrm{~A}>\mathrm{T}$ | VUS | g.129799957A > T | $\begin{aligned} & \text { r.(?)^ } \\ & \text { r.(spl?) } \end{aligned}$ | $\begin{gathered} \text { p.(Glu2524 } \\ \text { Val) }{ }^{\wedge} \text { p.(?) } \end{gathered}$ | - | Possibly affects function: <br> -Missense variant not tolerated in $3 / 4$ predictors used (all except MutPred2) <br> -Effect on splicing predicted in 3/4 of tools tested (all except GeneSplicer) | 102379 | F | United States | MDC1A | Partial deficiency | Het./ c.6588dupT /unknown | Likely pathogenic |

Notes. CMD: congenital muscular dystrophy; F: female; Het.: heterozygous; Hom.: homozygous; ID: identification; IHC: immunohistochemistry; M: male; MD: muscular dystrophy; MDC1A: congenital muscular dystrophy type 1A; n.a.: not applicable; U: unknown; VUS: variant of unknown (clinical) significance; [1]: Siblings; [2]: Variants identified through an anonymized screening performed in genetically uncharacterized LGMD/ EDMD patients; [a]: According to the ACMG guidelines. [b]: More detailed information available in Supporting Information Table S2. References sequences used to describe variants: NM_000426.3 and NC_000006.11.


FIGURE 1 Point variants recorded in the laminin- $\alpha 2$ (LAMA2)-LOVD. Top layer unveils the number of unique variants that originate premature termination codons (PTC): nonsense, out-of-frame (OF) deletions (DEL) or insertions (INS), per LAMA2 exon (black rectangles). Middle layer shows splice-site variants (SPL), also indicating the number of unique variants per region: intronic (grey) or exonic (light green). Laminin-211 protein domains: I to VI, and Laminin G-like (LG) are shown in light pink to red boxes, from the C-terminal (C) to N-terminal region (N). Bottom layer displays missense (MIS) changes or single codon in-frame (IF) deletions. Light blue triangles indicate substitution of a cysteine. Variants are clustered per exonic region, and numbers below each symbol indicate the total number of changes


FIGURE 2 Large deletions and duplications listed in the laminin- $\alpha 2$ (LAMA2)-LOVD. Large duplications (DUP) are shown in the top layer as yellow rectangles encompassing affected gene regions, and deletions (DEL) are shown in the bottom part of the picture as red rectangles. Black rectangles represent the LAMA2 gene exons. Grey boxes indicate undetermined breakpoints and numbers the total of entries in the database (otherwise only one entry is present)
the binding of laminin- $\alpha 2$ to the O-linked carbohydrate chains of $\alpha$-DG whereas LG2 and LG3 bind to integrin $\alpha 7 / \beta 1$. Rare missense variants and IF deletions pose a problem for genetic data interpretation. Four IF codon deletions have been reported so far and, since there are no functional analysis strategies currently available for laminin- $\alpha 2$ variants, their impact remains unclear. A second aspect to be considered, as highlighted before, is that some changes predicted to be missense may instead have an effect on mRNA splicing. In addition to the application of bioinformatics tools used to assess the pathogenicity of missense changes (such as those mentioned in 3.1), it is advisable to consider if their location coincides with the potential hotspots outlined here.

Since our previous assessment (Oliveira et al., 2014) only two novel large deletions have been reported (Bhowmik, Dalal, Matta, Sundaram, \& Aggarwal, 2016; Ding et al., 2016), totaling 17 deletions and two duplications (Figure 2). There are two apparent mutational "hotspots" for large deletions, the first region includes exons 3 and 4, and the sec ond is in the 3 ' end of LAMA2 gene (exons 56 to 65).

Considering the distribution of disease-associated variants, exons $14,21,22,26,27,36,38$, and 56 contain over 25 variant entries. In contrast, seven exons (namely 20, 28, 44, 45, 48, 53, and 58) have no disease-causing variants reported so far. Fourteen disease-
associated variants are among the most prevalent in the LAMA2-LOVD database, with at least 10 independent entries each (Table 3). The most frequent across different ethnical backgrounds are: c.2049_2050del (p.Arg683Serfs*21), c.3085C > T (p.Arg1029*), and c.3976C > T (p.Arg1326*). Interestingly, these variants are also represented in population variant databases such as gnomAD (ExAC), found in heterozygosity with frequencies ranging from 0.012 to $0.001 \%$. Other variants such as c.1854_1861dup (p.Leu621Hisfs*7), seem to be populationor ethnic group-specific, exhibiting a relatively high frequency (0.23\%) within control alleles from the "Latino" population (gnomAD).

## 5 | CLINICAL RELEVANCE: THE EXPANDING DISEASE SPECTRUM OF LAMA2-RELATED MD

## 5.1 | Genotype-phenotype correlations

The severest end of the spectrum of LAMA2-related MD-MDC1Acorresponds to a neonatal onset disease that gives rise to hypotonia and compromised normal motor development. In LAMA2-related MD

TABLE 3 List of the most frequent pathogenic variants in the LAMA2 LSDB (variants with 10 or more entries in LOVD)

| Exon/ Intron | DNA variant <br> (NM_000426.3) | DNA variant (hg19) | RNA variant | Predicted effect on protein | Number of independent entries in LOVD | Geographic origin of patients (LOVD) | gnomAD/ExAC data: population, nr alleles/ total alleles (frequency) |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 13 | c.1854_1861dup | $\begin{aligned} & \text { g. } 1295713288_{-} \\ & 129571335 d u p \end{aligned}$ | r.1854_1861dup | p.Leu621Hisfs*7 | 12 | France, Portugal, Brazil, Spain | Latino: 2/838 (0.23\%) |
| 14 | c.2049_2050del | $\begin{aligned} & \text { g. } 129573393-\quad 129573394 \mathrm{del} \end{aligned}$ | r.2049_2050del | p.Arg683Serfs*21 | 42 | Several countries | All populations except Ashkenazi Jewish: 34/277,008 (0.012\%) |
| 18 | c. $2461 \mathrm{~A}>\mathrm{C}$ | g. $129601216 \mathrm{~A}>\mathrm{C}$ | r.2461a>c | p.Thr821Pro | 18 | Portugal | - |
| 22 | c.3085C > T | g.129621928C > T | r.(?) | p.(Arg1029*) | 24 | Portugal, Spain, United States | Latino: 1/34,418 (0.003\%); <br> European (Non-Finnish): 1/126,676 (0.001\%) |
| $26 i$ | c. $3924+2$ T > C | g. $129637097 \mathrm{~T}>\mathrm{C}$ | r.3736_3924del | p.Leu1246_Glu1308del | 33 | Saudi Arabia, Sudan, United States | - |
| 27 | c. $3976 \mathrm{C}>\mathrm{T}$ | g. 129637234C > T | r.(?) | p.(Arg1326*) | 24 | Portugal, Sweden, United States, Spain, Denmark | European (Non-Finnish): <br> 14/126,524 (0.001\%); <br> European (Finnish): 1/25,788 (0.004\%) |
| 32 | c.4645C > T | g. $129674430 \mathrm{C}>\mathrm{T}$ | $\begin{aligned} & \text { r.[4645c>u, } \\ & \left.\quad 4580 \_4717 \mathrm{del}\right] \end{aligned}$ | $\begin{aligned} & \text { p.[Arg1549*, Cys1527_ } \\ & \text { Val1572del] } \end{aligned}$ | 10 | Australia, Italy, United States | South Asian: 2/30,780 (0.006\%) |
| $36 i$ | c. $5234+1 \mathrm{G}>\mathrm{A}$ | g. 129712799 C > | r.5072_5234del | p.Val1765Serfs*21 | 10 | Portugal, Canada, United States | Latino: 1/34,376 (0.003\%); <br> European (Non-Finnish): 2/126,266 (0.002\%) |
| 38 | c.5476C > T | g. $129722399 \mathrm{C}>\mathrm{T}$ | r.(?) | p.(Arg1826*) | 11 | China, Saudi Arabia, United Kingdom | East Asian: <br> 2/17,240 (0.012\%); <br> European (Non-Finnish): 5/111,398 (0.0045\%) |
| $38 i$ | c. $5562+5 \mathrm{G}>\mathrm{C}$ | g. 129722490 C > C | ```r.[5446_ 5562del, 5562_5563ins5562``` | p.[Lys1816_Asp1854del, Tyr1855Valfs*24] | 14 | United Kingdom, United States | $\begin{aligned} & \text { European } \\ & \text { (Non-Finnish): } \\ & 7 / 125,864 \\ & \text { (0.0056\%); } \\ & \text { European (Finnish): } \\ & 1 / 25,408 \\ & (0.0039 \%) \end{aligned}$ |
| 46 | c.6488del | g. 129774191 delA | r.(?) | p.(Lys2163Argfs*12) | 15 | Qatar, Saudi Arabia, United States | - |
| 55 | c.7732C > T | g. $129802567 \mathrm{C}>\mathrm{T}$ | r.(?) | p. (Arg2578*) | 12 | China, Denmark, Mexico, Russian Federation, United States | Latino: <br> 3/34,380 (0.0087\%); <br> European (Non-Finnish): $10 / 126,598$ (0.0079\%); <br> East Asian: 1/18,834 (0.0053\%); South Asian: 1/30,782 (0.0032\%) |
| $\begin{aligned} & 55 i_{-}^{i} \\ & 56 i \end{aligned}$ | $\begin{aligned} & \text { c. } 7750-1713- \\ & 7899-2154 \mathrm{del} \end{aligned}$ | $\begin{aligned} & \text { g. } 129805906- \\ & 129810892 \text { del } \end{aligned}$ | r.7750_7898del | p.Ala2584Hisfs*8 | 17 | Portugal | - |
| 58 i | c. $8244+1 \mathrm{C}>\mathrm{A}$ | g.129813629G $>$ A | r.8076_8244del | p.Pro2693Valfs*12 | 12 | Germany, Portugal, Tunisia, United States | $\begin{aligned} & \text { European } \\ & \text { (Non-Finnish): } \\ & 1 / 111,114 \\ & \text { (0.0009\%) } \end{aligned}$ |

Note. nr: number.
the locomotion attainment has been considered an important clinical measure of disease severity. In a series of 26 MDC1A patients only two had acquired independent locomotion (Oliveira et al., 2008). Interestingly, all patients that harbored variants inducing PTC in both disease alleles were unable to achieve independent walking. In con-
trast, the two patients that were able to walk had a missense or a single codon deletion in one of the disease genes. LSDB content and other studies reported in the literature (Geranmayeh et al., 2010) further corroborated our findings. However, there are exceptions to this rule; for example, a patient with a homozygous nonsense variant
(p.Arg1549*) was able to reach ambulation and even climb stairs (Geranmayeh et al., 2010). This particular variant was reported in association with partial deficiency of laminin- $\alpha 2$ in several unrelated patients (Di Blasi et al., 2000; Geranmayeh et al., 2010; Pegoraro et al., 1998). Here, an explanation for this discrepancy is the fact that the variant is located within exon 32 that undergoes alternative splicing (Pegoraro et al., 2000). The exon removal leads to an IF deletion at the mRNA level, thereby restoring the reading frame from the PTC created by the nonsense variant.

Geranmayeh et al. (2010) provided further genotype-phenotype correlations with prognostic clinical implications. Statistically significant differences were identified between patients with complete deficiency and those with partial deficiency of laminin- $\alpha 2$. Patients with absence of laminin- $\alpha 2$ had earlier onset ( $P=0.0073$ ), lack of independent ambulation ( $P=0.0215$ ), and were more prone to requiring artificial feeding ( $P=0.0099$ ) or respiratory support ( $P=0.0354$; Geranmayeh et al., 2010). Within MDC1A, there is a subset of patients with early onset phenotype but a "milder" disease progression and with partial laminin- $\alpha 2$ deficiency. This partial deficiency is often associated with missense variants, IF deletions, and splicing variants (leaky or inducing IF exon-skipping; Allamand \& Guicheney, 2002; Quijano-Roy et al., 2012). One of the earliest such cases reported had a homozygous variant (p.Cys996Arg) that affects domain IIb of laminin- $\alpha 2$ (Nissinen et al., 1996).

Despite the general consistency between phenotype, the type of variant and the IHC status, some exceptions have been documented in the literature. These include patients with complete laminin- $\alpha 2$ deficiency and missense variants that achieved independent locomotion (Geranmayeh et al., 2010), although this could be attributed to IHC sensitivity issues. Intrafamilial clinical variability has also been reported, such as that found among patients from one large Kenyan kindred of Asian ancestry. Here, patients shared the same genotype (homozygous missense variant located in the G-domain of laminin- $\alpha$ 2) but locomotion was not achieved in all cases (Geranmayeh et al., 2010).

As previously mentioned, a very small fraction of LAMA2-MD patients have brain structural defects, which are frequently associated with intellectual disability (ID) and/or refractory seizures (Geranmayeh et al., 2010, Vigliano et al., 2009). However, there are also reports of patients, with these structural defects who, apparently have no seizures or ID. The opposite also holds true in the case of seizures (and to a lesser extent ID) since they have been reported in patients without cerebral structural changes. Based on the reassessment of data available in the LOVD and reported in the literature, no association was found between epilepsy, cognitive function or brain anomalies, and a particular set of LAMA2 genotypes/variants. The variants found in these cases are diverse in terms of their impact, ranging from those causing PTC to missense changes, and are apparently dispersed with no obvious hotspot along the gene. Furthermore, phenotypical discrepancies have been found in patients sharing with same genotype. For example, two siblings reported by Di Blasi et al. (2001) and case \#2 from Nelson et al. (2015) share the same genotype (a homozygous nonsense variant p.Arg744*), but cortical polymicrogyria and lissencephaly were only reported in the latter patient. It is conceiv-
able that other genetic factors besides LAMA2 variants are contributing to these phenotypes.

Over the last few years there has been a significant increase in reports of late-onset LAMA2-related MD patients (Ding et al., 2016; Gavassini et al., 2011; Harris et al., 2017; Kevelam, van Engelen, van Berkel, Küsters, \& van der Knaap, 2014; Kim et al., 2017; Løkken, Born, Duno, \& Vissing, 2015; Marques et al., 2014; Nelson et al., 2015; Rajakulendran, Parton, Holton, \& Hanna, 2011). Most of these patients have heterozygous or homozygous missense or splice variants. Their clinical presentation is also variable but often overlapping with a childhood-onset LGMD, consisting of proximal muscle weakness and delayed motor milestones, but in all cases achieving independent ambulation. Rigid spine syndrome with joint contractures has been also reported in some patients (Nelson et al., 2015).

## 5.2 | Additional cases of late-onset LAMA2-related MD sharing the $p$.Thr821Pro variant

Phenotypic variability in LAMA2-related MD has been clearly underestimated so far, with only a limited number of patients with this lateronset phenotype reported in the literature. As for establishing further genotype-phenotype correlations, the cases are still relatively scarce and there is a vast diversity of genetic defects and/or genotypes, which makes it difficult to stratify patients into homogeneous groups.

To address some of these limitations, and resorting to our large LAMA-related MD patient cohort, the clinical and genetic characterization of six additional patients with a late-onset phenotype from four unrelated families is reported (Table 4). They all share the same missense variant: p.Thr821Pro. In five cases the genotype was similar in that, besides this missense substitution, the second allele was a truncating variant: c.7750-1713_7899-2154del (p.Ala2584Hisfs*8) in patients P 1 and $\mathrm{P} 2, \mathrm{c} .3976 \mathrm{C}>\mathrm{T}($ p.Arg1326*) in P3 and P4, and c.1854_1861dup (p.Leu621Hisfs*7) in P5. The sixth patient (P6) represents the first documented case with a homozygous p.Thr821Pro missense variant. Most of these patients were only diagnosed during adulthood, which reflects the diagnostic difficulties concerning nonMDC1A cases. All have a very mild muscle weakness (as compared with typical MDC1A) with lower limb weakness resulting in gait disturbances. In the oldest patient (P6) this weakness culminated in loss of ambulation during the sixth decade of life. In four patients brain MRI was performed ( $\mathrm{P} 1, \mathrm{P} 2, \mathrm{P} 3$, and P 6 ), revealing WMC like those usually found in LAMA2-related MD (Figure 3a-c). These findings were pivotal for conducting LAMA2 gene analysis in three of the cases. Patient P6, who developed dementia over the last 2 years, also had hypothalamus and pons alterations (data not shown). Five patients were subjected to a muscle biopsy. These showed myopathic or dystrophic features (Figure 3d-f), and IHC analysis for laminin- $\alpha 2$ revealed apparently normal labeling ( $n=3$, Figure 3 g -i) or partial deficiency ( $n=1$, data not shown).

## 5.3 | Prevalence of p.Trp821Pro variant in a genetically uncharacterized MD patient cohort

The missense variant p.Trp821Pro is one of the most frequent genetic causes of late-onset LAMA2-MD in a population-specific (Portuguese)
TAB LE 4 Additional LAMA2-related muscular dystrophy patients sharing the p.Thr821Pro variant

| Family patient \# | Genotype | Predicted effect on protein | Age (gender) | Age of first symptoms | Phenotype at onset | Pattern of muscle weakness; other clinical features | Cardiac involvement? | Contractures? | Independent locomotion? (age) | Loss of ambulation? (age) | CK <br> levels <br> (U/I) | Muscle biopsy | Laminin$\alpha 2$ IHC [1] | Brain changes (MRI) |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| F.I-P1 | $\begin{aligned} & c .2461 \mathrm{~A}>\mathrm{C}+ \\ & \text { c.7750- } \\ & 1713 \text { _7899- } \\ & 2154 \mathrm{del} \end{aligned}$ | $\begin{aligned} & \text { p.Thr821Pro + } \\ & \text { p.Ala2584 } \\ & \text { Hisfs*8 } \end{aligned}$ | 42 yrs (F) | Third decade | Migraine-like headaches for 8 mo . Complaints of limb weakness and walking difficulties | Mild generalized muscular atrophy and tetraparesis (4+/5 grade), feet dorsiflexion (4/5). Paresis of trunk and neck flexion, cannot do sit-ups or lift head while in supine position. | N | N | Y | N | n.p. | Moderate myopathic changes, with discrete endomysial fibrosis | Normal | WMC |
| F.I-P2 | $\begin{aligned} & c .2461 \mathrm{~A}>\mathrm{C}+ \\ & \text { c.7750- } \\ & 1713 \text { _7899- } \\ & 2154 \mathrm{del} \end{aligned}$ | p.Thr821Pro + p.Ala2584 Hisfs*8 | 50 yrs (M) | Forth decade | Running difficulties and progressive lower limb weakness | Proximal paresis (4/5 grade) in upper limbs, distal and proximal paresis in lower limbs. Paresis of trunk and neck flexion (grade 2 and 4+, respectively). Lordosis and myopathic gait with slight steppage. | N | N | Y | N | n.p. | Myopathic changes with fiber necrosis, also "ragged red fibers" and large number of COX negative fibers. | Normal | WMC |
| F.II-P3 | $\begin{aligned} & \text { c. } 2461 \mathrm{~A}>\mathrm{C}+ \\ & \mathrm{c} .3976 \mathrm{C}>\mathrm{T} \end{aligned}$ | p.Thr821Pro + p.Arg1326* | 15 yrs (F) | 18 mo | Running difficulties and stairs | Proximal paresis (2/5 grade) in upper limbs and in lower limbs (3/5 grade), neck flexion (grade 2/5). | N | Y | $\mathrm{Y}(15 \mathrm{mo})$ | N | 839 | Dystrophic changes | Normal | WMC |
| F.II-P4 | $\begin{array}{r} \text { c. } 2461 \mathrm{~A}>\mathrm{C}+ \\ \mathrm{c} .3976 \mathrm{C}>\mathrm{T} \end{array}$ | p.Thr821Pro + p.Arg1326* | 11 yrs (F) | 5 yrs | Facial fatigue | Proximal paresis ( $2 / 5$ grade) in upper limbs and in lower limbs (4/5 grade), neck flexion (grade 1/5). | N | N | Y (14 mo) | N | 2466 | n.p. | n.p. | n.p. |
| F.III-P5 | $\begin{aligned} & \text { c.2461A }>\mathrm{C}+ \\ & \text { c.1854_- } \\ & \text { 1861dup } \end{aligned}$ | p.Thr821Pro + p.Leu621 Hisfs*7 | 33 yrs (F) | Childhood | Running difficulties. Difficulty in getting out of a bed | Proximal paresis (4-3/5 grade) in lower limbs. Paresis of trunk and neck flexion (grade $3 / 5$ ). Lordosis and myopathic gait. | N | N | Y | N | $\sim 2,000$ | Dystrophic changes | Partial deficiency | n.p. |
| F.IV-P6 | c.2461A > C (hom.) | p.Thr821Pro | 71 yrs (M) | Childhood | Gait impairment | LGMD initially suspected. Proximal tetraparesis (grade 4/5, 4-/5 in lower limbs). Moderate intellectual disability (last 2 yrs ). | Y [2] | N | Y | Y (last 2 years) | 579 | Moderate dystrophic changes associated to angulated atrophic fibers and nuclear clumps | Normal | WMC, thalamus and pons involvement |

 Clone: Mer3/22B2 (Leica Biosystems, Newcastle upon Tyne, United Kingdom); [2]: Left ventricle hypokinesia of unknown cause, normal ejection fraction.


FIGURE 3 Brain magnetic resonance imaging (MRI) and muscle neuropathology results. Patient P1: (a) brain MRI (FLAIR) shows typical white matter changes (WMC) with normal structural cerebral cortex changes; (d) moderate myopathic changes with discrete endomysial fibrosis in hematoxylin eosin ( h \& e) staining, and (g) normal immunohistochemistry (IHC) for laminin- $\alpha 2$. Patient P2: (b) brain MRI (FLAIR) with WMC and normal cerebral cortex; (e) myopathic changes with necrotic fibers and several "ragged red fibers" (h \& e), and (h) normal IHC for laminin- $\alpha 2$. Patient P6: (c) WMC in brain MRI (FLAIR); (f) dystrophic changes (necrotic fibers under myophagocytosis, fiber splitting and hypersegmentation, and fat substitution) and mild neurogenic features (atrophic angulated fibers and nuclear clumps), and (i) normal IHC for laminin- $\alpha 2$
patient cohort. This missense substitution was initially identified in two patients with atypical presentations (Marques et al., 2014), prior to the six patients described above. A further three patients, from two unrelated families with Portuguese ancestry, have also been reported by other groups: one patient from Canada (with \#102482 in LAMA2-LOVD), and two brothers studied in France (Nelson et al., 2015). Thus far, all patients are reported to have milder muscle weakness and the majority were initially classified as possible LGMD or EDMD. To evaluate if this missense variant could account for additional uncharacterized cases, we screened an irreversibly anonymized group of 239 myopathic Portuguese patients with clinical presentation is compatible with LGMD or EDMD. Variant screening was performed by restriction fragment length analysis (RFLA, Supporting Information IV), since the c.2461A > C change creates a new restriction site for HpyCH4III (Supporting Information Figure S4). Positive samples were confirmed by Sanger sequencing. A total of seven patients carried this missense substitution (2.9\% of the cohort), three of which were homozygotes and four were heterozygotes ( $2 \%$ of all disease alleles). To further ascertain the genotype of the four patients carrying the c.2461A > C variant in heterozygosity, the entire coding sequence of the LAMA2 gene was sequenced. In all patients an
additional heterozygous variant was detected. Three were previously identified in other (MDC1A) patients: c.4739dup (p.Leu1581Profs*5; Oliveira et al., 2008), c.3372dup (p.Cys1125Metfs*4; patient \#103970 in Table 1) and a missense variant c.32T > C (p.Leu11Pro) listed in ClinVar (RCV000157587.1) as being disease associated. The fourth variant, c.6707G > A, is also new and was interpreted as a VUS; it predictably gives rise to a missense change (p.Arg2236Lys) and/or may have an effect on splicing (r.spl?; Table 2, patient \#103971).

Since the c.2461A > C variant was not listed in variant population databases, its prevalence was estimated in control individuals using the aforementioned RFLA-screening strategy. For this study, we randomly selected and irreversibly anonymized 1,100 out of a total of 11,000 samples previously analyzed in the laboratory. These were residual samples from genetic studies for diseases unrelated with neuromuscular disorders that are performed on a nationwide basis. The c.2461A > C variant was identified in one of these samples, in heterozygosity. Its allelic frequency in the general population was estimated as $0.0452 \%(1 / 2,200)$, which explains the relatively high prevalence of this variant among Portuguese patients with LAMA2-MD.

Overall, the presented data reinforces that it is diagnostically important to consider LAMA2 gene involvement not only in CMD


FIGURE 4 Different phenotypes in laminin- $\alpha 2$-related muscular dystrophy (LAMA2-MD) found in association with the $c .2461 A>C$ (p.Trp821Pro) variant. (a) Early-onset (classical muscular dystrophy type 1A [MDC1A]): no independent ambulation; muscle biopsy shows dystrophic features and no labeling for laminin- $\alpha 2$ in immunohistochemistry (IHC). Several patients reported in Oliveira et al. (2008). (b) Late-onset (Emery-Dreifuss muscular dystrophy [EDMD]/COL6-RD-like): rigid spine syndrome; cardiac involvement in some patients; walking difficulties; dystrophic features in MD, normal and irregular laminin- $\alpha 2$ staining in IHC. Pat.2-patient 2 (Marques et al., 2014); cases \#3, \#4 (Nelson et al., 2015); P5 (this work). (c) Late-onset (limb-girdle muscular dystrophy (LGMD)-like): slow progression; dystrophic features, normal and irregular laminin- $\alpha 2$ staining in IHC, walking difficulties later in life; P1-4, P6 (this work). (d) Atypical LAMA2-RD: predominant central nervous system (CNS) involvement, (occipital agyria, white matter changes (WMC), epilepsy); increased variability of muscle fiber diameter and irregular laminin- $\alpha 2$ staining in IHC. Pat.1-patient 1 in Marques et al. (2014). Genotype-phenotype correlations suggest that the classical MDC1A presentation is explainable by variants causing premature termination codons (PTC) in both disease alleles. While late-onset LAMA2-MD are more likely to be associated with missense (MS) substitutions
Note. •: homozygous; +: Heterozygous.
patients, but also as a possible cause of MD with onset beyond childhood and even in adulthood. The association between LAMA2 and this later onset phenotype was not fully established, considering the limited number of cases reported so far. Nonetheless, it is advisable to include LAMA2 in the list of candidate genes for MDs (LGMD or EDMD). The p.Trp821Pro missense variant constitutes an interesting genotype-phenotype linker, as it may give rise to different phenotypes depending on the variant found in the second allele (Figure 4).

## 6 | DIAGNOSTIC RELEVANCE

Molecular defects in the LAMA2 gene are the main genetic causes ( $\sim 30 \%$ ) of CMDs in most countries, except for Japan where Fukuyamatype CMD has the highest prevalence, due to a frequent founder mutation in the FKTN gene (Kobayashi et al., 1998). Besides the clinical examination, the clinical diagnostic workup of CMDs conventionally relies upon performing a muscle biopsy (Bönnemann et al., 2014). In addition to standard staining methods, muscle pathology analysis includes a panel of antibodies for IHC against proteins involved in MD (laminin- $\alpha 2$, sarcoglycans, dystrophin, and dysferlin). Three different commercial antibodies are currently available for laminin- $\alpha 2$ IHC studies: clone 5 H 2 detects the 80 kDa protein (C-terminal region),
clone Mer3/22B2 detects the 300 kDa product (N-terminal region), and clone 4H8-2 clone, which also recognizes the N-terminal domain. The diagnostic sensitivity of IHC is extremely high for typical MDC1A cases, where complete deficiency would be detectable regardless of the antibody used for analysis. The milder LAMA2-MD cases are more challenging as often only a partial deficiency is often documented. Moreover, depending on the underlying molecular defects, this IHC deficiency may not be consistent for the different antibodies (Cohn, Herrmann, Sorokin, Wewer, \& Voit, 1998). N-terminal antibodies usually have higher sensitivity for cases with partial laminin- $\alpha 2$ deficiency, as there was a relatively intact labeling with the antibody for the 80 kDa fragment, when compared with that using the other antibodies (Cohn et al., 1998). It is therefore advisable to include at least two different antibodies against laminin- $\alpha 2$ in order to increase IHC sensitivity. In a small fraction of CMD patients there is also irregular labeling or partial laminin- $\alpha 2$ deficiency. There is some degree of genetic heterogeneity among these patients, depending on whether it is a primary or a secondary deficiency. To distinguish between these two possibilities, antibodies against glycosylated residues of $\alpha$-DG and laminin$\alpha 4 / 5$ may be effective. If changes are detected in $\alpha$-DG, this would indicate a defective glycosylation pathway and involvement of other loci. On the other hand, normal $\alpha$-DG labeling and overexpression of laminin- $\alpha 4 / 5$ (a compensatory gene expression mechanism) is suggestive of a primary laminin- $\alpha 2$ deficiency.

Brain MRI performed beyond the first 6 to 12 months is also an important diagnostic resource for CMDs. As previously mentioned all LAMA2-related MD patients have brain WMC, consisting in bilateral hyperintensity signal on T2-weighted and FLAIR MRI, in periventricular areas and subcortical cerebral hemisphere (Quijano-Roy et al., 2012). These findings alone should be an indication to perform LAMA2 genetic testing. As demonstrated by this work and previously suggested by Gavassini et al. (2011), it is diagnostically relevant to perform brain MRI in uncharacterized LGMD patients. This could be performed even during adulthood, as these typical brain changes will persist throughout life. Brain MRI is especially relevant for "atypical" or mild MD cases where IHC for laminin- $\alpha 2$ has a lower diagnostic yield.

Considering the size and number of exons in the LAMA2 gene, its genetic analysis has been simplified, more than a decade ago, with the introduction of automatized sequencers (fragment analyzers) for Sanger sequencing and the use of universal-tailed primers. Gene sequencing is undoubtedly the approach with the highest sensitivity for LAMA2 analysis, detecting approximately $80 \%$ of diseaseassociated variants. Based on the variant data collected, there is a significant frequency ( $\sim 18 \%$ ) of large deletions and duplications. The genetic study should therefore be complemented with other molecular techniques such as MLPA or array-CGH.

Data available in the LSDB and population-specific cohorts can help to optimize the LAMA2 genetic analysis. This was exemplified in a Portuguese CMD patient cohort where a 3-tier genetic test was proposed (Oliveira et al., 2014): (a) sequencing a small set of selected exons where the majority of point mutations are located (based on a specific population or ethnical group variant data); (b) sequencing the remaining LAMA2 exons; and (c) MLPA analysis or array-CGH.

The introduction of next-generation sequencing technology (NGS, or massive parallel sequencing) has remodeled genetic analysis strategies, especially in genetically heterogeneous conditions such as the MDs. Distinct NGS applications such as gene panels or whole-exome sequencing (WES) can be extremely useful to address diagnostically difficult cases. In fact, novel cases with milder LAMA2-related phenotypes recently reported in the literature have been solved resorting to NGS (Dean, Rashid, Kupsky, Moore, \& Jiang, 2017; Ding et al., 2016; Harris et al, 2017; Kim et al., 2017).

The impact of NGS technology is also reflected in five patients described in this work: (ID\#s: 102662, 132012, 132013, 132015, 132025 in table I), whose disease-associated variants listed were identified by NGS gene panels. One further patient (ID\# 103207 in Table 1) demonstrates the utility of NGS to address genetic and clinical heterogeneity. This is a patient with an LGMD phenotype and ID, who has remained without genetic characterization for several years. The patient, currently 14 years of age, had delayed motor development (started walking at 31 months of age), lumbar lordosis, and elevated CK levels ( $\sim 1300$ U/I). Muscle biopsy performed at 6 years of age (in another clinical center where she was initially followed) revealed dystrophic features and normal IHC results for dystrophin and sarcoglycans. Genetic analysis of FKRP, CAPN3, LMNA, and DMPK genes were negative. The patient was studied by WES as previously reported in a similar research (Oliveira, Martins, Pinto Leite, Sousa, \& Santos, 2017). As a first approach, WES data analysis was restricted to a set of genes
known to be associated with muscle diseases (Supporting Information IV). Within the list of filtered-in variants, two heterozygous variants were identified in LAMA2 (Supporting Information Figure S5). The first was the c.1854_1861dup variant, previously reported as disease associated in several MDC1A patients, and the second was a novel splicing variant $\mathrm{c} .819+2 \mathrm{~T}>\mathrm{C}$ located in the donor splice-site of intron 5 (Table 1). To further characterize the effect of this splice-site variant, a muscle fragment available from patient ID\# 102735 (shares the same variant) was used (Supporting Information II). LAMA2 transcript analysis by RT-PCR showed the presence of multiple aberrant products that, upon sequencing, were attributed to multiple skipping events involving exons 5 to 7 (Table 1, Supporting Information Figures S1 and S2). Study of the patient's parents confirmed compound heterozygosity, as each progenitor carried a different LAMA2 variant. Brain MRI performed after WES analysis, revealed WMC but not configuring the typical pattern found in MDC1A cases. In this patient, axial T2 and FLAIR revealed small focal white matter hyperintensities in the subcortical part of brain, more specifically in the frontal, temporal-anterior, parahyppocampus, and insula regions with sparing of the internal capsule and corpus callosum (data not shown).

Finally, as demonstrated by five cases listed in Table 1 (ID\#s 102324, 102369, 102378, 132014, and 132015), a small percentage of patients were found to have only one heterozygous LAMA2 disease-causing variant. This could be attributed to deeply placed intronic variants affecting splicing or variants located in the gene's promotor region, both of which are not covered by conventional sequencing, gene panels, or even WES. Here, a more comprehensive NGS approach such as WES and/or RNA sequencing may ultimately provide a final answer to such cases with incomplete genotyping.

## 7 | FUTURE PROSPECTS

Although there is an increasing recognition of the involvement of LAMA2 disease-associated variants in the genetic etiology of muscular dystrophies, the incidence is probably still underestimated. To improve the diagnosis of these cases, it is necessary to include both brain MRI and to evaluate the expression of laminin- $\alpha 2$ in muscle by IHC. These two approaches are often not considered in the clinical workup of patients with (non-congenital) myopathies. NGS can also contribute toward the identification of further cases. However, the interpretation of variants from such studies often leads to their classification as VUS, which considerably limits the clinical utility of these genetic data.

As for further research, it is necessary not only to continue to document clinical data and LAMA2 variants to obtain further genotype-phenotype correlations, but also to develop strategies for functional analysis and validation of new variants, especially those predictably of the missense type. This task may be complex, as variants might affect several key aspects of the laminin-211 life-cycle: (a) posttranslational modification, (b) protein translocation and secretion process, (c) interaction with membrane-specific receptors, and (d) variety of molecular partners in the BM, possibly some yet to be identified. One strategy for functional analysis would imply obtaining a biological sample from the patient by an invasive procedure (e.g., muscle or
skin biopsy), expanding cells through in vitro culture, and performing protein-protein interaction studies, such as pull-down assays using a battery of different bait-proteins known to interact with laminin- $\alpha 2$. Failure to detect a particular interaction would indicate a deleterious effect. To enable such studies, further research should primarily focus in a comprehensive search for domain-specific interactions, which could be accomplished by high-throughput proteomics analyses. An assay for those variants specifically affecting domains involved in laminin polymerization has been reported (Cheng, Champliaud, Burgeson, Marinkovich, \& Yurchenco, 1997; Hussain, Carafoli, \& Hohenester, 2011). Basically, a mixture of wild-type with the mutated form of laminin would show a failure in establishing normal polymerization levels. Here, the limiting step would be generating and purifying sufficient amounts of proteins to conduct these in vitro studies.

As laminin- $\alpha 2$ is not confined to muscle or brain cells, in a transgenic mouse model with deficient laminin- $\alpha 2$ it was shown that the loss of this protein caused disruption of the apical ectoplasmic specialization-blood-testis barrier, and leading to male infertility (Häger, Gawlik, Nyström, Sasaki, \& Durbeej, 2005). The laminin- $\alpha 2$ in testis was further implicated in the regulation of an axis that functionally links the BM to the blood-testis barrier of Sertoli cells (Gao et al., 2017). Considering that human infertility has not been linked to laminin- $\alpha 2$, it would be relevant to evaluate male reproductive issues in late-onset LAMA2-MD patients.

One of the most important aspects concerning LAMA2-MD is the development of a suitable treatment for this condition. Several approaches have been proposed, developed, and tested in laminin$\alpha 2$-deficient mice and zebrafish models (reviewed by Durbeej, 2015; Wood \& Currie, 2014). One particularly effective approach targets extracellular matrix modulation as a way to ameliorate MDC1A. Here, strategies aim to improve muscle viability, through the augmentation of residual functionality within the cellular system, such as upregulation of other laminins ( $\alpha 4$ or $\alpha 1$ ) and integrin- $\alpha 7$ (Wood \& Currie, 2014). However, with laminin-411 there are some limitations for BM repair, since this laminin only forms a trimeric structure, lacking capacity to further self-polymerize into superstructures such as those derived from laminins-211 or -111. Overall there are some hurdles toward its applicability, namely the large size of laminins, which make its delivery to target locations extremely challenging. An effective way to address this problem is to use shorter engineered proteins, such as the chimeric laminin/nidogen protein or mini-agrin, shown to be effective in a LAMA2-MD mouse model ( $\mathrm{dy}^{\mathrm{w}} / \mathrm{dy}^{\mathrm{w}}$; McKee et al., 2017; Reinhard et al., 2017). Probably in a near future, we will witness a new generation of laminin-binding proteins that, depending on the underlying genetic defects, are able to replace defective domains of laminin and promote the assembly of a stable and fully functional BM.

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## CONFLICT OF INTEREST

The authors have no conflict of interest to declare.

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## SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section at the end of the article.

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