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REVIEW PAPER

Biohazards of Protein Biotoxins

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ABSTRACT

Biotoxins are toxic substances produced by a living organism that cause diseases in human beings, animals, or plants. The agent may be lethal or incapacitating. The new, emerging threat agents are biotoxins produced by animals, plants, fungi, and bacteria. Many types of organisms produce substances that are toxic to humans. Examples of such biotoxins are botulinum toxin, tetanus toxin, and ricin. Several bioactive molecules produced by the pharmaceutical industry can be even more toxic than the classical chemical warfare agents. Such new agents, like the biotoxins and bioregulators, often are called mid-spectrum agents. The threat to human beings from agents developed by modern chemical synthesis and by genetic engineering also must be considered, since such agents may be more toxic or more effective in causing death or incapacitation than classical warfare agents. By developing effective medical protection and treatment against the most likely chemical and mid-spectrum threat agents, the effects of such agents in a war scenario or following a terrorist attack can be reduced. Toxin-mediated diseases have made human beings ill for millennia. The use of biological agents as weapons of terror has now been realised, and separating naturally occurring disease from bioterroristic events has become an important public health goal.

Keywords: Biotoxin, bioterrorism, toxic protei, ricin, abrin, viscumin, volkenesin, modeccin, conotoxin, botulinum toxin, clostridium perfringens toxins, diphtheriae toxin, Staphylococcus toxin, shiga toxin, verotoxin, cholera toxin, tetanus toxin, microcystin, mid-spectrum agents

1. INTRODUCTION

Toxin-mediated diseases have made human beings ill for millennia and many of them had catastrophic¹ character. The use of biological agents as weapons of terror has been realised, and separating naturally occurring disease from bioterroristic events has become an important public health goal. The key to timely identification of such attacks relies on education of primary care physicians, first responders, and public health officials.

Natural toxins are toxic compounds produced by living organism^{1,2}. There are principle toxic substances of poisonous animals, plants, microorganisms and other forms of life. Such agents, like the biotoxins, often are called mid-spectrum agents³. The manner of their toxic effect can be different in individual substances. According to the organ toxicity, the neurotoxins, hepatotoxins, nephrotoxins, hemotoxins, etc. can be distinguished. For example neurotoxins are toxic agents or substances that damage or destroy the tissues of the nervous system, especially

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neurons. Neurotoxic effects can include behaviour changes, seizures, as well as wide range of effects, including death. According to the chemical structure, it is possible to divide all natural toxins (biotoxins) into nonprotein and protein compounds. This review covers peptide toxins which are on the control list of chemical weapons. Each toxin is briefly characterised chemically, pharmacologically, and toxicologically. Their natural sources, availability, stability, and military potential are discussed.

2. BIOTOXINS AND CHEMICAL WEAPONS CONVENTION

Biotoxins are used in nature for two primary reasons: (i) predation (spider, snake, jellyfish, wasp) and (ii) defence (honeybee, poison dart frog, deadly nightshade). All of these are very toxic compounds and many of them represent a risk for human health and the viability of organisms. That is why biotoxins can be employed in warfare and in terrorist attacks⁴. Chemical Weapons Convention (CWC) (1993) includes toxins as chemical agents, and specifically includes toxins in its control regime along with other highly toxic chemicals. Protein toxins are: Abrin, botulinum, *Clostridium perfringens*, *Corynebacterium diphtheriae*, microcystins, *Staphylococcus aureus*, ricin, shigatoxin, and tetanus. On the control list of the Australian Group, a voluntary association of 33 countries, are also conotoxins, verotoxin, cholera toxin, modeccin, volkensin and viscumin are enregistered.

Botulinum toxin produces a descending flaccid paralysis⁵. *Staphylococcal enterotoxin B* produces a syndrome of fever, nausea, and diarrhea and may produce a pulmonary syndrome if aerosolised. *Clostridium perfringens* epsilon-toxin could possibly be aerosolised to produce acute pulmonary edema. Ricin intoxication can manifest as gastrointestinal hemorrhage after ingestion, severe muscle necrosis after intramuscular injection, and acute pulmonary disease after inhalation⁶. All these biotoxins may be perilous if they will be misused by terrorists⁷.

3. PROTEIN BIOTOXIN CLASSIFICATION

All peptide toxins are created by one of more linear or cyclic polypeptide chains. Polypeptide is

constructed from more amino acids connected by means of peptide linkage.

Proteins are large macromolecules composed of one or more peptide chain. It is possible to say that polypeptides composed from less number of amino acids are designated as peptides or polypeptides, from the larger number as proteins. The crossover between peptides and proteins is not defined exactly and is unsubstantial from practice point of view.

The biological mechanism of biotoxin action as well as development of their toxic manifestation, clinical course, and therapy pretensions are very different.

3.1 PLANT TOXINS

Plant toxic proteins belong to a group of phytotoxins, which inhibit the protein synthesis of eukaryotic cells. The toxins of this group are glycoproteins with molecular weights of about 60 kDa which consist of two subunits linked into a dimer by a disulphide bond. One of the subunits is lectin with sites for carbohydrate binding (B-chain), and the other subunit is specific N-glycosidase (A-chain), which modifies the 28S rRNA-60S ribosomal subunit⁸.

The group of proteins known as the lectins was first recognised in plant seeds and they bind to specific sugars. Although lectins, in general, are not very toxic, there are some relationships between lectins and toxins⁹. They may also serve as recognition markers in cellular differentiation and act as immunotoxin. Only A-chain is toxic due to inhibiting protein synthesis. The A-chain catalytically inactivates 60S ribosomal subunit by removing adenine from positions 4 and 324 of 28S rRNA. Ribosome-inactivating proteins (RIP) have been identified in many plants and some of them are very toxic, but only in connection with B-chain¹⁰. The A-chain is carrier of toxicity, but B-chain binds to cell surface receptors and facilitates a transport of the A-chain across the cell membrane. The A-chain is not active until it internalised by the cell, where halts protein synthesis¹¹.

Each toxin molecule can disable approximately 2000 polysomes per minute, enough to eventually kill the cell. The most known plant toxic proteins

of this type are ricin, abrin, modessin, viscumin and volkensin^{12,13}.

3.1.1 Ricin

Ricin is a protein produced by the castor oil plant, *Ricinus communis*. It is native to tropical Africa, but has naturalised sub-tropical and temperate areas as well. The whole of the plant is poisonous, containing the toxin ricin, which reaches the highest levels in the seeds. The seeds also contain a purgative oil, the triglyceride of ricinoleic acid. The seeds have been used in folk medicine against many diseases (eg. remedies for abscess, anasarca, arthritis, asthma, boils, burns, cancer) for centuries.

Ricin is the only toxin to exist naturally in large quantities. It is a byproduct of castor oil production and ricin isolation and separation is simple and cheap. Easy preparation and low price might make this toxin attractive to a poor country. For nations or terrorists who lack the money to spend on nuclear weapons and other high-tech killing instruments, toxin warfare offers horrific appeal as biological/toxin weapons are cheap, easy to make, and simple to conceal. Even small amounts, if effectively used, could cause massive injuries and make many suffers¹⁴. Ricin's significance as a potential biological warfare toxin relates in part to its wide availability. Worldwide, one million ton of castor beans are processed annually to produce castor oil and in the waste there is five per cent ricin by weight. The toxin is also quite stable and extremely toxic by several routes of exposure, including the respiratory route¹⁵.

Ricin is the compound responsible for toxicity of the seeds of *Ricinus communis*¹⁶. Ricin has been known as a poison for years, usually through livestock deaths. One to three seeds may be fatal to a child; two to four may be poisonous to an adult, while eight may be fatal. A fatal ingested dose for human is about 1 µg/kg¹⁷. Because the alimentary tract destroys lots of ricin, it is much more potent when administered paraenterally.

The toxically active A-chain of ricin is a 267 amino acid globular protein and is classed as an N-glycosidase. The A-chain is protein containing 8 α -helices

and 8 β -sheets¹⁸. The B-chain is composed of 262 amino acid residues and is classed as a lectin. The B-chain has an affinity for binding to galactosides¹⁶ and possesses two galactose binding sites that are attracted to galactose containing glycoproteins at the cell surface. The A- and B-chain glycoproteins are linked by a disulphide bond located at residue 259 of the A-chain and residue 4 of the B-chain¹⁹.

Ricin is a glycoprotein with carbohydrate side chains in the form of mannose-rich N-linked oligosaccharides and particularly binds to mannose receptors of cells of the reticuloendothelial system. The specific sites with potential for binding of high mannose carbohydrate chains of ricin are at asparagines 10 and 236 of the A-chain, and asparagines 95 and 135 of the B-chain¹⁶.

The toxic effects of ricin are essentially the result of the action of the A-chain, which inactivates the ribosomes of the cell. It is thought that the B-chain serves to bind to galactose-containing and/or mannose-containing structures at the cell surface, thus allowing the A-chain to enter the cell²⁰. By this way, the protein is internalised, meaning that it is taken into the cell as a toxin-binding site complex in vesicles. The B-chain of ricin facilitates the escape of the A-chain by binding to the endosomal membrane allowing the A-chain to pass through the membrane. Then, the B-chain dissociates from the A-chain by breaking the disulphide bond. The A-chain is thus delivered to the cytoplasm where it is taken up by the golgi apparatus and transported to the endoplasmic reticulum²⁰.

At the endoplasmatic reticulum, ricin inactivates the cell ribosomes by elimination of adenine in specific RNA sequences on the 28S ribosomal subunit²¹. A single A-chain molecule is capable of deactivating every ribosome in the cell thus halting protein synthesis and culminating in cell death. It is important to note that the toxin is specific for eukaryotic ribosomes. A hairpin loop on the 28S rRNA containing the tetranucleotide loop GAGA is the most likely target for attack by ricin on the ribosome, however it is thought that the ribosome conformation is an important factor in recognition by the protein²¹. Therefore, ricin is not a nucleotide sequence-specific protein.

In biological warfare it is expected that ricin would be released as a toxic cloud. It could also be injected into specific persons as a terrorist or sabotage weapon. Additionally, ricin is easy to produce and is stable. The toxic effects of ricin occur because it kills the cells of the body that it contacts when it is taken into the body. Upon inhaling an adequate amount of ricin, death of persons affected would be expected in 36-48 h because of difficulty in breathing and circulatory system also gets effected. Ingested ricin is expected to cause internal bleeding, death of vital organs, and death of the individual. Injected ricin causes death by major organ failure.

The prediction of symptoms to be expected is based on animal studies and accidental human exposures, which were not fatal. Symptoms would probably vary depending on whether ricin was inhaled, ingested or injected. About three hours after inhaling ricin, the symptoms expected are cough, tightness of the chest, breathing difficulty, nausea, and muscle aches. This would progress to a severe inflammation of the lungs and airways, increased breathing difficulty, cyanosis, and death in 36-48 h from failure of the breathing and circulatory systems. Ingestion of ricin would be expected to cause nausea and vomiting, internal bleeding of the stomach and intestines, failure of the liver, spleen, and kidneys, and death of the individual by collapse of the circulatory vessels. No specific affects on the lungs and airways would be expected. If injected, ricin causes marked death of muscles and lymph nodes near the site of injection and probable failure of major organs and death of the individual²².

3.1.2 Abrin

Abrin is a potent toxin that has been isolated from the seeds of *Abrus precatorius* (or Rosary pea). Its use as a tool for research was described in 1972 by Sharon and Lis²³. Abrin exists in two forms, abrin a and abrin b. Both are composed of two chains, A-chain and B-chain. A disulphide bond between Cys247 of the A-chain and Cys8 of the B-chain links the A- and B-chains. The A-chain is 251 residues and is divided into 3 folding domains. The A-chain catalytically inactivates 60S ribosomal subunits by removing adenine from positions 4 and

324 of 28S rRNA, therefore inhibiting protein synthesis. The B-chain is a galactose specific lectin that facilitates the binding of abrin to cell membranes^{24,25}. The B-chain of both forms of abrin consists of 268 amino acid residues and shares 256 identical residues. Comparison of their sequences with that of the ricin's B-chain shows that 60 per cent of the residues of abrin's B-chain are identical to those of the ricin's B-chain and that two saccharide-binding sites in ricin B-chain identified by a crystallographic study are highly conserved in abrin B-chain²⁶.

The mechanism of toxic action of abrin is identical to that of ricin¹⁶ but the toxicity of abrin in mice is 75 times higher than that of ricin (0.04 µg/kg for abrin compared to 3 µg/kg for ricin). The diagnosis, clinical features, treatment, protection, prophylaxis and so on is the same as was described above for ricin intoxications²⁷. Recently, similar a highly toxic RIP protein from seeds of the *Abrus pulchelus*, entitled pulchellin, was isolated and characterised²⁸.

3.1.3 Viscumin

Viscumin (Mistletoe lectin I, ML I), inevitable to the family of Ribosome-inactivating proteins (RIP), was identified in the late 1980s as the main pharmacologically-active ingredient of mistletoe (*Viscum album*) extract and is largely responsible for its toxicity²⁹. It is comparable in toxicity to ricin and acts by the same mechanism. When viscumin binds to its target cell, protein synthesis in that cell is interrupted as a result of the A-chain's enzymatic activity, like a ricin. This interruption induces a cellular stress response, which triggers the release of cytokines by the target cell and, at high viscumin concentrations, apoptosis of the cell occurs. The association of A- and B-subunits is predominantly hydrophobic in nature³⁰. Viscumin has a concentration-dependent activity profile unique to plant toxins. It starts with lectin-dependent mitogenicity and then covers toxicity and cell agglutination, associated with shifts in the monomer/dimer equilibrium. Each lectin subunit harbors two sections for ligand contact³¹.

3.1.4 Volkensin

Volkensin is a lectin from *Adena volkensii* (the kilyambiti plant) that is comparable in toxicity

to ricin and that acts by the same mechanism (a ribosome-inactivating protein)³². The plant is a relatively unattractive and toxic succulent plant found in Africa that appears to be of little interest. However, it has proven to be useful as a research reagent in neurology because of its ability to be taken up and transported by certain types of nerves. There may be pressure to develop commercial sources for the research community²². Volkensin is probably the most potent plant toxin known, with an LD50 for rats of 50-60 ng/kg³³.

3.1.5 Modeccin

Modeccin is a lectin from the roots of *Adenia digitata* an African succulent plant that is comparable in toxicity to ricin³⁴ and acts by the same mechanism^{35,27}. The plant does not seem to have any significant uses, such as a food or as medicine, and so is not available in quantities comparable to abrin, let alone ricin. However, the seed does seem to be readily available. The subunits of modeccin were isolated (subsequently referred to as modeccin 4B), purified from the roots of *Adenia digitata* by affinity chromatography on Sepharose 4B³⁶. There is A-subunit (Mol Wt 26 000), which inhibits protein synthesis, and B-subunit (Mol Wt 31 000), which binds to cells. A second form of modeccin, not retained by Sepharose 4B, was purified by affinity chromatography on acid-treated Sepharose 6B: this form is subsequently termed modeccin 6B. Modeccin 6B has a molecular weight indistinguishable from that of modeccin 4B, and consists of two subunits of Mol Wts 27 000 and 31 000, joined by a disulphide bond. The subunits were not isolated because of their high insolubility in the absence of sodium dodecyl sulphate. As compared with modeccin 4B, modeccin 6B is slightly less toxic to animals, does not agglutinate erythrocytes, and is a more potent inhibitor of protein synthesis in a lysate of rabbit reticulocytes, giving 50 per cent inhibition at a concentration of 0.31 mg/ml³⁷.

3.2 ANIMAL TOXINS

There are numerous protein toxins produced by many different animal sources, such as snakes, scorpions, spiders, insects, frogs, sea anemones, etc. Only conotoxins, the toxic principle of marine

snails of the genus *Conus*, are presented on the control list of the Australian Group.

3.2.1 *Conus* Toxins (Conotoxins)

These are toxic peptides produced by the fish-hunting marine snails of the genus *Conus*. The conotoxins (a mixture of toxic substances produced by snails) are used to paralyse the fish being attacked³⁸. For centuries, members of the Conidae family have been collected for their unique and intricately designed shells. Only during the last few decades have cone shells become an exciting area for scientific research. Cone shells are marine snails and are found in reef environments throughout the world. They prey upon other marine organisms, immobilising them with unique venoms. They can be dangerous for human beings, too. There have been 30 recorded cases of human envenomation by fish-eating cone shells, in some cases fatal³⁹.

The composition of the venom differs greatly between species and between individual snails within each species, each optimally evolved to paralyse its prey. The active components of the venom are small peptide toxins, typically 12-30 amino acid residues in length. They have very tight conformations by multiple disulphide bridges. These patterns of disulphide bridge help to define a number of structural classes of conotoxin. Today several tens of conotoxins are known and these are divided into four groups. α -conotoxins are the shortest and have only two disulphide bridges. μ -conotoxins, ω -conotoxins, Δ -conotoxins, and κ -conotoxins contain three disulphide bridges between different cysteine residues of peptide chain⁴⁰.

The paralytic components of the venom, that have been the focus of recent investigation, are the α -, ω -, and μ -conotoxins. All of these conotoxins act by preventing neuronal communication, but each targets a different aspect of the process to achieve this. The α -conotoxins target nicotinic ligand-gated channels, the μ -conotoxins target the voltage-gated sodium channels and the omega conotoxins target the voltage-gated calcium channels.

Another class of peptides from *Conus venoms* are the conantokins, the first peptide antagonists which target to the major excitatory receptors in

the vertebrate central nervous system, glutamate receptors. The conantokins selectively inhibit a subtype of glutamate receptor, the N-methyl-D-aspartate (NMDA) receptor, which are ligand-gated *Ca* channels. The conantokins cause rather striking and till this time, little explored biological effects⁴¹.

Individual conotoxins, even within the same class, can vary greatly in lethality towards mammals. Some of the tremor-inducing ω conotoxins are not lethal, whereas others of the same group are lethal at low levels. However, they are only toxic in rats and mice when administered intracranially (into the brain). Some α conotoxins have lethal doses as low as 25 $\mu\text{g}/\text{kg}$ for mouse. This may be an overestimate of toxicity because it is determined from the dose required to kill a mouse in 20 min. In addition, it has to be borne in mind that the toxicity of the complex mixture of peptides that is cone snail venom, may be much greater than the sum of its parts because of the synergistic interaction between toxins acting on different aspects of neural function. Incidents of cone snails killing people are known to have occurred.

3.3 MICROBIAL TOXINS

Protein toxins represent numerous groups of toxic compounds, produced by different pathogenic bacteria. These bacterial toxins are: botulinum toxins, *Clostridium perfringens* toxins, *Corynebacterium diphtheriae* toxin, *Staphylococcus aureus* toxins, shigatoxin, verotoxin, cholera toxin, and tetanus toxin.

3.3.1 Botulinum Toxins

Botulinum toxin is very strong poison derived from the genus of anaerobic bacteria named Clostridia. Seven antigenic types of botulinum toxin exist⁴², designated from A through G. They can be identified based on antibody cross-reactivity studies. *Clostridium botulinum* is a familiar bacterium that causes botulism, a form of food poisoning⁴³. Naturally occurring botulism is the disease that results from the absorption of botulinum toxin into the circulation from a mucosal surface (gut, lung) or through a wound. It does not penetrate the intact skin. The toxin irreversibly binds to peripheral cholinergic synapses, preventing the release of the neurotransmitter acetylcholine

from the terminal end of motor neurons. This leads to muscle paralysis, and in severe cases, can lead to a need for mechanical respiration⁴⁴. Patients with botulism typically present with difficulty in speaking, seeing and/or swallowing. Prominent neurologic findings in all forms of botulism include ptosis, diplopia, blurred vision, dysarthria and dysphagia. Patients typically are afebrile and do not have an altered level of consciousness. Patients may initially suffer from gastrointestinal distress, nausea, and vomiting, preceding neurological symptoms⁴⁵.

3.3.2 Clostridium Perfringens Toxins

The gram-positive pathogen *Clostridium perfringens* is a major cause of human and veterinary enteric disease largely because this bacterium can produce several toxins when present inside the gastrointestinal tract. *Clostridium perfringens* food poisoning is one of the more common problem in the industrialised world⁴⁶. The enteric toxins of *Clostridium perfringens* share two common features: (i) they are all single polypeptides of modest (~25-35 kDa) size, although lacking in sequence homology, and (ii) they generally act by forming pores or channels in plasma membranes of the host cells. These enteric toxins include *Clostridium perfringens* enterotoxin (CPE), which is responsible for the symptoms of a common human food poisoning and acts by forming pores after interacting with intestinal tight junction proteins. Two other *Clostridium perfringens* enteric toxins, ϵ -toxin (a bioterrorism select agent) and β -toxin, cause veterinary enterotoxemias when absorbed by the intestine. *Clostridium perfringens* enterotoxin (β -toxin) has been shown to be the virulence factor responsible for causing the symptoms of *Clostridium perfringens* food poisoning. β -toxin is a single polypeptide chain with a molecular weight of 3.5 kDa that binds to receptors on the target epithelial cells. Through a unique four-step membrane action it finally causes a breakdown in normal plasma membrane permeability properties⁴⁷. Also β - and ϵ -toxins apparently act by forming oligomeric pores in intestinal or extra-intestinal target tissues. Other *Clostridium perfringens* toxins have different effect. *Clostridium perfringens* α -toxin is able to lyse erythrocytes via calcium channels activation⁴⁸. The main biological activity

of epsilon-toxin is the production of oedema in various organs⁴⁹ and cytoskeletal changes and plasma membrane functional alteration⁵⁰.

3.3.3 *Corynebacterium Diphtheriae* Toxin

Diphtheria toxin is an extracellular protein of *Corynebacterium diphtheriae* that inhibits protein synthesis and kills susceptible cells⁵¹. *Corynebacterium diphtheriae* is responsible for diphtheria. Diphtheria is a contagious, airborne, toxin-producing infection. It is considered as potential tools in bioterrorism⁵². It is characterised by the formation of a gray resistant pseudo-membrane in the lining of the mucous membrane of the upper respiratory tract as well as in the tonsils. Certain forms of the disease may be fatal. The global mortality rate for diphtheria is 5 per cent to 10 per cent and may reach 20 per cent among children under 5 and adults over 40. In 1888, Roux discovered the diphtheria toxin secreted by *Corynebacterium diphtheriae*. In 1890, the work of von Behring and Kitasato on antibodies to diphtheria antitoxins made it possible to envision their use in treating the disease⁵³. In 1897, Ehrlich established a standardised diphtheria toxin. These passive serum therapies would soon lead to active immunization. Diphtheria vaccines were first used in France in the 1920s. Mass immunization only began in the 1950s. The diphtheria vaccines used today throughout the world against the pathogenic and lethal effects of the diphtheria toxin are obtained by detoxification of the toxoid with formalin.

3.3.4 *Staphylococcus* Toxins

Staphylococcus aureus is a spherical bacterium (coccus) which on microscopic examination appears in pairs, short chains, or bunched, grape-like clusters. These organisms are gram-positive. Some strains are capable of producing a highly heat-stable protein enterotoxins, which range in size from 19 kDa to 26 kDa, that cause illness in human beings. Seven immunologically different forms of *Staphylococcus* enterotoxins is known: A, B, C1, C2, C3, D and E. These toxins are responsible for symptoms of food poisoning that follow consumption of food contaminated by *Staphylococcus* bacteria.

The onset of symptoms in staphylococcal food poisoning is usually rapid, and in many cases acute, depending on individuals susceptibility to the toxin, the amount of contaminated food eaten, the amount of toxin in the food ingested, and the general health of the victim. The most common symptoms are nausea, vomiting, retching, abdominal cramping, and prostration. Some individuals may not always demonstrate all the symptoms associated with the illness. In severe cases, headache, muscle cramping, and transient changes in blood pressure and pulse rate may occur. Recovery generally takes two days, however, it is not unusual for complete recovery to take three days and sometimes longer in severe cases. Infective dose - a toxin dose of < 1.0 µg in contaminated food will produce symptoms of *Staphylococcal* intoxication. This toxin level is reached when *Staphylococcus aureus* populations exceed 100,000/g.

Staphylococcus aureus also produces hemolytically-active toxins, so called hemolysins. Three hemolysins, α-toxin, β-toxin, and Δ-toxin, are known and each of them exist in more molecular forms. Their mechanism of toxic action is different⁵⁴.

α-toxin, also known as α-hemolysin, is one of the many virulence factors produced by *Staphylococcus aureus*⁵⁵. α-toxin is a single polypeptide chain known in four molecular forms with a molecular weight from 26 kDa to 39 kDa. The majority of *Staphylococcus aureus* strains isolated from human beings produce this toxin. α-toxin has been found to be lethal in animals, causing respiratory paralysis, vascular and smooth muscle spasms, and tissue necrosis⁵⁶.

β-toxin (β-hemolysin) is one of the several extracellular proteins produced by *Staphylococcus aureus*. It is a sphingomyelinase which disrupts the membranes of erythrocytes and other mammalian cells. Despite its characterised mechanism of action, the role of β-toxin in causing diseases in human beings and animals remains unclear⁵⁷.

Δ-toxin (Δ-hemolysin) is heat-stable peptide, having molecular weight 5 kDa, composed of 26 amino acid residues. Its molecules create amphiphatic

helix with hydrophilic amino acid residues on one side of long axis of molecule, hydrophobic residues on the other side and produced lesions in biological membranes very similar to those produced by Melittin and Triton X-100⁵⁵.

Staphylococcus toxins are suitable for use in warfare and terrorism in aerosolised form⁷.

3.3.5 Shiga Toxin

Shigella are the most important organisms that can cause dysentery. *Shigella dysenteriae* type 1 (Sd1) is the most virulent of the four serogroups of *Shigella*⁵⁸. The Sd1 is the only cause of epidemic dysentery. In addition to bloody diarrhoea, the illness caused by Sd1 often includes abdominal cramps, fever, and rectal pain. Less frequent complications of infection with Sd1 include sepsis, seizures, renal failure, and the haemolytic uraemic syndrome (HUS)⁵⁹. Altered arachidonic acid metabolism has been implicated in the pathogenesis of renal injury in the HUS caused by shigatoxin⁶⁰. Approximately 5-15 per cent of Sd1 cases are fatal. Shiga toxin works as enterotoxin, neurotoxin, and cytotoxin. It consists of two protein subunits. Subunit-A performs as N-glycosidase which splits adenin on ribosomes and inhibits protein synthesis⁶¹.

3.3.6 Verotoxin

The verotoxin, or shiga-like toxin family is a group of closely related toxins produced by certain pathogenic strains of *Escherichia coli*. Verotoxin-producing *Escherichia coli*, especially of serotype O157:H7, cause a zoonotic food or waterborne enteric illness that is often associated with large epidemic outbreaks as well as the HUS, the leading cause of acute renal failure in children⁶². These strains are a significant cause of human hemorrhagic colitis. In addition, they are both waterborne and foodborne and may also be transmitted from person-to-person by the oral-fecal route. In adults, illness caused by verotoxin may last several days. In children and the elderly, the illness can be fatal⁶³. Structure and mechanism of toxic action of verotoxin is equal to shigatoxin but recently new pieces of knowledge were obtained⁶⁴. It has been known, that following the intracellular routing of shigatoxin and/or verotoxin

to the endoplasmic reticulum and nuclear membrane, the toxins translocate into the cytoplasm and target ribosomes for the damage. However, numerous recent studies have shown that these toxins trigger programmed cell death, signaling cascades in intoxicated cells. The mechanisms of apoptosis induction by these toxins are emerging, and the data published to date suggest that the toxins may signal apoptosis in different cells types via different mechanisms⁶⁵.

3.3.7 Cholera Toxin

Vibrio cholerae is a gram-negative, curved rod bacteria. *Vibrio cholerae* is responsible for approximately seven pandemic infections of extreme diarrhea and dehydration across the globe, resulting in millions of death throughout the centuries. *Vibrio cholerae* as pathogen has been co-existing with mankind long before we were ever capable of detecting it, or understanding its mechanism of infection. Until recently, the methods to confront this pathogen, and begin to understand its function were not available.

The biochemistry of cholera toxin has been well characterised over the recent years^{66,67}. Cholera toxin is a heterohexameric protein, composed of five subunits B and one subunit A (AB₅, cholera toxin), responsible for the symptoms produced by *Vibrio cholerae* infection. In the first step of cell intoxication, the B-pentamer of the toxin binds specifically to the branched pentasaccharide moiety of ganglioside GM1 on the surface of target human intestinal epithelial cells.

The normal function of GM1 is not clearly understood but it has been implicated in numerous signal-transduction pathways. Cholera toxin and GM1 complex show no major conformational change, but it is theorised about that the A subunit of the cholera toxin is inserted into the cell. This translocation takes approximately 15 minutes and during this time it is theorised that the A subunit is cleaved along a disulphide bridge between A1 and A2, which keeps the protein inactive⁶⁸.

The A1-subunit of cholera toxin is the enzymatically active portion of the protein molecule, and it acts as an ADP-ribosyltransferase. It catalyses a transfer of

an ADP-ribose from an NAD⁺ to the arginine at location 187 in the α -chain of the regulatory protein⁶⁹ Gs. This ribosylation of Gs stabilises the GTP bound form of the protein, lower its GTPase activity creating a near-constitutively on signal for the generation of adenylyl cyclase, and therefore elevating cAMP levels. This high cAMP level results in the activation of the sodium pumps in the lumen of the cell through a cAMP-dependent kinase pathway, forcing out Na^+ ions. This resultant electrochemical imbalance drives out Cl^- and H_2O to balance the Na^+ release. The net flow of fluid is now out of balance and the cells attempt to compensate for the dehydration through removal of fluid from the blood. This is the biochemical process responsible for the clinical characteristics of epidemic cholera infection⁷⁰. Europe and America were largely clear of cholera cases.

3.3.8 Tetanus Toxin

The illness known as tetanus is caused by a neurotoxin produced by the anaerobic bacterium *Clostridium tetani*. It acts upon the presynaptic membranes of both central and peripheral nervous systems to block the release of neurotransmitters. Tetanus toxin is synthesised as a single polypeptide chain of 150 kDa, and undergoes proteolytic cleavage to produce a di-chain toxin consisting of the N-terminal 50 kDa fragment (light chain) linked by a disulphide bond to the 100 kDa carboxy terminal fragment (heavy chain). Fragment C (the C-terminal half of the heavy chain) retains ganglioside binding activity which is essential for the binding of the toxin to neuronal cells^{71,72}. Tetanus toxin is likewise botulinum toxin a neurotoxin and their molecular structures and mechanism of action are very similar⁷³. The LD₅₀ in unvaccinated human beings is estimated⁷⁴ at < 2.5 ng/kg. It is a powerful neurotoxin which may be fatal if inhaled or introduced through a wound. It causes muscle rigidity or spasms, paralysis, and death. If contact to this toxin occurs, flush eyes, skin or wounds thoroughly with water. Seek medical attention, since supportive therapy will be required if symptoms occur. Immune globulin may also be a part of the medical treatment.

4. CONCLUSIONS

Natural protein toxins represent a family of extremely potent toxic compounds, some of which, such as botulinum toxin, is the most potent toxin known till date. The use of toxins for terrorist affects aims in the form of aerosols is a perfectly credible eventuality. A number of toxins could be used in a terrorist attack, including ricin and botulinum toxins. Progress in molecular biology caused that it is easy to produce and can lead to massive destruction. It will be possible produce whatever toxin in a short time. Preparedness among physicians, first responders, and public health officials is especially important because these agents are very dangerous.

REFERENCES

1. Patocka, J. The toxins of cyanobacteria. *Acta Medica (Hradec Kralove)*, 2001, **44**, 69-75.
2. Mátlov, J.; Krejčí, V. & Patocka, J. Cyanotoxins and their effect on human health (Czech). *Kontakt*, 2004, **6**(1), 43-51.
3. Aas, P. The threat of mid-spectrum chemical warfare agents. *Prehospital Disaster Med.*, 2003, **18**, 306-12.
4. Slater, L.N. & Greenfield, R.A. Biological toxins as potential agents of bioterrorism. *J. Okla State Med. Assoc.*, 2003, **96**, 73-76.
5. Patocka, J.; Kuca, K. & Jun, D. Botulinum toxin: Bioterror and biomedical agent. *Def. Sci. J.*, 2006, **56**, 189-97.
6. Greenfield, R.A.; Brown, B.R. & Hutchins, J.B. Microbiological, biological, and chemical weapons of warfare and terrorism. *Am. J. Med. Sci.*, 2002, **323**, 326-40.
7. Zapor, M. & Fishbain, J.T. Aerosolized biologic toxins as agents of warfare and terrorism. *Respir Care Clin.*, 2004, **10**, 111-22.
8. Hartley, M.R. & Lord, J.M. Cytotoxic ribosome-inactivating lectins from plants. *Biochim. Biophys. Acta.*, 2004, **1701**, 1-14.

9. Wang, K.Y. & Xu, Q. Lectins and toxins. *Sheng Wu Hua Xue Yu Sheng Wu Wu Li Xue Bao* (Shanghai) 2000, **32**, 201-05.
10. Kozlov, J.V.; Sudarkina, O.I. & Karmanova, A.G. Ribosome-inactivating lectins from plants (Russian). *Molecular Biology* (Mosk), 2006, **40**, 711-723.
11. Stirpe F. Ribosome-inactivating proteins. *Toxicon*, 2004, **44**, 371-83.
12. Patocka, J. & Streda, L. Brief review of natural nonprotein neurotoxins. *ASA Newsletter*, 2002, **89**, 16-24.
13. Olsnes, S. The history of ricin, abrin and related toxins. *Toxicon*, 2004, **44**, 361-70.
14. Patocka, J. Toxicological characteristic of ricin (Czech). *Voj Zdrav Listy*, 19, **67**, 166-68.
15. Patocka, J. Abrin and ricin-two dangerous poisonous proteins. *ASA Newsletter*, 2001b, **85**, 205-08.
16. Rutenber, E. & Robertus, J.D. Structure of ricin B-chain at 2.5 Å resolution. *Proteins*, 1991, **10**, 260-69.
17. Clark, K. The chemical weapons convention: chemical and toxin warfare agents and disarmament. Royal Military College of Science, Cranfield University, August 1997.
18. Morris, K.N. & Wool, I.G. Analysis of the contribution of an amphiphilic α -helix to the structure and to the function of ricin A chain. *Proc. Natl. Acad. Sci.*, 1994, **91**, 7530-53.
19. Montfort, W.; Villafranca, J.E. & Monzingo, A.F. The three-dimensional structure of ricin at 2.8 Å. *J. Biol. Chem.*, 1987, **262**, 5398-403.
20. Olsnes, S. & Kozlov, J.V. Ricin. *Toxicon*, 2001, **39**, 1723-728.
21. Lord, J.M.; Roberts, L.M. & Robertus, J.D. Ricin: Structure, mode of action, and some current applications. *FASEB Journal*, 1994, **8**, 201-08.
22. Patocka, J. & Streda, L. Plant toxic proteins and their current significance for warfare and medicine. *J. Appl. Biomed.*, 2003, **1**, 141-47.
23. Sharon, N. & Lis, H. Cell-agglutinating and sugar-specific proteins. *Science*, 1972, **177**, 949-59.
24. Olsnes, S. & Pihl, A. Kinetics of binding of the toxic lectins abrin and ricin to surface receptors of human cells. *J. Biol. Chem.*, 1976, **25**, 3977-984.
25. Chen, Y.L.; Chow, L.P.; Tsugita, A. & Lin, J.Y. The complete primary structure of abrin-a B chain. *FEBS Letters*, 1992, **309**, 115-18.
26. Kimura, M.; Sumizawa, T. & Funatsu, G. The complete amino acid sequences of the B-chains of abrin-a and abrin-b, toxic proteins from the seeds of *Abrus precatorius*. *Biosci. Biotechnol. Biochem.*, 1993, **57**, 166-69.
27. Olsnes, S.; Sandvig, K.; Eiklid, K. & Pihl, A. Properties and action mechanism of the toxic lectin modeccin: interaction with cell lines resistant to modeccin, abrin, and ricin. *J. Supramol. Struct.*, 1978, **9**, 15-25.
28. Silva, A.L.; Goto, L.S.; Dinarte, A.R.; Hansen, D.; Moreira R.A.; Beltramini, L.M. & Araujo, A.P. Pulchellin, a highly toxic type 2 ribosome-inactivating protein from *Abrus pulchellus*. Cloning heterologous expression of A-chain and structural studies. *FEBS Journal*, 2005, **272**, 1201-210.
29. Krauspenhaar, R.; Eschenburg, S. & Perbandt, M. Crystal structure of mistletoe lectin I from *Viscum album*. *Biochem. Biophys. Res. Commun.*, 1999, **257**, 418-24.
30. Pryme, I.F.; Bardocz, S. & Pusztai, A. & Ewen S.W. Suppression of growth of tumour cell lines in vitro and tumours in vivo by mistletoe lectins. *Histol. Histopathol.*, 2006, **21**, 285-99.
31. Jimenez, M.; Andre, S.; Siebert, H.C.; Gabius, H.J. & Solis, D. AB-type lectin (toxin/agglutinin) from mistletoe: Differences in affinity of the two galactoside-binding Trp/Tyr-sites and regulation of their functionality by monomer/dimer equilibrium. *Glycobiology*, 2006, **16**, 926-37.
32. Chambery, A.; Di Maro, A.; Monti, M.M.; Stirpe, F. & Parente, A. Volkensin from *Adenia volkensii*

- Harms (kilyambiti plant), a type 2 ribosome-inactivating protein. *Eur. J. Biochem.*, 2004, **271**, 108-17.
33. Chambery, A.; Severino, V.; Stirpe, F. & Parente, A. Cloning and expression of the B chain of volkensin, type 2 ribosome inactivating protein from *Adenia volkensii* harms: Co-folding with the A chain for heterodimer reconstitution. *Protein Expr. Purif.*, 2006. Epub ahead of print.
 34. Olsnes, S.; Haylett, T. & Sandvig, K. The toxic lectin modeccin. *Methods Enzymology*, 1982, **83**, 357-62.
 35. Refsnes, K.; Haylett, T.; Sandvig, K. & Olsnes, S. Modeccin - a plant toxin inhibiting protein synthesis. *Biochem. Biophys. Res. Commun.*, 1977, **79**, 1176-83.
 36. Gasperi-Campani, A.; Barbieri, L.; Lorenzoni, E.; Montanaro, L.; Sperti, S. & Bonetti, E. Modeccin, the toxin of *Adenia digitata*. Purification, toxicity and inhibition of protein synthesis in vitro. *Biochemistry Journal*, 1978, **174**, 491-96.
 37. Barbieri, L.; Zamboni, M.; Montanaro, L.; Sperti, S. & Stirpe, F. Purification and properties of different forms of modeccin, the toxin of *Adenia digitata*. Separation of subunits with inhibitory and lectin activity. *Biochemistry Journal* 1980, **185**, 203-10.
 38. Lewis, R.J. Conotoxins as selective inhibitors of neuronal ion channels, receptors and transporters. *IUBMB Life*, 2004, **56**, 89-93.
 39. Fegan, D. & Andresen, D. *Conus geographus* envenomation. *Lancet*, 1997, **349**, 1672.
 40. Ohizumi, Y. & Matsunaga, K. Chemical structures and the mechanism of action of peptide toxins from cone shells (Article in Japanese). *Tanpakushitsu Kakusan Koso.*, 2001, **46**. (Suppl 14), 449-54.
 41. Castellino, F.J. & Prorok, M. Conantokins: inhibitors of ion flow through the N-methyl-D-aspartate receptor channels. *Curr. Drug Targets*, 2000, **1**, 219-35.
 42. Patocka, J. & Splino, M. Botulinum toxin: from poison to medicinal agent. *ASA Newsletter*, 2002, **88**, 14-19.
 43. Cherington, M. Botulism: update and review. *Semin Neurol.*, 2004, **24**, 155-63.
 44. Patocka, J.; Splino, M. & Merka, V. Botulism and bioterrorism: How serious is this problem? *Acta Medica (Hradec Kralove)*, 2005, **48**, 23-28.
 45. Robinson, R.F. & Nahata, M.C. Management of botulism. *Ann. Pharmacother*, 2003, **37**, 127-31.
 46. Smedley, J.G. 3rd; Fisher, D.J.; Sayeed, S.; Chakrabarti, G. & McClane, B.A. The enteric toxins of *Clostridium perfringens*. *Rev. Physiol. Biochem. Pharmacol.* 2004, **152**, 183-204.
 47. Brynestad, S. & Granum, P.E. *Clostridium perfringens* and foodborne infections. *Int. J. Food Microbiol.*, 2002, **74**, 195-202.
 48. Ochi, S.; Oda, M.; Nagahama, M. & Sakurai, J. *Clostridium perfringens* α -toxin-induced hemolysis of horse erythrocytes is dependent on Ca^{2+} uptake. *Biochim. Biophys. Acta*, 2003, **1613**, 79-86.
 49. Petit, L.; Gibert, M.; Gouch, A.; Bens, M.; Vandewalle, A. & Popoff, M.R. *Clostridium perfringens* epsilon toxin rapidly decreases membrane barrier permeability of polarized MDCK cells. *Cell Microbiol.*, 2003, **5**, 155-64.
 50. Donelli, G.; Fiorentini, C.; Matarrese, P.; Falzano, L.; Cardines, R.; Mastrantonio, P.; Payne, D.W. & Titball, R.W. Evidence for cytoskeletal changes secondary to plasma membrane functional alterations in the in vitro cell response to *Clostridium perfringens* epsilon-toxin. *Comp. Immunol. Microbiol. Infect. Dis.*, 2003, **26**, 145-56.
 51. Holmes, R.K. Biology and molecular epidemiology of diphtheria toxin and the tox gene. *J. Infect. Dis.*, 2000, **181**(Suppl 1), 156-67.
 52. Clarke, S.C. Bacteria as potential tools in bioterrorism, with an emphasis on bacterial toxins. *Br. J. Biomed. Sci.*, 2005, **62**, 40-46.

53. Haas, L.F. Emil Adolph von Behring (1854-1917) and Shibasaburo Kitasato (1852-1931). *J. Neurol. Neurosurg. Psychiatry*, 2001, **71**, 62.
54. Hildebrand, A.; Pohl, M. & Bhakdi, S. Staphylococcus aureus α -toxin. Dual mechanism of binding to target cells. *J. Biol. Chem.*, 1991, **266**, 17195-7200.
55. Freer, J.H. & Arbuthnott, J.P. Toxins of Staphylococcus aureus. *Pharmacol Ther.*, 1982, **19**, 55-106.
56. Thelestam, M. & Blomqvist, L. Staphylococcal α -toxin-recent advances. *Toxicon*, 1988, **26**, 55-65.
57. Low, D.K.; Freer, J.H.; Arbuthnott, J.P.; Mollby, R. & Wadstrom, T. Consequences of sphingomyelin degradation in erythrocyte ghost membranes by staphylococcal β -toxin (sphingomyelinase C). *Toxicon*, **19**, 12, 279-85.
58. Gyles, C.L. Shiga toxin-producing Escherichia coli: An overview. *J. Anim. Sci.*, 3 Nov. 2006, (Epub ahead of print).
59. Gordjani, N; Sutor, A.H.; Zimmerhackl, L.B. & Brandis, M. Hemolytic uremic syndromes in childhood. *Semin. Thromb. Hemost.*, 1997, **23**, 281-93.
60. Schmid, D.I. & Kohan, D.E. Effect of shigatoxin-1 on arachidonic acid release by human glomerular epithelial cells. *Kidney Int.*, 2001, **60**, 1026-036.
61. O'Brien, A.D.; Tesh, V.L. & Donohue-Rolfe, A. Shiga toxin: Biochemistry, genetics, mode of action, and role in pathogenesis. *Curr. Topics Microbiol. Immunol.*, 1992, **180**, 65-94.
62. Karmali, M.A. Infection by Shiga toxin-producing *Escherichia coli*: An overview. *Mol. Biotechnol.*, 2004, **26**, 117-22.
63. Gransden, W.R.; Damm, M.A. & Anderson, J.D. Further evidence associating hemolytic-uremic syndrome with infection by Verotoxinproducing *Escherichia coli* O157:H7. *J. Infect. Dis.*, 1986, **154**, 522-24.
64. Razzaq, S. Hemolytic uremic syndrome: An emerging health risk. *Am. Fam. Physician*, 2006, **74**, 991-96.
65. Cherla, R.P.; Lee, S.Y. & Tesh, V.L. Shiga toxins and apoptosis. *FEMS Microbiol Lett.*, 2003, **228**, 159-66.
66. Merritt, E.A.; Sarfaty, S.; van den, Akker F.; L'Hoir, C.; Martial, J.A. & Hol, W.G. Crystal structure of cholera toxin B-pentamer bound to receptor GM1 pentasaccharide. *Protein Science.*, 1994, **3**, 166-75.
67. De Haan, L. & Hirst, T.R. Cholera toxin: A paradigm for multi-functional engagement of cellular mechanisms (Review). *Mol. Membr. Biol.*, 2004, **21**, 77-92.
68. Merritt, E.A.; Kuhn, P.; Sarfaty, S.; Erbe, J.L.; Holmes, R.K. & Hol, W.G. The 1.25 Å resolution refinement of the cholera toxin B-pentamer: Evidence of peptide backbone strain at the receptor-binding site. *J. Mol. Biol.*, 1998, **282**, 43-59.
69. Kassis, S.; Hagmann, J.; Fishman, P.H.; Chang, P.P. & Moss, J. Mechanism of action of cholera toxin on intact cells. Generation of A1 peptide and activation of adenylate cyclase. *J. Biol. Chem.*, 1982, **257**, 12148-2152.
70. Enomoto, K. & Gill, D.M. Cholera toxin activation of adenylate cyclase: Roles of nucleoside triphosphates and a macromolecular factor in the ADP ribosylation of the GTP-dependent regulatory component. *J. Biol. Chem.*, 1980, **255**, 1252-228.
71. Helting, T.B.; Zwisler, O. & Wiegandt, H. Structure of tetanus toxin. II. Toxin binding to ganglioside. *J. Biol. Chem.*, 1977, **252**, 194-98.
72. Anderson, M.D.; Fairweather, N.; Charles, I.G.; Emsley, P.; Isaacs, N.W. & MacDermott. Crystallographic characterization of tetanus toxin fragment. *C. J. Mol. Biol.*, 1993, **230**, 673-74.

73. Turton, K.; Chaddock, J.A. & Acharya, K.R. Botulinum and tetanus neurotoxins: structure, function and therapeutic utility. *Trends Biochem. Sci.*, 2002, **27**, 552-58.
74. Gill, D.M. Bacterial toxins: A table of lethal amounts. *Microbiol. Rev.*, 1982, **46**, 86-94.

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