REVIEW PAPER

## Nutritional Requirement at High-altitude with Special Emphasis to Behaviour of Gastro-intestinal Tract and Hormonal Changes

M.B. Anusha, Naveen Shivanna<sup>\*</sup>, and K.R. Anilakumar

Department of Applied Nutrition, Defence Food Research Laboratory, Mysuru - 570 011, India \*E-mail: drnaveen@dfrl.drdo.in

#### ABSTRACT

When people are exposed to the extreme environmental conditions, such as high altitude where there is decrease in temperature and partial pressure of oxygen induces fatigue, insomnia, loss of appetite and increased cardiac output. Hence there is need to improve the appetite through the diet and digestion clout of the individual. In the present review paper the efficiency of digestion is compromised at high altitude is discussed. Also about, Hypoxia, resulting by decreased partial pressure of oxygen can be classified into acute hypoxia and chronic hypoxia based on the exposure time. There is increased formation of reactive oxygen species due to less oxygen available in the air at high altitude which leads to oxidative stress. Lipid peroxidation caused by oxidative stress. Hypoxia is mediated through hypoxia inducible factors which maintain oxygen haemostasis in the body. At high altitude diet rich in carbohydrates have been found to be beneficial as it increases glucose metabolism. Requirement of nutrients such as vitamin A, vitamin E and vitamin C as well as micronutrients such as zinc, iron, selenium, copper and manganese will be required at high altitude. Hypoxia effect on the intestine leads to malabsorption and the lipid storage is stimulated and lipid catabolism is inhibited through  $\beta$ -oxidation.

Keywords: High faltitude; Hypoxia; Oxidative stress; Acute mountain sickness; Hypoxia-inducible factors; Normoxia; Nutrition

#### 1. INTRODUCTION

Humans are able to survive in almost all the environmental extremes of Earth due to their physiological adaptability and/ or by modification of environment itself. Some of the extreme environments are deserts, humid forests, hot humid coastal regions and high altitude (HA). High-altitude presents an extreme environment with hypoxia, cold, high solar radiation as physical stresses beside the psychological stress<sup>1</sup>. HA is defined to begin at 2,400 m above sea level. The air pressure decreases as the altitude increases which leads to hypoxia or oxygen deprivation (Fig. 1).

The human body can adapt to high altitude through shortterm and long-term acclimatisation. Acute mountain sickness (AMS) occurs on short-term exposure to high-altitude due to low partial pressure. AMS can further lead to high altitude pulmonary edema (HAPE) and High altitude cerebral edema (HACE)<sup>2</sup>. Chronic mountain sickness (CMS) occurs after prolong stay at high altitude which leads to polycythemia<sup>3</sup> and hypoxemia<sup>4</sup>. Some of the common problem faced by people ascending high altitude are loss of appetite, fatigue, breathlessness, insomnia, abdominal pain, constipation, nausea, blisters in hands and feet<sup>5</sup> as shown in Fig. 2.

Hypoxia is a condition which occurs on oxygen depletion. Due to the lack of oxygen there is an increase in the breathing rate and sleep cycle is affected<sup>6</sup>. In addition, the heart rate increases and digestive efficiency of food is reduced, as the

Received : 10 March 2017, Revised : 12 April 2017

body suppresses the digestive system in favour of increasing its cardiopulmonary reserves; there is a decrease in the amount of blood flowing to digestive organs and increased blood to the brain, heart and lungs<sup>7</sup>. The glucose is metabolised by liver cells but is not able to utilise it. Associated with the depression of liver function will be a significant decrease in its ability to rid the body of metabolites or conjugate steroids. Very little work has been done on the effect of hypoxia on digestive system in a person ascending high altitude.

### 2. TYPES OF HYPOXIA

Hypoxia is a condition which arises by low oxygen tension (PO<sub>2</sub>) created either by environmental conditions like exposure to high altitude, or by pathological conditions such as chronic obstructive pulmonary disease (COPD), obstructive sleep apnea or severe anemia<sup>8</sup>. The traditional classification of hypoxia has only two subgroups based on empirical observations i.e., chronic and acute hypoxia (Fig. 3). Acute hypoxia response occurs within minutes of exposure to a hypoxic environment i.e., symptoms occur in first few hours–days. Some of the symptoms are hyperventilation, insomnia, fatigue, dizziness and gastrointestinal disturbances. In chronic hypoxia, the term 'chronic' is used to indicate the time interval which ranges over weeks and months. However long term adaptation to hypoxia has been studied in human populations in Himalayas, Ethiopian and Andes over generation<sup>9</sup>.

According to Best and Taylor<sup>10</sup> hypoxia is classified into four main type's i.e., Hypoxic hypoxia, anemic hypoxia, stagnant hypoxia and histotoxic hypoxia as shown in Fig. 4.

Accepted : 15 April 2017, Online published : 12 May 2017



Figure 1. Classification of high altitude and percentage of oxygen available.



Figure 2. Symptoms experienced at high altitude.

*Note:* Symptoms experienced at HA can be classified into two types, i.e. primary symptoms or acute mountain sickness (AMS) such as headache, difficulty in breathing, fatigue, sleeplessness, peripheral edema i.e. swelling of hands, feet and face, rapid pulse rate and decreased food intake. The secondary symptoms are life threatening. High altitude pulmonary edema (HAPE) is accumulation of fluid in the lungs and high altitude cerebral edema (HACE) is swelling of the brain which if not treated leads to coma or death.



Figure 3. Types of hypoxia.

Hypoxic hypoxia is caused when there is low oxygen tension in the inhaled air as a result of which the haemoglobin present in the erythrocyte cannot saturate the fully saturate with oxygen leading to lack of oxygen in arterial blood. This is one of the most serious forms of hypoxia. Anemic hypoxia is a less serious condition than hypoxic hypoxia and it may be caused by an insufficient transport function of the haemoglobin. Stagnant/ Circulatory hypoxia arises when the amount of oxygen reaching the tissue is inadequate. This turn leads to reduced rate of blood circulation thereby allowing accumulation of



Figure 4. Classification of hypoxia based on exposure.

carbon dioxide in the tissue. Histotoxic hypoxia is a condition where there is normal amount of oxygen in the blood and under normal tension but the cells are unable to accept oxygen from the capillaries. It may be produced by any agents that depress cellular respiration.

#### 3. HIGH ALTITUDE AND HYPOXIA

According to the observations made on humans a HA there is an initial weight loss and some of the factors that affects loss of weight are dysbarism, negative nitrogen balance, altered nutrient digestibility, increased water loss, increased energy expenditure, hypophagia, intestinal malabsorption<sup>11</sup> and change in hedonicity and taste perception<sup>12</sup>. There is an increase in metabolic rate at high altitude<sup>7</sup>. There are reports which confirmed by functional magnetic resonance imaging (fMRI) study that neural circuit for food craving on prolonged exposure to HA is reduced<sup>13</sup>.

Exposing the lowlanders to HA is a useful model in getting insight into physiological responses in humans to hypoxia. In normoxic condition, the heart produces an abundant supply of ATP for fat oxidation. While in hypoxic condition, there is a decreased proportion of energy reliance on fat oxidation and the use of carbohydrate increases as the energy metabolism turns anaerobic<sup>14</sup>. Hypoxia under severe condition has shown to stimulate glucose transport across plasma membrane<sup>15</sup>. There are studies which report that high carbohydrate diets are beneficial at high altitude due to its high respiratory coefficient (RQ) compared to protein and fat. Diets high in carbohydrates have shown to enhance glucose metabolism<sup>16</sup>. Fat malabsorption<sup>17</sup> is significant only at altitudes below 5000 m.

At HA there is increased formation of reactive oxygen and nitrogen species (RONS) due to decrease in pressure which leads to increased oxidative damage to macromolecules. The oxidative stress can further be enhanced through physical exercise at high altitude for which supplementation of antioxidant seems to prevent or decrease high altitude associated oxidative stress<sup>18</sup>. On the contrary, there are studies which suggest that supplementation of antioxidant supplementation does not attenuate HA related oxidative stress<sup>19-20</sup>. The human body on ascent to HA oxygen consumption increases to generate energy to meet body requirements. Due to low oxygen availability, reactive oxygen species (ROS) accumulates in mitochondrion as oxygen available is less to be reduced to water. ROS together with nitric oxide (NO) in vasculature combine to form reactive oxygen and nitrogen species (RONS). Exposure to high-altitude also leads to imbalance in the levels of vasoactive modulators which leads to generation of more ROS/RONS. This in turn leads to vascular dysfunction i.e. narrowing of lumen, smooth muscle proliferation and vasoconstriction exaggeration, which aggravates imbalance in vasoactive modulators and ROS/ RONS. Thus, this vicious cycle of oxidative stress goes on until the subject receives medical help (Fig. 5).

#### 4. HYPOXIA PATHWAY

When the level of oxygen in the air is low, the wall of Aorta (carry oxygenated blood) has chemoreceptor which detects oxygen level in the blood. There are sensory nerves



Figure 5. Oxidative stress at high-altitude.

which connect the aorta to the brain stem in medulla oblongata and carry information about the arterial blood. The carotid artery branches into internal and external carotid artery. The internal carotid artery carries blood to the brain while external carotid artery supplies blood to the neck and face. The carotid body receives blood from external carotid artery and detects the oxygen level. Internal carotid artery has chemoreceptors in carotid sinus which is more sensitive than aorta. The carotid sinus and the carotid body together sense the amount of oxygen and send the signal to medulla oblongata. The medulla oblongata contain sensors i.e., cardiac sensors, respiratory sensors and pulmonary vasculature which in increases the sympathetic outflow leading to increased heart rate, increased cardiac output and increased breath rate and constriction of peripheral vessels<sup>21</sup>.

The critical mediators of adaptive responses to hypoxia have been identified as the hypoxia-inducible factors (HIFs), which regulates the expression of genes responsible for growth, vascular development and metabolism. HIF-1 belongs to family of oxygen-sensitive transcription factors<sup>22</sup>. HIF-1 is found in all nucleated cells which are highly conserved and it is regulated by the oxygen available. HIF-1 is a heterodimer, composed of HIF-1 $\alpha$  and HIF-1 $\beta$  (Fig. 6(a)). It belongs to the PER-ARNT-SIM (PAS) subfamily of the basic helix-loophelix (bHLH) family of transcription factor. Under normoxic/ normal condition, HIF-1 dimer does not exist as the HIF-1a produced has a half life of less than 5 min. The HIF-1ß is constitutively present and HIF-1 $\alpha$  level is very low. HIF-1 $\alpha$  is degraded by proteosome system only in the presence of oxygen. In the absence of oxygen it cannot be degraded. HIF-1 $\alpha$  (826) amino acids) at the N-terminal consists of bHLH and PAS for heterodimerisation and DNA binding. It also has oxygen dependent degradation domain (ODDD) which contains two sub

domains N-ODDD and C-ODDD. They have two proline residues i.e. Pro 402 and Pro 564 which are hydroxylated in normoxia. At the C-terminal it has two terminaltransactivation domains (TAD) i.e. N-TAD and C-TAD which are involved in transcriptional activation during hypoxic condition (Fig. 6(b)). The hydroxylation of proline is catalysed by Prolyl hydroxylase domain (PHD) or hypoxia inducible factor -prolyl hydroxylase (HPH). To the hydroxylated proline residue von Hippel-Lindau (pVHL or VHL) tumor suppressor protein binds. VHL protein comprises of E3 ubiquitin ligase, which targets protein for proteosome degradation<sup>23</sup>.

Under hypoxic condition, the HIF-1 $\alpha$  is not degraded. HIF-1 $\alpha$  translocate to the nucleus to form a dimer with HIF-1 $\beta$ . In association with p300/ CBP (cAMP –response element- binding protein), it binds to

hypoxia response elements (HREs) in their upstream regulator region up regulating the expression of HIF-target genes such as vascular endothelial growth factor (VEGF), erythropoietin (EPO), and glucose transporters (GLUT) and key glycolytic enzymes, including hexokinase<sup>24</sup> as shown in Fig. 6(c).

The other pathway that affects the HIF-1 transcriptional activity is through the phosphatidylinositol 3-kinase/AKT pathway which influences HIF-1a levels through transcriptional regulation (in contrast to the proteasome degradation pathway) via the downstream effector mammalian target of rapamycin which inhibits FKBP12 rapamycin associated protein (FRAP)<sup>25</sup>. Genes that are up-regulates the increase of HIF-1 $\alpha$  production and/or stability are cobalt chloride (CoCl<sub>2</sub>), Human epidermal growth factor receptor 2 (HRE2), Insulin like growth factor (IGFR), Epidermal growth factor receptor (EGFR) and Protooncogene tyrosine-protein kinase (SRC). Genes that downregulate factors HIF-1a production and/or stability include Phosphatase and tensin homolog (PTEN) which inhibits Atk (Protein kinase B), Factor inhibiting HIF-1 (FIH-1) which inhibits HIF-1 transcription factor and specific drugs such as LY294002 which inhibits PI3K<sup>26</sup> as shown in Fig. 7.

## 5. NUTRITION AT HIGH ALTITUDE

The human habitation goes up to an altitude of 4300 m and the Indian soldiers are deployed to an altitude of 5800 m for fixed tenure. High altitude presents physical and psychological stress. The availability of drinking water is scarce and at the high altitude the vegetation is sparse. The boiling point of water is decreased due to reduced barometric pressure there by making preparation of food difficult. Studies have reported that there is reduction in meal size with increase in meal frequency and rapid satiety<sup>27</sup>. The diet rich carbohydrate are found to be beneficial at high altitude as they have a respiratory co-efficient



Figure 6. (a) Hypoxia inducible factor 1 (HIF-1) (b) Structure of HIF-1α (c) Effect of hypoxia.



Figure 7. Regulation of HIF-1a.

around 1 compared to fat (0.7) and protein (0.8-0.9). The diet rich in carbohydrate are found to enhance glucose metabolism and it serves as fuel for thermogenesis. The intake of food

may decrease due to anorexia but it does not affect absorption<sup>28</sup>. The diet rich in fat have been observed to increase cold endurance and tolerance in experimental animals<sup>29</sup>. At the initial stages of acclimatisation there is weight loss i.e. loss of body water due to decrease in food intake which causes a drastic reduction in lean body mass<sup>30-32</sup> and reduction of body fat due to the mobilisation of fatty acids from triglycerides pool of adipose tissue and it is measured by reduction in skin fold thickness<sup>33</sup>.

Exposure to altitude causes hypohydration, caused by increased diuresis and decreased water intake. Negative nitrogen balance have been reported at high altitude but this report is not reliable as the calorie intake was less<sup>34</sup>. To achieve a positive nitrogen balance the minimum intake of protein should be 0.96 g/kg for exercising men but there is increased excretion of protein and nitrogen at high altitude hypoxia<sup>35</sup>. During prolonged stay at an altitude of 3500 m - 4000 m, positive nitrogen balance was maintained with decreased amino acid excretion. The entry of amino acids into neural tissues and brain is modulated by the relative concentration of specific amino acids in the blood. So it would be ideal to design food with high tryptophan food to alleviate sleep disturbance, higher glutamic acid food for deterioration in cognitive function and higher phenylalanine/ tvrosine hindering for mood depression for high performance at high altitude<sup>36</sup>. Supplementation of branched chain amino acids such as leucine, isoleucine and valine have been reported to prevent muscle loss during trekking at high altitude<sup>37</sup>. In order to reduce lipid peroxidation caused by oxidative stress the supply of nutrients such as vitamin A, vitamin E and vitamin C as well as micronutrients such as zinc, iron, selenium, copper and manganese may be required in a greater amount at HA<sup>1</sup>.

# 6. GASTROINTESTINAL TRACT AT HIGH ALTITUDE

The primary functions of the gastrointestinal tract are

the absorption of ingested nutrients, removal of waste, fluid homeostasis and protection from pathogens<sup>38</sup>. The barrier function and the absorptive function of the epithelium of intestine is regulated based on oxygen availability as regulators of hypoxia i.e PHD and factor inhibiting HIF1 (FIH1) are expressed in the intestinal mucosal tissue<sup>39</sup>. At high altitude, as the core body temperature decreases the gastrointestinal smooth muscle motility decreases which further leads to distension of colon, reduced gastrointestinal secretion and free acid production. Hypothermic condition leads to decreased splanchnic blood flow and causes a catecholamine-induced vasoconstriction of blood vessels. As the catecholamine secretion decreases leading to vasodialation it results in reperfusion and extravasation of blood. The reperfusion and associated changes alter the gastric mucosa's protective mechanism, resulting in cellular damage induced by hydrochloric acid. The liver cells cannot utilise glucose but they continue to metabolise them. The depression of liver function leads to decrease in its ability to rid the body of metabolites, drugs, or conjugate steroids<sup>40</sup>.

There are reports which suggest that hypoxia effect on the intestine which leads to malabsorption<sup>41</sup> and there is no change in fat utilisation<sup>42</sup> up to an altitude of 4700 m. Up to an altitude of 3500 m the concentration of gastric acid, and total acid output, is reduced significantly in basal conditions. However; there is no change in maximal levels, using pentagastrin stimulation. The gastrointestinal function is not altered in terms of digestion and absorption of food components<sup>43</sup>. Reports on the gastrointestinal function on an altitude above 5000 m are sparse.

### 7. HORMONAL LEVEL AT HIGH ALTITUDE

The hypoxic effect at cellular level are thought to be mediated by the hypoxia inducible factor-1 (HIF-1) pathway<sup>44</sup>, and hypoxia-response elements (HRE) have been identified for erythropoietin (EPO), vascular endothelial growth factor-A (VEGF) and leptin<sup>45</sup>. Hypoxia-inducible factor is regulated through oxygen-dependent proteasomal degradation and it responds to variations in oxygen availability. VEGF expression in skeletal muscle increases after exercise and it is a potent stimulus for angiogenesis. Its expression at HA is crucial to promote muscle capillarity during training<sup>46</sup>. Leptin is produced by adipose tissue and is a major regulator of satiety and food intake47-48. Studies in humans at HA or mice exposed to hypobaric hypoxia have reported increased, unchanged or decreased leptin levels<sup>49</sup>. Little is known about changes in leptin after acute exercise at HA45. Literature on ghrelin in high altitude is sparse and inconsistent, with reports of both decreased ghrelin levels and no change in ghrelin at high altitude50. Studies report increased level of cholecystokinin (CCK) at high altitude<sup>51</sup>.

In most studies there have been increased level of thyroid hormones at high altitude, although report suggest that TSH secretion is not modified but the mechanisms involved in this process are still unclear<sup>52</sup>. The insulin level is increased during elevated glucose circulation and it functions to suppress the hepatic glucose output. The insulin level was found to be elevated in hypoxic condition<sup>53</sup>. Glucagon levels have been reported to be unaltered at HA<sup>54</sup>. Corticosteroids, have a well documented immunosuppressive effect which are released during cold stress, hypothermia, or both<sup>55</sup>.



Figure 8. Enzymes at hypoxic condition.

## 8. ENZYMES AT HIGH-ALTITUDE

Hypoxia, in general results in an increased level of blood haemoglobin and increased hematocrit values leading to increased oxygen capacity. As oxidative pathways are limited at altitude, there is a shift toward anaerobic energy sources. At any given work level, lactic acid production is higher than at sea level. The HIF-1 have been proved to (a) induce a variety of glycolytic enzymes and glucose transporters such as aldolase A and pyruvate kinase M, which help produce energy in hypoxic condition<sup>56-58</sup> (b) reduce mitochondrial oxygen consumption by activating pyruvate dehydrogenase kinase I(PDK1) and halts citric acid cycle<sup>59</sup> as shown in Fig. 8. HIF 1a encodes PDK1 which suppress oxygen consumption<sup>60</sup>. I Disaccharidase activity (lactase, at least) is HIF-responsive, which may protect carbohydrate absorption at moderate altitudes. In hypoxic condition the level of 2, 3-bisphosphoglycerate is found to be increased due to anaerobic glycolysis. At hypoxic condition the lipid storage is stimulated and lipid catabolism is inhibited through  $\beta$ -oxidation<sup>61</sup>.

## REFERENCES

- Selvamurthy, W.&Singh, S.N. Nutritional requirements for human adaptation in extreme environments. *Proceedings-Indian National Science Academy Part B*, 2003, 69(4), 485-506.
- Imray, C.; Wright, A.; Subudhi, A. & Roach, R. Acute mountain sickness: pathophysiology, prevention, and treatment. *Progress Cardiovascular Diseases*, 2010, 52(6), 467-484.

doi: 10.1016/j.pcad.2010.02.003

- Jefferson, J.A.; Escudero, E.; Hurtado, M.E.; Pando, J.; Tapia, R.; Swenson, E.R. & Johnson, R.J. Excessive erythrocytosis, chronic mountain sickness, and serum cobalt levels. *The Lancet*, 2002, **359**(9304), 407-408. doi: 10.1016/S0140-6736(02)07594-3
- Leon-Velarde, F.; Villafuerte, F.C. & Richalet, J.P. Chronic mountain sickness and the heart. *Progress Cardiovascular Diseases*, 2010, **52**(6), 540-549. doi: 10.1016/j.pcad.2010.02.012
- Wheatley, K.; Creed, M. & Mellor, A. Haematological changes at altitude. J. Royal Army Med. Corps, 2011, 157(1), 38-42. doi: 10.1136/jramc-157-01-07
- San, T.; Polat, S.; Cingi, C.; Eskiizmir, G.; Oghan, F. & Cakir, B. Effects of high altitude on sleep and respiratory system and theirs adaptations. *Sci. World J.*, 2013. doi: 10.1155/2013/241569
- 7. Westerterp, K.R. Energy and water balance at high altitude. *Physiology*, 2001, **16**(3), 134-137.
- Deldicque, L. & Francaux, M. Acute vs. chronic hypoxia: What are the consequences for skeletal muscle mass?. *Cellular Mol. Exercise Physiol.*, 2013, 2(1), e5. doi: 10.7457/cmep.v2i1.e5
- Xu, K. & LaManna, J.C. Chronic hypoxia and the cerebral circulation. J. Appl. Physiol., 2006, 100(2), 725-730. doi: 10.1152/japplphysiol.00940.2005
- Best, C. & Taylor, N. The physiological basis of medical practice, Ed. 3<sup>rd</sup>, Baltimore, Williams and Wilkins

Company, 1943, 591.

- Boyer, S.J. & Blume, F.D. Weight loss and changes in body composition at high altitude. *J. Appl. Physiol.*, 1984, 57(5), 1580-1585.
- Singh, D.S.N.; Sridharan, K. & Selvamurthy, W. Human nutrition at high altitude. *Nutrition Bulletin*, 1999, 24(4), 195-202.

doi: 10.1111/j.1467-3010.1999.tb00909.x

- Yan, X.; Zhang, J.; Gong, Q. & Weng, X. Appetite at high altitude: an fMRI study on the impact of prolonged high-altitude residence on gustatory neural processing. *Experimental Brain Res.*, 2011, 209(4), 495-499. doi: 10.1007/s00221-010-2516-8
- Purshottam, T.; Kaveeshwar, U. & Brahmachari, H. D. Changes in tissue glycogen stores of rats under acute and chronic hypoxia and their relationship to hypoxia tolerance. *Aviation, Space, Environmental Med.*, 1977, 48(4), 351-355.
- Fujii, N.; Jessen, N. & Goodyear, L.J. AMP-activated protein kinase and the regulation of glucose transport. *Am. J. Physiology-Endocrinology Metabolism*, 2006, **291**(5), E867-E877.

doi: 10.1152/ajpendo.00207.2006

- 16. Kayser, B. Nutrition and high altitude exposure. *Int. J. Sports Med.*, 1992, **13**(S 1), S129-S132.
- 17. Anand, A.C.; Sashindran, V.K., & Mohan, L. Gastrointestinal problems at high altitude. *Tropical Gastroenterology: Official J. Digestive Diseases Foundation*, 2005, **27**(4), 147-153.
- 18. Bakonyi, T., & Radak, Z. High altitude and free radicals. J. *Sports Sci. Med.*, 2004, **3**(2), 64-69.
- Subudhi, A.W.; Jacobs, K.A.; Hagobian, T.A.; Fattor, J.A.; Fulco, C.S.; Muza, S.R. & Friedlander, A.L. Antioxidant supplementation does not attenuate oxidative stress at high altitude. *Aviation, Space, Environmental Medicine*, 2004, **75**(10), 881-888.
- 20. Pandey, P. & Pasha, M.A.Q. Oxidative stress at high altitude: genotype-phenotype correlations. *Adv. Genomics Genet.*, 2014, **4**, 29-43.
- 21. De Geest, H.; Levy, M.N. & Zieske, H. Carotid chemoreceptor stimulation and ventricular performance. *Am. J. Physiology--Legacy Content*, 1965, **209**(3), 564-570.
- Shimoda, L.A. 55<sup>th</sup> Bowditch lecture: Effects of chronic hypoxia on the pulmonary circulation: role of HIF-1. *J. Applied Physiology*, 2012, **113**(9), 1343-1352. doi: 10.1152/japplphysiol.00843.2012
- Weidemann, A., & Johnson, R.S. Biology of HIF-1α. *Cell Death Differentiation*, 2008, **15**(4), 621-627. doi: 10.1038/cdd.2008.12
- Thomas, R. & Kim, M.H. Targeting the hypoxia inducible factor pathway with mitochondrial uncouplers. *Molecular Cellular Biochem.*, 2007, 296(1-2), 35-44. doi: 10.1007/s11010-006-9295-3
- Zhong, H.; Chiles, K.; Feldser, D.; Laughner, E.; Hanrahan, C.; Georgescu, M.M. & Semenza, G.L. Modulation of hypoxia-inducible factor 1α expression by the epidermal growth factor/ phosphatidylinositol 3-kinase/PTEN/

AKT/FRAP pathway in human prostate cancer cells: implications for tumor angiogenesis and therapeutics. *Cancer Research*, 2000, **60**(6), 1541-1545.

 Serganova, I.; Humm, J.; Ling, C. & Blasberg, R. Tumor hypoxia imaging. *Clinical Cancer Res.*, 2006, 12(18), 5260-5264.

doi: 10.1158/1078-0432.CCR-06-0517

- Westerterp-Plantenga, M.S.; Westerterp, K.R.; Rubbens, M.; Verwegen, C.R.; Richelet, J.P. & Gardette, B. Appetite at "high altitude"[Operation Everest III (Comex-'97)]: a simulated ascent of Mount Everest. *J. Appl. Physiol.*,1999, **87**(1), 391-399.
- 28. Kayser, B.; Acheson, K.; Decombaz, J.; Fern, E. & Cerretelli, P. Protein absorption and energy digestibility at high altitude. *J. Appl. Physiol.*,1992, **73**, 2425-2431
- 29. Templeton, H.A. & Ershoff, B.H. Comparative effect of carbohydrate, protein and fat when fed as single foods on the survival time of rats under conditions of accelerated metabolism. *Am. J. Physiol*, 1949, **159**, 33-39.
- Guilland, J.C. & Klepping, J. Nutritional alterations at high altitude in man. *European J. Appl. Phy. Occupational Physiol.*, 1985, 54(5), 517-523. doi: 10.1007/BF00422963
- Butterfield, G.E.; Gates, J.; Fleming, S.; Brooks, G.A.; Sutton, J.R. & Reeves, J.T. Increased energy intake minimizes weight loss in men at high altitude. *J. Appl. Physiol.*, 1992, **72**, 1741-1748
- Westerterp, K.R.; Kayser, B.E.; N.G.T.; Wouters, L.O.E.K.; Le Trong, J.L. & Richalet, J.P. Energy balance at high altitude of 6,542 m. *J. Appl. Physiol.*, 1994, 77(2), 862-866.
- 33. Gill, M.B. & Pugh, LGCE. Basal metabolism and respiration in man living at 5,800m (19,000 ft). *J. Appl. Physiol.*, 1964, **19**, 949-954
- Consalazio, C.; Johnson, H.L.; Krzywicki, H.J. & Daws, T.A. Metabolic aspects of acute altitude exposure (4300 m) in adequately nourished humans. *Am. J. Clin. Nutr.*,1968, 25, 23-29.
- Rennie, D.; Frayser, R.; Gray, G. & Houston, C. Urine and plasma proteins in men at 5400 m. *J. Appl. Physiol.*,1972, 32, 369-373.
- Srivastava, K.K. & Kumar, R. Human nutrition in cold and high terrestrial altitudes. *Int. J. Biometeorol.*, 1992, 36(1), 10-13.
  - doi: 10.1007/BF01208728
- Schena, F.; Guerrini, F.; Tregnaghi, P. & Kayser, B. Branched-chain amino acid supplementation during trekking at high altitude. *European J. Appl. Phys. Occupational Phy.*, 1992, **65**(5), 394-398. doi: 10.1007/BF00243503
- Taylor, C.T. & Colgan, S.P. Hypoxia and gastrointestinal disease. J. Molecular Med., 2007, 85(12), 1295-1300. doi: 10.1007/s00109-007-0277-z
- Colgan, S.P. & Taylor, C.T. Hypoxia: an alarm signal during intestinal inflammation. *Nature Rev. Gastroenterology Hepatology*, 2010, 7(5), 281-287. doi: 10.1038/nrgastro.2010.39
- 40. Pozos, R.S. & Danzl, D. Human physiological responses

to cold stress and hypothermia. *Med. Aspects Harsh Environments*, 2001, 1, 351-382.

- Pittman, J.G. & Cohen, P. The pathogenesis of cardiac cachexia. *New Eng. J. Med.*, 1964, **271**(9), 453-460. doi: 10.1056/NEJM196408272710908
- 42. Rai, R.M.; Malhotra, M.S.; Dimri, G.P. & Sampat Kumar, T. Utilization of different quantities of fat at high altitudes. *Am. J. Clin. Nutr.*, 1975, **28**, 242-245
- Sridharan, K.; Malhotra, M.S.; Upadhayay, T.N.; Grover, S.K. & Dua, G.L. Changes in gastro-intestinal function in humans at an altitude of 3,500 m. *Europ. J. Appl. Phys. Occupational Phys.*, 1982, **50**(1), 145-154. doi: 10.1007/BF00952253
- West, J.B. High-altitude medicine. Am. J. Respiratory Critical Care Med., 2012, 186(12), 1229-1237. doi: 10.1164/rccm.201207-1323CI
- 45. Morici, G.; Bonanno, A.; Licciardi, A.; Valli, G.; Passino, C.; Bonardi, D. & Bonsignore, M.R. Plasma leptin and vascular endothelial growth factor (VEGF) in normal subjects at high altitude (5050 m). *Archives Phy. Biochem.*, 2013, **119**(5), 219-224. doi: 10.3109/13813455.2013.814679
- Wagner, P.D. The critical role of VEGF in skeletal muscle angiogenesis and blood flow. *Biochem. Society Trans.*, 2011, **39**(6), 1556-1559. doi: 10.1042/BST20110646
- Kelesidis, T.; Kelesidis, I.; Chou, S. & Mantzoros, C.S. Narrative review: the role of leptin in human physiology: emerging clinical applications. *Annals Internal Med.*, 2010, **152**(2), 93-100. doi: 10.7326/0003-4819-152-2-201001190-00008
- Yingzhong, Y.; Droma, Y.; Rili, G. & Kubo, K. Regulation of body weight by leptin, with special reference to hypoxiainduced regulation. *Internal Medicine*, 2006, 45(16), 941-94.

doi: 10.2169/internalmedicine.45.1733 6.

- Sierra-Johnson, J.; Romero-Corral, A.; Somers, V.K. & Johnson, B.D. Effect of altitude on leptin levels, does it go up or down?. *J. Appl. Phys.*, 2008, **105**(5), 1684-1685. doi: 10.1152/japplphysiol.90679.2008
- Shukla, V.; Singh, S.N.; Vats, P.; Singh, V. K.; Singh, S.B. & Banerjee, P.K. Ghrelin and leptin levels of sojourners and acclimatized lowlanders at high altitude. *Nutritional Neurosci.*, 2013, 8(3), 161-165 doi: 10.1080/10284150500132823
- 51. Bailey, D.M.; Davies, B.; Milledge, J.S.; Richards, M.; Williams, S.R.P.; Jordinson, M. & Calam, J. Elevated plasma cholecystokinin at high altitude: metabolic implications for the anorexia of acute mountain sickness. *High Altitude Med. Biol.*, 2000, 1(1), 9-23. doi: 10.1089/152702900320649
- Barnholt, K.E.; Hoffman, A.R.; Rock, P.B.; Muza, S.R.; Fulco, C.S.; Braun, B. & Friedlander, A.L. Endocrine responses to acute and chronic high-altitude exposure (4,300 meters): modulating effects of caloric restriction. *Am. J. Physiology-Endocrinol. Metabolism*, 2006, **290**(6), E1078-E1088. doi: 10.1152/ajpendo.00449.2005

- Chen, C.H.; Liu, Y.F.; Lee, S.D.; Huang, C.Y.; Lee, W. C.; Tsai, Y.L. & Kuo, C.H. Altitude hypoxia increases glucose uptake in human heart. *High Altitude Med. Biol.*, 2009, **10**(1), 83-86. doi: 10.1089/ham.2008.1064
- 54. Kullmer, T.; Gabriel, H.; Jungmann, E.; Haak, T.; Morbitzer, D.; Usadel, K.H. & Kindermann, W. Increase of serum insulin and stable c-peptide concentrations with exhaustive incremental graded exercise during acute hypoxia in sedentary subjects. *Experimental Clinical Endocrinology Diabetes*, 1995, **103**(03), 156-161. doi: 10.1055/s-0029-1211344
- 55. Abbas, A.K.; Lichtman, A.H. & Pober, J.S. Antigen processing and presentation to T lymphocytes. *Cellular Molecular Immunol.*, 2003, 115-137.
- 56. Velasco, A.; Vongpatanasin, W. & Levine, B.D. Treating hypertension at high altitude: the quest for a magic bullet continues. *Eur. Heart J.*, 2014, **35**(44), 3083-4. doi: 10.1093/eurheartj/ehu366.
- 57. Carmeliet, P.; Dor, Y.; Herbert, J.M.; Fukumura, D.; Brusselmans, K.; Dewerchin, M. & Koch, C.J. Role of HIF-1 $\alpha$  in hypoxia-mediated apoptosis, cell proliferation and tumour angiogenesis. *Nature*, 1998, **394**(6692), 485-490.

doi: 10.1038/28867

- Vaupel, P. The role of hypoxia-induced factors in tumor progression. *The Oncologist*, 2004, 9(Suppl 5), 10-17. doi: 10.1634/theoncologist.9-90005-10
- Papandreou, I.; Cairns, R.A.; Fontana, L.; Lim, A.L.; & Denko, N.C. HIF-1 mediates adaptation to hypoxia by actively downregulating mitochondrial oxygen consumption. *Cell Metabolism*, 2006, 3(3), 187-197. doi: 10.1016/j.cmet.2006.01.012
- 60. Simon, M.C. Coming up for air: HIF-1 and mitochondrial oxygen consumption. *Cell Metabolism*, 2006, **3**(3), 150-151.

doi: 10.1016/j.cmet.2006.02.007

 Boström, P.; Magnusson, B.; Svensson, P.A.; Wiklund, O.; Borén, J.; Carlsson, L.M. & Hultén, L.M. Hypoxia converts human macrophages into triglyceride-loaded foam cells. *Arteriosclerosis, Thrombosis, Vascular Biology*, 2006, 26(8), 1871-18. doi: 10.1161/01.ATV.0000229665.78997.0b

#### CONTRIBUTORS

**Ms M.B. Anusha** pursuing her PhD at Applied Nutrition Division, Defence Food Research Laboratory, Defence Research and Development Organisation, Mysuru.

Contributed in writing the manuscript, Images for designing drawings for explanations, etc.

**Dr Naveen Shivanna**, received PhD (Biochemistry) from Mysuru University, in 2008, Presently working in the Applied Nutrition Department at Defence Food Research Laboratory, Mysuru. He has contributed for the development and evaluation of functional food to alleviate hyper cholesterolemic, fatigue, and stress and also isolated anti-diabetic and hypo-lipidemic novel compound from Decalepis. He is a recipient of *DRDO Technology Spinoff Award-2011, DRDO Technology Group Award-2010 and 2015.* 

Contributed in editing and formatting the manuscript.

**Dr K.R. Anilakumar** currently working as Scientist 'F' and Head, Applied Nutrition Division, Defence Food Research Laboratory, Mysuru. He is a recipient of *DRDO Laboratory Scientist of the Year Award-2006, DRDO Technology Group Award-2007 and 2015, DRDO Defence Technology spin-off Award-2011 and DRDO National Science Day Oration Award-2012.* Contributed in editing and finalising the manuscript.