

## Soybean leaf disease detection and severity measurement using multiclass SVM and KNN classifier

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### ABSTRACT

Soybean fungal diseases such as Blight, Frogeye leaf spot and Brown Spot are a significant threat to soybean plant due to the severe symptoms and lack of treatments. Traditional diagnosis of these diseases relies on disease symptom identification based on naked eye observation by pathologists, which can lead to a high rate of false-recognition. This work presents a novel system, utilizing multiclass support vector machine and KNN classifiers, for detection and classification of soybean diseases using color images of diseased leaf samples. Images of healthy and diseased leaves affected by Blight, Frogeye leaf spot and Brown Spot were acquired by a digital camera. The acquired images are preprocessed using image enhancement techniques. The background of each image was removed by a thresholding method and the Region of Interest (ROI) is obtained. Color-based segmentation technique based on Incremental K-means clustering is applied to the region of interest for partitioning the diseased region. The severity of disease is estimated by quantifying a number of pixels in the diseased region and in total leaf region. Different color features of segmented diseased leaf region were extracted using RGB color space and texture features were extracted using Gray Level Co-occurrence Matrix (GLCM) to compose a feature database. Finally, the support vector machine (SVM) and K-Nearest Neighbour (KNN) classifiers are used for classifying the disease. This proposed classifier system is capable to classify the types of blight, brown spot, frogeye leaf spot diseases and Healthy samples with an accuracy of 87.3% and 83.6% are achieved.

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## 1. INTRODUCTION

Soybean is one of the oldest crops of the world. Today, soybean is the largest source of oil for human consumption as well as protein for livestock feed. It is reported that soybean has helped in improving the social and economic conditions of a large number of small and marginal farmers in India. The major soybean-growing states are Madhya Pradesh, Maharashtra, Rajasthan, Karnataka, Andhra Pradesh, and Chattisgarh. These states together contribute to about 98% of the total soybean production in the country [1]. In recent decades due to some factors such as natural disasters, soil erosion etc., lead to the incidence of crop diseases. These diseases can result in significant reduction in both quality and quantity of agricultural yields. The consequence of this is it causes production and economic losses. Most of the time farmers come across great difficulties in identifying and controlling plant diseases. Thus, it is very important to diagnose the plant diseases at early or superior stages so that proper and suitable action can be taken by the farmers to avoid further loss.

Soybean Leaf diseases like Bacterial Leaf Blight, Septoria Brown Spot, and Soybean rust are cause significant reduction in yield loss and lead to affect quality of soybean Products [2], thus influence farmers life. An effective way to control soybean foliar diseases is by applying fungicides. The identification of disease with traditional expert system are always subjective and this will lead to an inaccurate diagnosis. In recent years there is rapid development of computer vision techniques and pattern recognition techniques, which can intelligently diagnose the crop diseases and accurately identify the types of diseases [3-8].

Image processing technique and computer vision system have been used for automatic detection and classification of plant disease from extracted color, texture and shape features [9-11]. Studied show that image processing and computer vision tools can be successfully used as automatic and accurate disease detection mechanism [12]. For this work, we had selected Bacterial Blight, Septoria Brown Spot and Frogeye Leaf spot disease samples, whose symptoms are specified in Table 1 and these disease samples, which are selected from our collected dataset of soybean diseases, are depicted in Figure 1.

Table 1. Symptoms of Soybean Leaf Diseases

Disease Name	Casual Pathogen	Symptoms	Conditions and Timing of Disease occurrence	Yield Loss in %
Bacterial Blight	<i>Pseudomonas syringae</i> pv. <i>glycinea</i> ,	<ul style="list-style-type: none"> <li>a) Causes small, angular, yellow-to-brown Spots.</li> <li>b) Surrounded by yellow halos.</li> <li>c) The angular spots enlarge and join together producing large, irregular dead areas.</li> <li>d) The centers of old dead areas tear away so infected leaves have a ragged appearance.</li> </ul>	<ul style="list-style-type: none"> <li>a) Plants can be infected at any time during the growing season.</li> <li>b) Cool, wet weather and rain storms favor disease development.</li> <li>c) It is spread by wind and rain and by cultivation when foliage is wet.</li> </ul>	severe defoliation
Cercospora Leaf Spot	<i>Cercospora sojina</i> .	<ul style="list-style-type: none"> <li>a) Lesions on leaves begin as small, dark, water-soaked spots.</li> <li>b) They develop into brown spots surrounded by a darker reddish-brown or purple ring.</li> <li>c) The center of spots may turn white with black specks visible</li> </ul>	<ul style="list-style-type: none"> <li>a) At any time during the growing season, but seems to typically occur after flowering.</li> <li>b) Young leaves on the tops of plants tend to be most susceptible.</li> <li>c) Infection and disease development is favored by warm, humid weather.</li> </ul>	severe defoliation
Brown Spot	<i>Septoria glycines</i> i	<ul style="list-style-type: none"> <li>a) The first symptoms usually appear on the lower leaves and then progresses to the mid-to-upper canopy throughout the summer.</li> <li>b) Initial symptoms are small dark brown spots.</li> <li>c) The brown spots often enlarge and grow together into irregular brown, yellow areas.</li> </ul>	<ul style="list-style-type: none"> <li>a) It is most common when conditions and leaves are wet and warm, ideally for extended periods of time. disease.</li> </ul>	5-8%



Figure 1. Soybean diseased leaf samples (a) Blight, (b) Frogeye leaf spot, (c) Brown spot

The various researchers investigated their methods for disease detection, classification and assessment key of disease severities for different plant diseases which are outlined as follows: Dheeb Al Bashish, Malik Braik, Sulieman [13], proposed a framework for detection of plant diseases present on leaves and stem. The proposed framework is composed of K-Means segmentation technique and the segmented images are classified using neural network classifier. Dhiman Mondal, Dipak Kumar Kole [14] proposed a method to detect and classify the presence of yellow vein mosaic virus disease of okra leaf with the aid of K-means and Naive Bayesian classifier. He has experimented on 79 standard diseased and

non-diseased okra leaf images. Classification showed an average accuracy of 87%.

Evy Kamilah Ratnasari [15] proposes a model to identify the severity of spot disease which appears on leaves. He used thresholding a\* component of L\*a\*b\* color space for disease spot detection. Then features of diseases spots are extracted using Support Vector Machine (SVM) classification techniques, first he uses L\*a\*b\* color space for its color features and then applied Gray Level Co-Occurrence Matrix (GLCM) to obtain texture features. Study revealed that proposed model is capable to determine the types of spot diseases with an accuracy of 80% and 5.73 error severity estimation average. It is reported that for severity measurement; percentage of the infected area estimated by quantifying number pixels in diseased symptoms area and number of pixels in total leaf area of the leaf has been often used in disease assessment techniques.

Also while using the manual assessment method it is found that such methods contain some degree of subjectivity; hence we may not be considered it as ground truth. Above discussion concludes that most of the researchers investigated disease segmentation method using K-means clustering techniques which has the limitation on cluster selection (local cluster selection). Therefore these issues along with enhancement of performance accuracies demand automatic and accurate method to detect plant diseases is of great realistic significance [2, 7, 9]. In this work, we present a novel Incremental K-means segmentation algorithm [16] (global cluster selection) for soybean leaf disease detection and classification using multiclass SVM and KNN which may be used to overcome these limits.

This paper is organized as follows: Section 2 methodology used for soybean leaf disease segmentation using novel Incremental means clustering and SVM and KNN Classifiers followed by disease identification, data acquisition, image processing technique, classification method. Section 3 reveals the experimental results and discussion using SVM and KNN Classifiers and finally, Section 4 presents the conclusion.

## 2. RESEARCH METHOD

This study reports on a framework for the detection, classification and severity measurement of soybean plant disease visual symptoms by the analysis of colored images. The framework was divided into two phases: (I) Training Phase and (II) Testing Phase. For both phases the first 4 stages are common that includes:

- Training Database image set and input test Image.
- Image pre-processing: to resizing the database images in to suitable form; to eliminate leaf object background regions; to specify a suitable color transformation that best highlighted the diseased regions shown in the image; image enhancement: to filtering unwanted noise in the input image and enhanced highlighted diseased regions considered targets (possible diseased area);
- Image segmentation: to partitioning the identified leaf regions in the image into 3 clusters that out of which one cluster were likely to qualify as diseased region;
- Feature Extraction: to extract the significant information from the given input sample; color and texture features were extracted and the same is used for further image analysis.
- Classification: Classifier is trained using extracted feature values and its respective target values using the SVM and KNN classifiers and then this trained classifiers is used to classify test images. A flowchart of the complete process is shown in Figure 2.

### 2.1. Image database

The set of images used in this study were obtained from the Department of Plant Pathology College of agriculture; Agriculture research Centre, Sub-mountain Zone, Kolhapur, affiliated to Mahatma Fule Krishi Vidyapeeth Rahuri, Maharashtra, India. The Department of plant pathology, at the college of agriculture, India supplied the sample of diseased soybean leaves. Then diseased soybean leaves are placed on the white base to remove complexity and then images are taken by using a digital camera which is used for both training and testing the system. In all cases, the standard JPG image format was used to store these images. Stored images include the soybean leaves infected by Septoria Brown Spot, Cercospora leaf spot, and Bacterial Blight.

### 2.2. Image preprocessing

The set of images from a database is given and then image preprocessing is done on gathered images for improving the image quality. The preprocessing of image includes: image resizing-in this stage due to the large size of the captured image the input image is first resized to size 256\*256; image smoothing and contrast enhancement—due to inadequate or non-uniform illumination input image may get blurred, so by adjusting the color map of image; contrast enhancement in R, G, B color plane in which the adjustment on

each plane (red, green, and blue) of the RGB intensity image RGB take place as shown in Figure 3. The result causes balance uneven contrast of the image and then any undesired distortion if there will be suppressed it out. After image enhancement, next stage is- image filtering: in which median filter is used to enhance highlighted diseased regions considered targets (possible diseased area) by better contrast and brightness balancing also by which noise elimination (background noise) is also removed. Next stage is Background Removal: in this stage mostly green colored pixels, are masked and based on it we computed threshold value of these pixels. Then green pixels are masked based on: if pixel intensity of the green component is less than the pre-computed threshold value, then zero value is assigned to the red components of this pixel and the region of interest (ROI) is obtained.

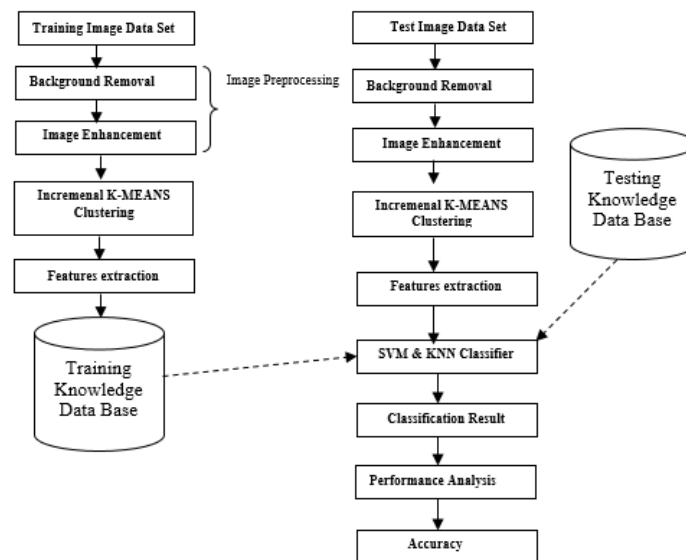


Figure 2. Framework for plant leaf disease detection and classification



Figure 3. ROI of blight diseased leaf sample: (a) Original image (b) ROI of original image

### 2.3. Leaf Disease Segmentation

We proposed a new system to detect and estimate the soybean leaf disease blight and leaf spots using threshold-based segmented spots image and classify the colors in (R, G, B) color space using incremental k-means clustering technique. The Otsu method is used for spot disease lesion segmentation from (R, G, B) channel of color space [17]. The threshold value of the pixels was computed according to the masked pixels. The green pixels were masked according to the following condition: If the pixel intensity of the green component was less than the pre-computed threshold value, a value of zero was assigned to the red components of this pixel and the region of interest (ROI) was obtained.

#### 2.3.1. Incremental k-means clustering algorithm

The conventional k-means clustering method is an extensively used clustering technique that seeks to minimize the average squared distance between points in the same cluster. Although this method offers no

accuracy guarantees, its simplicity and speed are very appealing in practice. By Incremental k-means with a simple, randomized seeding technique, we developed an algorithm that is competitive with the optimal K-means clustering [16]. Experimental results show our incremental k-means clustering improves both the speed and the accuracy of k-means, often quite intensely. Algorithm:

Let  $D(x)$  denote the shortest distance from a data point to the nearest center we have already chosen. Then, we define the following algorithm, which names as k-means++.

- 1a. Take one center  $c_1$ , chosen uniformly at random from  $X$ .
  - 1b. Take a new center  $c_i$ , choosing  $x \in X$  with probability  $D(x)^2 / \sum_{x \in X} D(x)^2$
  - 1c. Repeat Step 1b until we have taken  $k$  centers altogether.
  2. For each  $i \in \{1, \dots, k\}$ , set the cluster  $C_i$  to be the set of points in  $X$  that are closer to  $c_i$  than they are to  $c_j$  for all  $j \neq i$ .
  3. For each  $i \in \{1, \dots, k\}$ , set  $c_i$  to be the center of mass of all points in  $C_i$ :  
 $c_i = 1/|C_i| \sum_{x \in C_i} x$ .
  4. Repeat Steps 2 and 3 until  $C$  no longer changes euclidean distance.
- where  $d(i, j)$  is the distance between  $i^{\text{th}}$  and  $j^{\text{th}}$  pixels of corresponding cluster.

$$d(i, j) = \sqrt{|x_{i1} - x_{j1}|^2 + |x_{i2} - x_{j2}|^2 \dots + |x_{ip} - x_{jp}|^2} \quad (1)$$

An example of the output of incremental K-means clustering for a soybean leaf infected with Bacterial Blight depicted in Figure 4.

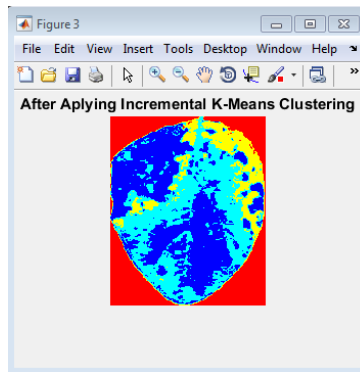


Figure 4. Output of incremental K-Means clustering for a bacterial blight leaf disease which is shown in 4 clusters

### 2.3.2. Soybean leaf disease detection and severity measurement system

System is composed of two subsystems a) Disease detection b) severity measurement.

#### a) Disease detection

The system for detection of soybean leaf disease is shown in Figure 5. It is used for disease detection, and concurrently provides the measurement of disease severity. The proposed system is divided into two stages: the calculation of the leaf disease severity and classification of leaf disease. The first stage is the measurement of disease severity which is performed only on testing data in a certain standard size of the leaves.

Whereas the second stage uses the data from the testing and training spot and blight disease (spot and blight disease use Otsu segmentation method from RGB color space which gives lesion region area (Diseased area-Ad) by keeping only disease pixels on also called lesion region extraction, this region (area) is computed by using regionprops Matlab command. Simultaneously total leaf area (Al) is calculated by keeping all pixels belongs leaf object high (1) and pixels belongs to background keeping it at low (0) using Otsu thresholding segmentation im2bw with greythresh Matlab command and this total leaf region (area) is computed by using regionprops Matlab command shown in Figure 6. Then after disease segmentation using incremental k-means clustering, whose extracted features from knowledge database are further used for disease classification; this is depicted in Figure 5 testing Phase system diagram. Classification result shows the type of disease class and its computed features.

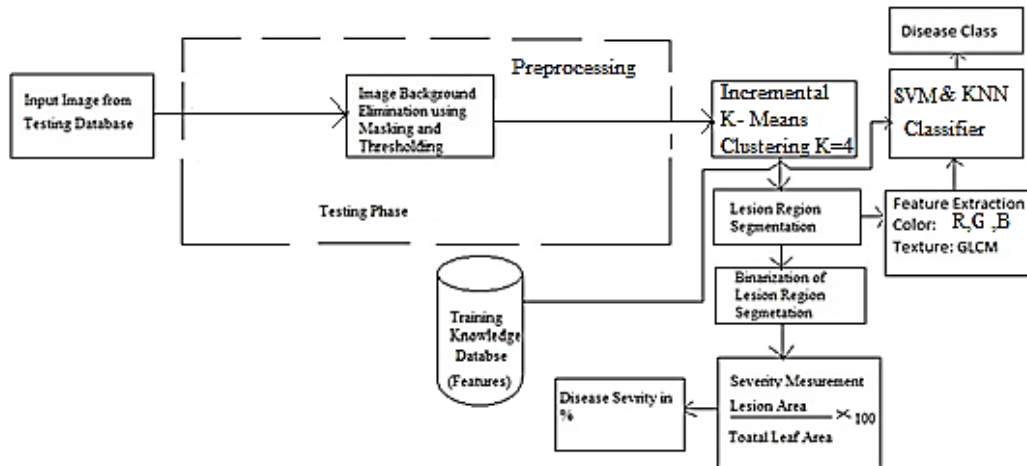


Figure 5. Soybean Leaf Disease Detection and Severity measurement system diagram at Training Phase

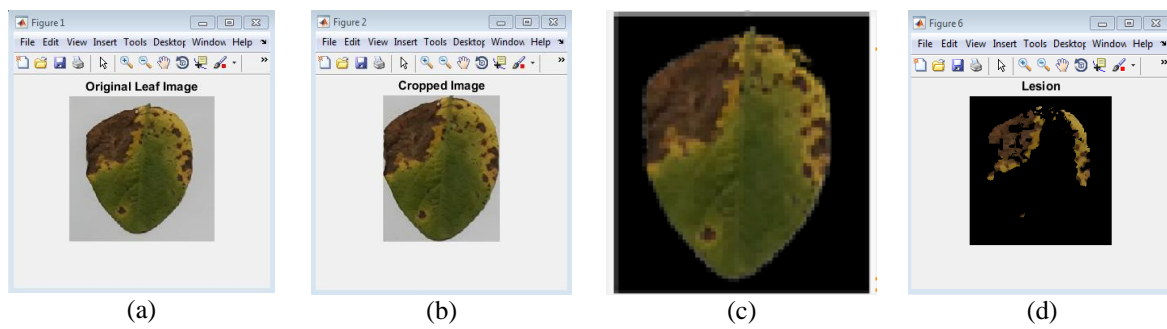


Figure 6. Otsu thresholding lesion segmentation: (a) Original leaf image, (b) cropped leaf image, (c) ROI of leaf image, and (d) Blight leaf disease lesion segmentation

#### b) Disease Severity Measurement

##### Algorithm:

- Image Acquisition.
- Images resize.
- Image preprocessing: Image correction, shadow and background removal
- Color image segmentation using Incremental k-means clustering
- Disease lesion region extracted and calculating disease area ( $A_d$ ).
- Convert the original image to B/W image.
- Calculate total leaf area ( $A_l$ )
- Calculating from extracted leaf region area ( $A_l$ ). (Binarization of lesion area)
- Applying formula to measure disease severity:

$$Ds = (A_d/A_l) * 100.$$

One of the objectives of this research work aims at a measurement of the severity of a soybean leaf disease. In this stage Otsu thresholding algorithm from RGB channel color space used to isolate the disease symptoms called lesion region ( $A_d$ ); from which features can be extracted and properly processed in order to provide an estimate for the severity of the disease. Disease severity is the lesion area of the leaves showing symptoms of spot disease and it is most often expressed as a percentage [15]. The disease severity of the soybean leaf is measured by quantifying the number of white (on) pixel belongs to lesion region called diseased area ( $A_d$ ) and comparing it with a quantified total number of leaf object pixels. In our work, the binarization method is used to calculate the total leaf area of soybean leaf object ( $A_l$ ) [18]. The severity of disease is given in percentage which is computed using (2). Adopting image processing method to measure disease severity can be expressed as the following formula.

$$DS = \frac{A_d}{A_l} = \frac{P \sum_{(x,y) \in R_d} 1}{P \sum_{(x,y) \in R_l} 1} = \frac{\sum_{(x,y) \in R_d} 1}{\sum_{(x,y) \in R_l} 1} \quad (2)$$

$A_d$  is Lesion Region Area;  $A_l$  is Leaf Region Area;  $P$  is Unit Pixel Expressed Area;  $R_d$  is Lesion Region;  $R_l$  is Leaf Region. Unit pixel in the same digital image represent the same size, so ratio DS can be obtained by segmenting lesion region from leaf Region and quantifying pixel number  $\sum_{(x,y) \in R_d} 1$  of Disease region and  $\sum_{(x,y) \in R_l} 1$  of leaf region in the cluster output of k-means segmented image.

### 2.3.3. Feature Extraction

In this research total of 8 features are extracted out of which 6 are color features such as Rmean, Gmean, Bmean, Rstd, Gstd, Bstd of R, G, B color space and 2 are texture feature such as skewness and kurtosis are extracted. For texture features extraction GLCM method is used [19].

- Color feature:

Color features such as Rmean, Gmean, Bmean, Rstd, Gstd, and Bstd of R, G, B channel color space are extracted these six features computed using following statistical expression:

i) Mean

$$\text{Mean: } X' = \frac{1}{N} \sum X_i \quad (3)$$

where  $X_i$  is the pixel intensity and  $N$  is the total number of pixels. Here mean is considered as one of the feature.

ii) Standard Deviation

Standard Deviation is the square root of the variance of the distribution. It is calculated using following formula:

$$\text{Standard Deviation } (\sigma) = \sqrt{\frac{1}{N} \sum_{i=1}^N (X_i - X')^2} \quad (4)$$

- GLCM texture feature

Texture feature in this research uses GLCM with two feature vectors:

i) Skewness

The skewness is used to judge the image surface. It is used to detect edges in dark objects on white background, having a sign change at luminance changes in images based on degree 3 and 4 moment, so these are termed higher order statistics [20].

$$\text{Skewness} = \frac{\frac{1}{N} \sum_{i=1}^N (X_i - X')^3}{\left(\frac{1}{N} \sum_{i=1}^N (X_i - X')^2\right)^{3/2}} \quad (5)$$

ii) Kurtosis

Kurtosis is a measure of whether the data are heavy-tailed or light-tailed relative to a normal distribution.

Then for univariate data  $X_1, X_2, \dots, X_N$ , the kurtosis is:

$$\text{kurtosis} = \frac{\sum_{i=1}^N (X_i - X')^4}{N s^4} \quad (6)$$

where  $X'$  is the mean,  $s$  is the standard deviation, and  $N$  is the number of data points. Note that in computing the kurtosis, the standard deviation is computed using  $N$  in the denominator rather than  $N - 1$ .

### 2.3.4. Multiclass Support Vector Machine (SVM)

In our research work a Multiclass Support Vector Machine is used for classification of leaf disease. This classifier method is used to identify the 3 different class of soybean leaf disease including 1 healthy class. Classification is based on feature extraction result of target class, which deals with associating a given input pattern with one of the defined class. The soybean diseased leaves samples were considered for testing and training data in this study is on segmented ROI, leaf spot image.

### 2.3.5. K-Nearest Neighbour Classifier (KNN)

The leaf sample from testing database is to be identified undergoes feature extraction process as the ones in training database. Then it will be compared to each leaf features extracted in the training dataset one by one. The sum of Euclidean distance between the leaf sample in the testing dataset and those in the training



database are calculated. Each ratio is then used as fetures in KNN classifier.The closest matches are estimated using KNN algorithm [14].

### 3. RESULTS AND ANALYSIS

#### 3.1. Image database

Classification task consists two phases, training phase and testing phase using the multiclass SVM. In the training phase, a SVM and KNN are trained using extracted leaf object feature values and its relative target class values. This trained SVM is then used to classify test images samples. Raw data of soybean diseased leaves were gathered from soybean fields in Agriculture Research Centre Kolhapur, and from Sangli, Kolhapur district Maharashtra, India and sankeshwar, karnataka, India. There are total 244 training samples of soybean leaf images as data set for defined disease class testing data set and 80 samples were considered for testing data set (including both early stage as well as Lateral stage images) are used. They cover 55 images with Blight disease, 49 images with frogeye leaf spot disease, and 40 images with Brown spot disease and 100 images with healthy leaf. Figure 1 shows the leaves sample of testing data. The details of the training database are shown in Table 2.

Table 2. Image Database for Training and Testing SVM and KNN classifier

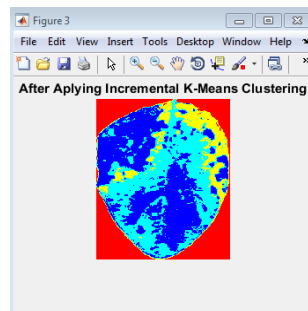
Leaf Species	Database Sample	Total Training Samples	Total Testing Samples
Soybean	Bacterial Blight	60	20
	Ftogeye Leaf Spot	60	20
	Brown Spot	50	20
	Healthy	100	20
Total		270	80

#### 3.2. Incremental K-means clustering

First step is ROI image shown in Figure 7(a) is partitioned into four clusters using Incremental K-means technique. Figure 7(b) shows 4 clusters formed using Incremental K-means techniue. Then second step is selection of diseased lesion rgion cluster containing ROI which is a final segmented image whose feature is extracted for disease analysis. Figure 8 shows diseased lesion rgion cluster containing ROI selection.



(a)



(b)

Figure 7. Output of Incremental K-Means clustering for a Bacterial Blight disease class which is shown in 4clusters, (a) ROI of leaf image, (b) Color image segmentation using Incremental k-means Clustering

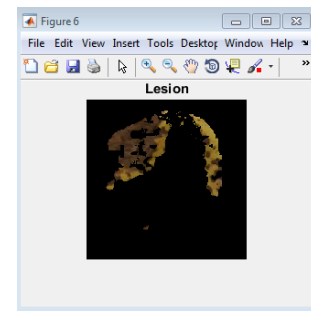


Figure 8. Diseased lesion rgion extraction

#### 3.3. Otsu thresholding and binarisation

The next step is a measurement of disease severity from extracted lesion region using binarization method applying over lesion region and total leaf area as shown in Figure 9. In this stage Otsu thresholding algorithm from R, G, B color space used to isolate the disease symptoms called lesion region (Ad); from which features can be extracted and properly processed in order to provide an estimate for the severity of the disease. Disease severity is the lesion area of the leaves showing symptoms of blight disease and it is most often expressed as a percentage.



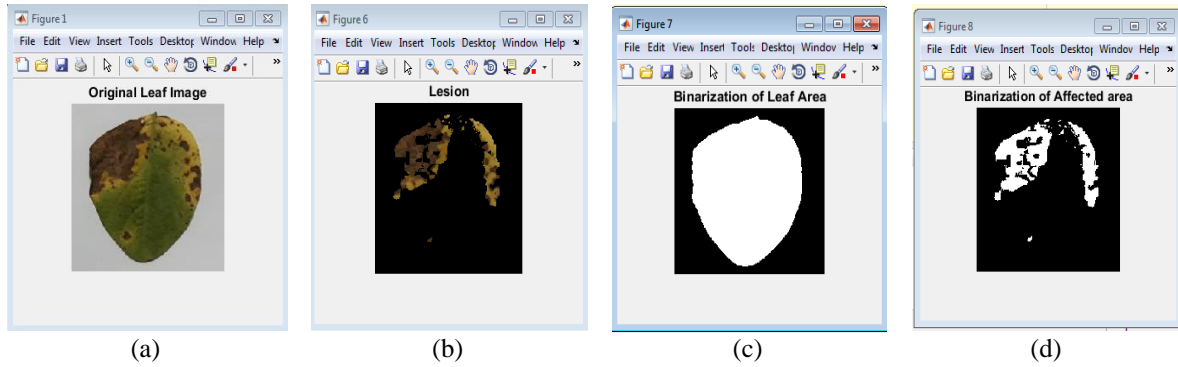



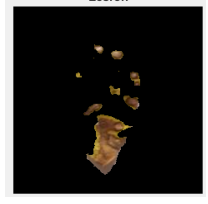




Figure 9. Example of binarisation method applied over blight disease. This shows (a) original image (b) Extracted lesion region (c) Binarization of Leaf Area (d) Binarisation of lesion region

### 3.4. Disease Severity Measurement

The disease severity of the soybean leaf is measured by quantifying the number of white (on) pixel belongs to lesion region called diseased area ( $A_d$ ) and comparing it with a quantified total number of leaf object pixels. In our work, the binarization method is used to calculate the total leaf area of soybean leaf object ( $A_l$ ). The lesion percentage of the leaf is computed using equation (2). After final clustering of ROI, the number of pixels  $\sum_{(x,y) \in R_d} 1$  in the disease region is **6220** and the number of pixels  $\sum_{(x,y) \in R_l} 1$  in the leaf region is **25798** for (**Bacterial Blight**). Thus it can be calculated that the ratio DS of the diseased and leaf area is **0.1460** and its severity is **24.11%**. Table 3 shows estimated soybean disease severities for Bacterial blight, Frogeye leaf spot, and Septoria brown spot disease.

Table 3. Disease severity measurement

Database Sample	Segmented Disease (Lesion) Region	( $A_d$ ) Total pixel in diseased area	( $A_l$ ) Total pixel in leaf area	( $DS$ ) Disease severity in%
 Bacterial Blight Original Leaf Image		6220	25798	24.11
 Frogeye leaf Spot Original Leaf Image		2947	12614	23.36
 Brown Spot		1328	33078	4.01

### 3.5. Disease classification

The final step is detecting the bacterial blight, frogeye leaf spot and a brown spot disease type of the segmented images using the multiclass SVM classifier. Furthermore segmented images were used to extract texture and color features. A total of 8 features are estimated for all three partitioned parts of single leaf image. This feature values, collectively called a feature vector, is given to trained multiclass SVM and KNN classifier which classifies the input leaf image into 4 classes Bacterial Blight, frogeye Leaf Spot, Septoria Brown spot and healthy, depending upon its feature values. We labeled black blight disease as class 1, Brown spot disease as class 2, Frogeye leaf spot disease as class 3, and healthy as class 4. Total 241 data samples for 4 data class are considered to train the system. From 241 data samples, there are 28 data misclassified, 10 data in class1 misclassified, 15 data with class2 misclassified and 3 data with class3 misclassified shown in Figure 8 of confusion matrix of SVM. So, classification accuracy for class 1 is 78.7%, class 2 is 55.9% class 3 is 95% and class 4 is 100% respectively for leaf with Bacterial Blight, leaf with Brown, spot and leaf with healthy. Average accuracy of SVM classifier is 88.38%.

Similarly for KNN- classifier out of 241 data samples, there are 37 data misclassified, 10 data in class1 misclassified, 16 data with class2 misclassified and 8 data with class3 misclassified shown in Figure 8 of confusion matrix of KNN. So, classification accuracy for class 1 is 78.7%, class 2 is 52.9% class 3 is 81.7% and class 4 is 100% respectively for leaf with Bacterial Blight, leaf with Brown, spot and leaf with healthy. Average accuracy of KNN classifier is 84.64%. Table 4 shows the result of multiclass SVM and KNN classifier out of which SVM perform extremely well with Bacterial black blight class and leaf with a brown spot of soybean leaves as compared with Septoria brown class.

Table 4. Performance analysis of SVM and KNN classifier

Image Database	Total Samples	Correctly Classified (SVM Classifier)	% Accuracy	Correctly Classified (KNN Classifier)	% Accuracy
Bacterial Blight	47	37	78.7	37	78.7
Septoria Brown Spot	34	19	55.9	18	52.9
Frog Leaf Spot	60	57	95	49	81.7
Healthy	100	100	100	100	100
Combined 4 Data Class	241	213	88.38	204	84.64

## 4. PERFORMANCE EVALUATION

### 4.1. Confusion Matrix and ROC for SVM Classifier

Figure 10 and Figure 11 shows confusion matrix of proposed disease classification system, it shows overall correctly classified and missclassified result of defined disease class, it shows percentage per true class including true positive rates (TPR) and False negative rates (FNR) [20]. The overall success rate of multiclass SVM classifier is 88.38% shown in Figure 12.

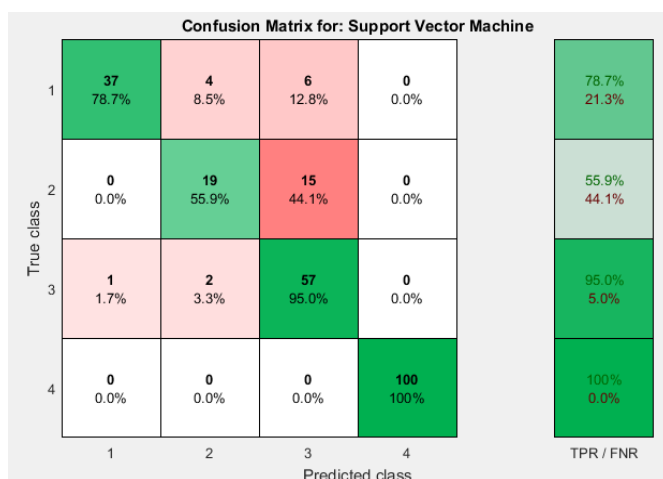


Figure 10. Confusion matrixes for SVM show percentage per true class

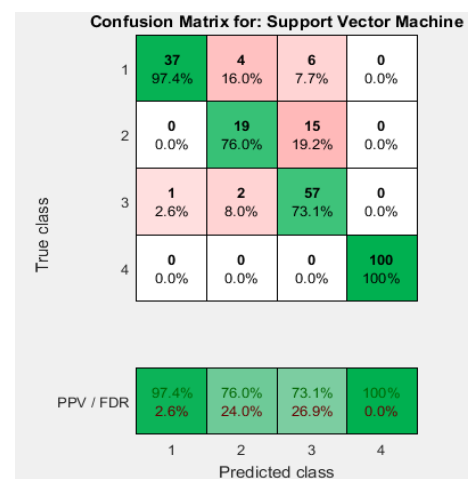


Figure 11. Confusion matrixes for SVM show percentage per predicted class

Figure 10 shows confusion matrix of proposed disease classification system, it shows overall percentage per predicted class including positive predictive values (PPV) and False Discovery Rates (FDR). Figure 12 shows confusion matrix of proposed disease classification system, it shows percentages over the entire confusion matrix.

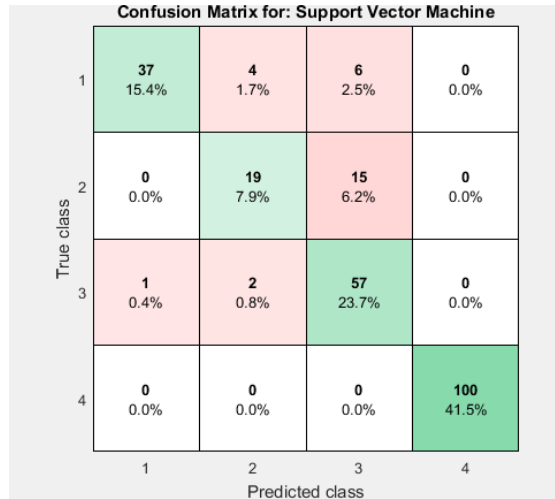


Figure 12. Percentage view of overall entire Confusion Matrix for SVM

**4.2. Confusion Matrix and ROC for KNN Classifier**

**4.2.1. Confusion Matrix**

Figure 13 and Figure 14 shows confusion matrix of proposed disease classification system, it shows overall correctly classified and missclassified result of defined disease class, it shows percentage per true class including true positive rates (TPR) and False negative rates (FNR) [20]. The overall success rate of multiclass SVM classifier is 84.64 % shown in Figure 15.

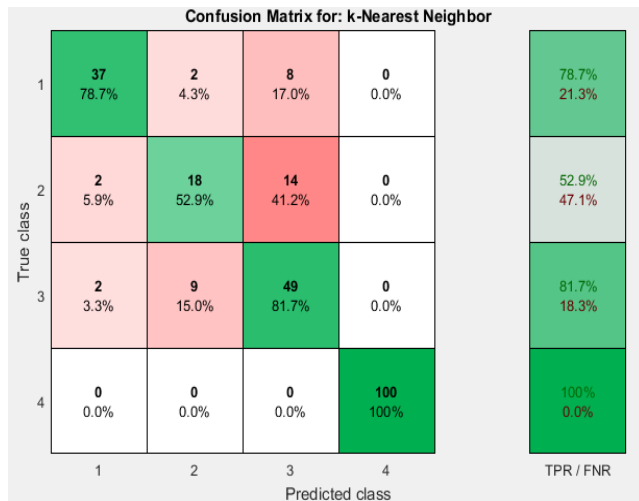


Figure 13. Confusion matrixes for KNN show percentage per true class

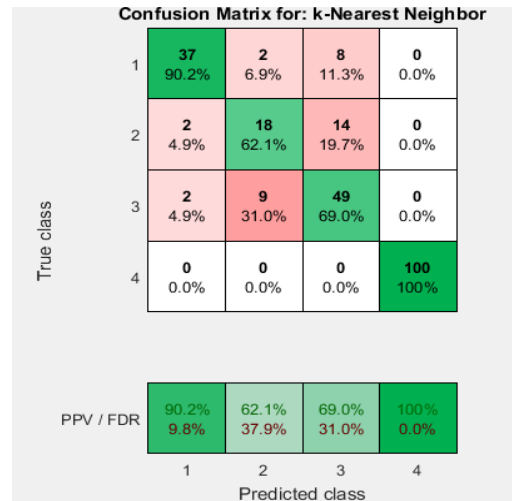


Figure 14. Confusion matrixes for KNN show percentage per predicted class

**Confusion Matrix for: k-Nearest Neighbor**

True class	1	37 15.4%	2 0.8%	8 3.3%	0 0.0%
	2	2 0.8%	18 7.5%	14 5.8%	0 0.0%
	3	2 0.8%	9 3.7%	49 20.3%	0 0.0%
	4	0 0.0%	0 0.0%	0 0.0%	100 41.5%
		Predicted class			
		1	2	3	4

Figure 15. Percentage view of overall entire Confusion Matrix for KNN

**4.2.2. ROC**

It shows Area Under curve for each defined disease class and healthy class. According to classification of positive class accuracy of each disease class area under curves varies which is shown in Figure 16 AUC positive class1, Figure 17 AUC positive class2, Figure 18 AUC positive class3 and Figure 19 AUC positive class4 respectively for SVM classifier.

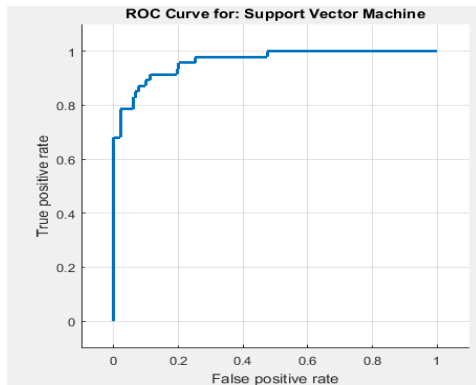


Figure 16. AUC positive class 1(0.963)

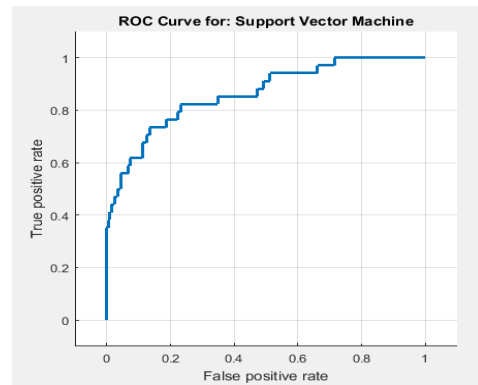


Figure 17. AUC positive class 2(0.863)

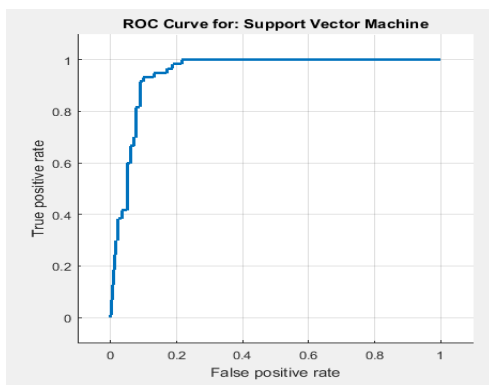


Figure 18. AUC positive class 3(0.947)

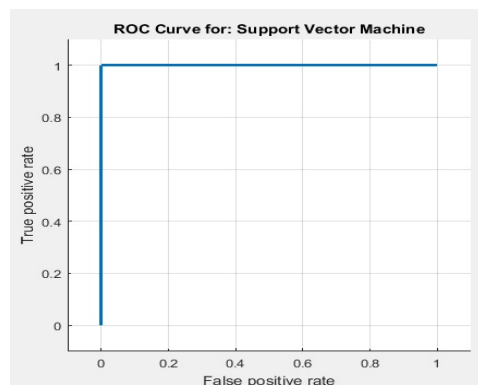


Figure 19. AUC positive class 4(1)

**4.3. ROC Curve for KNN-Classifier**

Similarly Area Under curve for KNN classifier which shows ROC characteristics for defined disease class and healthy class. According to classification of positive class accuracy of each disease class area under curves varies which is shown in Figure 20 AUC positive class1, Figure 21 AUC positive class2, Figure 22 AUC positive class3 and Figure 23 AUC positive class4 respectively for KNN classifier.

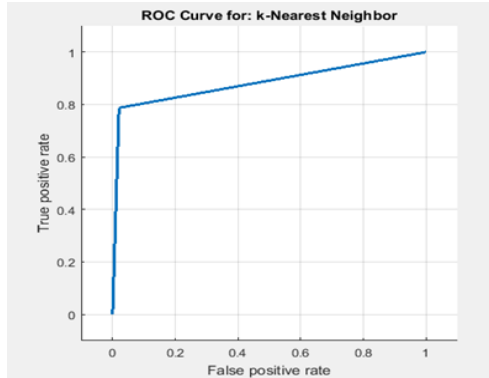


Figure 20. AUC positive class 1 (0.883)

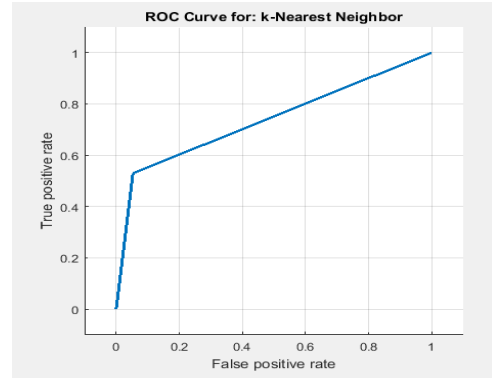


Figure 21. AUC positive class 2 (0.7381)

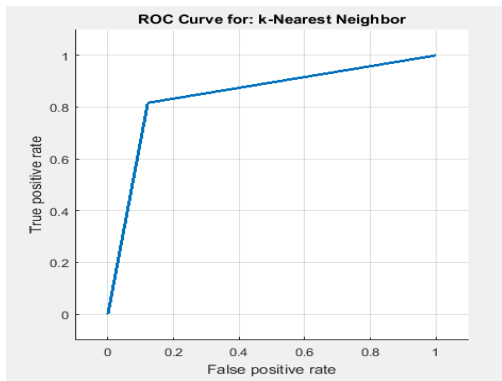


Figure 22. AUC positive class 3 (0.847)

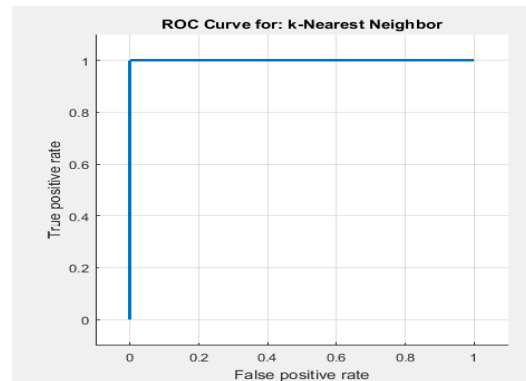


Figure 23. AUC positive class 4 (1)

**4.4. Scatter Plot for SVM and KNN classifier**

Figure 24 and Figure 25 shows scatter plot of disease classification system, for SVM and KNN classifiers [20] it shows classwise data samples correctly classified and misclassified for target group over train data.

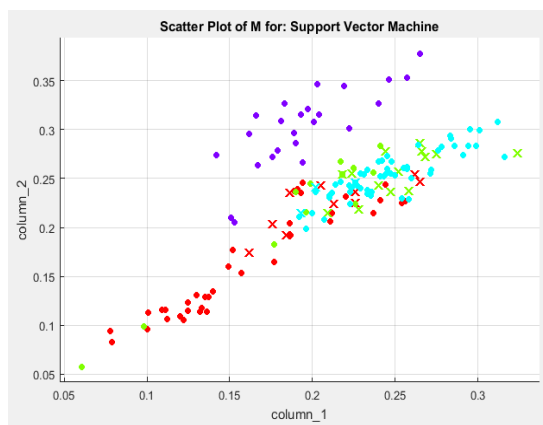


Figure 24. Scatter Plot for SVM classifier

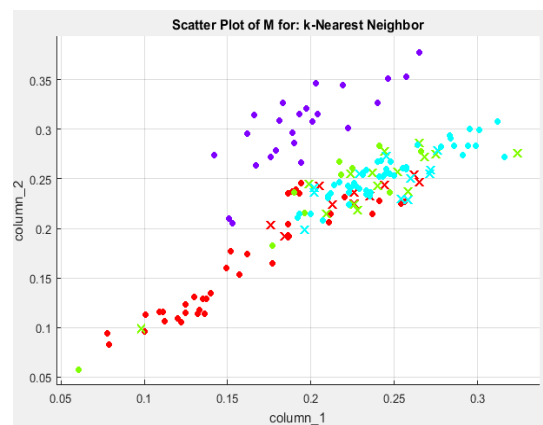


Figure 25. Scatter Plot for KNN classifier

The overall performance of SVM and KNN classifier is summarized into Figure 26 and Figure 27 respectively. Table 5 shows comparative study of our SVM classifier system with previous system.

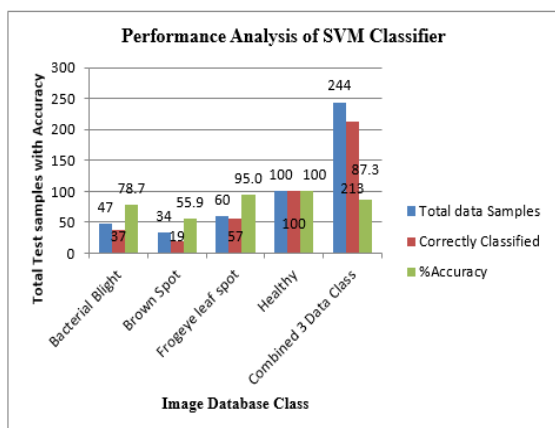


Figure 26. Performance analysis of SVM classifier

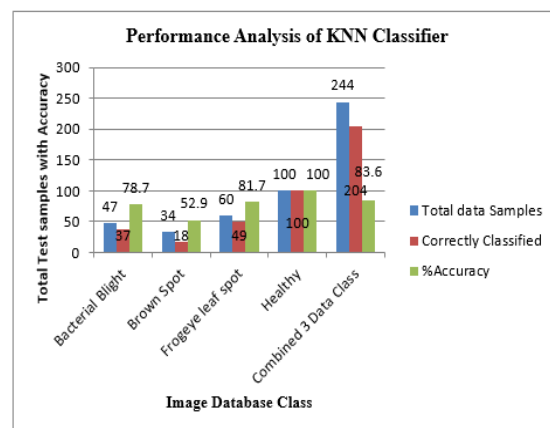


Figure 27. Performance analysis of KNN classifier

Table 5. Comparative study of Proposed SVM and KNN classifier system

Particulars	Previous Implemented System by Ratnasari et al. [7]	Proposed SVM and KNN Classifier System
Crop Species	Sugarcane	Soybean
Disease Detected Class	Leaf brown spot	Bacterial Blight, Brown spot & Cercospora Leaf spot
Feature extracted	Color & Texture	Texture & Color
Classifier	SVM	Multiclass Linear SVM
Accuracy	80%	87.3 % & 83.4%

## 5. CONCLUSION

We propose a system to identify soybean leaf disease including its severity estimation and classification. We test our algorithms on three disease class of soybean leaves were considered namely, Bacterial Blight, Frogeye Leaf Spot, and Septoria Brown Spot. The given system uses image resizing, and thresholding for image preprocessing. To segment the lesion leaf area, the Incremental K-means clustering technique is used then both color and texture features extraction are done using R,G,B color space and Gray Level Co-occurrence Matrix (GLCM) respectively. Then finally the SVM and KNN classification technique is used to detect the defined type of leaf disease. Our experimental results indicate that the SVM classifier algorithm outperforms over KNN algorithm with an accuracy of 87.3% and 83.4% respectively which can significantly support accurate and automatic detection including classification of leaf diseases. The performance of our proposed system is dependent on the size of a database. In future, we plan to increase random database and clean database to cover more disease types and still better result.

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