

STUDY ON ANTIPROLIFERATIVE
ACTIVITY OF *Hypsizygus tessellatus* AND
Flammulina velutipes EXTRACTS ADSORBED
ON SULPHATED ZIRCONIA
NANOPARTICLES AGAINST BREAST
CANCER CELL LINES

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C.I. Ukaegbu.
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ABSTRAK

Dalam kajian ini, aktiviti ‘*in vitro*’ biologi air, metanol, aseton, dan ekstrak etil asetat dari dua jenis *Hypsizygus tessellatus* (Buna shimeji coklat dan Bunapi shimeji putih) dan *Flammulina velutipes* (Enoki) telah dikaji. Topi dan batang cendawan telah diekstrak dengan pelarut yang berbeza dan telah dicirikan secara kimia dari segi jumlah kandungan fenol (TPC), jumlah kandungan flavonoid (TFC), aktiviti antioksidan (terhadap DPPH dan FRAP) dan pengklasifikasian fitokimia menggunakan UPLC-QTOF/MS (berdasarkan aktiviti antioksidan). Kemudian, “multiple correspondence analysis” (MCA) telah dilakukan ke atas fitokimia yang diekstrak menggunakan pelarut daripada cendawan. Kemudian, aktiviti antiproliferatif ke atas bahagian berpotensi dijalankan terhadap dua garisan sel kanser payudara (MCF-7 dan MDA-MB-231) dengan menggunakan kaedah MTT. Siasatan lanjut telah dijalankan ke atas aktiviti antiproliferatif bagi ekstrak cendawan yang dijerap pada nanopartikel zirconia sulfat (SZN). Keputusan kajian ini menunjukkan topi cendawan ekstrak mempunyai lebih TPC dan TFC berbanding ekstrak stem. Ekstrak topi Enoki mengandungi TPC dan TFC yang lebih tinggi berbanding ekstrak topi Buna shimeji dan Bunapi shimeji. Susunan melalui pemerhatian ke atas cendawan mengikut aktiviti antioksidan daripada ekstrak adalah: Enoki > Bunapi shimeji > Buna shimeji, dan untuk pelarut: air > methanol > aseton > etil asetat. TPC bagi ekstrak menunjukkan hubungan positif dengan aktiviti antioksidan keseluruhan analisis MCA itu menunjukkan hubungan yang positif antara pelarut pengekstrakan dan fitokimia yang diekstrak daripada cendawan. Kajian antiproliferatif daripada ekstrak menggunakan dua garisan sel kanser (MCF-7 dan MDA-MB-231) menunjukkan garisan sel kanser mempunyai tahap sensitiviti yang tinggi ke atas ekstrak air dan metanol Enoki dan topi Bunapi shimeji. Antara kesemua ekstrak, ekstrak topi Enoki dengan air menunjukkan aktiviti antiproliferatif yang tertinggi terhadap MCF-7 (nilai IC_{50} antara 14.42 - 24.84 $\mu\text{g/mL}$) dan MDA-MB-231 (nilai IC_{50} antara 151.57 - 227.99 $\mu\text{g/mL}$) selepas 72 h. SZN juga menunjukkan tahap yang agak aktiviti antiproliferatif yang tinggi terhadap bahagian-bahagian sel kanser pada kepekatan yang dikaji manakala sel-sel Normal (normal) kurang sensitif kepada (julat nilai IC_{50} daripada 130.7 - 134.1 $\mu\text{g/mL}$) SZN berbanding MCF-7 sel (IC_{50} julat nilai 36.5 - 37.0 $\mu\text{g/mL}$) dan MDA-MB-231 sel (IC_{50} nilai julat 68.9 - 70.9 $\mu\text{g/mL}$). Secara sinergi, penjerapan ekstrak pada SZN menambah baik aktiviti antiproliferatif mereka terhadap garisan sel kanser ($p < 0.05$), akan tetapi ekstrak topi Enoki dengan air menunjukkan aktiviti antiproliferatif yang terbaik selepas penjerapan pada SZN. Secara keseluruhan, ekstrak topi Enoki dan Buna shimeji dengan air dan metanol amat ketara menghalang percambahan ‘*in vitro*’ MCF-7 dan MDA-MB 231, dan ia boleh dikatakan aktiviti antiproliferatif mereka terhadap garisan sel kanser telah dipertingkatkan melalui penjerapan fitokimia pada nanopartikel zirconia sulfat.

ABSTRACT

In this study, the *in vitro* biological activities of water, methanol, acetone, and ethyl acetate extracts of two *Hypsizygus tessellatus* variants (brown Buna shimeji and white Bunapi shimeji) and *Flammulina velutipes* (Enoki) were investigated. The caps and stems of the mushrooms were extracted with different solvents and chemically characterized for total phenolic content (TPC), total flavonoids content (TFC), antioxidant activities (against DPPH and FRAP). Furthermore, the extracted phytochemicals from the mushrooms were identified using Ultra performance liquid chromatography quadrupole time of flight mass spectrometer (UPLC-QTOF/MS). A multiple correspondence analysis (MCA) was performed on the extracted phytochemicals from the mushrooms based on the solvents used during the extraction process. Then, the antiproliferative activity of the potent fractions were evaluated against two breast cancer cell lines (MCF-7 and MDA-MB-231) using MTT assay. Further investigations were carried out on the antiproliferative activity of the mushroom extracts adsorbed on sulphated zirconia nanoparticles (SZN). The results of this study showed the mushroom caps extracts to have more TPC and TFC compared to the stem extracts. Enoki cap extracts contained higher TPC and TFC compared to Buna shimeji and Bunapi shimeji cap extracts. The mushroom order of the observed antioxidant activity of the extracts was: Enoki > Bunapi shimeji > Buna shimeji, and in the solvent order: water > methanol > acetone > ethyl acetate. The TPC of the extracts showed a positive correlation with their antioxidant activities whole the MCA analysis showed a positive correlation between the extraction solvents and the extracted phytochemicals from the mushrooms. The antiproliferative study of the extracts using two cancer cell lines (MCF-7 and MDA-MB-231) showed a considerable level of sensitivity of the cell lines to water and methanol extracts of Enoki and Bunapi shimeji caps. Among the extracts, Enoki caps water extract showed the highest antiproliferative activity against MCF-7 (IC₅₀ value ranged from 14.42–24.84 µg/mL) and MDA-MB-231 (IC₅₀ value ranged from 151.57 – 227.99 µg/mL) after 72 h. SZN also showed a considerable level of antiproliferative activity against the cancer cell lines at the studied concentrations while the Normal (normal) cells were less sensitive to SZN (IC₅₀ value range of 130.7 – 134.1 µg/mL) compared to MCF-7 cells (IC₅₀ value range of 36.5 – 37.0 µg/mL) and MDA-MB-231 cells (IC₅₀ value range of 68.9 – 70.9 µg/mL). The adsorption of the extracts on SZN synergistically improved their antiproliferative activities against the cancer cell lines ($p < 0.05$), but Enoki caps water extract showed the best antiproliferative activity after adsorption on SZN. Conclusively, water and methanol extracts of Enoki and Bunapi shimeji caps significantly inhibited the *in vitro* proliferation of MCF-7 and MDA-MB 231, and it can be suggested that their antiproliferative activity against the cancer cell lines was enhanced through adsorption of the phytochemicals on the sulphated zirconia nanoparticles.

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LIST OF SYMBOLS

%	Percentage
IC ₅₀	50 % inhibitory concentration
µg	Microgram
<	Less than
>	Greater than
±	Plus/minus
*	Multiplication sign

LIST OF ABBREVIATIONS

ATCC	American Type Culture Collection
BC	Buna shimeji caps
BCA	Buna shimeji caps (acetone fraction)
BCEA	Buna shimeji caps (ethyl acetate fraction)
BCM	Buna shimeji caps (methanol fraction)
BCW	Buna shimeji caps (water fraction)
BHA	Beta-hydroxyl acid
BHT	Butylated hydroxytoluene
BPC	Bunapi shimeji caps
BPCA	Bunapi shimeji caps (acetone fraction)
BPCEA	Bunapi shimeji caps (ethyl acetate fraction)
BPCM	Bunapi shimeji caps (methanol fraction)
BPCW	Bunapi shimeji caps (water fraction)
BPSA	Bunapi shimeji stem (acetone)
BPC	Bunapi shimeji caps
BS	Buna shimeji stem
BPS	Bunapi shimeji caps
BPSEA	Bunapi shimeji stem (ethyl acetate)
BPSM	Bunapi shimeji stem (methanol)
BSA	Buna shimeji stem (acetone fraction)
BSEA	Buna shimeji stem (ethyl acetate)
BSM	Buna shimeji stem (methanol)
BSW	Buna shimeji stem (water)
CCS	Collision cross section
CNPs	Cerium nanoparticles
CO ₂	Carbon dioxide
DA	Diode array
DHT	Dihydrotestosterone
DMEM	Dulbecco's Modified Eagles Medium
DMSO	Dimethylsulfoxide
DNA	Deoxyribonucleic acid

DPPH	1, 1-diphenyl-2-picryl-hydrazil
EC	Enoki cap
ECA	Enoki cap (acetone fraction)
ECEA	Enoki cap (ethyl acetate fraction)
ECM	Enoki cap (methanol fraction)
ECW	Enoki cap (water fraction)
Eq	Equation
ES	Enoki stem
ESA	Enoki stem (acetone fraction)
ESEA	Enoki stem (ethyl acetate fraction)
ESM	Enoki stem (methanol fraction)
ESW	Enoki stem (water fraction)
FBS	Fetal bovine serum
FDA	Food and Drug Administration
FRAP	Ferric reducing antioxidant power
FTIR	Fourier Transform Infrared
GAE	Gallic acid equivalent
h	Hour
H ₂ O ₂	Hydrogen peroxide
LN	Lipid nanoparticle
MBC	Minimum bactericidal concentration
MCA	Multiple correspondence analysis
mg/g	Milligram per gram
MIC	Minimum inhibitory concentration
MRSA	Multi drug resistant Staphylococcus aureus
MTT	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium Bromide
NIST	National Institute of Standards and Technology
NK	Natural killer
NPs	Nanoparticles
OD	Optical density
PCA	Principal component analysis
PNPs	Platinum nanoparticles
QE	Quercetin equivalent

RES	Reticuloendothelial system
ROS	Reactive oxygen species
SD	Standard deviation
SEM-EDS	Scanning electron microscopy-energy dispersive spectroscopy
SWCNT	Single-walled carbon nanotubes
SZN	Sulphated zirconia nanoparticles
TCA	Trichloroacetic acid
TDM	Total dry material
TEM	Transmission electron microscopy
TGA	Thermogravimetric analysis
TFC	Total flavonoids content
TNF	Tumor necrosis factor
TPC	Total phenolics content
UPLC-QTOF/MS	Ultra-performance liquid chromatography quadrupole time of flight mass spectrometer
UV	Ultraviolet

REFERENCES

- Addai, Z.R., Abdullah, A., & Mutalib, S. A. (2013). Effect of extraction solvents on the phenolic content and antioxidant properties of two papaya cultivars. *Journal of Medicinal Plants Research*, 7, 3354–3359.
- Ae Mftah, Fatah H Alhassan, Mothanna Sadiq Al-Qubaisi, Mohammed Ezzat El Zowalaty, Thomas J Webster, Mohammed Sh-eldin, Abdullah Rasedee, Yun Hin Tautiq-yap, S. Samiur R., & Sh-eldin, M. (2015). Physicochemical properties, cytotoxicity and antimicrobial activity of sulphated zirconia nanoparticles. *International Journal of Nanomedicine*, 10, 765–774.
- Alam, M. N., Bristi, N. J., & Rafiquzzaman, M. (2013). Review on *in vivo* and *in vitro* methods evaluation of antioxidant activity. *Saudi Pharmaceutical Journal*, 21, 143–152.
- Alhassan, F., Rashid, U., Al-Qubaisi, M., Rasedee, A., & Yun H, T. (2014). The effect of sulfate contents on the surface properties of iron-manganese doped sulfated zirconia catalysts. *Powder Technology*, 253, 809–813.
- Al-Qubaisi, M., Rasedee, A., & Flaifel, M. (2013). Induction of apoptosis in cancer cells by NiZn ferrite nanoparticles through mitochondrial cytochrome C release. *International Journal of Nanomedicine*, 8, 4115–4129.
- Aneta, S., Wojciech, R., & Janusz, K. (2013). Antioxidant activities and phenolic content of *Flammulina velutipes* mushroom extracts. *Herba Polonica*, 59(3), 26–36.
- Ashokkumar, R., & Ramaswamy, M. (2014). Phytochemical screening by FTIR spectroscopic analysis of leaves extracts of selected Indian medicinal plants. *International Journal of Current Microbiology and Applied Sciences*, 3(1), 395–406.
- Aurelia, M., & Gheorghe, N. (2011). Methods for total antioxidant activity determination: A review. *Biochemistry and Analytical Biochemistry*, 1(1), DOI: 10.4172/2161–1009.1000106.
- Bailey-Downs, L., Thorpe, J., Disch, B., Bastian, A., Hauser, P., & Farasyn, T. (2014). Development and characterization of a preclinical model of breast cancer lung micrometastatic to macrometastatic progression. *PLoS One*, 9(5), e98624.
- Bao, H. N., Ushio, H., & Ohshima, T. (2008). Antioxidative activity and antidiscoloration efficacy of ergothioneine in mushroom (*Flammulina velutipes*)

- extract added to beef and fish meats. *Journal of Agricultural and Food Chemistry*, 56(21), 10032–40.
- Bao, H., Ushio, H., & Ohshima, T. (2009). Antioxidative activities of mushroom (*Flammulina velutipes*) extract added to bigeye tuna meat: dose-dependent efficacy and comparison with other biological antioxidants. *Journal of Food Science*, 74(2), C162–9.
- Bao-Hua, W., & Jing-Ping, O. (2005). Pharmacological actions of sodium ferulate in cardiovascular system. *Cardiovascular Drug Reviews*, 23(2), 161–172.
- Barros, L., Cruz, T., Baptista, P., Estevinho, L. M., & Ferreira, I. C. F. R. (2008). Wild and commercial mushrooms as source of nutrients and nutraceuticals. *Food and Chemical Toxicology*, 46(8), 2742–2747.
- Batterbury, M., Tebbs, C. A., Rhodes, J. M., & Grierson, I. (2002). *Agaricus bisporus* (edible mushroom lectin) inhibits ocular fibroblast proliferation and collagen lattice contraction. *Experimental Eye Research*, 74, 361–370.
- Bazzaz, B. S. F., Khayat, M. H., Emamic, S. A., Asili, J., Sahebkhara, A., & Neishabory, E. J. (2011). Antioxidant and antimicrobial activity of methanol, dichloromethane, and ethyl acetate extracts of *Scutellaria litwinowii*. *Science Asia*, 7(3), 327–334.
- Belton, V. & Stewart, T. (2002). *Multiple criteria decision analysis*. Boston, Kluwer Academic Publishers.
- Benzécri, J. P. (1979). Sur le calcul des taux d'inertie dans l'analyse d'un questionnaire. *Cahiers de l'Analyse Des Données*, 4, 377–378.
- Ben-Zion, Z., Solomon, P. W., & Eviatar, N. J. M. (2008). *Coprinus comatus* and *Ganoderma lucidum* interfere with androgen receptor function in LNCaP prostate cancer cells. *Molecular Biology Reports*, 35.(2), 107–117.
- Bhattacharyya, S., Kudgus, R., Bhattacharya, R., & Mukherjee, P. (2011). Inorganic nanoparticles in cancer therapy. *Pharmaceutical Research*, 28, 237–259.
- Bi, M., Li, H., Pan, W., Lloyd, W., & Davis, B. (1996). Thermal studies of metal promoted sulfated zirconia. *American Chemical Society Fuel Division Preprints*, 41, 77–81.
- Birben, E., Murat, U., Md, S., Sackesen, C., Erzurum, S., & Kalayci, O. (2015). Oxidative stress and antioxidant defense. *WAO Journal*, 5, 9–19.

- Biswal, B., Sulaiman, S. A., Ismail, H.C., & Zakaria, H. M. K. (2013). Effect of *Withania somnifera* (Ashwagandha) on the development of chemotherapy-induced fatigue and quality of life in breast cancer patients. *Integraed Cancer Therapy*, 12(4), 312–22.
- Bourbon, A. I., Cerqueira, M. A., & Vicente, A. A. (2016). Encapsulation and controlled release of bioactive compounds in lactoferrin-glycomacropeptide nanohydrogels: Curcumin and caffeine as model compounds. *Journal of Food Engineering*, 180, 110–119.
- Brown, & Hargreaves. (1999). Sulfated metal oxide catalysts. *Green Chemistry*, 1(1), 17–20.
- Butt, H., Cappella, B., & Kappl, M. (2005). Force measurements with the atomic force microscope: Technique, interpretation and applications. *Surface Science Reports*, 59(1–2), 1–152.
- Cásia, C., Thacyana, T., Miriam, S., Hohmann, F., Pinho, R., Victor, F., & Al., E. (2015). Vanillic acid inhibits inflammatory pain by inhibiting neutrophil recruitment, oxidative stress, cytokine production, and nfkb activation in micevanillic. *Journal of Natural Products*, 78(8), 1799–1808.
- Cha, J., Piao M. J., Kim K. C., Yao C. W., Zheng J., Kim S. M., Hyun C. L., & Ahn Y. S. (2014). The polyphenol chlorogenic acid attenuates UVB-mediated oxidative stress in human HaCaT keratinocytes. *Biomolecules and Therapeutics*, 22: 136–142.
- Chandel, N. S., & Tuveson, D. A. (2014). The promise and perils of antioxidants for cancer patients. *New England Journal of Medicine*, 371(2), 177–178.
- Chang, H.-H., Hsieh, K.-Y., Yeh, C.-H., Tu, Y.-P., & Fuu., S. (2010). Oral administration of an Enoki mushroom protein FVE activates innate and adaptive immunity and induces anti-tumor activity against murine hepatocellular carcinoma. *International Immunopharmacology*, 10, 239–246.
- Chavez, K., Garimella, S., & Lipkowitz, S. (2010). Triple negative breast cancer cell lines: One tool in the search for better treatment of triple negative breast cancer. *Breast Diseases*, 32(1-2), 35–48.
- Cheeseman, K. & Slater, T. (1993). An introduction to free radical biochemistry. *British Medical Bulletin*, 49, 481–493.
- Chen, X., Wang, W., Li, S., Xue, J., Fan, L., Sheng, Z., & Chen, Y. (2010).

- Optimization of ultrasound-assisted extraction of Lingzhi polysaccharides using response surface methodology and its inhibitory effect on cervical cancer cells. *Carbohydrate Polymers*, 80, 944–948.
- Choi, D. B., Cha, W. S., Kang, S. H., & Lee, B. R. (2004). Effect of *Pleurotus ferulae* extracts on viability of human lung cancer and cervical cancer cell lines. *Biotechnology and Bioprocess Engineering*, 9, 356–361.
- Circu, M. & Aw, T. Y. (2010). Reactive oxygen species, cellular redox systems and apoptosis. *Free Radical Biology and Medicine*, 48(6), 749–762.
- Clark, J. (2002). Solid acids for green chemistry. *Accounts of Chemical Research*, 35(9), 791–797.
- Clearfield, A., Serrette, G., & Khazi-Syed, A. (1994). Nature of hydrous zirconia and sulfated hydrous zirconia. *Catalysts Today*, 20(2), 295–312.
- Clogston, J. & Patri, A. (2011). Zeta potential measurement. *Methods in Molecular Biology*, 697, 63–70.
- Comşa, Ş., Ciuculescu, F., & Raica, M. (2012). Mesenchymal stem cell tumor cell cooperation in breast cancer vasculogenesis. *Molecular Medicine Reports*, 5, 1175–1180.
- Couvreur, P., Kante, B., Grislain, L., Roland, M., & Speiser, P. (1982). Toxicity of polyalkylcyanoacrylate nanoparticles II: Doxorubicin-loaded nanoparticles. *Journal of Pharmaceutical Science*, 71, 790–792.
- Cushnie, T., Cushnie, B., & Lamb, A. (2014). Alkaloids: An overview of their antibacterial, antibiotic-enhancing and antivirulence activities. *International Journal of Antimicrobial Agents*, 44(5), 377–386.
- Danladi, S., Wan-Azemin, A., Sani, Y. N., Mohd, K. S., Us, M. R., Mansor, S. M., & Dharmaraj, S. (2015). Phytochemical screening, antioxidant potential and cytotoxic activity of *Melastoma malabathricum* Linn. from different locations. *International Journal of Pharmacy and Pharmaceutical Sciences*, 7(7), 408–413.
- Dar, A. N., Savita, B. N., & Gulzar, S. (2016). Effect of storage period on physiochemical, total phenolic content and antioxidant properties of bran enriched snacks. *Journal of Food Measurement and Characterization*, 10, 755–761.
- Davies, C., Godwin, J., Gray, R., Clarke, M., & Cutter, D. (2011). Relevance of breast

- cancer hormone receptors and other factors to the efficacy of adjuvant tamoxifen: patient-level meta-analysis of randomized trials. *Lancet*, 378(9793), 771–84.
- Deborah, L. & Valerie, S. (2011). Choosing the right cell line for breast cancer research. *Breast Cancer Research*, 13, 215.
- Delahay, G., Ensuque, E., & Coq, B. F. F. (1998). Selective catalytic reduction of nitric oxide by n-decane on Cu/sulfated-zirconia catalysts in oxygen rich atmosphere: Effect of sulfur and copper contents. *Journal of Catalysis*, 175(1), 7–15.
- Demirel, Z., Yilmaz-Koz, F.F., Karabay-Yavasoglu, N.U., & Sukatar, A. (2011). Antimicrobial and antioxidant activities of solvent extracts and the essential oil composition of *Laurencia obtusa* and *Laurencia obtusa* var. *pyramidata*. *Romanian Biotechnological Letters*, 16, 5927–5936.
- Deshpande, R., Bhat, R., Ganachari, S., Bedre, M., Vasanth, H., & Manjunath, S. Y. (2011). Anti-cancer studies of noble metal nanoparticles synthesized using different plant extracts. *Cancer Nanotechnology*, 2(1-6), 57–65.
- Deters, M., Knochenwefel, H., Lindhorst, D., & Al., E. (2008). Different curcuminoids inhibit T-lymphocyte proliferation independently of their radical scavenging activities. *Pharmaceutical Research*, 25(8), 1822–1827.
- Devasagayam, T. P., Tilak, J., Bloor, K., Sane, K. S., Ghaskadbi, S. S., & Lele, R. (2004). Free radicals and antioxidants in human health: Current status and future prospects. *The Journal of the Association of Physicians of India*, 52, 794–804.
- Dilek, Y., Abel, U., Tulay, B., Aydin, A., & Ilkay, A. (2012). Fourier transform infrared (FTIR) spectroscopy for identification of *Chorella vulgaris* Beijerinck and *Scenedesmus obliquus* (Turpin) Kutzing 1833. *African Journal of Biotechnology*, 11(16), 3817–3824.
- Dilsad, O., Gokcen, Y. C., & Hikmet, K. I. N. (2015). Antimicrobial, antioxidant activities and chemical composition of *Lactarius deliciosus* (L.) collected from Kastamonu Province of Turkey. *Kastamonu University Journal of Forestry Faculty*, 15(1), 98–103.
- Ding, X., Hou, Y., & Hou, W. (2012). Structure feature and antitumor activity of a novel polysaccharide isolated from *Lactarius deliciosus* Gray. *Carbohydrate Polymers*, 89, 397–402.
- Dittmer, A., Hohlfeld, K., Lützkendorf, J., Müller, L., & Dittmer, J. (2009). Human mesenchymal stem cells induce E-cadherin degradation in breast carcinoma

- spheroids by activating ADAM10. *Cellular and Molecular Life Science*, 66, 3053–3065.
- Dorman, H.J., Bachmayer, O., Kosar, M., & Hiltunen, R. (2004). Antioxidant properties of aqueous extracts from selected Lamiaceae species grown in Turkey. *Journal of Agricultural Food Chemistry*, 52, 762–770.
- Dubost, N., Ou, B., & Beelman, R. (2007). Quantification of polyphenols and ergothioneine in cultivated mushrooms and correlation to total antioxidant capacity. *Food Chemistry*, 105, 727–735.
- Dulger, B., Fadime, Y., & Guçin, F. (2002). Antimicrobial activity of some *Lactarius* species. *Pharmaceutical Biology*, 40(4), 304–306.
- Eckhardt, S. G., Baker, S. D., Britten, C. D., Hidalgo, M., Siu, L., Hammond, L. A., Villalona-Calero, M. A., Felton, S., D., R., Kuhn, J. G., Clark, G. M., Smith, S. L., Macdonald, J. R., S., C., Moczygemba, J., Weitman, S., von Hoff, D. D., Rowinsky, E. K., & de la Rosa, L. (2000). Phase I and pharmacokinetic study of ifofulven, a novel mushroom-derived cytotoxin, administered for five consecutive days every four weeks in patients with advanced solid malignancies. *Journal of Clinical Oncology*, 18, 4086–4097.
- Ehsan, K., Oskoueian, E., Rudi, H., Armin, O., & Hawa, Z. (2012). Phenolic compounds characterization and biological activities of *Citrus aurantium* Bloom. *Molecules*, 17, 1203–1218.
- El Saghir, N. & Anderson, B. (2012). Breast cancer early detection and resources: Where in the world do we start? *Breast*, 21(4), 423–5.
- Elster, N., Collins, D., Toomey, S., Crown, J., Eustace, A., & Hennessy, B. (2015). HER2-family signalling mechanisms, clinical implications and targeting in breast cancer. *Breast Cancer Research and Treatment*, 149(1), 5–15.
- Endo, M., Beppu, H., Akiyama, H., Wakamatsu, K., Ito, S., & Kawamoto, Y., Shimpo, K., Sumiya, T., Koike, T., & Matsui, T. (2010). Agaritine purified from *Agaricus blazei* Murrill exerts anti-tumor activity against leukemic cells. *Biochimica et Biophysica Acta*, 1800, 669–673.
- Escofier, B. (1978). Analyse factorielle et distances répondant au principe d'équivalence distributionnelle. *Revue de Statistiques Appliquées*, 26, 29–37.
- Fadeel, B., Feliu, N., Vogt, C., Abdelmonem, A., & PJ, W. (2013). Bridge over troubled waters: Understanding the synthetic and biological identities of engineered

- nanomaterials. *WIREs Nanomed Nanobiotechnology* 2013, 5, 111–129.
- Fang, Y. Z., Yang, S., & Wu, G. Y. (2002). Free radicals, antioxidants, and nutrition. *Nutrition Journal*, 18, 872–879.
- Farrera, C. & Fadeel, B. (2015). It takes two to tango□: Understanding the interactions between engineered nanomaterials and the immune system. *European Journal of Pharmaceutics and Biopharmaceutics*, 95, 3–12.
- Feng, L., Weijun, D., & Shuge, T. (2014). Quantification of phytochemical constituents and *in-vitro* antioxidant activity of *Althaea rosea* seeds. *Journal of Chemical and Pharmaceutical Research*, 6(3), 1466–1471.
- Fisher, B., Redmond, C., Poisson, R., Margolese, R., Wolmark, N., & Wickerham, L. (1989). Eight-year results of a randomized clinical trial comparing total mastectomy and lumpectomy with or without irradiation in the treatment of breast cancer. *New England Journal of Medicine*, 320(13), 822–828.
- Forbes-hernández, T. Y., Giampieri, F., Gasparrini, M., Mazzoni, L., Quiles, J. L., Alvarez-suarez, J. M., & Battino, M. (2014). The effects of bioactive compounds from plant foods on mitochondrial function□: A focus on apoptotic mechanisms, *Food and Chemical Toxicology*, 68, 154–182.
- Fortes, R. C., Novaes, M., Recova, V. L., & Melo, A. L. (2009). Immunological, hematological, and glycemia effects of dietary supplementation with *Agaricus sylvaticus* on patients' colorectal cancer. *Experimental Biology and Medicine*, 234, 53–62.
- Friedman, R. (2011). Nano dot technology enters clinical trials. *Journal of the National Cancer Institute Monograph*, 103, 1428–1429.
- Gao, J. J., Min, B. S., Ahn, E. M., Nakamura, N., Lee, H. K., & Hattori, M. (2002). New triterpene aldehydes, lucialdehydes A–C, from *Ganoderma lucidum* and their cytotoxicity against murine and human tumor cells. *Chemical and Pharmaceutical Bulletin*, 50, 837–840.
- Gest, C., Joimel, U., Huang, L., Pritchard, L., Petit, A., Dulong, C., & Soria, C. (2013). Rac3 induces a molecular pathway triggering breast cancer cell aggressiveness: Differences in MDA-MB-231 and MCF-7 breast cancer cell lines. *BMC Cancer*, 13, 63.
- Ghasemzadeh, A., Jaafar, H.Z., & Rahmat, A. (2011). Effects of solvent type on phenolics and flavonoids content and antioxidant activities in two varieties of

- young ginger (*Zingiber officinale* Roscoe) extracts. *Journal of Medicinal Plants Research*, 5, 1147-1154.
- Gil, M., Tomas-Barberan, F., Hess-Pierce, B., & Kader, A. (2002). Antioxidant capacities, phenolic compounds, carotenoids and vitamin C contents of nectarine, peach and plum cultivars from California. *Journal of Agricultural Food Chemistry*, 50, 4976–4982.
- Gordon, M. ., & John, M. . (2016). Natural products as a vital source for the discovery of cancer chemotherapeutic and chemopreventive agents. *Medicinal Principles and Practices*, 25(suppl 2), 41–59.
- Greene, J. & Hennessy, B. (2014). The role of anthracyclines in the treatment of early breast cancer. *Journal of Oncology Pharmacy Practice*, 21(3), doi.org/10.1177/1078155214531513
- Griffiths, P. & Haset, J. (2007). *Fourier transform infrared spectrometry* (Second Edi). Hoboken, New Jersey: John Wiley & Sons, Inc.
- Grube, B. J., Eng, E. T., Kao, Y. C., Kwon, A., & Chen, S. (2001). White button mushroom phytochemicals inhibit aromatase activity and breast cancer cell proliferation. *Journal of Nutrition*, 131, 3288–3293.
- Guo, J. J., Zhu, T. B., Collins, L., Xiao, Z. X. J., Kim, S. H., & Chen, C. Y. (2007). Modulation of lung cancer growth arrest and apoptosis by *Phellinus linteus*. *Molecular Carcinogenesis*, 46, 144–154.
- Guotam, B., Dilip, G., & Rajiv, R. (2013). Argemone mexicana: chemical and pharmacological aspects. *Brazilian Journal of Pharmacognosy*, 23(3), 559–575.
- Hamid, A., Roziyahira, M., Mashitah, M., Nurul, A. A., & Ahmad, F. A. . (2016). Comparative analysis of antioxidant and antiproliferative activities of *Rhodomyrtus tomentosa* extracts prepared with various solvents. *Food and Chemical Toxicology*, 108(Part B), 451–457.
- Handa, S., Khanuja, S., Longo, G., & Rakesh, D. (2008). *Extraction technologies for medicinal and aromatic plants*. Trieste.
- Harhaji, L., Mijatovic, S., Maksimov-Ivanic, D., Stojanovic, I., Momcilovic, M., Maksimovic, V., Tufegdžic, S., Marjanovic, Z., Mostarica-Stojkovic, M., Vucinic, Z., & Stosic-Grujicic, S. (2008). Anti-tumor effect of *Coriulus versicolor* methanol extract against mouse B16 melanoma cells: *In vitro* and *in vivo* study. *Food and Chemical Toxicology*, 46, 1825–1833.

- Hirofumi, M. & Ikekawa, T. (2007). Immunomodulation and antitumor activity of a mushroom product, proflamin, isolated from *Flammulina velutipes* (W. Curt.: Fr.) Singer *Agaricomycetidae*. *International Journal of Medicinal Mushrooms*, 9(2), 109–122.
- Hodzic, Z., Pasalic, H., Memisevic, A., Scrabovic, M., Saletovic, M., & Poljakovic, M. (2009). The influence of total phenols content on antioxidant capacity in the whole grain extracts. *European Journal of Scientific Research*, 28, 471–477.
- Hong, K. J., Dunn, D. M., Shen, C. L., & Pence, B. C. (2004). Effects of *Ganoderma lucidum* on apoptotic and anti-inflammatory function in HT-29 human colonic carcinoma cells. *Phytotherapy Research*, 18, 768–770.
- Hyeon, H., Nan, L., & Byung, K. (2018). Dichloromethane extracts of *Geranium koreanum* Kom. alleviates esophagus damage in acute reflux esophagitis-induced rats by anti-inflammatory activities. *International Journal of Molecular Sciences*, 19(11), 3622.
- International Agency for Research on Cancer. (2019). *Incidence, mortality, and prevalence of cancer in Malaysia. Global cancer observatory (GLOBOCAN) report 2018*.
- Isabel, C. F, Lillian B., & Rui, M. V. (2004). Antioxidants in wild mushrooms. Instituto Politécnico de Bragança, Campus de Sta. *Journal of Agriculture and Food Chemistry*, 23, 1894–2845.
- Itoh, H., Ito, H., & Hibasami, H. (2008). Blazein of a new steroid isolated from *Agaricus blazei Murrill* (himematsutake) induces cell death and morphological change indicative of apoptotic chromatin condensation in human lung cancer LU99 and stomach cancer KATO III cells. *Oncology Reports*, 20, 1359–1361.
- Jake, L., Martin, K., & Naomi, A. (2017). Principal component analysis. *Nature Methods*, 14, 641–642.
- Jasril, L., Mooi, A., Ali, M., Sukari, A., Rahman, A., Othman, H., & Nakatani, K. (2003). Antioxidant and antitumor promoting activities of the flavonoids from *Hedychium thyrsoideum*. *Pharmaceutical Biology*, 41(7), 506–511.
- Jayaprakasha, G. ., Girenavar, B., & Patil, B. . (2008). Radical scavenging activities of Rio Red grapefruits and sour orange fruit extracts in different *in vitro* model systems. *Bioresource Technology*, 99, 4484–4494.
- Jentoft, F., Hahn, A., & Kröhnert, J. (2004). Incorporation of manganese and iron into

- the zirconia lattice in promoted sulfated zirconia catalysts. *Journal of Catalysis*, 224(1), 124–137.
- Jesus, J., Luis, S., Hugo, L., & Manuel, J. (2018). Quercetagenin and patuletin: Antiproliferative, necrotic and apoptotic activity in tumor cell lines. *Molecules*, 23(10), 2579.
- Jia, C., Jin-Hong, L., & Ji-Chang, X. (2018). Dehydroxylation of alcohols for nucleophilic substitution. *Chemical Communication*, 54, 7034–7037.
- Jiang, J. H., Grieb, B., Thyagarajan, A., & Sliva, D. (2008). Ganoderic acids suppress growth and invasive behavior of breast cancer cells by modulating Ap-1 and Nf-Kappa B signaling. *International Journal of Molecular Medicine*, 21, 577–584.
- Jin, G., Ying, F., Zhao, H., Xian, L., & Hong, Z. (2017). Correlations between antioxidant activity and alkaloids and phenols of Maca (*Lepidium meyenii*). *Journal of Food Quality*, 2017, doi.org/10.1155/2017/3185945.
- Jung, K., Kim, K., Yoon, D., Hong, Y., & Choi, C. (2012). A phase I trial to determine the maximum tolerated dose and evaluate the safety and pharmacokinetics (PK) of docetaxel-PNP, polymeric nanoparticle formulation of docetaxel, in subjects with advanced solid malignancies. In *ASCO Annual Meeting Proceedings*.
- Kalita, H., Prashanth, K., Konar, S., Tantubay, S., Mahto, M., Mandal, M., & Pathak, A. (2016). Sonochemically synthesized biocompatible zirconium phosphate nanoparticles for pH sensitive drug delivery application. *Materials Science and Engineering C*, 60, 84–91.
- Kato, K., Chin, K., Yoshikawa, T., Yamaguchi, K., & Tsuji, Y. (2012). Phase II study of NK105, a paclitaxel-incorporating micellar nanoparticle, for previously treated advanced or recurrent gastric cancer. *Investigational New Drugs*, 30, 1621–1627.
- Kay, M. (2010). Synthesis of naturally occurring (-)-1,3,5-tri-O-caffeoyquinic acid. *NIH Public Access*, 11(9), 1517–1519.
- Kenny, P., Lee, G., Myers, C., Neve, R., Semeiks, J., Spellman, P., & Bissell, M. (2007). The morphologies of breast cancer cell lines in three-dimensional assays correlate with their profiles of gene expression. *Molecular Oncology*, 1(1), 84–96.
- Khalaf, H. (2009). Textural properties of sulfated iron hydroxide promoted with aluminum. *Chemical Monthly*, 140(6), 669–674.

- Khandelwal, K. (2008). *Practical Pharmacognosy: Techniques and experiments*. (19th ed.). Pune: Nirali Prakashan.
- Kim, D., Han, K., Song, K., Lee, K., Jo, S., Lee, S., & Yoon, T. (2010). Activation of innate immunity by *Lepiota procera* enhances antitumor activity. *Korean Journal of Pharmacognosy*, *41*, 115–21.
- Kim, M., Seguin, P., Ahn, J., Kim, J., Chun, S., Kim, E., & Chung, I. (2008). Phenolic compound concentration and antioxidant activities of edible and medicinal mushrooms from Korea. *Journal of Agricultural and Food Chemistry*, *56*, 7265–7270.
- Ko, E., Kim, D., Roh, S. W., Yoon, W., Jeon, Y., Ahn, G., & Technopark, J. (2015). Evaluation on antioxidant properties of sixteen plant species from Jeju island in Korea. *EXCLI Journal*, *14*, 133–145.
- Kodama, N., Komuta, K., & Nanba, H. (2003). Activation of NK cells in cancer patients, *6*(4), 371–377.
- Kolhatkar R, & Lote A, K. H. (2011). Active tumor targeting of nanomaterials using folic acid, transferrin and integrin receptors. *Current Drug Discovery Technology*, *8*(3), 197–206.
- Konstantinos, D., Costas, D., Dimitra, A., & Thomas, M. (2000). Biological activity of myricetin and its derivatives against human leukemic cell lines *in vitro*. *Pharmacological Research*, *42*(5), 475–8.
- Kumar, P., Kulkarni, P., & Srivastava, A. (2015). Pharmaceutical application of nanoparticles in drug delivery system. *Journal of Chemical and Pharmaceutical Research*, *7*(8), 703–712.
- Kumar, S., Singh, A., & Kumar, B. (2017). Identification and characterization of phenolics and terpenoids from ethanolic extracts of *Phyllanthus species* by HPLC-ESI-QTOF-MS/MS. *Journal of Pharmaceutical Analysis*, *7*(4), 214–222.
- Kumari, S. & Chang, S. K. C. (2016). Effect of cooking on isoflavones, phenolic acids, and antioxidant activity in sprouts of prosoy soybean (*Glycine max*). *Journal of Food Science*, *81*(7), C1679–C1691.
- Labhassetwar, V. & Song, C. (1997). Nanoparticle drug delivery systems. *Advanced Drug Delivery Review*, *24*, 63–85.

- Lam, Y., Ng, T., & HX, W. (2001). Antiproliferative and antimutagenic activities in a peptide from puffball mushroom *Calvatia caelata*. *Biochemical and Biophysical Research Communications*, 289, 744–749.
- Lee, M. ., Shin, T., Utsuki, T., Choi, J., Byun, D., & Kim, H. (2012). Isolation and identification of phlorotannins from *Ecklonia stolonifera* with antioxidant and hepatoprotective properties in tacrine-treated HepG2 cells. *Journal of Agricultural Food Chemistry*, 60(21), 5340–5449.
- Leong, B., Chuah J. A, Kumar, V. M., Rohamini, S., & Siti, Z. S. (2009). Trends of breast cancer treatment in Sabah, Malaysia: a problem with lack of awareness. *Singapore Medicinal Journal*, 50(8), 772–6.
- Li, Y. & Somorjai, G. (2010). Nanoscale advances in catalysis and energy applications. *Nano Letters*, 10(7), 2289–2295.
- Liu, K., Xiao, X., Wang, J., Chen, C. O., & Hu, H. (2017). Polyphenolic composition and antioxidant, antiproliferative, and antimicrobial activities of mushroom *Inonotus sanghuang*. *LWT - Food Science and Technology*, 82, 154–161.
- Liu, M., Wangjinsong, Y., Yongfa, Z., Hui, L., Jianjun, Z., & Le, J. (2018). Characterization, antioxidant and antiinflammation of mycelia selenium polysaccharides from *Hypsizygus marmoreus* SK-03. *Carbohydrate Polymers*, 201, 566–574.
- Mahmood, A., Suzita, N., Vikineswary, S., Noorlidah, A., Wong, K. & Hapipah Mohd, A. (2008). Effect of culinary-medicinal lion's mane mushroom, *Hericium erinaceus* (Bull.: Fr.) Pers.(Aphyllphoromycetidae), on ethanol-induced gastric ulcers in rats. *International Journal of Medicinal Mushrooms*, 10, 325–330.
- Marijana, K., Branislav, R., & Stanojkovic, T. (2016). Evaluation of metal concentration and antioxidant, antimicrobial, and anticancer potentials of two edible mushrooms *Lactarius deliciosus* and *Macrolepiota procera*. *Journal of Food and Drug Analysis*, 24, 477 – 484.
- Masella, R., Santangelo, C., D'Archivio, M., Li Volti, G., Giovannini, C., & Galvano, F. (2012). Protocatechuic acid and human disease prevention: Biological activities and molecular mechanisms. *Current Medicinal Chemistry*, 19(18), 2901–17.
- Maskam, M.F., Mohamad, J., Abdulla, M.A., Afzan, A., & Wasiman, I. (2014). Antioxidant activity of *Rhodomyrtus tomentosa* (Kemunting) fruits and its effect on lipid profile in induced-cholesterol New Zealand white rabbits. *Sains Malaysia*, 43, 1673–1684.

- Masuda, Y., Kikuzaki, H., Hisamoto, M., & Nakatani, N. (2004). Antioxidant properties of gingerol related compounds from ginger. *Biofactors*, *21*(1-4), 293–296.
- Matsuzawa, T. H., Saitoh, M., Tomita, I. & Ohkawa, M. I. T. (1998). Studies on antioxidant effects of *Hypsizygus marmoreus*. *Yakugaku Zasshi*, *118*, 476 – 481.
- Mau, J., Lin, H., & Chen, C. (2002). Antioxidant properties of several medicinal mushrooms. *Journal of Agricultural Food Chemistry*, *50*, 6072–6077.
- Meda, A., Lamien, C. E., Romito, M., Millogo, J., & Nacoulma, O. G. (2005). Determination of the total phenolic, flavonoid and proline contents in Burkina Fasan honey, as well as their radical scavenging activity. *Food Chemistry*, *91*, 571–577.
- Mekhemer, G., Khalaf, H., Mansour, S., & Nohman, A. (2005). Sulfated alumina catalysts: Consequences of sulfate content and source. *Monthly Chemistry*, *136*(12), 2007–2016.
- Mftah, A., Fatah, H., Mothanna, S., Mohamed, E., Thomas, J., Mohammed, S., & Rashid, S. (2015). Physicochemical properties, cytotoxicity, and antimicrobial activity of sulphated zirconia nanoparticles. *International Journal of Nanomedicine*, *10*, 765–774.
- Ming-Yei, Y., Wen-Ching, K., & Li-Yun, L. (2014). Hypolipidemic and antioxidant activity of Enoki mushrooms (*Flammulina velutipes*). *BioMedical Research International*, 2014, doi.org/10.1155/2014/352385.
- Min-Young K, Philippe S, Joung-Kuk A, J.-J. K., Se-Chul C, Eun-Hye K, Su-Hyun S, E.-Y. K., Sun-Lim K, Yool-Jin P, & Hee-Myong R. O. (2008). Phenolic compound concentration and antioxidant activities of edible and medicinal mushrooms from Korea. *Journal of Agricultural Food Chemistry*, *56*, 7265–7270.
- Mohd, F. B. A., Pin, K., Zamree, M., Luqman, C., Soh, S., & Thomas, I. C. S. (2012). The effects of varying solvent polarity on extraction yield of *Orthosiphon stamineus* leaves. *Journal of Applied Sciences*, *12*, 1207–1210.
- Molyneux P. (2004). The use of the stable free radical diphenylpicrylhydrazyl (DPPH) for estimating antioxidant activity. *Songklanakarin Journal of Science and Technology*, *26*, 211–219.
- Monira, S., Haque, A., Muhit, A., Sarker, N. C., Alam, A. H. M. K., & Rahman, A.A. (2012). Antimicrobial, antioxidant and cytotoxic properties of *Hypsizygus tessulatus* cultivated in Bangladesh. *Research Journal of Medicinal Plants*.

<http://doi.org/10.3923/rjmp.2012>

- Moro, C., Palacios, I., Lozano, M., Arrigo, M. D., Guillamón, E., Villares, A., & García-lafuente, A. (2012). Anti-inflammatory activity of methanolic extracts from edible mushrooms in LPS activated RAW 264.7 macrophages. *Food Chemistry*, *130*, 350–355.
- Morterra, C., Cerrato, G., & Bolis, V. (1993). Lewis and Brønsted acidity at the surface of sulfate-doped ZrO₂ catalysts. *Catalysis Today*, *17*(3), 505–515.
- Muhammad, M., Mohd, F., & Alona, C. (2018). Malaysian medicinal plants' potential for breast cancer therapy. *Asian Journal of Pharmaceutical and Clinical Research*, *11*(6), 101–117.
- Murawa, P., Muwara, D., Adamczyk, B., & Polom, K. (2014). Breast cancer: Actual methods of treatment and future trends. *Reports of Practical Oncology and Radiotherapy*, *19*(3)(3), 165–72.
- National Center for Biotechnology Information. (2018). *Cytotoxicity of (Bavacin) Against Human A549 Cells After 48 Hrs By MTT Assay*.
- Nayeem, N., Asdaq, S. M. B., Salem, H., & Ahe, A. S. (2016). Gallic Acid: A promising lead molecule for drug development. *Journal of Applied Pharmacy*, *8*(2), Doi:10.4172/1920–4159.1000213.
- Nevcihan, G., Sarikurcu, C., Tepe, B., & Solak, H. M. (2010). Evaluation of antioxidant activities of 3 edible mushrooms: *Food Science and Biotechnology*, *19*(3), 691–696.
- Ng, T., Ngai, P., & Xia, L. (2006). An agglutinin with mitogenic and antiproliferative activities from the mushroom *Flammulina velutipes*. *Mycologia*, *98*(2), 167–71.
- Niestroy, J., Barbara, A., Herbst, K., Rode, S., van Liempt, M., & Roos, P. (2011). Single and concerted effects of benzo [a] pyrene and flavonoids on the AhR and Nrf2-pathway in the human colon carcinoma cell line Caco-2. *Toxicology in Vitro*, *25*(3), 671–683.
- Noguchi, M., Kakuma, T., Tomiyasu, K., Kurita, Y., Kukihara, H., & Konishi, F., Kumamoto, S., Shimizu, K., Kondo, R., & Matsuoka, K. (2008). Effect of an extract of *Ganoderma lucidum* in men with lower urinary tract symptoms: A double-blind, placebo-controlled randomized and dose-ranging study. *Asian Journal of Andrology*, *10*, 651–658.

- Oba, K., Teramukai, S., Kobayashi, M., Matsui, T., Koderu, Y., & Sakamoto, J. (2007). Efficacy of adjuvant immunochemotherapy with polysaccharide K for patients with curative resections of gastric cancer. *Cancer Immunology Immunotherapy*, *56*, 905–911.
- Oliviera, R., Mancini, M., Cabral, F., Oliviera, S., Passos, T., & Quilty, B. (2016). FTIR analysis and quantification of phenols and flavonoids of five commercially available plant extracts used in wound healing. *Revista Matera*, *12*(3), 767–779.
- Omar, A. P., Mar ía, L. V., Laura, Á. B., Rodríguez, L., Ang élica, M., & Ver ónica, R. (2016). Cytotoxicity, post-treatment recovery, and selectivity analysis of naturally occurring podophyllotoxins from *Bursera fagaroides* var. *fagaroides* on breast cancer cell lines. *Molecules*, *21*, 1–15.
- Otsuka, H. & Nagasaki, Y. K. K. (2012). PEGylated nanoparticles for biological and pharmaceutical applications. *Advanced Drug Delivery Reviews*, *3*, 23–27.
- Oyaizu, M. (1986). Studies on products of browning reaction prepared from glucosamine. *Japanese Journal of Nutrition*, *44*, 307 – 314.
- Ozsoy, N., Can, A., Yanardag, R., & Akev, N. (2008). Antioxidant activity of *Smilax excelsa* L. leaf extracts. *Food Chemistry*, *110*(3), 571–583.
- Panche, A., Diwan, A., & Chandra, S. (2016). Flavonoids: An overview. *Journal of Nutritional Science*, *5*, e47.
- Pandey, K. B., & Rizvi, S. I. (2009). Plant polyphenols as dietary antioxidants in human health and disease. *Oxidative Medicine and Cellular Longevity*, *2*(5), 270–278.
- Papakostas, D., Fiorenza, R., & Annika, V. (2011). Nanoparticles in dermatology. *Archives of Dermatology Research*, *303*, 533–550.
- Patel, S. & Goyal, A. (2012). Recent developments in mushrooms as anti-cancer therapeutics: A review. *Biotech*, *2*, 1–15.
- Pei-Feng, T., Chia-Yu, M., & James Swi-Bea, W. (2013). A novel glycoprotein from mushroom *Hypsizygus marmoreus* (Peck) Bigelow with growth inhibitory effect against human leukaemic U937 cells. *Food Chemistry*, *141*, 1252–1258.
- Perez-Vizcaino, F., Duarte, J., & Andriantsitohaina, R. (2006). Endothelial function and cardiovascular disease: Effects of quercetin and wine polyphenols. *Free Radicals Research*, *40*(10), 1054–65.

- Perrot-Applanat, M., & Di Benedetto, M. (2012). Autocrine functions of VEGF in breast tumor cells: Adhesion, survival, migration and invasion. *Cell Adhesion and Migration*, 6, 547–553.
- Policegoudra, R. S., Chandrashekhar, R.H., Aradhya, S. M., & Singh, L. (2011). Cytotoxicity, platelet aggregation inhibitory and antioxidant activity of *Curcuma amada Roxb.* extracts. *Food Technology and Biotechnology*, 49(2), 162–168.
- Qiaoping Wang, Haiyan Li, Zhen Sun, Lihua Dong, LingGao, & Chunlan Liu, X. (2016). Kukoamine A inhibits human glioblastoma cell growth and migration through apoptosis induction and epithelialmesenchymal transition attenuation. *Scientific Reports*, DOI: 10.1038/srep36543.
- Qin, Y., Zhang, W., Lin, Y., & Yang, M. (2013). Study on chemical constituents from the fruits of *Forsythia suspensa* and the antioxidant activity of Forsythoside D. *Chinese Journal of Experimental Traditional Medical Formulae*, 11, 333–345.
- Rajeshkumar, S. (2016). Anticancer activity of eco-friendly gold nanoparticles against lung and liver cancer cells. *Journal of Genetic Engineering and Biotechnology*, 14, 195–202.
- Ribeiro, B., Valentao, P., Baptista, P., Seabra, R., & Andrade, P. (2007). Phenolic compounds organic acids profiles and antioxidative properties of beefsteak fungus (*Fistulina hepatica*). *Food and Chemical Toxicology*, 45, 1805–1813.
- Rong, X., Cai, Z., Zheng, Y., He, G., Yu, F., & Qiong, D. (2009). Reversion of multidrug resistance by co-encapsulation of vincristine and verapamil in PLGA nanoparticles. *European Journal of Pharmaceutical Science*, 37(3-4), 300–305.
- Saitoh, H., Feng, W., Matsuzawa, T., & Ikekawa, T. (1997). Antitumor activity of *Hypsizigus marmoreus*. II. Preventive effect against lung metastasis of Lewis lung carcinoma. *Yakugaku Zasshi*, 117, 1006–1010.
- Salkind, N. (2007). Multiple correspondence analysis. In *Encyclopedia of Measurement and Statistics*. Thousand Oaks (CA): Sage.
- Sancheti, S., Um, B., & Seo, S. (2010). A cholinesterase inhibitor from *Terminalia chebula*. *South African Journal of Botany*, 76(2), 285–288.
- Saxena, N., Hartman, M., Bhoo-Pathy, N., Lim, J. N., & Aw, T. C. (2012). Breast cancer in South East Asia: Comparison of presentation and outcome between a middle income and a high income country. *World Journal of Surgery*, 36(12), 2838–2846.

- Serban, C., Anca, M., & Marius, R. (2015). The story of MCF-7 breast cancer cell line: 40 years of experience in research. *International Journal of Cancer Research and Treatment*, 35(6), 3147–3154.
- Sergio, C. & Gastao, C. (2016). Alternative methods to multiple correspondence analysis in reconstructing the relevant information in a burt's table. *Pesquisa Operacional*, 36(1), 23–44.
- Shaharudin, S., Sulaiman, S., Emran, N. A., & Shahril, M. R. (2011). The use of complementary and alternative medicine among Malay breast cancer survivors. *Alternative Therapies in Health and Medicine*, 17(1), 50–6.
- Shams, K., Abdel-azim, N., Saleh, I., Hegazy, M., El-missiry, M., Hammouda, F., & Tahrir, E. (2015). Green technology: Economically and environmentally innovative methods for extraction of medicinal and aromatic plants (MAP) in Egypt. *Journal of Chemical and Pharmaceutical Research*, 7(5), 1050–1074.
- Shashank, K., & Abhay, K. (2013). Chemistry and biological activity of flavonoids: An overview. *The Scientific World Journal*, 4, 32–48.
- Siddhuraju, P. & Becker, K. (2003). Antioxidant properties of various solvent extracts of total phenolic constituents from three different agroclimatic origins of drumstick tree (*Moringa oleifera Lam.*) leaves. *Journal of Agricultural Food Chemistry*, 51, 2144–2155.
- Simplice, J., Ndendoung, T., Jean de Dieu, T., Léopold, H., Dezső, C., & Peter, F. (2012). Antimicrobial and antioxidant activity of kaempferol rhamnoside derivatives from *Bryophyllum pinnatum*. *BMC Research Notes*, 5, 158.
- Siripireddy, B., Badal, K. M., Shivendu, R., Dasgupta, N., & Chidambaram, R. (2017). Nano-zirconia – Evaluation of its antioxidant and anticancer activity. *Journal of Photochemistry & Photobiology, B: Biology*, 170, 125–133.
- Sivakumar, K. & Rajagopal, S. V. (2011). Radical scavenging activity of green algal species. *Journal of Pharmaceutical Research*, 4, 723–725.
- Slinkard, K. & Slingleton, V. L. (1997). Total phenolic analyses: automation and comparison with manual method. *American Journal of Enology and Viticulture*, 28, 49–55.
- Soo, J. C., Su, C. K., Han, R., Sung, M. J., Young, H. L., Hye, S. K., & Jong, O. K. (2013). Antioxidant and tyrosinase inhibitory activity of *Hypsizygus marmoreus* (brown cultivar) methanol extracts. *Journal of Mushrooms*, 11(4), 254–260.

- Sova, M. (2012). Antioxidant and antimicrobial activities of cinnamic acid derivatives. *Mini Reviews in Medicinal Chemistry*, 12(8), 749–767.
- Stanisław, D., Piotr, P., Marek, H., Izabela, J.-M., Paweł, K., Anna, P., & Piotr, M. (2014). Do differences in chemical composition of stem and cap of *Amanita muscaria* fruiting bodies correlate with top soil type? *PLoS One*, 9(12), e104084.
- Suzuki, K., Nishiyama, I., Amano, T., Takahashi, T., Murayama, N., Oka, H., & Ogawa, N. (1992). Randomized, controlled study on adjuvant immunochemotherapy with PSK in curatively resected colorectal cancer. *Diseases of the Colon and Rectum*, 35, 123–130.
- Sven, B. (2015). A historic and scientific review of breast cancer: The next global healthcare challenge. *International Journal of Gynecology and Obstetrics*, 131, S36–S39.
- Sweeney, E., Mcdaniel, R., Maximov, P., Fan, P., & Craig, V. (2013). Models and mechanisms of acquired antihormone resistance in breast cancer: Significant clinical progress despite limitations. *Hormone Molecular Biology and Clinical Investigation*, 9, 143–163.
- Takahashi, H., Kosaka, M., Watanabe, Y., Nakade, K., & Fukuyama, Y. (2003). Synthesis and neuroprotective activity of bergenin derivatives with antioxidant activity. *Bioorganic and Medicinal Chemistry*, 11(8), 1781–1788.
- Thaipong, K., Boonprakob, U., Crosby, K., Cisneros-Zevallos, L., & Byrne, D. (2006). Comparison of ABTS, DPPH, FRAP, and ORAC assays for estimating antioxidant activity from guava fruit extracts. *Journal of Food Composition and Analysis*, 19, 669–675.
- Toriola, A. & Colditz, G. (2013). Trends in breast cancer incidence and mortality in the United States: Implications of prevention. *Breast Cancer Research and Treatment*, 138(3), 665–73.
- Ulrike, G., Martina, R., Elisabeth, W., Anja, H., Johannes, K., Vadim, M., Sandor, N., & Michaela, S. (2016). Discovery of prenylated flavonoids with dual activity against influenza virus and *Streptococcus pneumoniae*. *Scientific Reports*, 6, doi:10.1038/srep27156.
- US Breast Cancer Statistics. (2018). *Breast cancer reports 20018*, https://www.breastcancer.org/symptoms/understand_bc/statistics
- Usman, H., Abdulrahman, F., & Usman, A. (2009). Qualitative phytochemical

- screening and *in vitro* antimicrobial effects of methanol stem bark extract of *Ficus thonningii* (Moraceae). *African Journal of Traditional and Complementary Alternative Medicine*, 6(3), 289–295.
- Velloso, J. C., Regasini, L., Khalil, N., Da Silva Bolzani, V., Khalil, O. A., Manente, F.A., Netto, H., & De Faria Oliveira, O. M. (2011). Antioxidant and cytotoxic studies for keampferol, quercetin, and isoquercetin. *Eclética Química*, 36, 7–20.
- Veronesi, U., Salvadori, B., Luini, A., Greco, M., Saccozzi, R., & del Vecchio, M. (1995). Breast conservation is a safe method in patients with small cancer of the breast: Long-term results of three randomized trials on 1973 patients. *European Journal of Cancer*, 31A(10), 1574–9.
- Visht, S. & Chaturvedi, S. (2012). Isolation of natural products. *Current Pharmaceutical Research*, 2(584-599).
- Volman, J. J., Mensink, R. P., van Griensven, L. J., & Plat, J. (2010). Effects of alpha-glucans from *Agaricus bisporus* on *ex vivo* cytokine production by LPS and PHA-stimulated PBMCs: A placebo-controlled study in slightly hypercholesterolemic subjects. *European Journal of Clinical Nutrition*, 64, 720–726.
- Wan, J. M. F., Sit, W. H., & Louie, J. C. Y. (2008). Polysaccharopeptide enhances the anticancer activity of doxorubicin and etoposide on human breast cancer cells ZR-75-30. *International Journal of Oncology*, 32, 689–699.
- Wanmai, M., Supeecha, E., Nijsiri, R., & Wacharee, M. (2012). Effect of xanthoxylin on melanin content and melanogenic protein expression in B16F10 melanoma. *Asian Biomedicine*, 6(3), 413–422.
- Weng, C. J., Chau, C. F., Chen, K. D., Chen, D. H., & Yen, G. C. (2007). The anti-invasive effect of lucidenic acids isolated from a new *Ganoderma lucidum* strain. *Molecular Nutrition and Food Research*, 51, 1472–1477.
- Winefordner, J. (2009). *Liquid chromatography time-of-flight mass spectrometry*. Hoboken, New Jersey: John Wiley & Sons, Inc.
- Witasp, E., Shvedova, A., Kagan, V., & Fadeel, B. (2009). Single-walled carbon nanotubes impair human macrophage engulfment of apoptotic cell corpses. *Inhalation Toxicology*, 21, 131–136.
- Woo, E., Kwak, J., Kim, H., & Park, H. (1998). A new prenylated flavonol from the roots of *Sophora flavescens*. *Journal of Natural Products*, 61(12), 1552–4.

- Xiang, D. & Yiling, H. (2012). Structure feature and antitumor activity of a novel polysaccharide isolated from *Lactarius deliciosus* Gray. *Carbohydrate Polymers*, 89, 397–402.
- Xirui, H., Jiacheng, F., Linhong, H., & Jinhui, W. (2015). *Sophora flavescens* Ait.: Traditional usage, phytochemistry and pharmacology of an important traditional Chinese medicine. *Journal of Ethnopharmacology*, 172, 10–29.
- Xu, T., Beelman, R., & Lambert, J. (2012). The cancer preventive effects of edible mushrooms. *Anticancer Agents Medicinal Chemistry*, 12, 1255–63.
- Yamamoto, T., Tanaka, T., & Takenaka, S. (1999). Structural analysis of iron and manganese species in iron-and manganese-promoted sulfated zirconia. *Journal of Physical Chemistry B*, 103(13), 2385–2393.
- Yang, L., Zhang, A., & Zheng, X. (2009). Shrimp shell catalyst for biodiesel production. *Energy Fuels*, 23(8), 3859–3865.
- Yin, H. & Liao, L. F. J. (2014). Enhanced permeability and retention (EPR) effect-based tumor targeting: The concept, application and prospect. *JSM Clin Oncology Research*, 2(1), 1010.
- Ying, L., Bin, Z., Ibrahim, S., Shuang-Shuang, G., Hong, Y., & Wen, H. (2016). Purification, characterization and antioxidant activity of polysaccharides from *Flammulina velutipes* residue. *Carbohydrate Polymers*, 145, 71–77.
- Yip, C., Bhoo-Pathy, N., & Teo, S. (2014). A review of breast cancer research in Malaysia. *Medical Journal of Malaysia*, 69(Supplement A), 8–22.
- Youn, H., Jeong, J. C., Jeong, Y. S., Kim, E. J., & Um, S. J. (2013). Quercetin potentiates apoptosis by inhibiting nuclear factor-kappaB signaling in H460 lung cancer cells. *Biological and Pharmaceutical Bulletin*, 36, 944–951.
- Yu Huan, G. & Leonard, J. (2006). *In vitro* effects on proliferation, apoptosis and colony inhibition in ER-dependent and ER-independent human breast cancer cells by selected mushroom species. *Oncology Reports*, 15, 417–423.
- Yu, L., Fernig, D. G., Smith, J. A., Milton, J. D., & Rhodes, J. M. (1993). Reversible inhibition of proliferation of epithelial cell lines by *Agaricus bisporus* (edible mushroom) lectin. *Cancer Research*, 53, 4627–4632.
- Yue, G., Chan, B., Hon, P., Lee, M., Fung, K., & Leung, P. (2010). Evaluation of *in*

vitro anti-proliferative and immunomodulatory activities of compounds isolated from *Curcuma longa*. *Food and Chemical Toxicology*, 48, 2011–20.

Yumrutas, O., Serdar, O., Pehlivan, M., Ozturk, N., & Poyraz, I. (2015). Cell viability, anti-proliferation and antioxidant activities of *Sideritis syriaca*, *Tanacetum argenteum* sub sp. *argenteum* and *Achillea aleppica* subsp. *zederbaueri* on human breast cancer cell line (MCF-7). *Journal of Applied Pharmaceutical Science*, 5(3), 1–5.

Zhang, G., He, L., & Hu, M. (2011). Optimized ultrasonic-assisted extraction of flavonoids from *Prunella vulgaris* L. and evaluation of antioxidant activities *in vitro*. *Innovative Food Science & Emerging Technologies*, 12, 18–25.

Zhang, L., Yang, J., & Zhu, J. (2002). Properties and liquefaction activities of ferrous sulfate based catalyst impregnated on two Chinese bituminous coals. *Fuel*, 81(7), 951–958.

Zhang, M., Swarts, S., Yin, L., Liu, C., Tian, Y., Cao, Y., & Okunieff, P. (2011). Antioxidant properties of quercetin. *Advanced Experimental Medical Biology*, 701, 283–289.