NERC Open Research Archive



Article (refereed) - postprint

Hentley, William T.; Hails, Rosemary S.; Johnson, Scott N.; Jones, T. Hefin; Vanbergen, Adam J.. 2014. **Top-down control by Harmonia axyridis mitigates the impact of elevated atmospheric CO2 on a plant-aphid interaction.** *Agricultural and Forest Entomology*, 16 (4). 350-358. <u>10.1111/afe.12065</u>

© 2014 The Royal Entomological Society

This version available http://nora.nerc.ac.uk/508562/

NERC has developed NORA to enable users to access research outputs wholly or partially funded by NERC. Copyright and other rights for material on this site are retained by the rights owners. Users should read the terms and conditions of use of this material at http://nora.nerc.ac.uk/policies.html#access

This document is the author's final manuscript version of the journal article, incorporating any revisions agreed during the peer review

article, incorporating any revisions agreed during the peer review process. Some differences between this and the publisher's version remain. You are advised to consult the publisher's version if you wish to cite from this article.

The definitive version is available at http://onlinelibrary.wiley.com

Contact CEH NORA team at <u>noraceh@ceh.ac.uk</u>

The NERC and CEH trademarks and logos ('the Trademarks') are registered trademarks of NERC in the UK and other countries, and may not be used without the prior written consent of the Trademark owner.

1	Agricultural and Forest Entomology			
2				
3				
4	Top-down control by Harmonia axyridis mitigates			
5	the impact of elevated atmospheric CO ₂ on a plant-			
6	aphid interaction			
7				
8				
9				
10	WILLIAM T. HENTLEY ^{1,2,3} , ROSEMARY S. HAILS ¹ , SCOTT N.			
11	JOHNSON ⁴ , T. HEFIN JONES ³ , & ADAM J. VANBERGEN ⁵			
12				
13				
14	¹ CEH Wallingford, Maclean Building, Benson Lane, Crowmarsh Gifford,			
15	Wallingford, Oxfordshire, OX10 8BB, U.K.			
16	² The James Hutton Institute, Dundee, DD2 5DA, U.K.			
17	³ Cardiff School of Biosciences, Cardiff University, Cardiff, CF10 3AX, U.K.			
18	⁴ Hawkesbury Institute for the Environment, University of Western Sydney,			
19	Australia.			
20	⁵ Centre for Ecology & Hydrology (CEH), Bush Estate, Penicuik, Midlothian,			
21	EH26 0QB, U.K.			
22				
23				
24	Correspondence: William Hentley, Centre for Ecology & Hydrology,			
25	Maclean Building, Benson Lane, Crowmarsh Gifford, Wallingford, Oxfordshire,			
26	OX10 8BB, United Kingdom.			
27	E-mail: <u>wihen@ceh.ac.uk</u>			
28				
29				
30				
31				
32	Running title: Multi-trophic interactions in an elevated CO ₂ environment.			

33 Abstract

34

1) This study investigated the impact of elevated atmospheric CO_2 (390 or 650 μ mol/mol) on raspberry genotypes varying in resistance to the large raspberry aphid (*Amphorophora idaei*) and the subsequent impacts on the coccinellid predator *Harmonia axyridis*.

- 2) CO₂ enrichment promoted plant growth, ranging from 30% in the partially
 susceptible cultivar to over 100% increase for the susceptible cultivar.
- 3) Aphid abundance and colonisation (presence-absence) on the susceptible
 cultivars were not influenced by CO₂ enrichment. On the resistant cultivar,
 aphid colonisation increased from 14% in ambient CO₂ to 70% in elevated
 CO₂ with a subsequent increase in aphid abundance, implying a
 breakdown in resistance. Inclusion of the natural enemy on the resistant
 cultivar, however, suppressed the increase in aphid abundance at
 elevated CO₂.

48 4) This study highlights how crop genotypes vary in responses to climate
49 change; some cultivars can become more susceptible to aphid pests
50 under elevated CO₂. We do, however, demonstrate the potential for top
51 down control to mitigate the effect of global climate change on pest
52 populations.

- 53
- 54
- 55

56

57

58

59 Introduction

By 2100, atmospheric CO₂ concentrations are predicted to double pre-60 61 industrial levels of 280 µmol/mol (Meehl et al., 2007). There is growing 62 interest in understanding how insect herbivores found on crops will respond to 63 such global climate change, particularly in the context of achieving food 64 security (Gregory et al., 2009). While there is expanding literature on the 65 effects of elevated atmospheric CO₂ concentrations (eCO₂) on plant-66 herbivore interactions (Robinson et al., 2012; Zavala et al., 2013), only a few 67 studies have addressed crop cultivars with genetic resistance to insect pests 68 (e.g. Zavala et al., 2008; Sun et al., 2013). Moreover, to date these studies of 69 crop resistance have largely overlooked the indirect effects of eCO₂ on the 70 natural enemies of crop pests. It is these organisms which will ultimately 71 determine the net effect of eCO₂ on pest population dynamics (Robinson et 72 al., 2012). Given the need to increase food production by 50% by 2050 while 73 using less resources and pesticides (Royal Society, 2009), understanding 74 how climate change will affect ecosystem services such as predation of 75 herbivorous pests, and the underlying mechanisms, is of paramount 76 importance (A'Bear et al., 2014).

77 In the absence of trophic interactions, plants, which rely on CO₂ assimilation 78 for energy, generally respond positively to eCO₂, with 25-38% increases in 79 biomass being reported for C₃ plants (Stiling & Cornelissen, 2007; Robinson 80 et al., 2012). Within plant tissue, carbohydrates generally increase and 81 nitrogen content is either diluted due to increased carbohydrates or 82 reallocated, resulting in an average 19% increase in plant C:N ratio (Robinson 83 et al., 2012), ultimately altering many aspects of plant chemistry (Stiling & 84 Cornelissen, 2007). Plant resistance is multifaceted, involving direct (physical

and antibiotic) and indirect (volatile organic carbons to attract natural enemies) mechanisms (Turlings *et al.*, 1990; Schaller, 2008). Modification of plant defences in an enriched CO₂ atmosphere has been attributed to changes in plant chemistry (Zavala *et al.*, 2008).

89 The response of herbivores to the indirect effects of eCO₂ are modulated by 90 feeding guild and the plant species (Robinson et al., 2012). By feeding directly on the phloem, aphids can circumvent many of the plant defences associated 91 92 with feeding on plants (Raven, 1983). A meta-analysis by Robinson et al. 93 (2012) found only 15 studies investigating the response of phloem-feeding 94 insects to eCO₂, somewhat surprising given the significant damage they can 95 cause to host plants (Zvereva et al., 2010). Despite this, aphid abundance 96 and fecundity generally increases in eCO₂, suggesting a reduction in plant 97 resistance to aphid herbivory. Indeed several crop varieties have recently 98 been shown to become more susceptible to aphid herbivory under eCO_2 , via 99 manipulation of host plant chemistry and down regulation of the ethylene 100 pathway (Guo et al., 2013; Sun et al., 2013) In the present study, we 101 investigated the effects of eCO_2 on red raspberry (*Rubus idaeus* L.) 102 susceptibility to the European large raspberry aphid (Amphorophora 103 idaei Börner). Martin and Johnson (2011) demonstrated that this system is 104 affected by eCO₂; in particular the authors found that a partially resistant 105 cultivar became more susceptible to A. idaei. That study did not however, 106 include higher trophic groups, which have the potential to moderate these 107 effects (Martin & Johnson, 2011).

108 The inclusion of higher trophic levels within the community may mitigate the 109 breakdown of aphid resistance. The impact of eCO_2 on the plant may,

110 however, transfer to herbivores on the host plant. Aphids feeding on host 111 plants with low C:N ratio may have a high nutritional value for predators 112 (Couture et al., 2010), therefore in a high CO₂ environment, where the C:N 113 ratio is increased, predators may require greater numbers of prey to fulfil their 114 physiological demands. This is analogous to compensatory feeding seen in 115 herbivores (e.g. Watt et al., 1995) and detritivores (e.g. Dray et al., 2014). 116 There are, however, very few studies investigating the interacting effects of 117 bottom-up (host plant quality) and top-down (predation) on aphid abundance 118 in eCO₂, particularly for woody plants. By using a gradient of plant resistance 119 to aphid herbivory, this study aims to increase our understanding of how tritrophic interactions are impacted by an eCO₂ environment. We specifically 120 121 extend earlier research (Martin & Johnson, 2011) through inclusion of different 122 cultivars and also a natural enemy of the aphid. Since plant architecture and habitat complexity are important considerations for assessing the realistic 123 124 efficacy of natural enemies (Langelotto & Denno, 2004) our study also used larger, structurally complex plants compared to Martin and Johnson (2011). 125

126 We test the following hypotheses:

127 H_1) Raspberry plants, like most C_3 plants, respond positively to elevated 128 levels of atmospheric CO_2 . The magnitude of the response will be cultivar 129 specific, with the biggest increases in biomass in the partially resistant and 130 resistant cultivars (Martin & Johnson, 2011).

H₂) Aphid abundance will be distributed according to plant resistance with
more aphids on the susceptible cultivars. Under eCO₂ aphid abundance and
size will increase on less resistant cultivars (Martin & Johnson, 2011).

H₃) Predation levels will increase to compensate for changes in prey quality.
Consumption of prey from eCO₂ will increase development time and adult
mass of predators.

137

138 Materials and Methods

139 Chambers

140 Experiments were carried out in four controlled environment chambers 141 (approx. 4m x 10m) of the GroDome[™] climate change research facility at the 142 Centre for Ecology and Hydrology (CEH), Wallingford, UK. Chamber environments were maintained at $18 \pm 1^{\circ}$ C, 50-70% relative humidity. When 143 photosynthetic active radiation (PAR) dropped below 400µmol.s⁻¹.m⁻², 12 x 144 145 400W halide bulbs positioned approximately 1m above the plants 146 supplemented natural daylight in each chamber. A 16h photoperiod was 147 maintained. Chamber air cycled with outside air approximately four times 148 every hour, the industry standard (Buffington et al., 2013). Two of the 149 experimental chambers were maintained at ambient (390 \pm 50 μ mol/mol) and 150 two at elevated (650 \pm 50 μ mol/mol) atmospheric CO₂ levels. A CO₂ sensor 151 (Vaisala GMW22) was mounted in each chamber and connected to a controller unit (Mitsubishi Micro-controller AL2-24MR-D). Once CO₂ levels fell 152 153 below the target concentration (390 μ mol/mol and 550 μ mol/mol, 154 respectively), CO₂ gas (BOC) was injected for 1-second followed by 30-155 second delay, repeating until the target concentrations were reached.

156 Host plant

157 Three cultivars of European red raspberry (*R. idaeus*), varying in resistance to 158 aphid herbivory, were used in the experiment. Glen Ample possesses a

159 resistance gene (A₁), now largely ineffective following adaptation by aphid 160 biotypes (Birch et al., 2004) and thus represents the plant least resistant to 161 herbivory. Glen Clova has partial resistance to aphid herbivory underpinned 162 by multiple genes (multi-genic) (McMenemy et al., 2009). Octavia is highly 163 resistant to aphid herbivory, possessing two resistance genes (A_{10} and A_{k4a}) (Knight & Fernández-Fernández, 2008). Plants were grown from root-stock at 164 165 the James Hutton Institute (JHI), Dundee, UK. When approximately 1cm in 166 height, the plants were transferred to CEH where they were potted-out into 3L 167 pots filled with peat-based compost (Levington M3, no additional fertiliser) and 168 randomly allocated to CO₂ treatments. All plants were grown in ambient or 169 elevated CO₂ conditions for approximately five weeks prior to the experiment 170 commencing.

171 Aphids

172 The European large raspberry aphid (Amphorophora idaei) is a specialist 173 phloem-feeding herbivore, found only on the European red raspberry causing 174 direct and indirect (vectors four plant-viruses) economic damage to fruit crops 175 (McMenemy et al., 2009). Insect herbivore biotypes are populations that differ 176 in their ability to utilize a certain trait of a plant genotype/cultivar (Smith, 177 2005). The large raspberry aphid biotype (Biotype 2) used in this experiment 178 can survive on raspberry cultivars possessing A₁ resistance genes and is the 179 most common biotype found in the UK (McMenemy et al., 2009). The aphid 180 culture was initiated from field-collected aphids at JHI and maintained in the 181 laboratory for multiple generations. This aphid population was maintained 182 at 18 \pm 1°C, 16h photoperiod using the cultivar Malling Landmark (also A₁ 183 resistance) as a culture plant. The aphid population had been randomly

divided and maintained in either ambient or elevated CO₂ conditions for at
least five generations before the experiment.

186 Ladybirds

187 The aphidophagous harlequin ladybird (Harmonia axyridis Pallas), native to 188 Asia, was originally used throughout Europe and North America as a 189 biocontrol agent against aphids (Brown et al., 2008). Now established, it is 190 one of the most common ladybird species (Tedders & Schaefer, 1994; 191 Colunga-Garcia & Gage, 1998; Brown et al., 2008). Adult female ladybirds 192 were collected from lime trees (*Tilia* spp.) in Oxfordshire, UK. The population was maintained in clear acrylic cages (30cm x 20cm x 15cm) at 18 ± 1°C and 193 194 16hr photoperiod. In culture, H. axyridis populations were fed pea aphids 195 (Acyrthosiphon pisum Harris), but starved for 24 hours prior to the experiment.

196 Experiment 1: Trophic interactions

In a fully-factorial blocked design, 48 plants of each cultivar (susceptible, 197 198 partially-resistant and resistant) were randomly assigned to the two 199 atmospheric CO₂ (ambient and elevated) and subsequent predator (ladybird 200 present or absent) treatments. This gave 12 replicates per treatment 201 combination (cultivar x CO₂ x predator). The experiment was carried out 202 September 2011 - September 2012 over a series of four runs to avoid 203 psudoreplication of CO₂ treatment. Each run comprised of three full replicates 204 (n=36) of each treatment combination. Within each run the 18 plants were 205 randomly distributed along a single bench inside each chamber. To prevent 206 movement of flightless aphid nymphs between plants, individual pots were 207 secured on circular plinths (10cm diameter x 3cm height) and placed in 50cm

x 50cm plastic trays filled with water (four plants per tray), ensuring the pots
were above the water-line (see Johnson *et al.*, 2013 for details).

210 After five weeks growth in the CO₂ treatments, the height of each plant was 211 measured and three adult large raspberry aphids were placed on the first fully 212 unfurled leaf of each plant. Two weeks after aphid inoculation, the number of 213 nymphs and adult aphids on each plant was counted and then a single adult 214 female *H. axyridis* was introduced to the plants assigned to predator 215 treatment. All plants were then placed within individual insect cages 216 (25cm diam. x 65cm height, Insectopia, UK). The ladybirds remained on the 217 plants for 72 hours, after which they were removed and the aphid population 218 on each plant re-counted. Up to 10 adult aphids from each plant were 219 collected at random, snap-frozen and freeze-dried. All aboveground plant 220 material was destructively harvested and oven-dried for 48 hours at 70°C. 221 Aphid and plant dry mass were recorded. Total soluble protein was 222 determined from a subsample of the freeze-dried aphids using a protein assay kit (Thermo Scientific BCA Kit 23225) which used the Bradford (1976) 223 224 method.

225 Experiment 2: Ladybird development

To provide aphid prey, 32 plants of the susceptible and partially resistant cultivar were randomly assigned to two CO_2 treatments across four controlled environment chambers (2 x ambient, 2 x elevated). Plants were inoculated with large raspberry aphid as in Experiment 1 and after four weeks aphids were collected daily and used as prey for the ladybird larvae in the trial. Eggs were laid in a series of clutches over a 5-day period from three randomly selected mating pairs of Harlequin ladybirds. Each clutch (approximately 15 –

233 30 eggs) was collected and split randomly between the four diet treatments 234 (cultivar x CO₂). There were 30 individual ladybird replicates per treatment 235 combination, 120 in total. Eggs were placed individually into plastic pots (2cm 236 height x 3cm diameter) in a constant temperature room at 18°C, 16 hours 237 photoperiod. Upon eclosion from eqg, each larva was provided with 10 - 15238 aphids daily, any aphids not consumed from the previous day were removed. 239 Time to each larval instar was recorded. To establish the effect of diet 240 treatment on relative growth rate, a random sample of 11 individuals from 241 each treatment combination (44 in total) were selected and weighed every 242 day until pupation (Sartorius ME36S microbalance). Mean relative growth rate 243 (MRGR) was calculated following Gotthard et al. (1994):

244 MRGR = $(W_2 - W_1) / t$,

where W_1 is the initial weight, W_2 the final weight and t is the number of days for each life-stage. Mass of all individuals were recorded at pupation and emergence. Adult dry mass was recorded after emerged adults were snap frozen and freeze dried (Heto PowerDry PL3000).

249 Statistical analysis

All data were analysed using generalised linear mixed models (GLMM) using
 PROC GLIMMIX (SAS Institute, version 9.01).

252 Experiment 1

Hypotheses 1 and 2. Initial aphid abundance (counts) was modelled using a Poisson error distribution and log-link function. Aboveground plant dry mass, change (delta) in aphid abundance, aphid dry mass and total soluble protein content were modelled using a normal (Gaussian) error distribution with identity-link function. Random effects were experimental run and chamber

nested within run for all models. Models of aphid abundance had an additional, observation-level random effect fitted to account for overdispersion within the count data (Elston *et al.*, 2001). While chamber accounted for little variation in the data it represented an important structural random effect (i.e. CO_2 treatment was applied at the chamber level) and was thus retained in all models.

Potential explanatory variables included raspberry cultivar (susceptible (Glen Ample), partially resistant (Glen Clova), resistant (Octavia)), CO_2 treatment (ambient 390 μ mol/mol, elevated 650 μ mol/mol), predator treatment (ladybird present or absent) and plant biometrics (height, dry mass). Of the original 144 plants, 12 died at various stages during the experiment and were not included in the analysis. Aphid total soluble protein content was modelled separately using a normal (Gaussian) error distribution with identity-link function.

271 Experiment 2

272 Hypothesis 3. Relative growth rate, development time and pupal mass of 273 ladybirds were modelled using a normal (Gaussian) error distribution with 274 identity-link function. Random terms were parent identity and the experimental 275 chamber in which the aphid prey was reared. When repeated measures were 276 used (relative growth rate) an observation-level random effect was added to 277 the R-side of the random structure. Raspberry cultivar (susceptible and 278 partially resistant), CO₂ treatment (ambient 390 μ mol/mol and elevated 650 279 µmol/mol), sex upon emergence as adult and larval instars (relative growthrate only) were fitted as potential explanatory variables. 280

During the analysis of both experiments, explanatory variables were added in a forward stepwise fashion until a minimum adequate model was obtained (Crawley, 2002). F-ratio and p-values adjusted for other fitted terms (SAS type

III) are presented and, where multiple comparison tests (i.e. SAS Least-Square means) were used to test for treatment effects, a Bonferroni correction was applied. Two-way interactions (e.g. between cultivar, predator and CO_2 treatments in Experiment 1) are reported only when statistically significant (p<0.05).

- 289
- 290 Results
- 291 Experiment 1
- 292 Hypothesis 1 Plant responses

293 Aboveground biomass varied significantly among the raspberry cultivars 294 irrespective of CO₂ treatment (Table 1). Plants partially-resistant to aphid 295 herbivory had the greatest dry mass, followed by the resistant cultivar (Fig. 1). 296 The susceptible cultivar had the lowest dry mass, almost half that of the 297 partially resistant cultivar (Fig. 1). CO₂ treatment also influenced the plant 298 biomass, plants grown in eCO₂ achieving a greater dry mass compared to 299 plants grown in ambient CO₂ (Table 1). The susceptible cultivar was the most 300 responsive to eCO₂ with a 107% increase in dry mass compared to ambient 301 CO₂ (Fig. 1). There was an 85% increase of dry mass of the resistant cultivar 302 in eCO₂ compared to ambient. The partially susceptible cultivar was the least 303 responsive to eCO_2 , increasing in dry mass by 30%.

304 Hypothesis 2 - Aphid responses

There was a highly significant effect of cultivar on aphid abundance before the onset of the predation treatment (Fig. 2a, Table 1b). While there were similar numbers of aphids on the susceptible and partially-resistant cultivars, as expected, the aphid abundance on the resistant cultivar was lower by almost

309 a factor of 10 (Fig. 2a). Atmospheric CO₂ enrichment significantly affected 310 aphid abundance (Table 1b), but this varied between plant cultivars as 311 indicated by the significant $CO_2 \times CO_2 \times CO_2$ and $CO_2 \times CO_2 \times CO_2 \times CO_2$. Altered 312 population levels drove this effect of CO₂ enrichment on aphid abundance on 313 the resistant plant cultivar. On the resistant cultivar, elevation of atmospheric 314 CO₂ concentrations significantly increased the mean abundance of aphids 315 (Fig. 2a). Furthermore, aphid colonisation of the resistant cultivar was 316 markedly increased by CO₂ enrichment with 14% and 70% of plants 317 supporting aphids under ambient and eCO_2 conditions, respectively ($F_{1.5}$ = 318 7.9, p = 0.05). In contrast, aphid abundance on the susceptible and partially 319 resistant cultivars were unaffected by manipulation of the CO₂ environment 320 (Fig 2a, Table 1a).

321 The presence of a ladybird predator significantly reduced aphid abundance on 322 all cultivars (Fig. 2a versus Fig. 2b; Table 1c). Moreover, while CO₂ 323 enrichment increased aphid herbivore colonisation and abundance on the 324 resistant cultivar, once ladybird predation was introduced this CO₂ effect was 325 nullified (Table 1c, Fig. 2b). On the susceptible and partially-resistant 326 cultivars, the number of aphids consumed by the ladybird did not significantly 327 vary with CO₂ treatment (Fig. 2b). CO₂ treatment did not affect adult aphid dry 328 mass or total protein content ($F_{1,2} = 0.25$, p = 0.667 and $F_{1,2} = 1.44$, p = 0.353, 329 respectively). Aphid total soluble protein was greater when reared on the 330 susceptible cultivar than the partially resistant cultivar ($F_{1,100} = 11.6$, p = 331 0.001).

332 Experiment 2

333 Hypothesis 3. The mean relative growth rate over the full duration of ladybird 334 development was not affected by the prey source environment (CO₂: $F_{1,2}$ = 335 1.03, p = 0.42 and cultivar: $F_{1.24} = 0.78$, p = 0.38). Relative growth rate was 336 stage-specific with the earlier instars having a much lower mean growth rate 337 than the later instars. When fed aphids from the partially resistant cultivar, the 338 mean relative growth rate of fourth instar ladybird was significantly increased 339 (Fig. 3, Table 2a). When fed aphids reared on the partially resistant cultivar, 340 fourth instar ladybird larvae had significantly higher relative growth rate 341 compared to their siblings fed aphids reared on the resistant cultivar (Fig.3). 342 There was no significant effect of CO₂ treatment on relative growth rate of 343 ladybird larvae (Table 2.a). Despite the significant effect of cultivar on fourth 344 instar larval growth rate, duration of development from egg to adult was not 345 affected by the cultivar or CO₂ treatment ($F_{1.80} = 0.29$, p = 0.59 and $F_{1.2} =$ 0.61, p = 0.44, respectively) aphid prey was reared in. Similarly, pupal mass 346 347 and adult mass were not affected by the rearing conditions of the aphid prey (Table 2b). Pupal and adult mass was, however, affected by adult sex: 348 349 females were significantly heavier than males (Table 2b).

350

351 **Discussion**

The fertilising effect of CO_2 enrichment is predicted to increase plant biomass and productivity (Ainsworth & Long, 2005; Robinson *et al.*, 2012), particularly for woody plants (Curtis & Wang, 1998). This study confirms this, with all three raspberry cultivars showing increased biomass in response to elevated atmospheric CO_2 . This was also seen for the raspberry cultivars investigated by Martin and Johnson (2011) (summarised in Table 3), suggesting that this response is common to the species as a whole. On the two susceptible

cultivars, aphid populations were unaffected by the increased plant biomass associated with elevated CO_2 . Aphid colonisation and subsequent abundance was greater on the resistant cultivar grown in elevated CO_2 , suggesting a reduction in resistance to aphid herbivory in the novel environment.

363 Plant biomass in ambient conditions was not correlated with resistance to 364 herbivory; the partially resistant cultivar had the greatest biomass, followed by 365 the resistant and susceptible cultivars. The extent to which plant biomass 366 increased under CO₂ enrichment varied with cultivar. The partially resistant 367 cultivar, with the greatest biomass in ambient CO₂, was the least responsive 368 (30% increase in biomass), suggesting that it is already close to its maximum 369 growth capacity under ambient CO₂. The 85% increase in biomass of the 370 resistant cultivar under eCO_2 suggests this cultivar to be particularly 371 responsive to eCO₂. In eCO₂ aphid colonisation was significantly higher on 372 the resistant cultivar, but aphid numbers remained very low despite a 373 significant increase from ambient conditions. The resistant cultivar used in this 374 study, Octavia, is the successful crossing of two aphid resistance genes, A₁₀ 375 and A_{k4a} . Previous work by Martin and Johnson (2011) found the A_{10} was 376 robust to changes in CO₂ concentrations. This implies that CO₂ enrichment may be modifying the function of the A_{k4a} resistance gene. This, however, 377 378 remains an untested hypothesis and is only one possible explanation. 379 Raspberry cultivars possessing the A₁₀ resistance gene can show significant 380 variation in minor genes associated with aphid resistance, which may modify 381 the responses to elevated CO₂ (Hall, 2009). Even in cultivars possessing the 382 same resistance gene, it seems their genotypic background can modify 383 resistance expression at elevated CO₂. For example with two cultivars 384 possessing the A₁ resistance gene either becoming more susceptible to

aphids (Table 3, Martin & Johnson, 2011) or unaffected, as reported here. Similarly, expression of anti-herbivore defences among individuals from the same population of common milkweed (*Asclepias syriaca* L) vary considerably when grown in elevated CO_2 (Vannette & Hunter, 2011).

389 Without a detailed mechanistic understanding of raspberry resistance to A. 390 idaei, the reason why aphid numbers increased in elevated CO₂ remains speculative. Resistance to A. idaei in raspberry is thought to be the result of 391 392 antibiosis reducing colonisation and antixenosis reducing individual 393 performance (Mitchell, 2007). The observed increase in colonisation rate suggests the former defence may be impaired in elevated CO₂. Increasing 394 395 CO₂ levels have been shown to suppress the production of jasmonates and 396 increase the production of salicylic acid, affecting specific signalling pathways related to plant defence (Zavala et al., 2013). In particular, the down 397 398 regulation of jasmonates has been linked to increased aphid abundance in 399 elevated CO₂ (Sun et al., 2013).

400 Top-down regulation of agricultural pest species by natural enemies is becoming increasingly important as use of conventional chemical pesticides 401 402 becomes progressively more difficult under stricter legislation (such as 403 European Union Regulation (EC) No 1107/2009) (Van Driesche, 2008). This 404 is particularly true for crops grown under cover (e.g. glasshouse, polytunnel), 405 an increasingly common practice for enhancing productivity (Johnson et al., 406 2010, 2012, Wittwer & Castilla, 1995), since natural enemies work more 407 effectively in closed environments (McMenemy et al., 2009). Even in non-408 covered agricultural crops, top-down regulation of herbivore populations is,

409 however, important and increasingly encouraged (Stiling & Corneilissen,
410 2005, Van Driesche, 2008).

Predation by a natural enemy mitigated the breakdown of resistance to aphid 411 412 herbivory, returning the aphid population to its "ambient" state. There was no 413 evidence for a transfer of bottom-up effects across multiple trophic levels. The 414 CO₂ environment host plants were exposed to, did not affect the size or total 415 protein content of aphids living on them. Moreover, the CO₂ environment their 416 prey had been reared in did not influence the development of the next 417 generation of ladybirds. Similar to other studies, we found the effect of 418 elevated CO₂ on prey quality weak or non-existent (Salt et al., 1995; Stacey & 419 Fellowes, 2002, Chen et al., 2005) and subsequent predator generations were 420 also unaffected (Chen et al., 2005).

421 The influence of bottom up processes, such as the effect of plant genotype on 422 prey quality, had a much more significant effect than eCO₂ on ladybird 423 development. Plant cultivar significantly affected total protein content in 424 aphids. When reared on the susceptible cultivar, aphids had a greater total protein content than aphids reared on the partially resistant cultivar. 425 426 Unexpectedly, the opposite was observed for the mean relative growth rate of ladybird larvae. Larvae fed aphid prey from the susceptible cultivar had 427 428 significantly lower growth rate than larvae fed aphids from the partially 429 susceptible cultivar. The underlying reason for this remains unclear, but the 430 higher protein content of aphids on the susceptible cultivar may reflect greater 431 fitness and behavioural responsiveness of these individuals in addition to their 432 nutritional value as prey. These individuals may be able to better resist attack 433 by ladybirds using behavioural strategies (e.g. kicking, evasion) and thereby

impose extra fitness costs (e.g. handling time) on ladybirds (Dixon, 2000).
Mitchell *et al.* (2010) reported that *A. idaei* showed less 'dropping behaviour'
and suggested they may rely more on such behavioural resistance to
parasitoid attack when feeding on susceptible cultivars, so this explanation is
at least credible.

439 Confining aphids on plants necessitated use of potted plants in closed 440 chambers, which may be argued to give artificially high plant growth 441 responses to CO_2 (Ainsworth *et al.*, 2008). Given, however, that > 90% of 442 raspberry production takes place in closed polytunnels which buffer 443 environmental fluctuations (Johnson et al., 2010; Johnson et al., 2012), this is 444 perhaps a less relevant concern in this system as chambers have similar effects. Moreover, our use of large pots and potting media minimised 445 446 restrictions to root growth the potential for hypoxic conditions, as advocated 447 by Passioura (2006).

448 This study highlights the importance of considering multiple trophic levels 449 when trying to understand pest dynamics and ecosystem responses to future climates. Increasing atmospheric CO₂ has the potential to impair plant 450 451 defences against herbivory which may have important implications for agroecosystems. We demonstrate that higher trophic levels may, however, partly 452 453 mitigate this reduction in plant defences by controlling herbivore numbers on 454 the affected plants. The longer-term effects of elevated CO₂ on tri-trophic 455 interactions remain however little understood. This study provides an 456 empirical demonstration of how the net level of plant herbivory under elevated 457 CO₂ depends on both the interaction between the herbivore and the natural 458 enemy.

459	
460	
461	
462	
463	Acknowledgements
464	We thank Alison Dobson, Emily Shaw and Scott McKenzie for technical
465	advice and support, and Adam Butler for statistical advice. The Natural
466	Environment Research Council funded this research.
467	
468	
469	
470	
471	
472	
473	
474	
475	
476	
477	
478	
479	
480	
481	
482	
483	
484	
485	
486	
487	
488	
489	
490	
491	
492	

493 494 495 496 497 References 498 A'Bear, A.D., Johnson, S.N. & Jones, T.H. (2013) Putting the 'upstairs-downstairs' 499 into ecosystem service: What can aboveground-belowground ecology tell us? 500 Biological Control, http://dx.doi.org/10.1016/j.biocontrol.2013.10.004 501 Ainsworth, E.A. & Long, S.P. (2005) What have we learned from 15 years of free-502 air CO₂ enrichment (FACE)? A meta-analytic review of the responses of 503 photosynthesis, canopy properties and plant production to rising CO₂. New 504 *Phytologist*, **165**, 351-71. 505 Ainsworth, E.A., Leakey, A.D.B., Ort, D.R. & Long, S.P. (2008) FACE-ing the facts: 506 inconsistencies and interdependence among field, chamber and modeling studies 507 of elevated CO₂ impacts on crop yield and food supply. New Phytologist, 179, 5-508 9. 509 Bale, J.S., van Lenteren, J.C. & Bigler, F. (2008) Biological control and sustainable 510 food production. *Philosophical Transactions of the Royal Society B: Biological* 511 *Sciences*, **363**, 761-776. 512 Birch, A.N.E., Gordon, S.C., Fenton, B., Malloch, G., Mitchell, C., Jones, A.T., Griffiths, 513 D.W., Brennan, R., Graham, J. & Woodford, J.A.T. (2004). Developing a sustainable 514 IPM system for high value Rubus crops (raspberry, blackberry) for Europe. In 515 Proceedings of the Euro Berry Symposium - Cost 836 Final Worskhop (ed D.W. 516 Simpson), pp. 289-292. International Society Horticultural Science, Leuven 1.

- 517 Bradford, M.M. (1976) Rapid and sensitive method for quantitation of microgram
- 518 quantities of protein utilizing principle of protein-dye binding. *Analytical*
- 519 *Biochemistry*, **72**, 248-254.
- 520 Brown, P.M.J., Adriaens, T., Bathon, H., Cuppen, J., Goldarazena, A., Hägg, T., Kenis,
- 521 M., Klausnitzer, B.E.M., Kovář, I., Loomans, A.J.M., Majerus, M.E.N., Nedved, O.,
- 522 Pedersen, J., Rabitsch, W., Roy, H.E., Ternois, V., Zakharov, I.A. & Roy, D.B. (2008)
- 523 *Harmonia axyridis* in Europe: spread and distribution of a non-native coccinellid.
- 524 *BioControl*, **53**, 5-21.
- 525 Buffington, D.E., Bucklin, R.A., Henley, R.W. & McConnell, D.B. (2013).
- 526 Greenhouse Ventilation. University of Florida, Institute of Food and Agricultural
- 527 Sciences.
- 528 Chen, F., Ge, F. & Parajulee, M.N. (2005) Impact of elevated CO₂ on tri-trophic
- 529 interaction of *Gossypium hirsutum*, *Aphis gossypii*, and *Leis axyridis*.
- 530 Environmental Entomology, **34**, 37-46.
- 531 Colunga-Garcia, M. & Gage, S.H. (1998) Arrival, establishment, and habitat use of
- the multicolored Asian lady beetle (Coleoptera: Coccinellidae) in a Michigan
- 533 landscape. *Environmental Entomology*, **27**, 1574-1580.
- 534 Couture, J.J., Servi, J.S. & Lindroth, R.L. (2010) Increased nitrogen availability
- 535 influences predator-prey interactions by altering host-plant quality.
- 536 *Chemoecology*, **20**, 277-284.
- 537 Crawley, M.J. (2002) Statistical Computing: An Introduction to Data Analysis Using
- 538 S-PLUS. John Wiley & Sons
- 539 Curtis, P.S. & Wang, X. (1998) A meta-analysis of elevated CO₂ effects on woody
- plant mass, form, and physiology. *Oecologia*, **113**, 299-313.

- 541 Dixon, A.F.G. (2000) *Insect Predator-Prey Dynamics*. Cambridge University Press,
 542 Cambridge.
- 543 Dray, M.W., Crowther, T.W., Thomas, S.M., A'Bear, A.D., Godbold, D.L., Ormerod,
- 544 S.J., Hartley, S.E. & Jones, T.H. (2014) Effects of elevated CO₂ on litter chemistry
- 545 and subsequent invertebrate detritivore feeding responses. *PLoS ONE*, **9**, e86246.
- Elston, D.A., Moss, R., Boulinier, T., Arrowsmith, C. & Lambin, X. (2001) Analysis
- 547 of aggregation, a worked example: numbers of ticks on red grouse chicks.
- 548 *Parasitology*, **122**, 563-569.
- 549 Gotthard, K., Nylin, S. & Wiklund, C. (1994) Adaptive variation in growth-rate-
- 550 life-history costs and consequences in the speckled wood butterfly, *Pararge*
- 551 *aegeria Oecologia*, **99**, 281-289.
- 552 Gregory, P.J., Johnson, S.N., Newton, A.C. & Ingram, J.S.I. (2009) Integrating pests
- and pathogens into the climate change/food security debate. *Journal of*
- 554 *Experimental Botany*, **60**, 2827-2838.
- 555 Guo, H., Sun, Y., Li, Y., Tong, B., Harris, M., Zhu-Salzman, K. & Ge, F. (2013) Pea
- aphid promotes amino acid metabolism both in *Medicago truncatula* and
- bacteriocytes to favor aphid population growth under elevated CO₂. *Global*
- 558 *Change Biology*, **19**, 3210-3223.
- Hall, H.K., Hummer, K.E., Jamieson, A.R., Jennings, S.N. & Weber, C.A. (2009).
- 560 Raspberry Breeding and Genetics. In *Plant Breeding Reviews*, pp. 39-353. John
- 561 Wiley & Sons, Inc.
- Johnson, S.N., Petitjean, S., Clark, K.E. & Mitchell, C. (2010) Protected raspberry
- 563 production accelerates onset of oviposition by vine weevils (*Otiorhynchus*
- 564 sulcatus). Agricultural and Forest Entomology, **12**, 277-283.

- Johnson, S.N., Young, M.W. & Karley, A.J. (2012) Protected raspberry production
- alters aphid-plant interactions but not aphid population size. *Agricultural and*
- 567 *Forest Entomology*, **14**, 217-224.
- Johnson, S.N., Mitchell, C., McNicol, J.W., Thompson, J. & Karley, A.J. (2013)
- 569 Downstairs drivers root herbivores shape communities of above-ground
- 570 herbivores and natural enemies via changes in plant nutrients. *Journal of Animal*
- 571 *Ecology*, **82**, 1021-1030.
- 572 Knight, V.H. & Fernández-Fernández, F. (2008) New summer fruiting red
- 573 raspberry cultivars from East Malling Research. *Acta Horticulture* **777**, 173-176.
- 574 Langellotto, G.A. & Denno, R.F. (2004) Responses of invertebrate natural enemies
- 575 to complex-structured habitats: a meta-analytical synthesis. *Oecologia*, **139**, 1-
- 576 10.
- 577 Martin, P. & Johnson, S.N. (2011) Evidence that elevated CO₂ reduces resistance
- 578 to the European large raspberry aphid in some raspberry cultivars. *Journal of*
- 579 *Applied Entomology*, **135**, 237-240.
- 580 McMenemy, L.S., Mitchell, C. & Johnson, S.N. (2009) Biology of the European large
- 581 raspberry aphid (*Amphorophora idaei*): its role in virus transmission and
- resistance breakdown in red raspberry. *Agricultural and Forest Entomology*, **11**,
 61-71.
- 584 Meehl, G.A., Stocker, T.F., Collins, W.D., Friedlingstein, P., Gaye, A.T., Gregory, J.M.,
- 585 Kitoh, A., Knutti, R., Murphy, J.M., Noda, A., Raper, S.C.B., Watterson, I.G., Weaver,
- 586 A.G. & Zhao, A.C. (2007). Global Climate Projections. In *Climate Change 2007: The*
- 587 Physical Basis. Contribution of Working Group I to the Fourth Assessment Report
- 588 *of the Intergovernmental Panel on Climate Change* (eds S. Solomon, D. Qin, M.

- 589 Manning, Z. Chen, M. Marquis, K.B. Averyt, M. Tignor & H.L. Miller), pp. 747–845.
- 590 Cambridge University Press, Cambridge, UK.
- 591 Mitchell, C. (2007) Novel approaches to the development of intragrated pest
- 592 management in UK raspberry production, University of Dundee.
- 593 Mitchell, C., Johnson, S.N., Gordon, S.C., Birch, A.N.E. & Hubbard, S.F. (2010)
- 594 Combining plant resistance and a natural enemy to control *Amphorophora idaei*.
- 595 *Biocontrol*, **55**, 321-327.
- 596 Passioura, J.B. (2006) The perils of pot experiments. *Functional Plant Biology*, 33,
 597 1075-1079.
- 598 Raven, J.A. (1983) Phytophages of Xylem and Phloem a comparison of animal
- and plant sap-feeders. *Advances in Ecological Research*, **13**, 135-234.
- 600 Robinson, E.A., Ryan, G.D. & Newman, J.A. (2012) A meta-analytical review of the
- 601 effects of elevated CO₂ on plant–arthropod interactions highlights the
- 602 importance of interacting environmental and biological variables. New
- 603 *Phytologist*, **194**, 321-336.
- 604 Salt, D.T., Brooks, G.L. & Whittaker, J.B. (1995) Elevated carbon dioxide affects
- 605 leaf-miner performance and plant growth in docks (*Rumex* spp.). Global Change
- 606 *Biology*, **1**, 153-156.
- 607 Schaller, A. (2008) Induced Plant Resistance to Herbivory. Springer.
- 608 Smith, C.M. (2005) Plant Resistance to Arthropods. Springer Press, The
- 609 Netherlands, Dordrecht.
- 610 Stacey, D.A. & Fellowes, M.D.E. (2002) Influence of elevated CO₂ on interspecific
- 611 interactions at higher trophic levels. *Global Change Biology*, **8**, 668-678.

- 612 Stiling, P. & Cornelissen, T. (2005) What makes a successful biocontrol agent? A
- 613 meta-analysis of biological control agent performance. *Biological Control*, 34,
- 614 236-246.
- 615 Stiling, P. & Cornelissen, T. (2007) How does elevated carbon dioxide (CO₂) affect
- 616 plant-herbivore interactions? A field experiment and meta-analysis of CO₂-
- 617 mediated changes on plant chemistry and herbivore performance. *Global Change*
- 618 *Biology*, **13**, 1823-1842.
- 619 Sun, Y., Guo, H., Zhu-Salzman, K. & Ge, F. (2013) Elevated CO₂ increases the
- 620 abundance of the peach aphid on Arabidopsis by reducing jasmonic acid
- 621 defenses. *Plant Science*, **210**, 128-140.
- 622 Tedders, W.L. & Schaefer, P.W. (1994) Release and establishment of *Harmonia*
- 623 axyridis (Coleoptera, Coccinellidae) in the southeastern United States. Entomol
- 624 *News*, **105**, 228-243.
- 625 The Royal Society (2009). Reaping the benefits: science and the sustainable
- 626 intensification of global agriculture, *The Royal Society*, London, UK.
- 627 Turlings, T.C.J., Tumlinson, J.H. & Lewis, W.J. (1990) Exploitation of herbivore-
- 628 induced plant odors by host-seeking parasitic wasps. *Science*, **250**, 1251-1253.
- 629 Van Driesche, R.G. (2008) Control of Pests and Weeds by Natural Enemies an
- 630 *introduction to biological control,*. Blackwell Publishing, Oxford, UK.
- 631 Vannette, R.L. & Hunter, M.D. (2011) Genetic variation in expression of defense
- 632 phenotype may mediate evolutionary adaptation of Asclepias syriaca to elevated
- 633 CO₂. *Global Change Biology*, **17**, 1277-1288.
- 634 Watt, A.D., Whittaker, J.B., Docherty, M., Brookes, G. & Salt, D.T. (1995). The
- 635 impact of elevated CO₂ on Insect herbivores. In *Insects in a Changing*
- 636 Environment (eds R. Harrington & M.E. Stork), pp. 197-217. Academic press,

- 637 Sandiego, California.
- 638 Wittwer, S.H. & Castilla, N. (1995) Protected cultivation of horticultural crops
- 639 worldwide. *HortTechnology*, **5**, 6-23.
- 640 Yuan, J.S., Himanen, S.J., Holopainen, J.K., Chen, F.J. & Stewart, C.N. (2009)
- 641 Smelling global climate change: mitigation of function for plant volatile organic
- 642 compounds. *Trends in Ecology & Evolution*, **24**, 323-331.
- 643 Zavala, J.A., Casteel, C.L., DeLucia, E.H. & Berenbaum, M.R. (2008) Anthropogenic
- 644 increase in carbon dioxide compromises plant defense against invasive insects.
- 645 Proceedings of the National Academy of Sciences of the United States of America,
- 646 **105**, 5129-5133.
- 647 Zavala, J.A., Nabity, P.D. & DeLucia, E.H. (2013) An Emerging Understanding of
- 648 Mechanisms Governing Insect Herbivory Under Elevated CO₂. Annual Review of
- 649 *Entomology*, **58**, 79-97.
- 650 Zvereva, E.L., Lanta, V. & Kozlov, M.V. (2010) Effects of sap-feeding insect
- herbivores on growth and reproduction of woody plants: a meta-analysis of
- experimental studies. *Oecologia*, **163**, 949-960.

653

654

655

- Table 1. GLIMMIX results summary for a) plant dry mass, b) initial aphid
- abundance and c) change in aphid abundance-post predation in relation to
- 659 biotic and abiotic environment. Significant variables in bold retained in final
- 660 model. MPE = multiple parameter estimates.
- 661

Response variable	Explanatory variables	Estimate	$F_{(ndf, ddf)}$	Р
a) Plant dry mass	CO₂ Ambient Elevated	12.830 20.671	12.85 _(1,3)	0.0377
Random effect estimate: Chamber (experimental run) = 28.73 ± 27.512 Experimental run = 6.28 ± 7.84	Cultivar Susceptible Partially resistant Resistant Initial aphid abundance	14.940 25.221 20.671 20.666	20.37 _(2,121) 0.33 _(1,123)	<0.0001 0.568
b) Initial aphid abundance	CO₂ Ambient Elevated	-1.555 0.71	14.49 (1,2)	0.063
Random effect estimate: Replicate = 0.46±0.081 Chamber (experimental run) = 0	Cultivar Susceptible Partially resistant Resistant	4.203 4.197 0.71	148.67 _(2,128)	<0.0001
Experimental run = 0.13 ± 0.12	Dry mass	0.888	0.88 _(1,121)	0.349
	CO ₂ * Cultivar	MPE	8.89 _(2,128)	0.0002
c) Delta aphid abundance Random effect estimate: Chamber (experimental run) = 0	CO ₂ Ambient Elevated	-0.678 -4.747	0.3(1,2)	0.639
Experimental run = 468.36±429.36	Cultivar Susceptible Partially resistant Resistant	-1.99 6.21 -2.84	10.64 _(1,125)	<0.0001
	Dry mass	2.062	0.6 _(1,123)	0.439
	Predator treatment Control Ladybird	MPE 0.04 -2.84	33.55 _(1,125)	<0.0001
	Cultivar * predator treatment	MPE	7.37 _(2,1.25)	0.0009

- Table 2. GLIMMIX results summary for ladybird responses (a) relative growth
- rate and b) pupal mass), in relation to rearing conditions of their aphid prey
- 666 (CO₂ and raspberry cultivar), larval instar and gender. Significant variables in
- 667 bold retained in final model. MPE = multiple parameter estimates
- 668

Response variable	Explanatory variables	Estimate	$F_{(ndf, ddf)}$	Р
a) Relative growth rate	CO ₂ Ambient Elevated	4.793 4.715	0.79 _(1,2)	0.385
Random effect estimate: Parent = 0.001±0.007 Roplicato = 0.252±0.125	Cultivar Susceptible Partially resistant	5.195 4.670	3.07 _(1,23)	0.093
Chamber = 0	Larval instar 1 st 2 nd 3 rd 4 th	MPE 0.022 0.727 2.033 4.67	637.22 _(3.60)	<0.0001
	Larval instar * Cultivar	MPE	3 _(3,60)	0.038
b) Pupal mass	CO ₂ Ambient Elevated	35.725 35.846	0.03(1,2)	0.884
Random effect estimate: Parent = 1.463±1.863 Chamber = 0	Cultivar Susceptible Partially resistant	35.831 35.767	0.02 _(1.92)	0.898
	Sex Male Female	0.868 1.603	26.30 _(1.93)	<0.0001

669			
670			
671			
672			
673			
674			
675			
676			
677			
678			
679			

Table 3. Comparison of plant and aphid responses to elevated atmospheric CO₂ (eCO₂) found by Martin & Johnson, 2012 and the

- 681 findings of this study.

	Martin & Johnson		Hentley et al	
Cultivar	eCO ₂ impacts on plants	eCO ₂ impacts on aphids	eCO ₂ impacts on plants	eCO ₂ impacts on aphids
Malling Jewell (susceptible)	197% increase growth rate	None		
Glen Lyon – A ₁	41% increase in growth rate	Increase in abundance and adult mass		
Glen Ample – A ₁			107% increase in dry mass	None
Glen Clova – multi			30% increase in dry mass	None
Glen Rosa – A ₁₀	186% increase in growth rate	None		
Octavia – A_{10} and A_{K4}			85% increase in dry mass	Increase in aphid colonization and abundance

Figure 1. Aboveground plant dry mass of three raspberry cultivars in response to ambient and elevated CO_2 . Data are least square mean \pm S.E.

687

688	Figure 2. The effect on aphid abundance of cultivar resistance, CO_2 treatment
689	and presence a) or absence b) of ladybird predation. Ambient (white bars)
690	and elevated (grey bars) atmospheric CO_2 levels. Letters above bars denote
691	significant differences. Aphid abundance for resistant cultivar scaled using a
692	second y-axis to make treatment effects clearer. Data are mean \pm S.E.
693	
694	Figure 3. Least square mean for relative growth rate of larval stages of the
695	ladybird <i>H. axyridis</i> fed aphid prey from susceptible (dashed line and triangle)
696	or partially resistant (solid line and circle) raspberry cultivars. Data are least
697	square mean ± S.E.
698	
699	
700	
701	
702	
703	
704	
705	
706	
707	
708	
709	
710	
711	
712	
713	





