NERC Open Research Archive



Article (refereed) - postprint

Everest, David J.; Shuttleworth, Craig M.; Stidworthy, Mark F.; Grierson, Sylvia S.; Duff, J. Paul; **Kenward, Robert E.** 2014. **Adenovirus: an emerging factor in red squirrel Sciurus vulgaris conservation.** *Mammal Review*, 44 (3-4). 225-233. <u>10.1111/mam.12025</u>

Copyright © 2014 Crown Copyright

This version available http://nora.nerc.ac.uk/509027/

NERC has developed NORA to enable users to access research outputs wholly or partially funded by NERC. Copyright and other rights for material on this site are retained by the rights owners. Users should read the terms and conditions of use of this material at http://nora.nerc.ac.uk/policies.html#access

This document is the author's final manuscript version of the journal article, incorporating any revisions agreed during the peer review process. Some differences between this and the publisher's version remain. You are advised to consult the publisher's version if you wish to cite from this article.

The definitive version is available at http://onlinelibrary.wiley.com

Contact CEH NORA team at <u>noraceh@ceh.ac.uk</u>

The NERC and CEH trademarks and logos ('the Trademarks') are registered trademarks of NERC in the UK and other countries, and may not be used without the prior written consent of the Trademark owner.

- 1 Adenovirus: An emerging factor in red squirrel Sciurus vulgaris
- 2 conservation
- 3
- 4 David J. EVEREST\* AHVLA-Weybridge, New Haw, Addlestone, Surrey KT15
- 5 3NB, UK. E-mail: david.everest@ahvla.gsi.gov.uk
- 6 Craig M. SHUTTLEWORTH Red Squirrels Trust Wales, Plas Newydd Country
- 7 House, Llanfairpwll, Anglesey LL61 6DQ, UK.
- 8 *E-mail:* craig.shuttleworth@rsst.org.uk
- 9 Mark F. STIDWORTHY International Zoo Veterinary Group, Station House,
- 10 Parkwood Street, Keighley, West Yorkshire BD21 4NQ, UK.
- 11 *E-mail: m.stidworthy@izvg.co.uk*
- 12 Sylvia S. GRIERSON AHVLA-Weybridge, New Haw, Addlestone, Surrey KT15
- 13 3NB, UK. E-mail: sylvia.grierson@ahvla.gsi.gov.uk
- 14 J. Paul DUFF AHVLA-Penrith, Diseases of Wildlife Scheme (AHVLA DoWS)
- 15 Calthwaite, Penrith, Cumbria CA11 9RR, UK. E-mail: paul.duff@ahvla.gsi.gov.uk
- 16 Robert E. KENWARD Centre for Ecology and Hydrology, Benson Lane,
- 17 Crowmarsh Gifford, Wallingford OX10 8BB, UK. E-mail: reke@ceh.ac.uk
- 18
- 19 \*Correspondence author.
- 20
- 21

# 22 ABSTRACT

24	1.	Adenovirus is an emerging threat to red squirrel Sciurus vulgaris			
25		conservation, but confirming clinically-significant adenovirus infections in			
26		red squirrels is challenging. Rapid intestinal autolysis after death in wild			
27		animals frequently obscures pathology characteristic of the disease in			
28		animals found dead.			
29	2.	We review the available literature to determine current understanding of			
30		both sub-clinical and clinically significant adenovirus infections in free-			
31		living wild and captive red squirrel populations.			
32	3.	Benefits of scientific testing for adenovirus incorporating both			
33		transmission electron microscopy (TEM) and polymerase chain reaction			
34		(PCR) technologies are compared and contrasted. We favour viral			
35		particle detection using TEM in animals exhibiting enteropathy at post			
36		mortem and the use of PCR to detect sub-clinical cases where no			
37		enteric abnormalities are observed.			
38	4.	Adenoviral infections associated with re-introduction studies are			
39		evaluated by examination of sporadic cases in wild populations and of			
40		data from captive collections used to service such studies.			
41	5.	The paucity of data available on adenovirus infection in grey squirrel			
42		Sciurus carolinensis populations is documented and we highlight that			
43		although sub-clinical virus presence is recorded in several locations in			

- Britain and Italy, no clinically-significant disease cases have been
  detected in the species thus far.
- 6. Current speculation for potential inter-specific infection between sciurids
  and other woodland rodents such as wood mice *Apodemus sylvaticus* is
  examined. Where sub-clinical adenovirus presence has been detected
  in sympatric populations occupying the same point food sources,
  husbandry methods may be used to diminish the potential for crossinfection.
- 52
   7. Our findings highlight the importance of controlling disease in red
   53 squirrel populations by using clearly defined scientific methods. In
   54 addition, we propose hypothetical conservation benefits of restricting
   55 contact rates between red squirrels and sympatric grey squirrels and of

56 limiting competition from other woodland rodent species.

57

58 KEY WORDS: adenovirus infection, conservation, disease, grey squirrel,

- 59 red squirrel
- 60
- 61
- 62
- 63
- 64

65

# 68 INTRODUCTION

69

70	Historically, disease was not recognized as a mechanism by which red squirrels
71	Sciurus vulgaris were replaced by grey squirrels S. carolinensis in a landscape.
72	Indeed, it was unclear initially whether the larger grey squirrel was directly
73	involved at all, was taking advantage of space vacated by natural fluctuations in
74	red squirrel population, or ultimately was better adapted to a larger range of
75	habitats (Middleton, 1931). Disease enzootics that were recorded in red squirrel
76	populations were notable for encompassing areas where grey squirrels were
77	absent (Shorten, 1954). Gurnell, (1987) noted "no evidence that grey squirrels
78	brought with them a disease which is causing the downfall of the red".
79	
80	By the 1990s, research had focussed heavily upon resource competition (Gurnell
81	and Pepper, 1993), including inter-specific differences in the relative efficiency
82	with which some tree seed is digested (Kenward and Holm, 1993). Inter-specific
83	resource competition, (Wauters et al. 2005), negative impacts on red squirrel
84	juvenile recruitment rates (Gurnell et al. 2004) and the effects of seed cache

piracy (Wauters *et al.* 2002) are today recognised as major contributors to red
squirrel extinction in sympatric populations.

87

However, progressive advances in viral research subsequently established that
grey squirrels carry the Squirrel pox virus (SQPV) as a sub-clinical infection, and

90 that inter-specific infection in sympatric red squirrels leads to epizootic disease 91 which is a significant factor in regional population declines in the UK (Rushton et 92 al. 2006; Sainsbury et al. 2008; Carroll et al. 2009; Bruemmer et al. 2010). Even 93 more recently, adenovirus infection has been identified increasingly as a cause of 94 mortality in free-living wild and captive red squirrel populations. An expanding 95 geographic distribution of cases has been revealed, affecting not only wild 96 populations, but increasingly being associated with high mortalities in captive collections used as both breeding stock and for use in wild population re-97 98 enforcement programmes. Additionally, grey squirrels have now been identified 99 as a sub-clinical carrier of the adenovirus among sympatric populations (Everest 100 et al. 2009; Romeo et al 2014).

101

102 Blood analyses, using enzyme linked immuno sorbent assay (ELISA) and tissue polymerase chain reaction (PCR) techniques, are routinely applied to determine 103 104 SQPV infection in both squirrel species. Parallel transmission electron 105 microscopy (TEM) screening of skin lesion material can be used to confirm the 106 presence of pox viral particles in typical advanced red squirrel cases. However, in 107 contrast to the detection of SQPV, the detection of adenovirus-associated or 108 clinically-significant adenovirus cases among red squirrels is challenging, due to 109 an absence or non-specificity of external clinical signs of disease. Until relatively 110 recently little was known about this infection in either squirrel species, or its 111 significance in red squirrel declines. Due to increasing scientific activity, both as a 112 retrospective exercise and proactive surveillance, a wider picture is gradually

113 emerging of the evolving impact that this virus is having with respect to both 114 sporadic disease cases in free-living wild squirrels across Great Britain and in red 115 squirrel re-introduction and captive breeding programmes. However, the mere 116 presence of adenovirus in the body does not signify disease. The virus may 117 indeed be present as a clinically significant infection, causing the death of the 118 animal; in this case viral particles can be detected by TEM in faecal material or 119 viral DNA can be amplified from tissue material such as the spleen. Adenovirus 120 can also be present as an asymptomatic infection or transience presence, 121 causing no apparent disease signs or indications of ill health, and the animal may 122 be outwardly healthy. Subsequent death due to an unrelated problem could then 123 show the presence of the amplified viral DNA by PCR analysis, whereas TEM 124 would fail to detect any viral particles.

125

126 Adenovirus infection damages the villi in the red squirrel intestinal mucosa, but 127 autolysis within hours of death typically confounds histological examination, by 128 precluding detection of characteristic adenovirus inclusion bodies (Erdélyi and 129 Duff, 2012), as seen in Fig. 1. By TEM on ultra-thin sections, these inclusions 130 have been shown to contain abundant viral particles (Fig. 2, arrowed). The 131 findings of enteropathy or diarrhoea are non-specific and are associated with 132 several other diseases (Everest et al. 2010a). While it is difficult to obtain 133 histologically-adequate gut wall samples prior to autolysis, experience shows that 134 gross pathological changes indicative of enteropathy, such as liquid intestinal 135 content, correlate strongly with gut viral particle detection by TEM (Fig. 3). The

136	presence of viral particles is therefore considered strongly suggestive of				
137	clinically-significant infection (Everest et al. 2012b). Nonetheless, in the most				
138	autolysed wild squirrel cases, pathologists may assume intestinal material to be				
139	of such limited value that it is not retained, even though archival samples of other				
140	tissues such as liver or spleen may be. Our understanding of the temporal and				
141	spatial scope of clinical adenovirus infection (Fig. 4) has recently been improved				
142	through more frequent proactive and reactive post mortem screening, in				
143	particular with TEM application.				
144					
145	We review the current understanding of infection and disease in red squirrels,				
146	grey squirrels and other small rodents such as wood mice Apodemus sylvaticus,				
147	with particular reference to the UK, and highlight key areas for future adenovirus				
148	infection research that have particular relevance to the applied conservation of				
149	the red squirrel.				
150					
151	RED SQUIRRELS				
152					
153	The geographical distribution of adenovirus infection				
154					
155	The first reports in the literature of adenovirus in free-living wild red squirrels from				
156	Great Britain were recorded from Suffolk (Sainsbury et al. 2001) and Cumbria,				

- 157 England (Duff *et al.* 2007), then from Wales (Everest *et al.* 2008), Scotland
- 158 (Everest *et al.* 2010a) and finally from Northern Ireland (Everest *et al.* 2012a),

159 demonstrating a wide geographical distribution (Fig. 4: Table 1). Retrospective 160 national surveillance of red squirrel mortalities across Great Britain, reported by 161 Martínez-Jiménez et al., (2011) revealed that 60 (12%) of 493 cases showed 162 enteric signs. Of these 60, 13 animals, all of which were exhibiting diarrhoea, 163 were selected for analysis by TEM. Of these 13, two animals (15%; Table 1), one 164 from Cumbria, the other from Lancashire, England were confirmed as adenovirus 165 cases by the TEM detection of viral particles. In another retrospective study, 166 adenovirus particles were identified by TEM in 10 (14%) of 70 free-living wild red 167 squirrels where enteropathy was suspected, from Cumbria, Lancashire and 168 Northumberland, England and Anglesey, Wales (Everest *et al.* 2010b; Table 1). 169 However, given the opportunistic sampling of post mortem cases and the paucity 170 of data from living animals, it is difficult to interpret the importance of adenovirus 171 as an overall contributor to mortality from these studies alone.

172

173 Sainsbury et al., (2001) and Martínez-Jiménez et al., (2011) both reported on a 174 population re-enforcement study at Thetford Chase (Suffolk, England) in the late 175 1990s, with animals that had been trans-located from Cumbria and had 176 contracted the infection and died in 1997 (Table 1). These animals may have been under stress that could have influenced the course of the disease. 177 178 Diarrhoea was associated with each of 10 adenovirus cases recorded in red 179 squirrels and intestinal haemorrhage or inflammation was observed in seven 180 cases. The extant Thetford Chase wild red squirrel population at that time was 181 judged to consist of 10 to 20 individuals (no more than 40, Gurnell et al. 1997),

and consequently adenovirus infection was a notable factor in the study.

183 Subsequent research, (Everest *et al.* 2012b; Table 1) has revealed adenovirus

184 infection to be associated with a high proportion of deaths in squirrels housed in

185 captive collections in Wales, indicating that viral epizootics can be locally

186 significant.

187

188 Of 13 captive deaths sampled from the re-introductions on the island of 189 Anglesey, situated off the North Wales coast, 12 (92%) were confirmed as 190 positive for the virus (three detecting viral particles by TEM and nine amplifying 191 viral DNA by PCR). Samples from 16 captive deaths at the Welsh Mountain Zoo, 192 Colwyn Bay, Wales (TWMZ) revealed viral DNA amplified by PCR in 14 (88%) 193 cases (Everest et al. 2012b). In a further 24 captive deaths originating from 194 zoological collections in England, for which tissue, faecal or intestinal content 195 samples were available, 20 (83%), were observed to be positive for adenovirus 196 (Everest et al., unpublished; Table1).

197

Analyses performed on 31 free-living wild red squirrels found dead on Anglesey revealed that 13 (42%) were positive for the virus. Of these positive cases, seven (54%), originated from within Newborough Forest and from these, five (71%), were identified as positive by PCR analyses, three of which also tested negative by TEM. One (14%), contained viral particles when analysed by TEM only, and one case was confirmed by both tests. The remaining six cases were from other Anglesey coniferous and broad-leaved woodlands; all were detected as viral

205 DNA carriers by PCR, but negative for viral particles by TEM (Everest *et al.*206 2012b).

207

208 In the latest published report of adenovirus in red squirrels from Great Britain, 209 Everest et al. (2013) record that nine (45%) of 20 animals were identified as 210 positive for the virus through amplification of viral DNA by PCR. These animals 211 derived from locations on the Isle of Wight, Jersey and Brownsea Island, all 212 islands off Great Britain without grey squirrels. Intestinal content samples from 12 213 of these 20 animals were originally examined by TEM and found to be negative 214 for virus particles (Everest *et al.* 2010b). This shows the benefit of using parallel 215 TEM and PCR screening to determine sub-clinical virus presence, which can 216 easily go undetected.

217

218 There are very limited reports of adenovirus outbreaks in red squirrels from 219 outside the UK. One, involving three deaths, was from a captive collection in 220 Germany (Peters et al. 2011); in the other, 77 road traffic accident carcasses 221 from Italy were examined by a combination of TEM and PCR analyses(Romeo et 222 al. 2014). Twelve (16%) were positive for amplified viral DNA by PCR (Table 1). 223 As with the outbreak in Germany (Peters et al. 2011), and unlike the situation in 224 most of Great Britain, viral presence was detected in red squirrel populations 225 from areas where the grey squirrel was not known to be present in the immediate 226 landscape.

227

# 228 Adenovirus presence determined in deaths by other causes

230	Traumatic deaths, such as drowning and road traffic accidents, have revealed
231	animals positive for amplification of viral DNA by PCR, but negative for faecal
232	viral particle detection by TEM. These cases occur in animals which lack enteric
233	abnormalities at post mortem examination. These findings suggest that sub-
234	clinical infections are present and may be widespread within wild British
235	populations of red squirrels (Everest et al. 2012b; Table 1).
236	
237	Adenovirus strain speciation
238	
239	Phylogenetic analysis demonstrates that adenovirus sequences from squirrel
240	samples cluster with mastadenoviruses but are distinct from other adenoviruses
241	within the genus (Sainsbury et al. 2001, Peters et al. 2011), although squirrel
242	adenovirus has not yet been approved as a species (King et al. 2011).
243	Sequencing has revealed a lack of adenovirus strain variability. The identity of
244	the adenovirus in a partial fragment of the hexon gene from the German outbreak
245	(GU735084) described by Peters et al., (2011) was identical to the putative
246	Suffolk strain (Sainsbury et al. 2001). In contrast, in those cases described by
247	Everest et al. (2012b), sequences were detected which were identical to those
248	found in Cumbrian cases from 2007 (JN205244.1). Everest et al. (2012b) used a
249	partial fragment of the polymerase gene, which in turn identified cases that were
250	genetically identical to the grey squirrel cases detected on Anglesey (Everest et

251 al. 2009). This is remarkable, as the cases were separated both spatially and 252 temporally. It has been suggested, therefore, that the viruses involved in each of 253 these cases are very closely related, or perhaps identical (Peters *et al.* 2011). 254 255 In general, the samples described above have not been randomly sourced and 256 case selection was influenced by carcass suitability and value in terms of post 257 mortem examination. This is particularly true for captive collections, where the 258 prevailing close confinement within enclosures would have allowed for easy 259 spread of the virus between individual animals, thus accounting for the 260 apparently high incidence of infection in such collections. 261 262 263 **GREY SQUIRRELS** 264 265 Given the role that grey squirrels play in SQPV infection in red squirrel 266 populations, it is natural to investigate whether sympatric grey squirrel 267 populations are also a source of inter-specific adenovirus infection. 268 Romeo et al., (2014) found PCR amplified adenoviral DNA in only two (1%) of 269 232 grey squirrels from Italy. Screening of tissues from wild adult grey squirrels 270 (n=18) trapped and euthanased at the Welsh Mountain Zoo in 2011 failed to

- 271 reveal viral particles in the gut by TEM (which would have suggested clinically-
- significant infection), yet 10 of these 18 animals (56%) were positive by PCR
- analyses on spleen tissue, (Everest et al. unpublished) and were hence

determined as adenovirus carriers. Although the numbers of animals were small
in this study, the PCR figure is very similar to the 60% sero-prevalence reported
by Greenwood and Sanchez, (2002) using murine adenovirus ELISA tests for
antibodies in grey squirrels from the same zoo; a location where dead captive red
squirrels have been found with enteric symptoms and viral particles in the
intestinal tract.

280

281 At the Newborough Forest re-introduction site on Anglesey, adenovirus DNA was 282 detected by PCR analysis from two grey squirrels caught in 2006 (Everest et al. 283 2009). Wider PCR screening of archived and proactively-sourced blood 284 sampling, involving over 200 samples and thus forming a study larger than that 285 reported by Romeo et al. (2014), was subsequently undertaken and reported by 286 Everest et al. (2012b, Table 2) for both Anglesey locations and woodland in 287 Gwynedd, North Wales, within a few kilometres of the Menai Straits. Spleen 288 tissue collected from the Gwynedd site was examined in 2012 (Everest et al. 289 unpublished), and amplification of DNA revealed a much higher percentage of 290 positives (54%) than in blood (7%, see Table 2).

291

292 The 2012 Gwynedd result (Everest *et al.* unpublished) was further confirmed,

when both spleen and blood were available for analysis from each of 14 adult

grey squirrels trapped at the Welsh mountain Zoo. Adenovirus DNA was detected

from spleen tissue in eight cases (57%), but there were no positive results from

the 14 blood samples from the same animals (Everest *et al.* unpublished), thus

demonstrating that source tissue type is an important consideration in adenovirusscreening.

299

Historically, assessing infection in grey squirrels is challenging, as previously reported blood based testing was serologically based. Thus exposure to the virus could result in potentially long-lasting sero-conversion, although this may wane with age. In contrast, an animal may be viraemic (and therefore PCR-positive) for only a short period, meaning PCR analyses have only a small time window to be effective for viral diagnosis. In this context, serologically- based ELISA analyses may be more sensitive in nature than PCR techniques.

307

Although evidence of infection has been found, no clinically-significant cases of adenovirus have been identified to date in grey squirrels and viral particles have been absent from intestinal content examined by TEM studies of grey squirrels from Cumbria (*n*=36), Wales (*n*=58, Everest *et al.* unpublished), Thetford Chase study (*n*=10, Martínez-Jiménez *et al.* 2011) and Italy (*n*=3, Romeo *et al.* 2014).

314

## 315 SMALL RODENTS

316

Peters *et al.*, (2011), documented adenovirus infection by TEM in a captive red
squirrel collection from Germany, and red squirrel infections have been recorded
on both the Isle of Wight and Jersey (Everest *et al.* 2013), all of which are regions

where the grey squirrel is absent. Additionally, Romeo *et al.*, (2014) documented infections in red squirrels in areas where the grey squirrel was not known to be present. This means that alongside intra-specific and potential grey squirrel to red squirrel infections, inter-specific infection from other small rodents such as wood mice is possible.

325

326 In order to investigate this potential infection route, Everest *et al.*, (2013) 327 examined the spleens of wood mice trapped on the Island of Anglesey for the 328 presence of adenovirus by PCR analyses. Adenoviral DNA was amplified from 329 three of 15 mice (20%), trapped at two woodland sites which had red squirrel 330 feeding stations and where cases of clinically-significant adenovirus infection of 331 wild red squirrels had been recorded. Two of 24 (8%) mice trapped in north 332 Wales within woodland enclosures housing captive red squirrels also tested 333 positive for adenovirus by PCR. Our results therefore demonstrate the potential 334 for adenovirus infection in sympatric communities of grey squirrel, red squirrel 335 and wood mice.

336

The PCR primers used to test the wood mice samples had been designed based
on a sequence of the adenoviral DNA polymerase gene from squirrel samples
(JN205244.1; Everest *et al.* 2012). However, further investigations into whether
these primers would detect other adenoviruses were not undertaken.
It is therefore unclear at present whether the strain detected in mice is identical to
that detected in squirrels, and so further molecular sequencing is required.

343 Greenwood and Sanchez (2002) used an ELISA derived for the serological

344 detection of murine adenoviruses to detect adenovirus in grey squirrels.

345 Therefore, it is possible that either cross reactivity exists between species-

346 specific viruses from the two species, or an identical virus infects both.

347

348

# 349 **DISCUSSION**

350

351 Retrospective study of archived tissue and blood samples (Everest et al. 2010; 352 2012b; Martínez-Jiménez et al. 2011) has advanced our understanding of both 353 the temporal and spatial distribution of adenovirus infection within red squirrel 354 populations. Recent examination of trauma deaths has also revealed sub-clinical 355 infections in wild individuals at the time of death, namely negative TEM results for 356 adenovirus particles in faecal and intestinal samples but positive results for viral 357 DNA from tissues by PCR analysis (Everest et al. 2012b; Romeo et al. 2014). 358 Much however, remains unclear about the epizootiology, in particular, the roles of 359 squirrel population density and stress. Currently, much of our understanding is 360 based upon captive collections and there is therefore also a pressing research 361 need to investigate the distribution and effects of the infection among wild red 362 and grey squirrel populations. Opportunities for application of a qPCR technique 363 to quantitate virus load in faeces, tissues and blood in order to partition 364 pathological from asymptomatic infections would also be beneficial. 365

Research in North Wales (Everest *et al.* 2009; 2012b; Greenwood and Sanchez,
2002), and in Italy by Romeo *et al.*, (2014) has demonstrated adenovirus
infection or exposure in grey squirrels, but whether this has any clinical
significance in these populations remains unknown. To this end, a controlled

370 challenge experiment in grey and red squirrels using the same virus isolate would

also help to advance our understanding.

372

373 There is also a paucity of data on adenovirus prevalence within regional grey

374 squirrel populations in the UK. An annual survey combining PCR and TEM

analyses was limited solely to squirrel populations in North Wales. Additional

376 regional studies of this type would therefore be useful

377

378 Given that grey squirrels appear to be infected with both adenovirus and SQPV, 379 the accepted management practice of removing grey squirrel populations to 380 control SQPV infections would also mitigate the potential for adenovirus infection. 381 Additionally, conservation managers could potentially evolve protocols to combat 382 potential infection pathways involving other woodland rodents such as wood 383 mice, although there may be a significant cost implication to this approach. On 384 Anglesey, adenovirus infection risk was highlighted as a major difficulty faced 385 during the re-introduction of red squirrels to Newborough Forest (Shuttleworth et 386 al. 2008). Release protocols have been modified with animals now housed for 387 only a few weeks, during which faecal and blood screening is undertaken for 388 adenovirus (Shuttleworth, 2010). More widely, it has been recommended that

hygiene protocols at supplemental feeding hoppers routinely focus upon limiting
adenovirus infection via faecal-oral routes (Everest *et al.* 2012b). Given our
recent findings and because of the potential for transmission of other rodentborne infections, this should encompass mouse control.

393

394 If wood mice act as an infection reservoir, there are obvious implications for 395 scenarios that concentrate their activities at point food sources such as garden 396 bird tables or supplemental feed hoppers also visited by red squirrels. It may 397 therefore be prudent to place red squirrel supplemental feed hoppers on posts 398 with cone shaped baffles near the base to prevent mice from accessing the 399 hopper above, instead of, as is common practice, fixing hoppers to tree trunks, 400 which allows mice easy access. Accumulation of discarded shells and food 401 remains beneath hoppers should be minimised. Trapping protocols should 402 include regular disinfection of all traps, not only those that have contained grey 403 squirrels, so as to limit any potential mouse to red squirrel inter-specific virus 404 transmission.

405

406

#### 407 CONCLUSIONS

408

This review of the available evidence within the published literature, coupled with recent findings, lead us to conclude that adenovirus should be regarded as a

411 serious disease threat to the various red squirrel re-introduction and captive

412 breeding programmes, and to red squirrel populations in places where grey 413 squirrels, red squirrels and wood mice can interact at point food sources. We also 414 conclude that TEM, while excellent at detecting clinically-significant infection from 415 intestinal samples, is not as sensitive as PCR for detecting sub-clinical 416 adenovirus cases, and that spleen tissue is a better material to screen by PCR 417 than blood. ELISA-based assay on blood samples is the only test available for 418 live animals at present. To address practical and potentially also welfare 419 considerations, alternative assay platforms should be investigated for live animal 420 testing. We would also recommend that disease investigations and adenoviral 421 infection surveillance extend to all three rodent species identified in this review, 422 and where possible, include parallel PCR and TEM sample testing of tissue and 423 intestinal content samples, respectively.

424

425

## 426 ACKNOWLEDGEMENTS

427

The authors extend grateful thanks to those individuals and zoological collections that provided red and grey squirrel sample material; to the Animal Health and Veterinary Laboratories Agency Diseases of Wildlife Scheme (AHVLA DoWS) and the International Zoo Veterinary Group who both provided veterinary examination services input; and to Defra for the provision of funding for some of the analyses undertaken as part of their scanning surveillance programme via the AHVLA DoWS. Research in North Wales was funded by the Nineveh Trust,

435	Environment Wales, Red Squirrels Survival Trust and Red Squirrels Trust Wales
436	whereas research in Italy was funded by the European Squirrels Initiative.
437	Thanks must go to Alex Schock of AHVLA Lasswade for kindly permitting us to
438	reproduce her image for Figure 1 and to Simon O'Hare of Red Squirrels Northern
439	England for kindly producing the squirrel adenovirus case map (Fig. 4).
440	
441	
442	REFERENCES
443	
444	Bruemmer CM, Rushton SP, Gurnell J, Lurz PWW, Nettleton P, Sainsbury AW,
445	Duff JP, Gilray J, McInnes CJ (2010) Epidemiology of squirrel pox virus in grey
446	squirrels in the UK. Epidemiology and Infection 138: 941-950
447	
448	Carroll B, Russell P, Gurnell J, Nettleton P, Sainsbury AW (2009) Epidemics of
449	squirrel pox virus disease in red squirrels (Sciurus vulgaris): temporal and
450	serological findings. Epidemiology and Infection 137: 257-265
451	
452	Duff J, Higgins R, Farrelly S (2007) Enteric adenovirus infection in a red squirrel
453	(Sciurus vulgaris). Veterinary Record 160: 384
454	
455	Erdélyi K, Duff JP. Adenovirus infection in squirrels. In: Gavier-Widen D, Duff JP
456	and Meredith A (eds) Infectious diseases of wild animals and birds in Europe
457	Wiley, UK

- 459 Everest DJ, Butler H, Blackett T, Simpson VR, Shuttleworth CM (2013)
- 460 Adenovirus infection in red squirrels in areas free from grey squirrels. Veterinary
- 461 *Record* **173**: 199-200
- 462
- 463 Everest DJ, Grierson SS, Meredith AL, Milne EM (2010a) Adenovirus in a red 464 squirrel (*Sciurus vulgaris*) from Scotland. *Veterinary Record* **167**: 184
- 465
- 466 Everest DJ, Grierson SS, Stidworthy MF, Shuttleworth C (2009) PCR detection of
- adenovirus in grey squirrels on Anglesey. *Veterinary Record* **165**: 482

- 469 Everest DJ, Griffin J, Warnock ND, Collins L, Dick J, Reid N, Scantlebury M,
- 470 Marks N, Montgomery I (2012a) Adenovirus particles from a wild red squirrel
- 471 (Sciurus vulgaris) from Northern Ireland. Veterinary Record 170:188
- 472
- 473 Everest DJ, Shuttleworth CM, Grierson SS, Duff JP, Jackson N, Litherland P,
- 474 Kenward RE, Stidworthy MF (2012b) A systematic assessment of the impact of
- 475 adenovirus infection on a captive re-introduction project for red squirrels (Sciurus
- 476 vulgaris). Veterinary Record **171**: (7)176
- 477
- 478 Everest DJ, Stidworthy MF, Milne EM, Meredith AL, Chantrey J, Shuttleworth
- 479 CM, Blackett T, Butler H, Wilkinson M, Sainsbury AW (2010b) Retrospective
- 480 detection by negative contrast electron microscopy of faecal viral particles in wild

481	red squirrels	(Sciurus	vulgaris) v	vith suspected	enteropathy in	Great Britain.
		<b>`</b>	<b>J</b> ,			

- 482 Veterinary Record **167**: 1007-1010
- 483
- 484 Everest DJ, Stidworthy MF, Shuttleworth C (2008) Adenovirus-associated
- 485 mortalities in red squirrels (*Sciurus vulgaris*) on Anglesey. *Veterinary Record*
- 486 **163: 430**
- 487
- 488 Greenwood AG and Sanchez S (2002) Serological evidence of murine pathogens
- in wild grey squirrels (*Sciurus carolinensis*) in north Wales. *Veterinary Record*
- 490 **150:** 543-546
- 491

492 Gurnell J. (1987) The natural history of Squirrels. p 162 Christopher Helm,

- 493 London.UK
- 494
- 495 Gurnell J. and Pepper H. (1993). "A critical look at conserving the British Red

496 Squirrel (Sciurus vulgaris)." Mammal Review 23: (3-4): 127-137

- 498 Gurnell J, Sainsbury AW, Venning T. (1997) Conserving the red squirrel in
- 499 Thetford Forest. p 59 English Nature Research Report, Peterborough, UK
- 500
- 501 Gurnell J, Wauters L, Lurz PWW, Tosi G (2004) Alien species and inter-specific
- 502 competition: effects of introduced eastern grey squirrels on red squirrel
- 503 population dynamics. *Journal of Animal Ecology* **73:** 26-35

Kenward RE. and Holm JL. (1993). On the replacement of the Red Squirrel in
Britain: A phytotoxic explanation. Proceedings of the Royal Society Series B **251:** 187-194
King AMQ, Adams MJ, Carstens EB, Lefkowitz EJ (eds; 2011) *Virus Taxonomy:Classification and Nomenclature of Viruses*. Ninth Report of the

511 International Committee on Taxonomy of Viruses. Elsevier, Amsterdam, The

512 Netherlands.

513

514 Martínez-Jiménez D, Graham D, Couper D, Benkö M, Schöniger S, Gurnell J,

515 Sainsbury AW (2011) Epizootiology and pathologic findings associated with a

newly described adenovirus in the red squirrel, (*Sciurus vulgaris*). Journal of

517 Wildlife Diseases 47: 442-454

518

519 Middleton AD. (1931) The Grey Squirrel. p78-80 Sidgwick and Jackson Ltd,

520 London. UK

521

522 Peters M, Vidovszky MZ, Harrach B, Fischer S, Wohlsein P, Kilwinski J (2011)

523 Squirrel adenovirus type 1 in red squirrels (*Sciurus vulgaris*) in Germany.

524 *Veterinary Record* **169:** 182

525

526	Romeo C, Ferrari N, Rossi C, Everest DJ, Grierson SS, Lanfranchi P, Martinoli A,
527	Saino N, Wauters LA, Hauffe HC. (2014) Ljungan virus and an adenovirus in
528	Italian squirrel populations. Journal of Wildlife Diseases 50: (2) in press, DOI:
529	10.7589/2013-10-260
530	
531	Rushton SP, Lurz PWW, Gurnell J, Nettleton P, Bruemmer C, Shirley MDF,
532	Sainsbury AW (2006) Disease threats posed by alien species: the role of a
533	poxvirus in the decline of the native red squirrel in Britain. Epidemiology and
534	Infection <b>134:</b> 521-533
535	
536	Sainsbury AW, Adair B, Graham D, Gurnell J, Cunningham AA, Benko M, Papp
537	T (2001) Isolation of a novel adenovirus associated with splenitis, diarrhoea and
538	mortality in trans-located red squirrels, (Sciurus vulgaris). Verhandlungs Bericht
539	über die Erkrankung der Zootiere <b>40:</b> 265-270
540	
541	Sainsbury AW, Deaville R, Lawson B, Cooley WA, Farrelly SS, Stack MJ, et al.
542	(2008) Pox viral disease in red squirrels (Sciurus vulgaris) in the UK: spatial and
543	temporal trends of an emerging threat. Ecohealth 5: 305-316
544	
545	Shorten MR. (1954) Squirrels. p 71 Collins, London UK
546	
547	Shuttleworth CM (2010) Turning the grey tide: progress in red squirrel recovery.
548	Ecos <b>31</b> : 27-35

550	Shuttleworth CM, Kenward RE, Jackson N (2008) Re-introduction of the red
551	squirrel into Newborough forest on the island of Anglesey, UK. In: Soorae PS
552	(ed) Global re-introduction perspectives: Re-introduction case-studies from
553	around the globe. 163-166 IUCN/SSC Re-introduction Specialist Group, Abu
554	Dhabi UAE
555	
556	Wauters LA, Tosi G, Gurnell J, (2002) Inter-specific competition in tree squirrels:
557	do introduced grey squirrels (Sciurus carolinensis) deplete tree seeds hoarded by
558	red squirrels (Sciurus vulgaris)? Behavioral Ecology and Sociobiology 51: 360-
559	367
560	
561	Wauters LA, Tosi G, Gurnell J, (2005) A review of the competitive effects of alien
562	grey squirrels on behavior, activity and habitat use of red squirrels in mixed
563	deciduous woodland in Italy: <i>Hystrix Italian Journal of Mammalogy</i> (n.s.) <b>16:</b> (1)
564	27-40
565	
566	
567	
568	
569	
570	
571	

**Table 1.** Test results for adenovirus from tissues or intestinal or faecal content

573 material from red squirrels *Sciurus vulgaris*.

Reference	Study location	Number	Number (%)
		Tested	Positive
Duff <i>et al</i> . 2007	Cumbria wild	2	2/2(100%)
Everest <i>et al.</i> 2008	Anglesey captive	3	3/3 (100%)
Everest <i>et al</i> . 2010a	Scotland wild	1	1 (100%)
Everest <i>et al</i> . 2012a	Northern Ireland wild	2	1/2 (50%)
Martínez-Jiménez <i>et al</i> . 2011	Great Britain wild	13	2/13 (15%)
Martínez-Jiménez <i>et al</i> . 2011	Suffolk captive	10	10/10 (100%)
Everest <i>et al</i> . 2010b	Great Britain wild	70	10/70 (14%)
Sainsbury <i>et al</i> . 2001	Suffolk captive	6	3/6 (50%)
Everest <i>et al.</i> 2012b	Anglesey captive	13	12/13 (92%)
Everest <i>et al</i> . 2012b	Zoo captive Wales	16	14/16 (88%)
Everest <i>et al</i> . 2012b	Anglesey wild	31	13/31 (42%)
Everest, <i>et al.</i> unpublished	England captive	24	20/24 (83%)
Everest <i>et al.</i> 2013	Isle of Wight/	20	9/20 (45%)
	Jersey wild		
Peters <i>et al</i> . 2011	Germany captive	3	3/3 (100%)
Romeo <i>et al.</i> 2014	Italy wild	77	12/77 (16%)

**Table 2.** Positive PCR test results for adenovirus DNA from blood and spleen
tissue from grey squirrels Sciurus carolinensis in North Wales. Total number
tested and percentage positive are shown.

	Anglesey		Gwynedd (Bangor Area)	
	Blood	Spleen	Blood	Spleen
2007 <sup>1</sup>	0% (55)	-	-	-
2010 <sup>1</sup>	23% (26)	-	21% (39)	-
2011 <sup>2</sup>	-	-	10% (48) <sup>3</sup>	-
2012 <sup>2</sup>	25% (4)	-	7% (15)	54% (35)

<sup>583</sup> <sup>1</sup> Everest *et al.* (2012b); <sup>2</sup> Everest *et al.* unpublished.

<sup>585</sup> <sup>3</sup>10% (n=48), adults were 14% (n=28) and juveniles 5% (n=20)

- **Figure 1.** Haematoxylin and Eosin histology image of a section of red squirrel
- 593 Sciurus vulgaris small intestine, showing intra-nuclear virus inclusion bodies
- 594 (arrowed) and extensive damage to the villi, findings consistent with adenovirus
- 595 infection. x600 magnification.



- **Figure 2.** Micrograph of adenovirus particles (arrowed) detected in an ultra-thin
- 600 section of enterocytes from a red squirrel *Sciurus vulgaris* large intestine. Bar
- 601 (bottom right) = 500 nm.



- **Figure 3.** Micrograph of adenovirus particles detected in contents from the large
- 604 intestine of a captive red squirrel *Sciurus vulgaris*. *Bar* = 100nm.



- **Figure 4.** Location of adenovirus positive free-living wild (●) and captive (□) red
- 607 squirrel *Sciurus vulgaris* cases from Great Britain, as analysed by PCR and TEM.



## All cases of adenovirus in red squirrels in the UK to June 2013

Crown Copyright 2013. Northumberland WildlifeTrust. OS Licence No. AL100035023.