- 1 Coastal waters contamination by mining tailings: what triggers the stability of iron in the
- 2 dissolved and soluble fractions?

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- 4 Cybelle Menolli Longhini^{1*}
- 5 Léo Mahieu²
- 6 Fabian Sá³
- 7 Constant M G van den Berg⁴
- 8 Pascal Salaün⁵
- 9 Renato Rodrigues Neto⁶

- ¹Postgraduate program in Environmental Oceanography, Federal University of Espírito Santo,
- 12 Fernando Ferrari Avenue, 514, Goiabeiras, Vitória, ES, 29055-460, Brazil; *Corresponding
- 13 author; e-mail: cybelle.longhini@ufes.br
- ² Department of Earth, Ocean and Ecological Sciences, School of Environmental Sciences,
- 15 University of Liverpool, L69 3GP, Liverpool, United Kingdom; e-mail:
- 16 Leo.Mahieu@liverpool.ac.uk
- ³ Department of Oceanography and Ecology, Federal University of Espírito Santo, Fernando
- Ferrari Avenue, 514, Goiabeiras, Vitória, ES, 29055-460, Brazil; e-mail: fabian.sa@ufes.br
- 19 ⁴ Department of Earth, Ocean and Ecological Sciences, School of Environmental Sciences,
- 20 University of Liverpool, L69 3GP, Liverpool, United Kingdom; e-mail:
- 21 vandenberg@liverpool.ac.uk
- ⁵ Department of Earth, Ocean and Ecological Sciences, School of Environmental Sciences,
- 23 University of Liverpool, L69 3GP, Liverpool, United Kingdom; e-mail: salaun@liv.ac.uk

- ⁶ Department of Oceanography and Ecology, Federal University of Espírito Santo, Fernando
- 25 Ferrari Avenue, 514, Goiabeiras, Vitória, ES 29055-460, Brazil; e-mail: renato.neto@ufes.br

- 27 Corresponding author: Cybelle Menolli Longhini, Francisco Eugênio Mussielo Street, 1104,
- 28 apt 403, Jardim da Penha, Vitória, ES 29060-290, Brazil, Telephone number: +55 73
- 29 991267557, e-mail: cybelle.longhini@ufes.br
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Abstract

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34 The solubility of iron (Fe) in seawater is greatly enhanced by complexation with organic 35 ligands, predominantly occurring as humic substances (HS) in coastal areas. Mining 36 exploitation is believed to change the natural biogeochemical cycle of Fe in coastal waters, 37 even though its impacts on the physical and chemical speciation of the Fe fractions are not 38 known. Here we show that dissolved and soluble Fe concentrations in coastal waters affected 39 by a mining catastrophe (Fundão dam, Southeast Brazil) remain very high, even almost 3 40 years later, with concentrations of dissolved Fe up to 2.8 μ M (0.45 μ m filtration) or 700 nM 41 $(0.22 \,\mu\text{m} \text{ filtration})$, and soluble Fe $(0.02 \,\mu\text{m})$ up to 40 nM. Levels of humic substances can 42 only explain the binding of 2 % and 10 % (median values) of dissolved Fe (0.22 μ m) and 43 soluble Fe concentrations, respectively, which shows that processes other than complexation 44 with humic substances are at play to maintain such high Fe concentrations. We hypothesize 45 that the colloidal phase that dominates the dissolved Fe fraction occurs as Fe(III) 46 oxyhydroxides while the soluble fraction is comprised of Fe(III) complexes with amine 47 compounds (widely used in the ore extraction process). Mass balance of dissolved Fe in the 48 water column suggests that sediment resuspension on the continental shelf is by far the 49 dominant process delivering dissolved Fe to coastal and shelf waters. Estimates of dissolved 50 Fe yearly fluxes highlight the sheer magnitude of this catastrophe that might provide a non-51 negligible amount of dissolved Fe to the open ocean.

Introduction

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53 Iron (Fe) is a bio-essential element that is limiting primary productivity in 40 % of the 54 world oceans (Moore et al. 2013). It is entering the marine system through multiple external 55 sources. The atmospheric dust is dominant to the Fe supply at the low latitudes whereas 56 continental margin and hydrothermal inputs are more important for Fe inventory at the high 57 latitudes (Tagliabue et al. 2014). Nevertheless these natural inputs, dissolved Fe (dFe) 58 residence time in the water column is much shorter than other nutrients due to its low 59 solubility (Liu and Millero 2002) and its propensity to adsorb on any reactive surface, thus 60 removing it from the water column (Boyd and Ellwood 2010). Understanding the distribution 61 of Fe between different size fractions is a key factor to describe the chemical reactivity and 62 bioavailability of Fe in seawater and its biogeochemical behaviour (Tagliabue et al. 2017). 63 Speciation is usually done in one of the three operationally-defined size fractions: particulate 64 (pFe; > 0.45 or $0.2 \mu m$), dissolved Fe (dFe; < 0.45 or $0.22 \mu m$) and to a much lesser extent, 65 soluble Fe (sFe; $< 0.02 \mu m$). 66 In the dissolved fraction, Fe speciation has been focused on the complexation with organic 67 compounds as Fe bound to natural organic ligands represents more than 99 % of the dissolved 68 Fe concentrations in marine ecosystems (Gledhill and van den Berg 1994). Organic 69 complexes increase the dissolved Fe solubility by reducing its removal by precipitation from 70 the water column (Liu and Millero 2002; Boyd and Ellwood 2010; Gledhill and Buck 2012). 71 In the absence of organic ligands, Fe concentrations in natural oxygenated seawater with a pH 72 around 8.0 would not be more than 0.01 nM (Liu and Millero 2002), due to extensive 73 precipitation. The presence of organic ligands, their reactivity and own biogeochemical 74 cycling, thus defines the distribution patterns of dissolved Fe concentrations in the ocean 75 (Hassler et al. 2017).

Natural Fe-binding ligands in seawater are thought to be mostly siderophores, strong chelators released by heterotrophic bacteria and cyanobacteria as a biochemical strategy to assimilate dissolved Fe, exopolymeric substances from cellular exudates and refractory organic matter like humic substances (Gledhill and Buck 2012). These compounds are different in their origin, conditional stability and dispersal patterns from coastal to offshore oceanic regions (Hassler et al. 2017). In estuarine and coastal waters, there is mounting evidence that humic substances dominate the bulk of organic ligands (e.g. Laglera and van den Berg 2009). These humic substances have been showed to be mostly of terrestrial origin (e.g. Laglera and van den Berg 2009) and although they undergo extensive removal within estuaries through coagulation and flocculation processes (Sholkovitz et al. 1978), a significant proportion make it through the estuarine trapping (Muller 2018). Speciation of the soluble Fe fraction in terms of organic ligands has not been largely addressed but the presence of soluble chelates from microbial degradation of settling biogenic particles (e.g. faecal pellets) has been suggested to explain the levels of soluble Fe in the North Atlantic and North Pacific waters (Wu et al. 2001). Recent studies using both the size partitioning method and ligands determination by cathodic stripping voltammetry showed the presence of two soluble organic ligand classes (stronger L₁ and weaker L₂) occurring in excess in surface waters of the Atlantic Ocean (Fitzsimmons et al. 2015). Although the type of ligands has not been specified, the range of ligand conditional stability constants K observed by Fitzsimmons et al. (2015) for L_1 fall into siderophores (log $K_1 > 12$) (Hassler et al. 2017) while log K₂ values for L₂ can represent a wide class of ligands with high similarity in complex stability (e.g. humic and exopolymeric substances) (Hassler et al. 2017). In addition to the stabilization of soluble Fe by organic complexation, recent studies have suggested that stability of soluble Fe in the vicinity of hydrothermal vents is mostly related to the presence of

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small colloids present as inorganic nanoparticles of pyrite FeS₂, Fe_xSi_y (Gartman et al. 2014) and Fe oxy-hydroxide; FeO_x(OH)_y (Fitzsimmons et al. 2017).

Besides the natural processes controlling Fe cycle and speciation, anthropogenic inputs have strongly modified metal distribution and biogeochemistry in coastal seas. Extraction and processing of ores have impacted several coastal environments, modifying the dispersal of metals by various physical and chemical processes that are fundamentally different than those that disperse natural inputs (Koski 2012). In this context, in November 2015, the collapse of the Fundão dam in South-eastern Brazil spilled more than 55 million m³ of Fe ore tailings that travelled 600 km along the Doce River before reaching the adjoining coastal waters. Even though this event is considered as one of the largest dam tailings failure worldwide, its impacts in the processes controlling the Fe cycle on the marine affected areas are still unknown (Longhini et al. 2019).

The aim of this study is to evaluate the impact of this catastrophic dam failure on the distribution and physicochemical forms of Fe in the adjoining coastal waters as a result of a massive and sustained increase in ambient Fe concentrations. We determined the distribution of three size-fraction of Fe (dissolved Fe: $< 0.45~\mu m$ and $< 0.22~\mu m$, and soluble Fe: $< 0.02~\mu m$) and their relation with humic substances. This study brings new insights into the biogeochemical factors related to the Fe fluxes and tailings deposition in coastal areas impacted by mining activities and also presents the first direct measurements of humic substances in marine area directly affected by mining tailings.

- Materials and methods
- 121 Study area

The Doce River watershed (Southeast Brazil) comprises two states (Minas Gerais and Espírito Santo). In Minas Gerais, this watershed is located in the Quadrilátero Ferrífero

mineral province, the largest Fe ore complex in Latin America. One of the most important ore deposits exploited in this region is the Itabirite, a low-grade Fe ore that requires several treatment steps to reach a higher mass recovery. The flotation method is mostly used to reduce the presence of quartz and other impurities in Fe ore concentrates (Filippov et al. 2014). Tailings sludge with high Fe content is generated as a waste product of the Fe ore processing and stored in dams, structures built of earthfill to contain the tailings (https://www.samarco.com/en/barragens/ accessed 03 May 2020). The activities of exploitation and processing of Fe ore in this region were known to affect the metal levels in water bodies near the tailings dam including a coastal lagoon located in Espírito Santo state (Pereira et al. 2008). The object of this study is the continental shelf adjacent to the Doce River mouth (Espírito Santo state) (Fig. 1) which has been impacted by the tailings sludge deposition after the Fundão mining disaster. The hydrological and oceanographic conditions (winds/waves regimes) are known to affect the Doce River discharge conditions, defining the flow of the turbidity plume (Rudorff et al. 2018), the sedimentary processes (Quaresma et al. 2015) and consequently the dispersion pattern of the tailings along the Doce River continental shelf. The average Doce River flow is 541.5 m³ s⁻¹, with lower median streamflows occurring around 368 m³ s⁻¹ during the dry season (from April to October), while higher median streamflows of about 965 m³s⁻¹ occurs during the wet season (between November and March) (Oliveira and Quaresma 2017). Flooding events occur with highest frequencies in January and December and they usually exceed 2,379 m³s⁻¹ with a maximum flood record of approximately 9,000 m³ s⁻¹ (Oliveira and Quaresma 2017).

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Sampling strategy

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147 Sampling was carried out in August 2018, approximately two years and nine months after 148 the Fe tailings reached the coastal area. Three transects were established perpendicular to the 149 shoreline: North (N), Central (C) and South (S), each composed of three stations from 10 to 150 30 m isobaths. Seawater samples were also collected 6 days (the 27th of November 2015) and 151 3 months (February 2016) after the Fundão dam failure to evaluate the geochemical 152 conditions during the acute phase of the impact. Two locations were sampled in November 153 2015 and three in February 2016, as denoted by stars and crosses in Fig. 1, respectively. 154 Coastal water samples from surface (≈ 50 cm below the water surface) and bottom (≈ 2 m 155 above the bottom) waters were collected using a 5 L Niskin horizontal bottle sampler 156 (General Oceanics) (n = 4 in November 2015; n = 6 in February 2016; n = 18 in August 2018) 157 into 500 mL acid-cleaned LDPE bottles. Temperature and seawater pH were measured on-158 board using a portable pH meter (Metrohm model 826). The pH was calibrated using 159 Metrohm buffer solutions of 4.01, 7.00 and 9.00 pH (NBS scale). Salinity was measured with 160 a CTD instrument (11 plus Sea-Bird in Nov 2015 and 9 plus Sea-Bird in Feb 2016). In Aug 161 2018 this parameter was measured using a refractometer. 162 Hydroclimatic conditions during the samplings 163 The Fundão dam rupture occurred in the context of an extreme hydrological drought in the region (Rudorff et al. 2018). After the dam failure on the 5th of November 2015, the Fe ore 164 165 tailings reached the coast on the 21st of November 2015 by low streamflow conditions (267.5 m³s⁻¹) (Fig. 2). The samplings were conducted under low Doce River discharge which ranged 166 from 291.3 m³ s⁻¹ in Nov 2015, 457.8 m³ s⁻¹ in Feb 2016 and 231.8 m³ s⁻¹ in Aug 2018 (Fig. 2). 167 168 A flooding event occurred towards the end of January 2016 reaching a maximum of 3,667.1 m³ s⁻¹ on the 22nd of January (Fig. 2). 169

The winds predominantly came from N, NE and S in Nov 2015 causing the Doce River turbidity plume to disperse southward, southwest-ward and northward. In Feb 2016 the plume was dispersed mainly to the southern region while in Aug 2018 it was concentrated parallel to the coast and moved southwest-ward (MODIS-Aqua imagery).

Processing and preservation of samples

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The samples were filtered once back at the laboratory, immediately after collection. Unfiltered seawater was passed either through 0.45 μ m or through 0.22 μ m cellulose acetate membrane (Millipore®) using a vacuum pump system to evaluate the Fe concentration in two dissolved size fractions: dissolved Fe (0.45 μ m) and dissolved Fe (0.22 μ m), respectively. The aliquots filtered through $0.22 \mu m$ membrane were also used to determine humic substances. The filtrates for dissolved Fe (0.45 μ m) were acidified to reach pH < 2.0 by adding HNO₃ 65 % (previously distilled using sub-boiling procedure; boiling sub-distill acid mark Berghof BSB-939-IR model) and stored at 4°C until analysis. Samples for dissolved Fe (0.22 μ m) determination were stored in a frozen state and transported in a thermic box filled with dry ice from Brazil to the University of Liverpool (Liverpool, UK). The soluble fraction was obtained by filtration in the laboratory after thawing of 0.22 μ m filtrate samples through acid-cleaned 0.02 µm Anotop filter syringes (25 mm diameter). Samples were always analysed within 24 h of thawing. Fe was thus determined in three different size fractions: a) dissolved Fe $(0.45 \,\mu\text{m})$ by ICP-MS which corresponds to the total dissolved Fe; b) dissolved Fe (0.22 μ m) and soluble Fe $(0.02 \mu m)$ by voltammetry. Humic substances were measured by voltammetry in the $0.22 \mu m$ fraction.

Chlorophyll-*a* pigments were concentrated by filtration on board through Glass Microfibre Filter (Sartorius) of 25 mm diameter. The filters were kept frozen until the extraction procedure. The suspended particulate material (SPM) was determined by

gravimetric method after filtration through pre-weighted glass fibre membranes (porosity of 0.45 μ m).

Chemical analyses

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Mass spectroscopy

Dissolved Fe (0.45 μ m) in seawater samples was determined by Inductively Coupled Plasma – Mass Spectroscopy (ICP-MS, Agilent, 7500 cx). The pH in the pre-acidified aliquots was adjusted using 1 M NaOH to a final pH around 7.0. The samples were then preconcentrated by passing through a cationic resin column (Chelex[®]). The trace metal extracts were eluted from the resin column with HNO₃-2 % solution. The compounds were quantified according to the EPA 6020A method for multi-element determination using ICP-MS (U.S. EPA 1998). The fluctuations in the signals during the measurements were corrected by using a multi-element internal standard (Internal Standard Mix - Bi, Ge, In, Li, Sc, Tb and Y, Agilent Technologies). Multi-element standards (ICP multi-element standard solution XXI for MS, CentiPUR ® MERCK, Darmstadt - Germany) were used to carry the calibration curve (concentration range from 0 to 300 μ g L⁻¹). The dissolved Fe (0.45 μ m) concentrations were determined by the linear regression (y = ax + b) plotted as a result of the calibration curve. Three analytical modes were used (no gas, Helium and Hydrogen) and the optimum one, as determined by the method of recovery from spiked samples, was used. All solutions were prepared with ultra-pure water processed using a PURELAB ultra (Model Ultra an MKZ, the brand Elgar).

214 Adsorptive cathodic stripping voltammetry

The voltammetric system used for dissolved Fe (0.22 μ m), soluble Fe and humic substances determination was a μ Autolab(III) potentiostat connected to a VA663 Stand (Metrohm, Switzerland) and a 663 IME interface and controlled with the GPES software. The

working, counter and reference electrodes were a static mercury drop electrode, a glassy carbon rod and an Ag/AgCl//KCl (3 M), respectively. The voltammetric cell was an acid cleaned polytetrafluoroethylene cell placed into a home-made acrylic holder to allow analysis of small sample volume (5 mL). The stirrer, a specially made rotating polytetrafluoroethylene rod, was set to a fixed stirring speed of 5. All the materials used during the analytical procedure and for storing samples and reagents were cleaned by 1 M HCl (1 week), 0.1 M HCl (1 week), 0.01 M during storage and rinsed using Milli-Q water between steps and prior to usage. Dissolved Fe (0.22 μ m) and soluble Fe determination: Diluted Fe standard solution (1 μ M and 10 μ M Fe) was prepared from a 1,000 mg L⁻¹ standard (SpectrosoL®) diluted with ultrapure water (Millipore, UK, $18 \text{ M}\Omega \text{ cm}^{-1}$ resistivity) and acidified to pH 2 using 9 M HCl (10 μL 9 M HCl/10 mL). HCl trace analysis grade (Fisher Scientific), and ammonia trace metal grade (Fisher Scientific) were employed. The buffer was 0.2 M HEPPS (3-[4-(2hydroxyethyl)-1-piperazinyl]propanesulfonic acid; Merck, ≥ 99.0 %) prepared in ultra-pure water and the pH was adjusted using adequate amount of solid sodium hydroxide (Fisher, 99.2 % trace reagent grade) to a final pH_{NBS} of 8.15. The HEPPS buffer was purified by two overnight equilibrations with 100 μ M MnO₂, removed by filtration using 0.2 μ m cellulose acetate membrane (Whatman) (Obata and van den Berg 2001). The added ligand was 2 mM DHN (2,3-dihydroxynaphthalene) which was prepared by dissolving DHN (Fluka, $\geq 98.0 \%$) in Milli-Q water. The voltammetric procedure used to determine Fe levels in dissolved Fe (0.22 μ m) and in soluble Fe fraction was the method of Caprara et al. (2015), based on the reduction of the electroactive complex Fe-DHN by adsorptive cathodic stripping voltammetry, but slightly modified to suit the high Fe concentrations present in our samples. Standard additions were

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used to determine Fe in each sample, with a minimum of 2 additions and a minimum of 3 voltammetric scans per addition. Prior to the analysis, the samples were acidified to pH < 2.0by addition of HCl (Fisher, Trace Analysis) (3.6 µL of 30 % HCl/3 mL sample) and UVdigested for 2 hours using a home-built system with a 100-W, high-pressure, mercury vapour lamp in 30 mL quartz tubes. Then 1 M NaOH (21 μ L of 1 M NaOH/3 mL sample) was added to adjust the pH to around 8.0 before measurements. For dissolved Fe $(0.22 \mu m)$ determination, the samples were diluted using oceanic seawater (dissolved Fe concentration was 1 ± 0.1 nM; n = 6) which was UV-irradiated (1 hour) freshly prior to use. This oceanic seawater is referred below as the background seawater. The deposition time and the dilution factor were defined by a series of calibration curves using 30 s, 15 s, 10 s and 5 s deposition time and a range of Fe concentrations from 5 nM to 40 nM. The best linear range was achieved using 5 s deposition time until 14 nM Fe concentration. Subsequently the dilution factor for dissolved Fe (0.22 μ m) samples was adjusted so that a maximum of 14 nM was present in the cell at the end of the standard addition (minimum of 2 additions). soluble Fe determinations were mostly made in undiluted samples. Regarding dissolved Fe (0.22 μ m) analysis, 5 mL of the background seawater was pipetted into the polytetrafluoroethylene cell then DHN (final concentration $10 \mu M$) and HEPPS buffer (final concentration 10 mM) were added. The solution was air-purged using cylinder air (1 bar pressure) for 90 s and five voltammetric scans were made to determine the background peak height. Then the sample was added to the background seawater with a dilution factor of at least 30 to ensure an increase of the signal of between 2 and 3 times the background seawater signal. The solution was air-purged again for 90 s and six more voltammetric scans were made. The first Fe standard addition was made sufficient to

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approximately double the original peak and a minimum of three voltammetric scans were carried out. A second Fe standard addition was made to confirm the sensitivity. The sensitivity (nA/nM) was calculated from the slope of the increase in peak height by the Fe standard additions and used to calculate the Fe concentration in the sample. The Fe concentration in the background seawater was calculated in the same way and subtracted from those of the samples. A number of samples were analysed from repeated measurements to determine the precision (standard deviation) of the method. For samples of soluble Fe no dilution was required because the Fe concentrations were sufficiently low. The sample (5 mL) was pipetted directly into the polytetrafluoroethylene cell and the same procedure as that described for dissolved Fe (0.22 μ m) analyses was performed. The method used for soluble Fe determination was validated by using the sample C-10 Bottom (Aug 2018) as control which was previously analysed in triplicate to confirm its concentration. This control sample was measured several times along the days of analysis. Only the certified results for soluble Fe are given here (n = 14, from a total of n = 28) because of a contamination issue resulting in higher soluble Fe values and non-validation from the control sample. Measurement errors ranged around 5.1 % when dilution was required (dissolved Fe (0.22 μ m) analyses) and around 4.4 % for non-diluted samples (soluble Fe determination). The polytetrafluoroethylene cell was rinsed five times using Milli-Q water and then with Milli-Q water plus 10 µM DHN between the analyses to avoid adsorption on the cell wall. Measurements were made using square wave cathodic stripping voltammetry with a deposition time of 5 s at -0.1 V, a 5 s equilibration time (no stirrer) and stripping from -0.35 to -0.75 V (5 mV step, 50 mV amplitude and 10 Hz frequency). Humic substances determination: The borate buffer (boric acid, BDH, UK; 99.8 %) and

the oxidant (potassium bromate, Fisher) were prepared in a mixed solution of 0.2 M borate

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and 0.4 M potassium bromate. The pH of this mixture was adjusted with ~0.08 M ammonia to a final pH_{NBS} of 8.30. The boric acid solution was UV-irradiated for 1 h to ensure that the solution was free of organic contamination. The borate/bromate mixture was purified twice by overnight equilibration with $100~\mu\text{M}$ MnO₂, as previously described (Obata and van den Berg 2001). Fulvic acid stock solutions of Suwannee River Fulvic Acid (SRFA; 0.2 g SRFA L⁻¹) were prepared in ultra-pure water from standard reagents (Suwannee River, IHSS reference material, 2S101F) and stored on dark and refrigerated conditions. The SRFA standard solution was saturated with Fe prior to use to ensure the voltammetric signal reflected the contribution of all Fe-binding groups (Sukekava et al. 2018). Previous work (Laglera and van den Berg 2009) has shown that the binding capacity of SRFA is 16.7 nM Fe /mg FA. We used this value as a guide to saturate the SRFA standard.

Concentrations of humic substances were determined on dissolved samples ($< 0.22 \, \mu m$) using the method described by Laglera et al. (2007) which includes a step of saturation of the sample with Fe in high concentration, equilibration time for 2 to 16 h followed by catalytic analysis and internal calibration with Suwannee River Fulvic Acid (SRFA) standard. To confirm that the concentration of added Fe was sufficient to saturate all the humic binding groups, experiments were performed to evaluate the reaction kinetic in two samples (higher and lower Fe concentration) after two Fe spikes of 100 nM (Sukekava et al. 2018). Immediately after the first Fe standard addition, a rapid increase followed by a slower decrease of the Fe signal was observed (Fig 3). This decrease corresponds to the precipitation of excess, unbound Fe(III) as Fe(III) oxyhydroxides while the signal at equilibrium is due to Fe-humic complexes. After the second Fe addition of 100 nM, the catalytic signal decreased back to its original value after 2 h 30 min confirming that the first addition of 100 nM was enough to saturate all humic binding groups (see Fig. 3). These results led us to adopt the

following protocol. An aliquot of 10 mL of each sample was pipetted into sterilin tubes (Sterilin, polyethylene, Fisher Scientific), saturated with 100 nM Fe and 500 μ L of 0.2 M borate/0.4 M bromate mixed solution were added (final concentration of 10 mM borate buffer/20 mM bromate). The solution was left to equilibrate for 2 h 30 min to allow Fe in excess to compete with other cations and then saturate all the humic binding groups. The sample was transferred to the polytetrafluoroethylene cell for measurements and the oxygen was removed by purging the solution with nitrogen for 300 s. The catalytic analysis consisted in five voltammetric scans followed by internal calibration with two additions of Fe-saturated SRFA standard solution (with a minimum of 3 voltammetric scans per addition) to determine the original concentration of humic substances in µg SRFA L⁻¹ units. The instrumental parameters were: sampled direct current mode at 50 mV s⁻¹, deposition potential -0.1 V, 60 s deposition time, 10 s equilibration time (no stirrer), scan from 0 to -1.0 V.

326 Biological parameters - chlorophyll-a and phaeophytin

> Filters were left overnight into centrifuge tubes filled with 10 mL of 90 % acetone (HPLC grade acetone) on dark and refrigerated conditions to allow the extraction of the pigments (APHA 2012). Extracts were centrifuged at 5,000 rpm for 10 minutes. The absorbance in the extract were determined spectrophotometrically (BEL Photonics UV-M51-Vis spectrophotometer) at 665 nm, 647 nm, 630 nm and 750 nm wavelength before and after acidification with 0.1 mL of 1 N HCl. Chlorophyll-a and phaeophytin (phytoplanktonic detrital material) concentrations were calculated by the equations from the monochromatic method (Lorenzen 1967). The detection limit (DL) was $0.01 \mu g L^{-1}$ for both analytes.

Statistical analyses

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Shapiro-Wilk normality test was used to test the normality of the data distribution, and indicated a departure from normality (p < 0.05). The non-parametric Kruskal-Wallis test was used to identify significant differences between the sampling periods (Nov 2015; Feb 2016; Aug 2018), the isobaths (10, 20 and 30 m) or the inshore-offshore coastal transects (North, Central and South sectors). When the result of the Kruskal-Wallis test was p < 0.05, the Mann-Whitney U test was then used to reveal which pairs of samplings/isobaths/transects differed from each other, and how. The Mann-Whitney U test was also applied to reveal the depth stratification for the physico-chemical, biological parameters, dissolved and soluble Fe fractions and humic substances concentrations. Spearman rank order correlation test was used to test the correlation of Fe size-fractions with humic substances, suspended particulate material and biological parameters. All statistical analyses were based on $\alpha = 0.05$. Statistics analyses were performed using PAST software (version 3.26).

348 Results

Physico-chemical and biological parameters

Over the course of the 3 sampling campaigns, there were no significant differences with depth, neither for the physico-chemical nor the biological parameters (Mann-Whitney, p > 0.05). Significant differences in the temporal distribution were only observed for suspended particulate material (Kruskal-Wallis, p < 0.05), which reached the highest values in Nov 2015 (from 30 to 300 mg L⁻¹) (Table 1). Because of the low quantity of sampling stations in Nov 2015 and Feb 2016 we were not able to test statistically spatial changes between the isobaths and inshore-offshore coastal transects separately for these samplings. In Aug 2018, the levels of suspended particulate material decreased with distance from the Doce River mouth (Kruskal-Wallis, p < 0.05), ranging from 16.5 to 25.3 mg L⁻¹ at the 10 m isobaths (i.e. near the coast) and from 14.1 to 17.3 mg L⁻¹ at the 30 m isobaths (i.e. far from the coast) (Table 1).

waters when compared to surface (Table 1). There was no trend of depth stratification for the

salinity (Mann-Whitney, *p* > 0.05) which ranged from 36.0 to 36.6 at bottom and from 34.5 to 36.8 at surface (Table 1). The pH was measured between 8.00 and 8.15 in the last sampling campaign; it was not recorded in Nov 2015, nor in Feb 2016.

Chlorophyll-a levels were comparable to phaeophytin levels in Nov 2015 and Feb 2016 but markedly lower in Aug 2018 (Table 1). Most of the Chlorophyll-a concentrations in surface waters were below detection limit. Near the bottom, concentrations averaged 1.5 ± 1.6 μ g L⁻¹ for Chlorophyll-a (n = 9) and 1.3 ± 1.1 μ g L⁻¹ for phaeophytin (n = 12) (Table 1). Distribution of dissolved Fe (0.45 μ m), dissolved Fe (0.22 μ m), soluble Fe and humic substances

Over the course of the 3 sampling campaigns, the parameters dissolved Fe (0.45 μ m), dissolved Fe (0.22 μ m), soluble Fe and humic substances presented homogeneous distributions along the water column depth (Mann-Whitney, p > 0.05) and distance from the Doce River mouth (Kruskal-Wallis, p > 0.05).

The dissolved Fe (0.45 μ m) concentrations in surface and bottom waters were similar, ranging from 784 to 2,784 nM at the surface, and between 739 and 2,570 nM for the bottom waters (Table 2). During the acute phase of the impact (Nov 2015), the highest dissolved Fe (0.45 μ m) values in both the surface and bottom waters were found at station C-10, located in front of the Doce River mouth (Fig. 4). For the Central and North sections, dissolved Fe (0.45 μ m) concentrations decreased with time at similar rates, while the South section showed a different behaviour, with concentrations increasing from Feb 2016 to Aug 2018 (Fig. 4a). In Aug 2018, it is remarkable that the Central and North measurements were within a narrow concentration range (863 \pm 90 nM on North stations; 1,166 \pm 166 nM on Central stations) while much higher concentrations were observed for the South stations (2,010 \pm 248 nM) (Kruskal-Wallis, p < 0.05) (Fig. 4a and Table 2). The same pattern was observed for

sedimentary Fe distribution (Supplementary, Table S1; Fig. S2) indicating a trend of material accumulation on the south sector over time.

The concentrations of dissolved Fe (0.22 μ m) were higher during the acute phase of the disaster (Nov 2015; Kruskal-Wallis, p < 0.05) and increased again in Aug 2018 (Kruskal-Wallis, p < 0.05) (Fig. 4b). The levels ranged from 30 to 245 nM in surface waters, whereas the values in bottom waters were found to vary between 30 and 698 nM. In Aug 2018 the highest values occurred at station N-10 for the surface waters and N-20 at bottom. The sediment collected at station N-20 in Aug 2018 presented a marked reddish colour characteristic of the presence of Fe oxide. Processes such as resuspension, remobilisation and diffusion may thus explain the elevated value of dissolved Fe (0.22 μ m) concentration measured in the water column just above the sediment. The lowest dissolved Fe (0.22 μ m) concentrations were observed in Feb 2016 at the isobaths of 30 meters (station C-30 for the surface samples and N-30 for those at bottom) (Fig. 4b; Table 2).

The concentrations of soluble Fe ranged from 10 to 25 nM (stations S-30 and N-20, respectively) in the surface waters and varied between 10 and 37 nM at bottom, with the lowest concentrations at station C-30 in Aug 2018 and the highest occurring at station C-30 in Feb 2016 (Fig. 4c; Table 2).

Similar distribution patterns were observed for humic substances and dissolved Fe (0.22 μ m) in surface waters. In general, the concentrations of dissolved Fe (0.22 μ m) and humic substances showed a trend of decrease with the distance from the Doce River mouth in the surface samples and increased again in the 30 m isobaths (Fig. 4d; Table 3). The highest value of humic substances was found at station N-10 in Aug 2018 (350 μ g SRFA L⁻¹) and the lowest occurred at station C-30 in Feb 2016 (15 μ g SRFA L⁻¹) (Fig. 4d; Table 3). In bottom waters, the highest concentration of humic substances occurred at station C-30 in Feb 2016

411 L^{-1}). The dissolved Fe (0.22 μ m) concentrations increased from North to the South sectors 412 whereas the humic substances decreased until the station C-20 and increased again from the 413 station S-20 (Fig. 4d; Table 3). 414 Size-fractionated Fe proportion and Fe-binding by humic substances 415 The concentrations of dissolved Fe (0.22 μ m) were up to 98 % smaller than those of 416 dissolved Fe (0.45 μ m) (Table 2; Fig. 5) suggesting that most Fe is present in a large (> 0.22 μ m) colloidal form. Similarly, ultrafiltration (0.02 μ m) of some pre-filtered samples (0.22 417 418 μ m) caused significant removal, from 39 to 95 % of the Fe from dissolved Fe (0.22 μ m) to the 419 soluble fraction (Table 2; Fig. 5), also suggesting that a significant proportion of dissolved Fe 420 (0.22 µm) fraction was present in a colloidal form higher than 0.02 µm. The high presence of 421 orange particulate matter retained on 0.22 μ m membrane is an additional evidence of colloidal 422 dominance in our samples (Supplementary, Fig. S1). 423 Assuming a ratio of 16.7 nmol Fe (mg FA)⁻¹ for the binding capacity of humic substances 424 (Laglera and van den Berg 2009), the maximum concentration of Fe-binding by humic substances would have been 0.2 to 5.8 nmol L⁻¹ (Table 3). Based on previous studies, most of 425 426 dissolved Fe would have been anticipated to occur as organic complexes, mainly with humic 427 substances (Laglera and van den Berg 2009). In reality, the binding capacity of humic 428 substances could only explain a maximum of between 0.2 and 13 % (median is 2 %) of the 429 dissolved Fe (0.22 μ m) (Table 3). We did not measure levels of humic substances in the 430 soluble fraction but assuming that those are the same as in the $< 0.22 \,\mu m$ fraction, they could 431 only bind from 0.2 to 43 % (median is 10 %) of the soluble Fe levels (Table 3). These 432 percentages correspond to the maximum binding that would occur if there was no competition

from other metals. However, when carrying out our measurements of humic substances (Fig.

 $(241 \mu g SRFA L^{-1})$ while the lowest value occurred at station N-20 in Nov 2015 $(44 \mu g SRFA L^{-1})$

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3), it was clear that humic substances present in the sample were not saturated with Fe since a significant increase (52 % for the sample shown in Fig. 3) of the signal of Fe-humic complexes was observed upon additions of Fe (light grey and black markers in Fig. 3). This increase can be explained by the presence of competing metals for complexation with humic substances (Sukekava et al. 2018). It is thus clear that levels of humic substances measured in this study cannot explain the Fe dissolved and soluble concentrations, indicating that there are other processes and/or other compounds that maintain the Fe in the dissolved/soluble fraction. Correlation analyses No significant correlation was observed between dissolved Fe (0.45 μ m) and suspended particulate material, neither with Chl-a nor Phaeophytin (Supplementary, Table S2). For the dissolved Fe $(0.22 \, \mu \text{m})$, on one hand, a positive correlation with humic substances was found in surface waters (Supplementary, Table S2; Fig. 6a; R = 0.62; p = 0.02; n = 12) suggesting that these variables originate from the same source. The coeficient of correlation in these samples was relatively low due to three outliers (open circles in Fig. 6a). Exclusion of these points resulted in a strong linear fit $(R = 0.98; r^2 = 0.97; p = 0.00001; n = 9)$, as shown in the inserted diagram (Fig. 6a). On the other hand, no significant correlation was found between humic substances and dissolved Fe (0.22 μ m) concentrations in bottom waters (R = -0.44; p =0.15; n = 11). The dissolved Fe (0.22 μ m) concentrations increased with the levels of suspended particulate material in surface waters (Supplementary, Table S2; R = 0.60; p =0.02; n = 14). A weak positive correlation was also found between phaeophytin and dissolved Fe (0.22 μ m) (Supplementary, Table S2; Fig. 6b; R = 0.40, p = 0.04; n = 25) which may be

due to the export of phytoplanktonic debris to the marine environment.

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Dissolved Fe concentrations and sources

The results of this study demonstrate that the concentrations of dissolved Fe in coastal waters impacted by the Fundão dam collapse are very high across all size fractions, even almost three years after the disaster, without showing any obvious signs of decrease. The dissolved Fe (0.45 μ m) levels found here are much higher than those reported in other marine environments affected by mining tailings (Supplementary, Table S3). Only the lowest value recorded in this study is comparable to the mean concentration of dissolved Fe $(0.22 \,\mu\text{m})$ observed in coastal systems impacted by abandoned lead mining (Tovar-Sánchez et al. 2016), while the range of dissolved Fe (0.45 μ m) concentrations found here is similar to the values recorded in coastline impacted by acidic metal-rich fluids from abandoned mines (Koski et al. 2008) (Supplementary, Table S3). The fluxes of dissolved Fe to the coastal areas vulnerable to mining impacts are highly intensified by the Fe inputs from mining tailings. These loads can reach the coastal zone by riverine inputs, pore-water exchange and submarine groundwater discharge (Longhini et al. 2019). In our case, the spatial distribution pattern of dissolved Fe (0.45 μ m) and dissolved Fe (0.22 µm) concentrations indicates that there are two main sources of dissolved Fe to consider: the fluvial input of suspended particulate material from the Doce River and the resuspension of sediment/tailings particles deposited on the continental shelf. The positive correlation between dissolved Fe (0.22 µm) and suspended particulate material (Supplementary, Table S2) in surface waters is in agreement with the substantial increase in the Doce River transport of Fe rich suspended particulate material observed by Hatje et al. (2017) after the Fundão dam disaster (maximum values of suspended particulate material was 33 g L⁻¹ and particulate Fe reached up to 48 mg Fe g⁻¹). Iron in Fundão dam tailings is mainly present as crystalline oxyhydroxides (89 ± 2.3 %) in forms of stable goethite (Almeida et al. 2018; Queiroz et al. 2018), which possibly reduce the estuarine removal of dissolved Fe. This suggests a limited estuarine trapping with particulate and colloidal organic matter (Muller 2018) and then a high stability of the dissolved Fe forms that reached the coastal area after the dam failure.

The transport of tailings mud from the river to the coastal sea in the acute stage of the disaster occurred as a massive loading of sediment/tailings highly enriched with Fe (44.3%; Pereira et al. 2008) of finer grain size compared to pre-disaster conditions (Gomes et al. 2017). This modified sediment deposited on the continental shelf remains fluid and subject to resuspension by high winds/waves dynamics (Rudorff et al. 2018). The tailings mud transport at the bottom then explains the increase of sedimentary Fe and dissolved Fe (0.45 μ m) concentrations on South stations over time and higher values of bottom dissolved Fe (0.22) μm) at those stations. These increasing concentrations are in agreement with the bottom sediment transport (Quaresma et al. 2015) and with the southward trajectory of the Doce River plume when trade winds are prevalent (Marta-Almeida et al. 2016). Mobility tests using the Fundão dam tailings confirm this sedimentary/tailings source as they have shown that Fe (and other metals like Ba, Sr, Mn and Al) can enter the dissolved phase from mud particles, especially in environments with high calcium content, like seawater (Segura et al. 2016). This phase transfer leads to an increased Fe bioavailability from the sediment during the transport from the riverine portion to the estuarine mouth, which has been quantified as a 16 % increase by Hatje et al. (2017).

Concentrations and sources of humic substances

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Our study is the first one to inform about the role of humic substances on Fe transport in waters impacted by Fe ore tailings. Even if humic substances show the same distribution

pattern as dissolved Fe (0.22 μ m) in surface waters and show higher concentration near the coast, attesting of their continental input from the Doce River, dissolved Fe-humic complexes can explain only from 0.2 to 13 % of the dissolved Fe (0.22 μ m) levels. To compare with studies not subject to ore tailing, humic substances represented from 23 to 58 % of the ligand bulk in North Atlantic (Whitby et al. 2020b), while they could account for almost the all Febinding ligands in Liverpool Bay (Laglera and van den Berg 2009) and in the Arctic (Sukekava et al. 2018). Furthermore, humic-binding capacity of Fe usually shows a good agreement with dissolved Fe (0.22 μ m) concentrations (Sukekava et al. 2018) and dissolved Fe-humic complexes accounts for 80 % of the dissolved Fe speciation (Laglera et al. 2019).

The imbalance that we observe between humic substances and the dissolved Fe concentrations is likely caused by the abnormal processes that mobilize the dissolved Fe from the tailings material, whereas humic substances are delivered to the area from natural sources. Likewise, the lack of significant correlation between humic substances and dissolved Fe (0.22 μ m) concentrations in the bottom waters might be because different processes are decoupling those components, as dissolved Fe shows substantial inputs from the sediment/tailings ressuspension. In natural systems, humic substances and Fe are known to be subject to coprecipitation in the estuarine mixing zone reaching a removal rate ranges from 90 % to 95 % (Boyle et al. 1977; Sholkovitz et al. 1978). At our location, dissolved Fe (0.22 μ m) concentrations reached 21.8 \pm 46.1 μ M in continental water bodies affected by the tailings (Segura et al. 2016) and part of this material was deposited during the transport, probably carrying downward the humic substances and other compounds.

Our results also suggest some influence of humic substances from marine sources as their concentrations increased at the 30 m isobaths (i.e. far from the coast). The high concentrations of phaeophytin against the very low Clh-a levels are proxies of the presence of

phytoplanktonic debris/senescence which can be transformed by microorganisms to marine humic-like compounds and possibly contribute to the bulk of humic substances in our system. Indeed, marine-derived humic substances produced by microbial transformation of biogenic particles has been recently described as the most important Fe-binding ligand in surface waters of the Mediterranean Sea and Southern Ocean (Whitby et al. 2020a). In coastal waters of Western Antarctic Peninsula and Drake Passage, humic-like compounds was attributed to the mucus production (as exopolymeric substances) by phytoplanktonic assemblages (Trimborn et al. 2015) confirming the marine origin of these compounds. Senescence of biological material is also a source of exopolymeric substances released to the water column as cellular exudates, which is believed to be included in the pool of humic-like compounds (Norman et al. 2015). The kinetic experiments revealed the presence of humic under-saturated by Fe as the signal of humic substances in the sample N-20 bottom (Aug 2018) shown in Fig. 3 was increased by 52 % after the saturation with 100 nM Fe(III). Similarly, Sukekava et al. (2018) observed an increase of about 67 % on signal of humic substances after the saturation with 60 nM Fe(III) in seawater samples from Arctic Ocean. This result could indicate that the behaviour of humic substances to outcompete Fe and other metals may be similar, irrespective of the origin of the seawater, as our samples are fundamentally different than those analysed by Sukekava et al. (2018) in terms of metal concentrations, physico-chemical conditions and possibly organic ligands contents. So, the increase of the initial signal after Fe saturation can be explained by two hypotheses: 1) the dissolved Fe is already complexed by other compounds or stabilized as colloidal forms (see next item); and/or 2) humic substances are

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bound by other metals.

The co-existence of multiple metals should change the Fe organic speciation due to the differences on binding capacity of specific organic compounds with different metals (Hassler et al. 2017). Humic substances are also important ligands for Cu, Zn, Co and Al in seawater (Yang and van den Berg 2009). Considering the levels of dCu (0.45 μ m) were lower than dissolved Fe (0.45 μ m) (Supplementary, Table S4), competition with Cu does not seem to be a plausible explanation. On the other hand, dAl (0.45 μ m) reached levels as high as 9.9 to 12.5 μ M in Nov 2015, from 0.39 to 9.2 μ M in Feb 2016 and from 3.4 to 14.8 μ M in Aug 2018 (Supplementary, Table S4) suggesting that Al could potentially outcompete Fe for binding to humic substances in this coastal area. Similarly, dZn (0.45 μ m) was a possible candidate for humic competition in the acute phase of the impact as its concentrations occurred between 0.7 and 1.1 μ M in Nov 2015 (Supplementary, Table S4).

Forms of dissolved Fe and soluble Fe

The rapid change in Fe concentrations across the size spectrum indicates that a very significant portion of Fe occurs in colloidal forms. Indeed, Fe in seawater was shown to be primarily in the colloidal size fraction, considering the size range from 0.02 to 0.4 μ m (Wu et al. 2001) and from 10 kDa to 0.2 μ m (Fitzsimmons et al. 2015). In our case, these colloids, highly modified by unnatural chemical and physical processes (mineral treatment for Fe ore pre-concentration), may be the main reason for the high dissolved Fe levels in the < 0.45 μ m and < 0.22 μ m size fractions.

This is also in agreement with changes to flocculation processes that were demonstrated by settling experiments performed with mud sediment fraction ($< 63 \mu m$) from the continental shelf before and after the disaster. The post-disaster decrease in the settling velocity of fine flocs was suggested to be a result of changes of the sediment composition, like the presence of porous Fe oxide goethite (Grilo et al. 2018). In addition, adsorptive characterisation of tailings

from the Fundão dam was consistent with a monolayer model with low adsorption capacity and low interaction among adjacent molecules (Almeida et al. 2018) which means that this material was probably less prone to processes of flocculation.

Humic substances played a minor role in stabilizing the soluble Fe fraction. Therefore, soluble Fe speciation can be assumed to be mostly comprised of various small organic complexes, nanoparticles (essentially Fe oxy-hydroxide) and a minute amount of free metal ions (Tagliabue et al. 2017). Considering that humic substances are generally the most important organic ligands in coastal waters affected by riverine inputs (Laglera and van den Berg 2009), it is therefore likely that in our case, processes other than complexation by humic substances are at play to stabilise soluble Fe in the water column. Even though other natural organic substances, such as siderophores, are important ligands to soluble Fe in marine systems (Gledhill and Buck 2012), these compounds are unlikely to be the predominant ligand in view of the low levels of Chl-a that were consistently measured in our study.

The complexation of soluble Fe with unnatural organic ligands is one hypothesis to be considered. Organic compounds are frequently used during the flotation stage employed to concentrate the mineral product as a richer Fe ore pulp (Filippov et al. 2014). Reverse cationic flotation using amine-based collectors is the most common method employed by the Fe ore processing industry (Filippov et al. 2014). Amine adsorb on silicates (mainly quartz) surface and also on metal oxides causing the flotation of quartz and other associated mineral (Filippov et al. 2014), such as goethite. Querol et al. (1999) and Alzaga et al. (1999) were the first authors to reference aromatic amine compounds as a geochemical tracer of mine spillage after the Aznalcóllar dam disaster (Los Frailes, SW Spain). Similarly to the Fundão, ore processing in Aznalcóllar generated waste products with very high Fe content and other chemical contaminants, like amine compounds from flotation reagents, were deposited as tailings

(Querol et al. 1999). Aromatic amine was widespread recorded either in atmospheric particulate matter (Querol et al. 1999) or in water bodies affected by that mining disaster (Alzaga et al. 1999). Likewise, infrared spectroscopy analyses confirmed the presence of primary and secondary amines and amides in the Fundão tailing composition (Almeida et al. 2018) whereas ether amine was quantified in high concentrations (up to 57.8 mg kg⁻¹) when comparing sediments contaminated by those tailings against preserved areas (Santos et al. 2019). According to these authors both amine and Na are the main toxic compounds found in the Fundão tailings and both are related to reverse flotation procedure (Santos et al. 2019). These compounds would appear to be highly stable as aromatic amine have been frequently identified in seawater samples collected on the continental shelf adjacent to the Doce River mouth since the disaster (R.R. Neto unpubl.). Stability of the complex soluble Fe-amine and concentrations of that amine ligand could be high enough to compete with the binding from other natural ligands, such as humic substances, but this is presently unknown. The presence of colloidal Fe species smaller than $0.02 \mu m$ is also a likely hypothesis to support the levels of soluble Fe in our study area. Analyses of sedimentary metal partitioning in areas affected by the Fundão dam tailing showed that pyrite accounts for only 1 % of all metals whereas crystalline Fe oxyhydroxides are the most important solid phases for Fe and other metals (Queiroz et al. 2018). Fitzsimmons et al. (2015) reported that inorganic Fe occurring as nanoparticles was probably ubiquitous in the colloidal phase (10 kDa < cFe < 0.2 μm) of samples collected in surface oceanic waters. Inorganic colloids as nanoparticulate pyrite (FeS₂) (Gartman et al. 2014), Fe-containing silicate particles (Gartman et al. 2014) and nanoparticulate Fe(III) oxyhydroxides (Fitzsimmons et al. 2017) have been identified to encompass the bulk of dissolved Fe in fluids leaking from hydrothermal vents. These

inorganic entities fall into the colloidal and soluble size classes as their aggregates reach from

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3 nm to 200 nm in size (Gartman et al. 2014; Fitzsimmons et al. 2017). They are characterized by a relatively long half-life, slow settling rates and thus a potential to travel long distances (Gartman et al. 2014). Another process controlling the stability of nanoparticles is the reversible scavenging mediated by desorption/sorption ligand exchanges from particulate Fe (pFe) to the colloidal phase which protect it from precipitation and settling (Fitzsimmons et al. 2017). These processes are expected to occur in ocean regions with high particulate Fe inputs, including continental margins (Fitzsimmons et al. 2017).

Taking all the above factors into consideration, the different dissolved Fe size fractions occur as a continuum of inorganic and organic components regulating the dissolved Fe speciation (Tagliabue et al. 2017). The major dissolved Fe form in the study area is colloidal Fe, mostly as nanoparticulate Fe(III) oxyhydroxides which may be stabilised by amine or humic compounds. Amine compounds in particular, are thought to strongly bind Fe, thus controlling its solubility in the coastal waters off the Doce River (Fig. 7).

How much Fe can be transported to the open ocean?

In view of the continuing input and high stability of dissolved Fe and soluble Fe in those waters, it is worth considering the possibility of Fe transport from the continental shelf to the open ocean. Modelling study evaluating the dispersion of contaminated Doce River plume two months after the disaster showed a net flow mainly southward along the outer shelf, shelf break and reaching the open ocean about 200 km from the coast (Marta-Almeida et al. 2016). The riverine water residence time on the shelf is a key factor to determine the transport of inorganic N and P to the oceans and similar behaviour is expected for trace metals (Sharples et al. 2017). Shelves with short residence time of riverine plume (essentially located at low latitudes) deliver higher loads of nutrients to the open ocean (Sharples et al. 2017). In this context, the mean shelf residence time at Abrolhos (17.5 S, 37.5 W; 250 km off the Doce

647 River mouth) was estimated to be 7 days, with the result that 84 % and 97 % of the dissolved 648 inorganic nitrogen and phosphorus (DIN and DIP, respectively) reached the open ocean, 649 which is also believed to occur for metals (Sharpes et al. 2017). Considering that Abrolhos is 650 a wider shelf (250 km) than that at our study site (50 km), the residence time on the 651 continental shelf adjacent to the Doce River mouth is expected to be lower. 652 Using a conservative residence time of 7 days, a 12 x 15 km impacted area (area 653 monitored in this study; Fig. 1), an average depth of 20 m and an average dissolved Fe (0.45 654 μ m) of 1.44 μ M (average concentration in Feb 2016 and Aug 2018; Table 2), the total Fe flux to the open ocean is estimated at c.a. 260 Mmoles y⁻¹, which represents approximately 655 between 1 and 5% of the net Fe fluvial worldwide flux in the oceans $(5.4 - 23 \text{ Gmoles v}^{-1} -$ 656 657 Chester 2003), highlighting the sheer magnitude of this event. Furthermore, this 260 Mmoles y⁻¹ Fe flux is a lower approximation since: 1 - the impacted area might be significantly larger 658 659 than the area considered here (the Doce River turbidity plume extended more than 75 km into 660 the continental shelf during the acute phase of the impact according to Rudorff et al. 2018); 661 and 2 - the sampling campaigns were conducted during conditions where remobilisation 662 processes were probably at their lowest (low river discharge; Fig. 2; and no cold front passage). In addition, using an average Doce River flow of 540 m³ s⁻¹ (Oliveira and Quaresma 663 664 2017) and an average dissolved Fe (0.45 μ m) concentration of c.a. 1.6 μ M (mean 665 concentration found in Feb 2016 in comparable river discharge conditions; Table 2) a net dissolved Fe flux from the Doce River is estimated at c.a. 27 Mmoles v⁻¹ suggesting that in 666 667 post-disaster conditions almost c.a. 90 % of dissolved Fe present in the water column is 668 coming from continuous remobilisation of the sediment deposited on the continental shelf (Fig. 7). This contribution of the Doce River for the dissolved Fe fluxes to the continental 669 670 shelf is also in agreement with dissolved Fe fluxes calculated by Hatje et al. (2017) which

reached 58.8 mg dFe s^{-1} (c.a. 35 Mmoles y^{-1}) in river discharge conditions of about 1,000 m³ s^{-1} .

Issues to be further discussed: the fate of submarine tailings disposal

Considering the huge fluxes of dissolved Fe estimated here and the main role of sediment/tailings resuspension on the continental shelf, it is also worth to consider the role of submarine tailings disposal and deep-sea tailings placement for dissolved metal release and consequent ocean enrichment (Ramirez-Llodra et al. 2015). Tailings disposal in marine shore was used in the 70s and has been recently discussed as an alternative strategy to deal with land tailing disposal problems, including catastrophic rupture (Kwong et al. 2019). Currently, there are fifteen mines in activity using submarine tailings disposal or deep-sea tailings placement whereas most of operations started in the past were closed and generated abandoned mine wastes along the coast (Ramirez-Llodra et al. 2015). The accumulation of tailings in marine systems are object of concern since most of the impacts, including chemical processes of dissolved and particulate metals released from the submarine tailings, are still unknown (Ramirez-Llodra et al. 2015). Our study suggests that mining tailings deposited on marine bottom might be non-negligible sources of stable forms of dissolved Fe and soluble Fe to the oceans and may affect the global Fe ocean budget considering all the coastal areas affected by mining activities worldwide (Longhini et al. 2019).

Conclusion

The Fe ore tailings spillage after the Fundão dam disaster lead to huge and stable concentrations of dissolved and soluble Fe in the adjoining coastal areas even three years later. This dissolved and soluble Fe stability over time suggests that processes are at play to help Fe remobilization, e.g. complexation or stabilisation of Fe rich colloids through adsorption of organic molecules. While humic substances have been previously identified as

the main Fe ligands in coastal waters, the Fe-humic concentrations observed in our study indicate that other processes are able to stabilise Fe in the water column. Colloidal particles are the main forms encompassing the bulk of dissolved Fe fractions, while other organic substances (here supposed to be amine compounds) are probably acting to stabilise small colloids as nanoparticulate Fe(III) oxyhydroxides. Further studies should evaluate the relation between dissolved Fe and synthetic organic ligands used during Fe ore processing (specifically amine groups). Microscopy studies to identify the presence of nanoparticulate colloidal species could also shed light on the processes at play that maintain Fe in the soluble fraction.

At present, the main source of dissolved Fe to the water column is the resuspension of fine mud on the continental shelf which has a high content of goethite, whereas the offshore transport of aquatic colloids by the Doce River and the remobilisation of accumulated material from the river bed seem to be less representative. The Fe fluxes estimated here are comparable to 1-5% of the net Fe fluvial worldwide inputs to the oceans (as calculated by Chester 2003) highlighting the sheer magnitude of this catastrophe. The role of mining tailings as sources of stable forms of dissolved and soluble Fe in a global context is currently unknown but this study suggests that it should be investigated further.

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886	Figure legends
887	Fig. 1. Distribution of sampling stations on the continental shelf adjacent to the Doce River
888	mouth (Southeast, Brazil) after the Fundão dam rupture. N, C and S correspond to the sectors
889	North, Central and South, respectively; 10, 20 and 30 indicate the isobaths 10, 20 and 30
890	meters.
891	Fig. 2. Daily average of the Doce River discharge (m ³ s ⁻¹) for the months November 2015,
892	December 2015, January 2016, February 2016 and August 2018. Black dashed arrow
893	represents the day that the iron ore tailings reached the coastal waters after the Fundão dam
894	rupture; Light grey arrows indicate the sampling days. Data from Agência Nacional de Água
895	- ANA (Colatina station): https://www.snirh.gov.br/.

Fig. 3. Catalytic adsorptive cathodic stripping voltammetry used for determination of humic

897 substances. Peak heights were obtained from the sample N-20 bottom (Aug 2018) before and 898 after two iron additions of 100 nM and kinetic stabilization for 2 h 30 min. Light grey markers 899 define the original Fe-humic complexes in the sample; black markers are Fe-humic complexes 900 after saturation of all humic substances binding with iron; triangle markers: internal 901 calibration with iron-saturated SRFA standard to determine the humic substances 902 concentration. Dissolved Fe (0.22 μ m) concentration in the sample was 698 \pm 44 nM. 903 Procedure as described by Sukekava et al. (2018). SRFA: Suwannee River Fulvic Acid. 904 Fig. 4. Spatial and temporal distribution of dissolved Fe (0.45 μ m), dissolved Fe (0.22 μ m), 905 soluble Fe $(0.02 \,\mu\text{m})$ and humic substances concentrations (average between the surface and 906 bottom data) of the continental shelf adjacent to the Doce River mouth. dFe: dissolved Fe; 907 sFe: soluble Fe; HS: humic substances. 908 Fig. 5. Spatial distribution of dissolved Fe (0.45 μ m) (dark grey boxplots), dissolved Fe (0.22 909 μ m) (light grey boxplots) and soluble Fe (0.02 μ m) (white boxplots) to highlight the 910 difference in concentrations between the size-factions (see scale breaks) in each transect over 911 the course of the three samplings (Nov 2015, Feb 2016 and Aug 2018). dFe: dissolved Fe; 912 sFe: soluble Fe. 913 Fig. 6. Scatterplot diagrams between a) humic substances and dissolved Fe $(0.22 \, \mu \text{m})$ 914 concentrations in surface seawater, R = 0.62, p = 0.04, n = 12 (excluding the points out of the 915 linear fit as showed in the inserted diagram: R = 0.98, p = 0.00001, n = 9); b) Phaeophytin and 916 dissolved Fe (0.22 μ m) concentrations including surface and bottom results, R = 0.40, p =917 0.04, n = 25. Dashed lines represent the upper and lower 95% confidence limit ($\alpha = 0.05$). dFe: dissolved Fe; HS: humic substances. 918

Fig. 7. Schematic presentation of the dissolved and soluble Fe speciation in the water column of continental shelf in the vicinity of the Doce River. ^a dissolved Fe fluxes and the relative importance of Doce River flow *vs* sediment resuspension for dissolved Fe inputs were estimated as described in the text. dFe: dissolved Fe.

Table 1. Physico-chemical (temperature, salinity, pH and suspended particulate matter - SPM) and biological parameters (chlorophyll-*a* and phaeophytin) recorded in the continental shelf adjacent to the Doce River mouth at two depths (surface and bottom) and at three different times after the environmental disaster (November 2015, February 2016; August 2018). ND: not determined; BDL: below detection limit.

SAMPLE	T (°C)		Salinity		Hd	4	SPM (mo L. ¹)	0	$\mathbf{Chl-}a\;(\boldsymbol{\mu}\mathbf{g}\;\mathbf{L}^{\text{-}1})$		Phaeophytin	$(\mu {f g} {f L}^{-1})$
SA	Surface	Bottom	Surface	Bottom	Surface	Bottom	Surface	Bottom	Surface	Bottom	Surface	Bottom
Nov 2015												
N-20	26.5	21.8	36.5	36.6	ND	ND	30.0	42.0	0.62	5.35	0.68	BDL
C-10	24.8	24.4	36.4	36.5	ND	ND	85.0	300.0	BDL	BDL	1.87	3.03
Feb 2016												
N-25	26.7	20.9	36.7	36.5	ND	ND	2.93	1.20	1.23	BDL	BDL	0.50
C-30	26.1	21.1	36.8	36.5	ND	ND	1.60	0.27	0.45	BDL	0.18	0.67
S-20	24.0	21.9	34.5	36.5	ND	ND	1.07	62.0	1.78	1.78	0.30	3.21
Aug 2018												
N-10	24.8	25.1	36.0	36.5	8.07	8.09	16.5	25.3	BDL	1.91	2.51	BDL
C-10	24.5	25.3	36.0	36.5	8.11	8.11	23.3	24.7	BDL	1.49	2.86	0.59
S-10	25.0	24.6	36.0	36.5	8.09	8.07	18.3	21.4	BDL	BDL	2.27	3.70
N-20	23.6	26.5	35.0	36.0	8.13	8.07	16.6	16.9	BDL	0.41	1.70	1.01
C-20	25.4	25.0	36.5	36.5	8.09	8.07	14.1	15.8	BDL	0.41	2.96	0.73

S-20	25.7	24.5	36.0	36.0	8.14	8.12	16.9	16.2	2.03	0.89	BDL	0.98
N-30	24.0	23.6	36.0	36.5	8.15	8.12	17.3	14.1	BDL	BDL	1.42	0.85
C-30	25.0	25.3	36.5	36.5	8.06	8.04	14.3	15.7	BDL	0.45	2.67	0.49
S-30	25.1	24.7	36.5	36.5	8.00	8.02	14.4	15.5	BDL	0.42	1.42	1.06
Min	23.6	20.9	34.5	36.0	8.00	8.02	1.07	0.27	0.45	0.41	0.18	0.49
Max	26.7	26.5	36.8	36.6	8.15	8.12	85.00	62.00	2.03	5.35	2.96	3.70
Average	25.1	23.9	36.1	36.4	8.09	8.08	19.44	20.86	1.22	1.45	1.74	1.25
SD	0.9	1.8	0.6	0.2	0.05	0.03	20.50	16.20	0.69	1.58	0.97	1.11

Table 2. Iron concentrations for dissolved dFe ($< 0.45 \ \mu m$), dissolved Fe ($< 0.22 \ \mu m$) and soluble Fe ($< 0.02 \ \mu m$) fractions in the continental shelf adjacent to the Doce River mouth. The dissolved Fe ($0.45 \ \mu m$) concentrations were determined by ICP-MS; dissolved Fe ($0.22 \ \mu m$) and soluble Fe were analysed by cathodic stripping voltammetry. The standard deviation of the dissolved Fe ($0.22 \ \mu m$) indicated for some samples was obtained from replicate measurements. The percentages shown in parentheses indicate the proportion of dissolved Fe ($0.45 \ \mu m$) found in the $< 0.2 \ \mu m$ or $< 0.02 \ \mu m$ fraction. dFe: dissolved Fe; sFe: soluble Fe; ND: not determined.

Sample	dFe (0.45	μm) nM	dFe (0.22 μ	m) nM	sFe $(0.02 \mu\mathrm{m}) \mathrm{nM}$		
Sample	Surface	Bottom	Surface	Bottom	Surface	Bottom	
Nov 2015							
N 20	1 126	1,177	$204 \pm 11 \; (n=3)$	109	10	NID	
N-20	N-20 1,136		(17%)	(10%)	(1%)	ND	
C 10	2.607	2,570	182	ND	15 ± 1	ND	
C-10	C-10 2,607		ND (6%)		(1%)	ND	
Feb 2016							
N 05	1.756	2 104	$35 \pm 2 \ (n=2)$	30	ND	ND	
N-25	1,756	2,184	(2%)	(2%)	ND	ND	
G 20	1.666	1,550	30	61	ND.	38 ± 2	
C-30	C-30 1,666		(3%)	(5%)	ND	(3%)	
G 20	1.022	1 225	97	71	11 ± 0.4	ND	
S-20	1,032	1,337	(6%)	(5%)	(1%)	ND	

Aug 2018

N-10	964	739	245	75	ND	11 ± 0.4
IN-10	904	139	(26%)	(26%) (10%)		(1%)
C-10	990	964	137	91	16 ± 1	29 ± 1
C-10	990	904	(14%)	(9%)	(2%)	(3%)
S-10	1 6/11	1 776	148	206	9 ± 1	ND
5-10	1,641	1,776	(9%)	(12%)	(1%)	ND
N 20	784	025	117	$698 \pm 44 \; (n=2)$	25 ± 1	ND
N-20	704	925	(15%)	(76%)	(3%)	ND
C 20	1 220	1 201	64	51	ND	17 ± 1
C-20 1,238	1,281	(5%)	(4%)	ND	(1%)	
g 20	2 170	1,975	102	$194 \pm 13 \; (n=2)$	ND	ND
3-20	S-20 2,170		(5%)	(10%)	ND	ND
N-30	837	927	125	$176 \pm 5 \ (n=2)$	18 ± 1	28 ± 1
N-30	037	921	(15%)	(19%)	(2%)	(3%)
C-30	1,140	1 202	155	185	ND	10 ± 1
C-30	1,140	1,383	(14%)	(13%)	ND	(1%)
g 20	2 794	1 716	$186 \pm 7 \; (n=3)$	95	10 ± 0.4	ND
S-30	2,784	1,716	(7%)	(7%) (6%)		ND
Min	784	739	30	30	9	10
Max	2,784	2,570	245	698	25	38

Table 3. Humic substances concentrations in 0.22- μ m filtered samples, and corresponding Fe binding capacity of humic calculated according to Laglera and van den Berg (2009). % dFe (0.22 μ m)-HS and % sFe-HS indicate the maximum contribution that Fe-humic complexes can make to the dissolved Fe and soluble Fe pool, respectively. dFe: dissolved Fe; sFe: soluble Fe; HS: Humic substances; SRFA: Suwannee River Fulvic Acid; ND: not determined.

	HS (μ g SRFA L ⁻¹) (< 0.22 μ m)		(a)Fe bo	ound by	%dFe	e (0.22	0/ -E- 115(b)		
Sample			HS (nmol L ⁻¹)		μm)-HS		%sFe-HS ^(b)		
	Surface	Bottom	Surface	Bottom	Surface	Bottom	Surface	Bottom	
Nov 2015									
C-10	219	ND	4	ND	2	ND	24	ND	
N-20	247	44	4	1	2	1	43	2	
Feb 2016									
S-20	ND	129	ND	2		3	ND	ND	
C-30	15	241	0.2	4	1	7	0.2	11	
N-25	ND	229	ND	4		13	ND	8	
Aug 2018									
N-10	350	196	6	3	2	4	15	31	
C-10	328	91	6	2	4	2	34	5	
S-10	ND	79	ND	1	ND	1	ND	2	
N-20	127	65	2	1	2	0.2	8	3	
C-20	56	68	1	1	2	2	1	7	

Max	350	241	6	4	4	13	43	31
Min	15	44	0.2	1	0.4	0.2	0.2	2
S-30	47	ND	1	ND	0.4	ND	8	ND
C-30	189	99	3	2	2	1	4	16
N-30	103	141	2	2	1	1	10	8
S-20	257	181	4	3	4	2	6	13

^{941 &}lt;sup>-(a)</sup>Normalized with 16.7 nmol Fe (mg FA)⁻¹ according Laglera and van den Berg (2009).

^{942 (}b)Assuming levels of humic substances measured in dissolved Fe ($< 0.22 \mu m$).