

REVIEW ARTICLE

Role of Growth Factor Signaling in Cancer

Richa Shrivastava[#], and Smrati Bhadauria^{*}[#]Division of Toxicology, Central Drug Research Institute, Lucknow, 226 031, India^{*}Academy of Scientific and Innovative Research, New Delhi - 110 025, India^{*}E-mail: smraticdri@gmail.com

ABSTRACT

Growth factors may be defined as any group of protein that stimulate the growth of specific tissues and play an important role in promoting cellular differentiation and cellular division. Growth factors impart one of the important hallmark of cancer i.e sustaining proliferative signaling. They may act through paracrine, autocrine and endocrine signaling to effect growth and proliferation of cancer cells. They may act through various signaling cascades like MAPK, PI3K/AKT, JAK/STAT etc to activate their downstream mediators affecting various pathological and physiological functions. Abrupt signaling patterns of growth factors can induce oncogenic transformations. An enhanced understanding of these pathways can help targeting neoplastic transformation at an early stage. This review summarizes various mechanisms for targeted therapeutics against growth factor in cancer and their future prospective.

Keywords: Growth factors, cancer, paracrine, autocrine, signaling, ligands, kinases, inhibitors

1. INTRODUCTION

Growth factors may be defined as any group of protein that stimulate the growth of specific tissues and play an important role in promoting cellular differentiation and cellular division¹. Growth factors generally act through paracrine and autocrine signaling. Contrary to original belief, they may also act in endocrine manner. Among these autocrine mechanism is thought to play a significant role in growth of cancer cells^{2,3}. Growth factors have diverse mode of action but mostly act through tyrosine kinase receptor pathway. Tyrosine kinase receptors are membrane bound complexes having intrinsic kinase activity on cytoplasmic domain. Upon binding of specific growth factors (ligand) their kinase activity is activated and they phosphorylate their downstream targets on their tyrosine and serine residues to recruit other molecules into signaling cascades³.

1.1 Growth Factors: Role in Imparting an Important Hallmark of Cancer

Cancer is defined as a cell that grows uncontrollably and hence sustaining chronic proliferation is the most important characteristic of cancer⁵. This excellent proliferative property is imparted largely by growth factors.

Literature suggests link between oncogenes and growth factors⁶:

1. Protooncogene *C-sis* codes for B-chain of PDGF⁷
2. *C-erb* codes for EGF receptor⁸

3. *C-fms* oncogene is similar to CSF-1⁹.

Secondly, evidences suggest that growth factors can increase transcription of certain proto-oncogene (*myc* and *fos*)¹⁰. Cancer cells may produce growth factors themselves or may send signals to stimulate normal cells to secrete growth factors⁵.

2. CLASSIFICATION OF GROWTH FACTORS

Growth factors are generally classified in following classes⁴:

2.1 Platelet Derived Growth Factor Family

Platelet derived growth factor (PDGF) is initially released from alpha-granules of platelets and act as a chemoattractant for fibroblasts and as mitogen for these

Table 1. Growth factors are generally classified

S No.	Growth factors
1.	Platelet derived growth factor family
2.	Vascular endothelial growth factor family
3.	Epidermal growth factor family
4.	Fibroblast growth factor family
5.	Transforming growth factor-B family
6.	The Insulin family
7.	Hepatocyte growth factor family
8.	Neurotrophin family
9.	The Ephrin family
10.	Angiopoietins

cells¹¹. PDGF stimulates production of collagenase by fibroblasts causing remodeling of matrix required for tissue repair¹². It is also released from activated macrophages¹³. Platelet derived growth factor (PDGF) family of growth factor consists of 5 different disulphide linked dimers PDGF-AA, PDGF-BB, PDGF-AB, PDGF-CC and PDGF-DD that act via 2 receptors PDGFR α and PDGFR β ¹⁹. Platelet derived growth factor receptors (PDGFR) are receptors with intrinsic tyrosine kinase activity that regulates several functions in normal cells¹⁴. PDGFR play a role in development of lungs, heart, CNS and kidney¹⁵. In addition to physiological functions, PDGF play pathological roles in disease such as atherosclerosis¹⁶, glomerulonephritis¹⁷ and cancer¹⁸.

In context of cancer, PDGF help in blood vessel formation, cell migration and metastasis. Overexpression of V-sis oncogene product PDGF enhances transformation¹⁹. PDGF signaling is critical for cell-cell communication as PDGF ligands are expressed and secreted by epithelial or endothelial cells to recruit and activate PDGF receptors in stromal components such as smooth muscle cells, pericytes and fibroblasts. Cancer cells express PDGF ligands and its cognate receptors, hence inducing both autocrine and paracrine signaling which helps in cancer progression¹⁸.

Expression of PDGFR α has been reported in breast carcinoma. According to clinical studies on human breast cancer samples 39.2% of invasive ductal carcinoma PDGFR α is present which attributes aggressiveness as it metastasizes to lymph nodes and shows HER2 and Bcl2 expression¹⁹. Simultaneous expression of PDGF-A and PDGFR- α in epithelium, stroma and endothelium of invasive breast carcinoma suggests possibility of autocrine and paracrine signaling which promotes angiogenesis. Hence PDGF α has emerged as a potential target of breast cancer²⁰. PDGFR β is upregulated and activated in prostate cancer of which it is overexpressed in 88% of primary prostate cancer and 80% of metastasized bone lesions²¹. PDGF-D was identified as a ligand for PDGFR β and the serine protease matrilysin as its extracellular proteolytic activator in prostate cancer²². PDGF-BB isoform promotes growth of human oesophageal carcinoma cell lines and prevents apoptosis of cancer cells²³. Additionally overexpression of PDGFR β is associated with tissues of esophageal cancer²⁴. PDGF-C and PDGF-D ligands play a role in development of brain tumor and PDGF autocrine signaling regulates survival and mitogenic pathway in glioblastoma¹⁸. In coordination with these studies it has also been found that antitumor effect of chemotherapeutic agents can be enhanced by inhibiting PDGF receptor signaling in tumor stroma.

2.2 Vascular Endothelial Growth Factor Family

Human vascular endothelial growth factor (VEGF) family consists of VEGF-A, VEGF-B, VEGF-C, VEGF-D and Placental Growth factor²⁵. There are 3 receptors which are regulated by protein kinase for VEGF family of ligands: VEGFR1, VEGFR2, VEGFR3. And two non-enzymatic receptors: Neuropilin1 and Neuropilin-2²⁶. VEGF is secreted by any cell that encounters hypoxia²⁷. VEGF acts

as a mitogen thereby being important survival factor for endothelial cells and monocyte motility²⁸. VEGF changes permeability of endothelial cells by causing injury to help angiogenesis²⁹.

Major factors regulating VEGF includes growth factors, local environmental hypoxia, hormones and cytokines³⁰. The key regulator of hypoxia induced angiogenesis is transcription factor Hypoxia-induced-Factor (HIF-1)³¹. It was very early proposed that inhibiting angiogenesis can be effective antitumor strategy because tumor growth required for blood vessel formation³². VEGF mRNA is expressed in neoplastic cells where as endothelial cells express VEGFR1 and VEGFR2 mRNA and proteins³³. The increase in blood vessel formation helps tumor to gain necessary oxygen and nutrient. Tumor angiogenesis is a hall mark of cancer which supports tumor growth and metastasis³⁴.

VEGFA binds to progenitor cells which express VEGFR1 and induce suppression of NF- κ B activation and signal transduction pathway³⁵. In Breast cancer, VEGF-A expression is associated with increase in microvessel density³⁶. Clinical trials confirm that VEGF expression is an indicator for relapse-free survival^{32,37}. It has also been found that VEGF is essential for initial subcutaneous growth of breast carcinoma cells³⁸. Recent studies also suggest that upregulated VEGF-A is associated with poor prognosis in non-small cell lung carcinoma³⁹. Low VEGF-D to VEGF-C ratio in tumors correlates with both lymphnode metastasis and lymphatic invasion by cancer cell⁴⁰. In colorectal cancer, VEGFA expression is stepwise upregulated for MVD and VEGF-C correlates with extent of lymph node metastasis^{36,40}. In prostate cancer VEGFA expression is reported to play a role in advancing focal carcinoma, invasion and metastasis³².

Because of above vital roles VEGF plays in cancer via upregulating angiogenesis there is lot of focus on targeting VEGF for advanced anti-cancer strategies. Antibody based strategies to target VEGF are extensively being studied. Currently more than 20 monoclonal antibodies are approved by FDA for therapeutic use⁴¹. Example:

1. Monoclonal antibodies
 - a. Rituximab against CD20 for non-hodgkin lymphoma⁴²
 - b. Trastuzumab for breast cancer⁴³
 - c. Cetuximab for metastatic colorectal cancer and head and neck cancer⁴⁴
2. Antibodies with fusion protein
 - a. Bevacizumab for metastatic colorectal cancer and non-small cell lung cancer and metastatic breast cancer⁴⁵.

2.3 Epidermal Growth Factor Family

Epidermal growth factor family (EGF) is a complex network that modulates growth of cells. EGF is released by cells and then either by autocrine signaling i.e. stimulates its own growth or paracrine signaling i.e. stimulate growth of neighboring cells⁴⁶. Ligands known to bind to EGFR are Epidermal growth factor (EGF), Transforming Growth Factor- α (TGF- α), amphiregulin, heparin-binding EGF-

like growth factor, Betacellulin and Epiregulin⁴⁷. EGFR are Receptor tyrosine kinases and they belong to ErbB family which consists of Erb-1 (EGFR), ErbB-2 (HER2 or Neu), ErbB-3, Erb4⁴⁸

EGF has been known to be mitogenic for mesenchymal and epithelial cells⁴⁹. EGF stimulus to normal cells causes them to transform into neoplastic cells by increasing the level of phosphotyrosine in proteins⁵⁰ and increase in sugar and aminoacid metabolism. Expression of *c-fos* and *c-myc* is upregulated by EGF⁵¹. EGF has also been found to play a vital role in viral carcinogenesis as it enhances viral transformation of cells⁵². Chemical carcinogenesis of methylcholanthrene in skin is enhanced by EGF⁵³. EGF phosphorylates tyrosine residues of src, erb, abl, yes, fgr, ros, fes (fps) and fms⁵⁴.

Human breast cancer cell line like MCF-7 was found to synthesize and secrete EGF-like immunoreactive factor in culture⁵⁵. Prostate cancer cell lines DU-145 were also found to secrete EGF-like polypeptides into serum-free culture medium⁵⁶. Overexpression of TGF- α and EGFR by carcinomas are correlated to chemotherapeutic resistance, metastasis, and poor prognosis⁵⁷. EGF-like peptides and erbB receptor induce transformation of cells in transgenic mice models. Evidences support that expression of rodent p-185c-neu and EGFR in Nh-373 cells is necessary for neoplastic transformation⁵⁸. EGF overexpression has been found in metastatic breast cancer and aggressive form of uterine cancer⁵⁹. Erb3 is associated in human mammary cancers. ErbB1/EGFR cases have accelerated intraepithelial proliferation^{48,60}.

Therapeutic interventions against EGF includessuramine which inhibits the binding of EGF to its recetor⁶¹ and gefinitib which targets ErbB1 tyrosine kinase activity which causes inhibition of autophosphorylation⁶². Cetuximab, a monoclonal antibody which prevents ErbB1 signaling by binding to the ligand binding domain is now being clinically used⁶³.

2.4 Fibroblast Growth Factor Family

In humans, FGF family has 23 polypeptide encoding genes. Fibroblast growth factor family includes FGF1 i.e. acidic FGF, FGF-2 i.e. basic FGF and FGF-6 and FGF8. FGF receptors comprises of FGFR1-4 which are involved in both autocrine and paracrine signaling⁶⁴. FGF are mitogenic for epithelial and mesenchymal cells. FGFs were first angiogenic factors to be indentified⁶⁵ and hence posses high angiogenic activity⁶⁶ in addition to enhancing motility and invasiveness of cells⁶⁷.

FGF signaling is a must for sustained self-renewal and pluripotency of human embryonic stem cells (HESCs)⁶⁸. Duringhaematopoiesis,FGFstimulatesgrowthofprogenitor cells⁶⁹. FGF-2 stimulus leads to neoplastic transformation of cells⁷⁰. Elevated levels of FGF2 in micro-environment of metastatic prostate cancer help cells to evade the anti-proliferative effect of chemotherapy⁷¹. Myeloma-associated oncogene FGFR3 is upregulated in cancer cells from patients with Chronic Myeloid leukemia (CML)^{67,72}.

Transgenic mice with both epithelial FGF3

overexpression and FGFR1 activation lead to epithelial proliferation and invasion of lesions⁷³. FGFR2IIIB knockout mice fail to develop branching morphogenesis in breast which suggests that FGFR2 expression predisposes to breast cancer⁷⁴. Single Nucleotide Polymorphisms occurring in FGFR2 and FGFR4 have been linked to their critical role in pathogenesis of breast cancer⁷⁵. FGFR2 amplification occurs both in breast cancer and gastric cancer and is associated with poor prognosis⁷⁶. Evidences indicating FGFR activating mutations specifically in FGFR2 are found in endometrium cancer⁷⁷ and in FGFR3 are present in bladder cancer⁷⁸.

FGF2 is present at increased levels in prostate cancer. FGF1 is found to be upregulated in 80% of prostate cancer which can be confirmed from IHC⁷⁹. FGF1 has also been found to increase in prostatic intra-epithelial neoplasia (PIN)⁸⁰. Transgenic mice with activated FGFR1 kinase develop PIN⁸¹. FGFR4 plays a significant role in prostate cancer initiation as confirmed by the presence of homozygosity for FGFR arg388 is associated occurrence of prostate cancer⁸².

A lot of efforts are being put to FGF/FGFR inhibitors. Data from phase I/II clinical trial proved that FGFR inhibitors exhibit antitumor activity. For example:

1. Nonselective FGFR TKIs-Donovitinib⁸³ inhibits FGFR, PDGFR and VEGFR. Nintedanib⁸⁴ potentially blocks VEGFR, PDGFR, FGFR. And also targets SRC, LYN,LLK.Poratinib, an oral multikinase inhibitors inhibits BCR-ABC⁸⁵.
2. Selective anti-FGF-TKIs- AZD4547⁸⁶ pan FGFR selective causes desregulation of FGFR expression and hence shows anticancer response.BGJ398 selectively inhibits FGFR1-3 and is in phase I clinical trial⁸⁷.

2.5 Transforming Growth Factor- β Family

Transforming growth family - β is a secreted cytokine that critically influences proliferation and cellular differentiation. It plays an important role in immunity, cancer, bronchial asthma, lung fibrosis, heart diseases, diabetes etc⁸⁸. Ligands of TGF- β family are bone morphogeneic proteins (BMPs), growth and differentiation factors (GDFs), anti-mullerian hormone (AMH, activin, Nodal and TGF- β , TGF- β includes- TGF- β 1, TGF- β 2, TGF- β 3⁸⁹.

There are 8 types of SMAD proteins divided in 3 classes⁹⁰:

- i. Receptor regulated SMAD (R-Smad) -Smad 1,2,3,5 and 8.
- ii. Co-mediator SMAD (Co- Smad) - smad 4
- iii. Inhibitory SMAD (I-smad) - smad 6,7.

TGF- β signaling negatively regulates growth and inhibition of this pathway contributes to tumorigenesis⁹¹. As TGF- β negatively regulates growth, inactivating mutations in TGFBR1, TGFBR2, SMAD2, SMAD4 are commonly found in human cancers. In addition to this there is also loss of expression of TGF- β and Smad in many cancers. Disruption of TGF- β activated Smad pathway is sufficient to change gene expression which accelerates tumor formation. SMAD proteins play important role in TGF- β

associated growth inhibition and apoptosis. Rho proteins and PI3K regulate cell shape, loss of adherens junction and motility⁹². Colon, gastric, biliary, pulmonary and ovarian cancer witness biallelic inactivation of TGFBR1, which leads to inhibition of kinase domain of the receptor⁹³. Some proteins called as 'ligand traps' trap TGF- β family members so that their membrane access can be stopped. Example Follistatin trap BMPs and it is found to be overexpressed in hepatocarcinoma⁹⁴ and breast cancer bone metastasis⁹⁵. Germin-1 dysregulated expression is associated with skin and other cancer⁹⁶.

Intragenic mutation in SMAD4 occurs in colorectal cancer⁹⁷. Gastric cancer and T-cell lymphoblastic leukemia has been characterised by Loss of Smad3 expression⁹³. Smad2 and Smad3 protein expression is decreased in epithelial human cancers and rat prostatic carcinomas⁹⁸ and expression of Smad6 and Smad7 is increased⁹⁹.

2.6 Insulin-like Growth Factor Family

Insulin-like growth factors are associated with regulation of metabolism, growth and survival¹⁰¹. The signaling pathway utilized by IGF includes phosphoinositide-3-kinase (PI3K) and Akt or Ras and MAPK, which mediates response to many stimuli¹⁰². IGF family is constituted of two polypeptide ligands (IGF-1 and IGF-2), two membrane bound receptors (IGF-IR, IGF-IIR) and six binding proteins (IGFBP-1,2,3,4,5,6)¹⁰³. IGF-I increases cellular uptake of amino acids and glucose and stimulates glycogen and protein synthesis hence exerting an anabolic effect on protein and carbohydrate metabolism¹⁰⁴. IGF-I plays role in cell proliferation, differentiation and apoptosis. IGF-1 stimulates expression of Bcl proteins and hence blocks the initiation of apoptosis¹⁰⁵. IGF-II has mitogenic and anti-apoptotic action and also regulates cell proliferation and differentiation¹⁰⁶.

IGFBPs have both IGF-dependent and IGF-independent functions. When IGFBPs bind to IGFs, they help in transporting IGFs, protecting IGFs from proteosomal degradation, and regulate the interaction between IGFs and IGF-IR¹⁰⁷.

IGF-I and IGF-II are associated with variety of cancer like sarcoma, leukemia, prostate, breast, lung, colon, stomach, esophageal, liver, pancreas, kidney, thyroid, brain, ovary and uterus¹⁰⁸. Cancer cells having high metastatic potential have higher expression of IGF-II and IGF-IR. IGF-IR elimination from cell inhibits the signal transduction, abolishing the mitogenic activation of IGFs in cancer cells¹⁰⁹. IGF-IIR antagonizes effect of TGF-II and loss of function of IGF-IIR is associated in cancer and cancer cells which lack IGF-II degrading ability have good growth profile¹¹⁰. The action of IGFs is regulated by IGFBPs in cancer¹¹¹.

Oh¹¹², et al. found that IGFBP-3 inhibits breast cancer cell growth and induces apoptosis in both breast cancer and prostate cancer¹¹³. This happens because of presence of IGFBPs proteases. It was emphasized that increased levels of IGF-I is associated with increased breast cancer risk¹¹⁴. Increased levels of IGF-I and II are reported in colorectal cancer and hepatocarcinoma^{115,116}. In pancreatic cancer

pathway, mTOR is the point intersection, which allows IGF-IR/GPCR interaction to potentiate cell growth and proliferation¹¹⁷. IGFs can cause direct HIF-1 α expression through MAPK/PI3K pathway and IGF-IR synthesis occurs during hypoxia leading to neoangiogenesis¹¹⁸.

IGF pathway is an interesting target for cancer therapeutics. The mainstay of IGF target therapy is:

1. Small molecule inhibitors which inhibits tyrosine kinase domain of IGF-IR. (eg. Tyrostophosphins, Picropodopyllins, INSM-18, BNS-754807¹¹⁹).
2. Monoclonal antibodies (mAb) directed at IGF-IR. (eg. Ganitumab, AVE-1642, MK-0646, cixutumunab)¹²⁰.

OS7-906 is a small molecule TKIs that attaches to ATP binding site of tyrosine kinase receptor and causes dual inhibition of IR and IGF-IR¹²¹. Picropodophyllins inhibit IGF-IR. Exelixis are multitarget TKIs inhibit IGF-IR and Bcr-abl kinase¹²². Epigallocatechins is a catechinpolyphenolic component of green tea. It phosphorylates and activates TK and activates TK and inhibits IGF-IR by autophosphorylation of IGF-IR tyrosine kinase which causes cell cycle arrest¹²³. Ganitumab is monoclonal antibody with IgG1 back bone and inhibits ligand binding of IGF-1 and -2 which inhibits IGF-IR phosphorylation¹²⁴. AVE-1642 bind to IGF-IR and inhibits binding of IGFs. Cixutumunab is a fully humanised antibody blocking IGF-Rs and is in clinical trial phase II for breast cancer¹²⁵.

2.7 Hepatocyte Growth Factor Family

Hepatocyte growth factor (HGF) is also known as Serum factor (SF, HGF mediates its action through binding to a specific receptor site c-Met¹²⁶. HGF binds to extracellular α -chain of c-Met receptor leading to tyrosine phosphorylation of terminal kinase domain and progression of downstream pathways¹²⁷. HGF is expressed mainly by mesenchymal cells. Activation of HGF includes Hepatocyte growth factor Activator (HGFA)¹²⁸. Matriptase, matrix degrading serum protease converts pro-HGF to biologically active HGF (also Heparin, Factor XIII, tissue plasminogen activator (t-PA) and Urokinase Plasminogen activator (u-PA))¹²⁹.

HGF is a mitogenic regulating cell growth and death¹³⁰. HGF functions includes inhibition of apoptosis¹³¹, angiogenesis¹³², morphogenesis¹³³ and regulator of organ development¹³⁴.

Met is mainly expressed in epithelial cells whereas HGF is predominantly secreted by mesenchymal cells. This represents a cross-talk between epithelial cells and stromal cells^{135,136}. The autocrine activation of Met occurs in osteosarcoma and glioblastomas. In breast cancer, it is activated by paracrine signaling¹³⁷. Many carcinomas overexpress met as it is required for tumor growth and survival¹³⁸. In hepatocytes, Met binds to Fas and inhibits Fas-induced apoptosis. Mutations in Met have been evident in juxtamembrane in gastric and lung cancer^{139,140}. Interaction between c-Met and RTKs is linked to development of resistance to cancer therapies¹⁴¹. C- Met interacts directly with epidermal growth factor (EGFR), allowing activation of c-Met after stimulation of cells with EGFR

ligands EGF or TGF- α ¹⁴². Mutation in c-Met kinase domain were reported in human renal papillary carcinomas¹⁴³. Genetically transmitted cancers might be due to presence of heterozygous mutations in c-CBL binding site¹⁴⁴. Hypoxia, a usual oxygen deficient state in tumors activate c-Met transcription via HIF1 α stabilisation¹⁴⁵.

HGF has emerged as potentially good therapeutic target against cancer and following anti-cancer strategies have been devised:

- (a) Against HGF: neutralizing antibodies, anti-sense oligonucleotides, ribozyme siRNA and HGF regulators¹⁴⁶.
- (b) Against c-Met: HGF antagonist, antibodies, small molecule inhibitors, antisense oligonucleotides, ribozymes, siRNA as non-specific inhibitors¹⁴⁷.
- (c) Against c-Met signaling events¹⁴⁸.
- (d) HGF activation inhibitors¹⁴⁹.
- (e) SRC inhibitors have also shown good primary results in treatment of cancer cells.
- (f) HIF can also be good target as its stabilization leads to transcription of c-Met¹⁵⁰.

2.8 Neurotrophin Family

It was found in 1990 that some secreted proteins were important to normal growth of neurons and dendritic cells. Later they were named as neurotrophins¹⁵¹. The first neurotrophin discovered was NGF (Nerve growth factor)¹⁵². And next was brain derived neurotrophic factor (BDNF). In mammals two more neurotrophins were discovered, neurotrophin 3 and neurotrophin 4 and 5.

Receptors for Neurotrophin are transmembrane receptor of two different classes i.e. Trks and neurotrophin receptor P75. The cell survival is mediated by Trk receptor whereas P75 induces cell death. Trk i.e. (tropomyosine receptor kinases) are receptor tyrosin kinases and they are further categories into 3 types receptor types, TrkA¹⁵³ TrkB and TrkC. The preferred ligand for TrkA is NGF, where as for TrkB preferred ligand is BDNF and NT4/5 and TrkC binds preferentially to NTB¹⁵⁴. P75 is a bundle of six short α -helixes spanning of 90 amino acids forming a fold but do not possess any intrinsic catalytic activity¹⁵⁵.

Spingomyelin is hydrolysis to ceramide by P75¹⁵⁶ and after ligand binding causes dopamine release¹⁵⁷. P75 interacts with caveolin following TNF- α and NFKB activation¹⁵⁸. NFG was discovered from sarcoma while human colon cancer biopsy revealed TrkA and P75 was isolated from human melanoma cell lines¹⁵⁹. In breast cancer there is increase in NGF expression leading to phosphorylation mitogenesis, invasion, metastasis and angiogenesis via activation of all the three pathways i.e. MAPK, PI3K/AKT and PLCy¹⁶⁰. In melanomas, NGF-mediated paracrine signaling promotes proliferation and invasion¹⁶¹. In prostate cancer there is loss of P75 protein in basal epithelium cells due to mRNA instability. Loss of function of P75 leads to cell survival, proliferation and metastasis¹⁶². In pancreatic cancer NGF expression is increased which enhances proliferation, invasion and tumorigenicity¹⁶³.

In neuroblastoma TrkAIII isoforms potentiates

survival via activation of PI3K-AKT pathway¹⁶⁴. Whereas in glioblastoma NGF induces causes cell death by autophagy¹⁶⁵ and in medulloblastoma TrkA expression has good prognosis¹⁶⁶. TrkA signaling can be inhibited by tyrosine kinase inhibitors such as indocarbazole¹⁶⁷. Tamoxifen inhibits NGF mediated TrkA phosphorylation independent of its action on estrogen receptor¹⁶⁸.

In phase I clinical trial neurotrophin receptor-linked tyrosine kinase receptor inhibitor, CEP-701 was found to be orally safe and has entered phase II clinical trial¹⁶⁹. The selective Trk inhibitor AZ613 was found active against neurotrophin factor-mediated proliferation and signaling of neuroblastoma cells¹⁷⁰. The Trk tyrosine kinase inhibitor CEP-701 exhibited anti-tumor efficacy in xenograft models of human pancreatic ductal adenocarcinoma¹⁷¹. It has found that Lestaurtinib enhances the anticancer efficacy of chemotherapy in murine xenograft model of neuroblastoma via inhibition of TrkB activation and has promoted the clinical trial of Lestaurtinib¹⁷².

2.9 The Ephrin family

Ephrin expression play regulatory role in development and tissue homeostasis, along with the formation of tissue boundaries, assembly formation of neuronal mesh work, remodeling of blood vessels and organ size¹⁷³. Eph receptors are largest family of receptor tyrosine kinase. They bind GPI-linked and transmembrane ephrin ligands, generating bidirectional signaling at site of cell-cell contacts. Eph receptors are divided into two groups EphA and EphB, depending on two types of ligands that bind. EphA are further divided into 10 sub-groups EphA (EphA1-10) and EphB kinase (EphB1-6, Similarly there are two types of ephrin ligands: EphrinsA (A1-A6) and EphrinsB (B1-B3, EphrinA ligands are tethered to the cell membrane through glycosyl phosphatidylinositol (GP1) anchor. Whereas EphrinB ligands are transmembrane proteins possessing a cytoplasmic region and a PDZ-binding motif¹⁷⁴.

Expression of Ephs and Ephrins is frequently altered in human cancers. Eph receptors promote both tumorigenesis and tumor suppression¹⁷⁵. EphA2 overexpression has been found to cause oncogenic mutations and promotes metastasis in murine breast cancer models¹⁷⁶. EphB2 provides survival advantage to breast cancer by attenuating the inherent cell death pathways and upregulated anti-apoptotic proteins. EphB4 knockdown inhibit breast cancer cell viability, migration and invasion¹⁷⁷. EphA1 is elevated in colorectal cancer cases. It is mostly detected in stage II¹⁷⁸. Signaling EphA2 and EphrinA1 were more common in early stages of cancer. High expression of EphA3 is associated with lower survival rate¹⁷⁹. It has been found that EphA2 is overexpressed in human prostate cancer and it is linked with metastasis¹⁸⁰. EphA2 overexpression is associated with invasiveness in glioblastoma and is now important in targeted therapeutics against glioblastomas¹⁸¹. That overexpression of EphA4 enhances cell migration and proliferation through promoting the FGF signaling pathway¹⁸². EphB2 expression is higher in

invasive glioblastomas¹⁸³. Metastatic melanoma cells are characterised by EphA2 expression¹⁸⁴. EphrinA1, serves as growth factor, angiogenic signal and chemoattractant for melanoma cells but also angiogenic and chemoattractant for epithelial cells¹⁸⁵. EphA2 promotes angiogenesis¹⁸⁶.

Therapeutic targets against ephrins and Ephs are classified as :

- (i) Inhibition of receptor-ligand interactions¹⁸⁷:
Eg a) EphA2-Fc, EphB3-Fc are used in breast cancer and pancreatic cancer that targets ephrinA.
b) 2,5-dimethyl pyrrolyl benzoic acid derivatives target EphA4 to inhibit angiogenesis.
- (ii) Activation of Eph forward signaling¹⁸⁸:
Eg EA2, B233 antibody against EphA2 in breast cancer.
- (iii) Kinase inhibitors: E.g. Dasatinib against EphA2 in prostate cancer.
- (iv) Inhibition of Eph expression: E.g. EphA2 siRNA against pancreatic cancer and ovarian cancer¹⁸⁹.

2.10 Angiopoietins

Various growth factors are linked to physiological as well as pathological angiogenesis¹⁹⁰. Angiopoietin family of growth factor is composed of four members that bind to same TIE-2 tyrosine kinase receptor leading to different consequences. Angiopoietins was discovered by Davis et al., in 1996¹⁹¹. On the basis of structure Angiopoietins (ANG) are classified as¹⁹²:

1. ANG-1
2. ANG-2
3. ANG-3
4. ANG-4.

ANG-1 function is to stabilise and mature the blood vessels¹⁹³. Whereas ANG-2 is produced by endothelial cells at the site of vascular remodeling and destabilizes the vessel by lossening the endothelial cell junctions. ANG-2 acts as antagonist of ANG-1 because they compete with each other for TIE-2 receptor binding site¹⁹⁴. ANG-3 acts in a similar way as ANG-1 promoting phosphorylation of TIE-2 receptortyrosine residues. ANG-4 effects are similar to ANG-2 acts as a antagonist of ANG-1¹⁹². Upregulated ANG-2 is correlated to metastasis in breast cancer and lung cancer¹⁹⁵. Apart from cancer, ANG-2 upregulation is frequently found in diseases such as macula degeneration, rheumatoid arthritis¹⁹⁶, osteoarthritis¹⁹⁷, and psoriasis¹⁹⁸. Major therapeutics targeting ANG-2 in clinical trial are:

- (i) AMG-386: which is Fc-fusion protein blocks interaction between ANG-1 and ANG-2 and TIE-2 receptor. It is proposed to be effective in multiple cancers like ovarian, breast, fallopian tube cancer¹⁹⁹.
- (ii) CVX-060: it selectively blocks ANG-2 and TIE-2 interaction and has a very long half-life. It is in trials in combination with sunitinib, sorafenib, bevacizumab and irinotecan²⁰⁰.
- (iii) CEP-11981: it is found to inhibitor of both VEGF and TIE-2 receptor tyrosine kinase²⁰¹.
- (iv) MED13617: it is human anti-ANG-2 mAb. It has been shown to decreases angiogenesis and retard tumor

growth in mouse models in combination with mAb bevacizumab²⁰².

3. PERSPECTIVE

Growth factors are necessary for normal growth of non-transformed cells. These cells perform their committed physiological functions via availability of growth factors. But the signaling in normal cells is limited as the growth factors are depleted at the site by various degrading pathways. But as discussed by Hanahan and Weinberg, the first hallmark of cancer is self-sufficiency of growth signals, which has two advantages. Firstly, because of continued availability of growth factors the check points of cell cycle can be overcome. Secondly, the growth suppressor signaling is bypassed. Hence abrupt signaling can induce oncogenic transformations. An enhanced understanding of these pathways can help targeting these neoplastic transformation at an early stage.

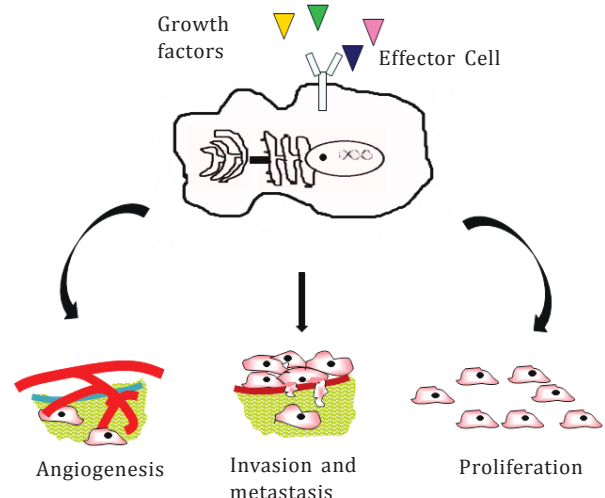


Figure 1. Schematic representation of Growth factor mediated signaling leading to malignant transformation of non-malignant cell.

Conflict of Interest : None

REFERENCES

1. Growth factor. *In* Encyclopædia Britannica. 2016, <http://www.britannica.com/science/growth-factor> (Retrieved on 24 April 2016)
2. Chen, S.C.; Chou, C.K.; Wong, F.H.; Chang, C. & Hu, C.P. Overexpression of epidermal growth factor and insulin-like growth factor-I receptors and autocrine stimulation in human esophageal carcinoma cells. *Cancer Research*, 1991, **51**(7), 1898-1903.
3. Maryam Zare, Mehdi Moghanibashi & Ferdous RastgarJazii Growth Factors, Signal Transduction Pathways, and Tumor Suppressor Genes in Esophageal Cancer. *In* Esophageal Cancer - Cell and Molecular Biology, Biomarkers, Nutrition and Treatment, *edited* by Prof. Ferdous Rastgar Jazii, 2012, ISBN: 978-953-51-0223-6,
4. Bafico, A. & Aaronson, S.A. Classification of Growth Factors and Their Receptors. 2003,

5. Hanahan, D. & Weinberg, R.A. Hallmarks of cancer: the next generation. *cell*, 2011, **144**(5), 646-674.
6. Heldin, C.H. & Westermark, B. Growth factors: mechanism of action and relation to oncogenes. *Cell*, 1984, **37**(1), 9-20. doi: 10.1016/0092-8674(84)90296-4
7. Johnsson, A.; Heldin, C.H.; Wasteson, A.; Westermark, B.; Deuel, T.F.; Huang, J.S. & Scrace, G. The c-sis gene encodes a precursor of the B chain of platelet-derived growth factor. *The EMBO journal*, 1984, **3**(5), 921.
8. Yamamoto, T.; Nishida, T.; Miyajima, N.; Kawai, S.; Ooi, T. & Toyoshima, K. The erbB gene of avian erythroblastosis virus is a member of the src gene family. *Cell*, 1983, **35**(1), 71-78. doi: 10.1016/0092-8674(83)90209-X
9. Sherr, C.J.; Rettenmier, C.W.; Sacca, R.; Roussel, M.F.; Look, A.T. & Stanley, E. R. The c-fms proto-oncogene product is related to the receptor for the mononuclear phagocyte growth factor, CSF 1. *Cell*, 1985, **41**(3), 665-676. doi: 10.1016/S0092-8674(85)80047-7
10. Sotiropoulos, C.; Neo, S. Y.; McShane, L. M.; Korn, E. L.; Long, P. M.; Jazaeri, A. & Liu, E. T. Breast cancer classification and prognosis based on gene expression profiles from a population-based study. *Proceedings of the National Academy Sci.*, 2003, **100**(18), 10393-10398. doi: 10.1073/pnas.1732912100
11. Sporn, M.B. & Roberts, A. B. Peptide growth factors and inflammation, tissue repair, and cancer. *J. Clinical Investigation*, 1986, **78**(2), 329. doi: 10.1172/JCI112580
12. Grotendorst, G.R.; Seppä, H.E.; Kleinman, H.K. & Martin, G.R. Attachment of smooth muscle cells to collagen and their migration toward platelet-derived growth factor. In Proceedings of the National Academy Science, 1981, **78**(6), 3669-3672. doi: 10.1073/pnas.78.6.3669
13. Shimokado, K.; Raines, E.W.; Madtes, D.K.; Barrett, T.B.; Benditt, E.P. & Ross, R. A significant part of macrophage-derived growth factor consists of at least two forms of PDGF. *Cell*, 1985, **43**(1), 277-286. doi: 10.1016/0092-8674(85)90033-9
14. Carvalho, I.; Milanezi, F.; Martins, A.; Reis, R. M. & Schmitt, F. Overexpression of platelet-derived growth factor receptor alpha in breast cancer is associated with tumour progression. *Breast Cancer Res.*, 2005, **7**(5), R788-95. doi: 10.1186/bcr1304
15. Najy, A.J.; Won, J.J.; Movilla, L.S. & Kim, H. R. C. Differential tumorigenic potential and matriptase activation between PDGF B versus PDGF D in prostate cancer. *Molecular Cancer Research*, 2012, **10**(8), 1087-1097. doi: 10.1158/1541-7786.MCR-12-0071
16. Ross, R.; Masuda, J.; Raines, E. W.; Gown, A. M.; Katsuda, S.; Sasahara, M. & Sato, H. Localization of PDGF-B protein in macrophages in all phases of atherosclerosis. *Science*, 1990, **248**(4958), 1009-1012. doi: 10.1126/science.2343305
17. Johnson, R.J.; Raines, E.W.; Floege, J.; Yoshimura, A.; Pritzl, P.; Alpers, C. & Ross, R. Inhibition of mesangial cell proliferation and matrix expansion in glomerulonephritis in the rat by antibody to platelet-derived growth factor. *J. Experimental Medicine*, 1992, **175**(5), 1413-1416. doi: 10.1084/jem.175.5.1413
18. Lokker, N.A.; Sullivan, C.M.; Hollenbach, S.J.; Israel, M.A. & Giese, N.A. Platelet-derived Growth Factor (PDGF) Autocrine Signaling Regulates Survival and Mitogenic Pathways in Glioblastoma Cells Evidence That the Novel PDGF-C and PDGF-D Ligands May Play a Role in the Development of Brain Tumors. *Cancer Research*, 2002, **62**(13), 3729-3735.
19. Tekmal, R.R.; Ramachandra, N.; Gubba, S.; Durgam, V.R.; Mantione, J.; Toda, K. & Dillehay, D.L. Overexpression of int-5/aromatase in mammary glands of transgenic mice results in the induction of hyperplasia and nuclear abnormalities. *Cancer Research*, 1996, **56**(14), 3180-3185.
20. De Jong, J.S.; van Diest, P. J.; van der Valk, P. & Baak, J. Expression of growth factors, growth-inhibiting factors, and their receptors in invasive breast cancer. II: Correlations with proliferation and angiogenesis. *The J. Pathology*, 1998, **184**(1), 53-57. doi: 10.1002/(SICI)1096-9896(199801)184:1<53::AID-PATH6>3.0.CO;2-7
21. Siegel, R.; Ward, E.; Brawley, O. & Jemal, A. Cancer statistics, 2011. *CA: a cancer journal for clinicians*, 2011, **61**(4), 212-236. doi: 10.3322/caac.20121
22. Ustach, C. V.; Huang, W.; Conley-LaComb, M. K.; Lin, C.Y.; Che, M.; Abrams, J. & Kim, H. R. C. A novel signaling axis of matriptase/PDGF-D/ β -PDGFR in human prostate cancer. *Cancer Research*, 2010, **70**(23), 9631-9640. doi: 10.1158/0008-5472.CAN-10-0511
23. Liu, Y. C.; Chen, S. C.; Chang, C.; Leu, C. M. & Hu, C. P. Platelet-derived growth factor is an autocrine stimulator for the growth and survival of human esophageal carcinoma cell lines. *Experimental Cell Res.*, 1996, **228**(2), 206-211. doi: 10.1006/excr.1996.0318
24. Zhang, X.; Rong, T. H.; Zhang, Y.; Long, H.; Fu, J. H.; Ling, P. & Su, X. D. [Expression and significance of C-kit and platelet-derived growth factor receptor-beta (PDGFRbeta) in esophageal carcinoma]. *Ai zheng= Aizheng= Chinese J. Cancer*, 2006, **25**(1), 92-95.
25. Ferrara, N. Vascular endothelial growth factor: basic science and clinical progress. *Endocrine Reviews*, 2004, **25**(4), 581-611. doi: 10.1210/er.2003-0027
26. Roskoski, R. Vascular endothelial growth factor (VEGF) signaling in tumor progression. *Critical Rev. Oncology/Hematology*, 2007, **62**(3), 179-213. doi: 10.1016/j.critrevonc.2007.01.006
27. Forsythe, J.A.; Jiang, B. H.; Iyer, N. V.; Agani, F.; Leung, S. W.; Koos, R. D. & Semenza, G.L. Activation of vascular endothelial growth factor gene transcription by hypoxia-inducible factor 1. *Molecular Cellular Bio.*, 1996, **16**(9), 4604-4613. doi: 10.1128/MCB.16.9.4604
28. Alon, T.; Hemo, I.; Itin, A.; Pe'er, J.; Stone, J. & Keshet, E. Vascular endothelial growth factor acts as a survival factor for newly formed retinal vessels and has implications for retinopathy of prematurity. *Nature medicine*, 1995, **1**(10), 1024-1028. doi: 10.1038/nm1095-1024
29. Dvorak, H. F. Angiogenesis: update 2005. *J. Thrombosis Haemostasis*, 2005, **3**(8), 1835-1842. doi: 10.1111/j.1538-7836.2005.01361.x
30. Roberts, E.; Cossigny, D.A. & Quan, G.M. The role of vascular endothelial growth factor in metastatic prostate cancer to the skeleton. *Prostate Cancer*, 2013. doi: 10.1155/2013/418340
31. Pugh, C.W. & Ratcliffe, P.J. Regulation of angiogenesis by hypoxia: role of the HIF system. *Nature Medicine*,

- 2003, **9**(6), 677-684. doi: 10.1038/nm0603-677
32. Lawler, J. Thrombospondin-1 as an endogenous inhibitor of angiogenesis and tumor growth. *J. Cellular Molecular Medicine*, 2002, **6**(1), 1-12. doi: 10.1111/j.1582-4934.2002.tb00307.x
 33. Hoeben, A.; Landuyt, B.; Highley, M.S.; Wildiers, H.; Van Oosterom, A.T. & De Bruijn, E.A. Vascular endothelial growth factor and angiogenesis. *Pharmacological Reviews*, 2004, **56**(4), 549-580. doi: 10.1124/pr.56.4.3
 34. Hanahan, D. & Folkman, J. Patterns and emerging mechanisms of the angiogenic switch during tumorigenesis. *Cell*, 1996, **86**(3), 353-364. doi: 10.1016/S0092-8674(00)80108-7
 35. Dikov, M. M.; Oyama, T.; Cheng, P.; Takahashi, T.; Takahashi, K.; Sepetavec, T. & Gabrilovich, D. I. Vascular endothelial growth factor effects on nuclear factor- κ B activation in hematopoietic progenitor cells. *Cancer Research*, 2001, **61**(5), 2015-2021.
 36. Toi, M.; Inada, K.; Suzuki, H. & Tominaga, T. Tumor angiogenesis in breast cancer: its importance as a prognostic indicator and the association with vascular endothelial growth factor expression. *Breast Cancer Res. Treatment*, 1995, **36**(2), 193-204. doi: 10.1007/BF00666040
 37. Linderholm, B. K.; Lindahl, T.; Holmberg, L.; Klaar, S.; Lennerstrand, J.; Henriksson, R. & Bergh, J. The expression of vascular endothelial growth factor correlates with mutant p53 and poor prognosis in human breast cancer. *Cancer Research*, 2001, **61**(5), 2256-2260.
 38. Yoshiji, H.; Harris, S.R. & Thorgeirsson, U.P. Vascular endothelial growth factor is essential for initial but not continued in vivo growth of human breast carcinoma cells. *Cancer Research*, 1997, **57**(18), 3924-3928.
 39. Giatromanolaki, A.; Koukourakis, M.; O'byrne, K.E.N.; Fox, S.; Whitehouse, R.; Talbot, D.C. & Gatter, K.C. Prognostic value of angiogenesis in operable non-small cell lung cancer. *Journal Pathology*, 1996, **179**(1), 80-88. doi: 10.1002/(SICI)1096-9896(199605)179:1<80::AID-PATH547>3.0.CO;2-X
 40. Maeda, K.; Chung, Y.S.; Ogawa, Y.; Takatsuka, S.; Kang, S.M.; Ogawa, M. & Sowa, M. Prognostic value of vascular endothelial growth factor expression in gastric carcinoma. *Cancer*, 1996, **77**(5), 858-863. doi: 10.1002/(SICI)1097-0142(19960301)77:5<858::AID-CNCR8>3.0.CO;2-A
 41. Scolnik, P.A. mAbs: a business perspective. *MAbs* 2009; **1**: 179-84; PMID: 20061824. doi: 10.4161/mabs.1.2.7736
 42. Robak, T.; Wierzbowska, A. & Robak, E. Recent clinical trials of cladribine in hematological malignancies and autoimmune disorders. *Reviews Recent Clinical Trials*, 2006, **1**(1), 15-34. doi: 10.2174/157488706775246102
 43. Slamon, D.J.; Leyland-Jones, B.; Shak, S.; Fuchs, H.; Paton, V.; Bajamonde, A. & Baselga, J. Use of chemotherapy plus a monoclonal antibody against HER2 for metastatic breast cancer that overexpresses HER2. *New England J. Medicine*, 2001, **344**(11), 783-792. doi: 10.1056/NEJM200103153441101
 44. Bonner, J. A.; Harari, P.M.; Giralt, J.; Cohen, R.B.; Jones, C.U.; Sur, R.K. & Yousoufian, H. Radiotherapy plus cetuximab for locoregionally advanced head and neck cancer: 5-year survival data from a phase 3 randomised trial, and relation between cetuximab-induced rash and survival. *The Lancet Oncology*, 2010, **11**(1), 21-28. doi: 10.1016/S1470-2045(09)70311-0
 45. Johnson, D.H.; Fehrenbacher, L.; Novotny, W.F.; Herbst, R.S.; Nemunaitis, J.J.; Jablons, D.M. & Holmgren, E. Randomized phase II trial comparing bevacizumab plus carboplatin and paclitaxel with carboplatin and paclitaxel alone in previously untreated locally advanced or metastatic non-small-cell lung cancer. *J. Clinical Oncology*, 2004, **22**(11), 2184-2191. doi: 10.1200/JCO.2004.11.022
 46. Goodsell, D. S. The molecular perspective: epidermal growth factor. *The Oncologist*, 2003, **8**(5), 496-497. doi: 10.1634/theoncologist.8-5-496
 47. Groenen, L.C.; Nice, E.C. & Burgess, A. W. Structure-function relationships for the EGF/TGF- α family of mitogens. *Growth Factors*, 1994, **11**(4), 235-257. doi: 10.3109/08977199409010997
 48. Sasaki, T.; Hiroki, K. & Yamashita, Y. The role of epidermal growth factor receptor in cancer metastasis and microenvironment. *Bio. Med. Res. Int.*, 2013. doi: 10.1155/2013/546318
 49. Shipley, G.D.; Childs, C.B.; Volkenant, M.E. & Moses, H.L. Differential effects of epidermal growth factor, transforming growth factor, and insulin on DNA and protein synthesis and morphology in serum-free cultures of AKR-2B cells. *Cancer Research*, 1984, **44**(2), 710-716.
 50. Cooper, J.A. & Hunter, T. Similarities and differences between the effects of epidermal growth factor and Rous sarcoma virus. *J. Cell Biology*, 1981, **91**(3), 878-883. doi: 10.1083/jcb.91.3.878
 51. Müller, R.; Bravo, R.; Burckhardt, J. & Curran, T. Induction of c-fos gene and protein by growth factors precedes activation of c-myc. *Nature*, 1983, **312**(5996), 716-720. doi: 10.1038/312716a0
 52. Fisher, P.B.; Bozzone, J.H. & Weinstein, I.B. Tumor promoters and epidermal growth factor stimulate anchorage-independent growth of adenovirus-transformed rat embryo cells. *Cell*, 1979, **18**(3), 695-705. doi: 10.1016/0092-8674(79)90124-7
 53. Reynolds, V.H.; Boehm, F.H. & Cohen, S. Enhancement of chemical carcinogenesis by an epidermal growth factor. In *Surgical forum*, 1965, **16**, p. 108,
 54. Heldin, C. H. & Westermark, B. Mechanism of action and in vivo role of platelet-derived growth factor. *Physiological Reviews*, 1999, **79**(4), 1283-1316.
 55. Mori, K.; Kurobe, M.; Furukawa, S.; Kubo, K. & Hayashi, K. Human breast cancer cells synthesize and secrete an EGF-like immunoreactive factor in culture. *Biochem. Biophys. Res. Comm.*, 1986, **136**(1), 300-305. doi: 10.1016/0006-291X(86)90909-5
 56. Connolly, J. M. & Rose, D. P. Secretion of epidermal growth factor and related polypeptides by the DU 145 human prostate cancer cell line. *The Prostate*, 1989, **15**(2), 177-186. doi: 10.1002/pros.2990150211
 57. Herbst, R. S. Review of epidermal growth factor receptor biology. *Int. J. Radiation Oncology Bio. Phys.*, 2004, **59**(2), S21-S26. doi: 10.1016/j.ijrobp.2003.11.041
 58. Normanno, N.; De Luca, A.; Bianco, C.; Strizzi, L.; Mancino, M.; Maiello, M. R. & Salomon, D. S. Epidermal growth factor receptor (EGFR) signaling in cancer. *Gene*,

- 2006, **366**(1), 2-16. doi: 10.1016/j.gene.2005.10.018
59. Santin, A.D.; Bellone, S.; Siegel, E.R.; Palmieri, M.; Thomas, M.; Cannon, M. J. & Pecorelli, S. Racial differences in the overexpression of epidermal growth factor type II receptor (HER2/neu): a major prognostic indicator in uterine serous papillary cancer. *Am. J. Obstetrics Gynecology*, 2005, **192**(3), 813-818. doi: 10.1016/j.ajog.2004.10.605
 60. Carraway, K.L. & Cantley, L.C. A new acquaintance for erbB3 and erbB4: a role for receptor heterodimerization in growth signaling. *Cell*, 1994, **78**(1), 5-8. doi: 10.1016/0092-8674(94)90564-9
 61. Coffey, R. J.; Leof, E. B.; Shipley, G. D. & Moses, H. L. Suramin inhibition of growth factor receptor binding and mitogenicity in AKR-2B cells. *J. Cellular Physiology*, 1987, **132**(1), 143-148. doi: 10.1002/jcp.1041320120
 62. Kris, M.G.; Natale, R.B.; Herbst, R.S.; Lynch Jr, T.J.; Prager, D.; Belani, C.P.; & Albain, K.S. Efficacy of gefitinib, an inhibitor of the epidermal growth factor receptor tyrosine kinase, in symptomatic patients with non-small cell lung cancer: a randomized trial. *Jama*, 2003, **290**(16), 2149-2158. doi: 10.1001/jama.290.16.2149
 63. Wheeler, D.L.; Huang, S.; Kruser, T.J.; Nechrebecki, M.M.; Armstrong, E.A.; Benavente, S. & Harari, P.M. Mechanisms of acquired resistance to cetuximab: role of HER (ErbB) family members. *Oncogene*, 2008, **27**(28), 3944-3956. doi: 10.1038/onc.2008.19
 64. Kwabi-Addo, B.; Ozen, M. & Ittmann, M. The role of fibroblast growth factors and their receptors in prostate cancer. *Endocrine-related cancer*, 2004, **11**(4), 709-724. doi: 10.1677/erc.1.00535
 65. Folkman, J. & Shing Y. Angiogenesis. *J. Bio. Chem.*, 1992, 267 10931-10934.
 66. Folkman, J. & Klagsbrun, M. Angiogenic factors. *Science*, 1987, **235**(4787), 442-447. doi: 10.1126/science.2432664
 67. Kamura, S.; Matsumoto, Y.; Fukushi, J.I.; Fujiwara, T.; Iida, K.; Okada, Y. & Iwamoto, Y. Basic fibroblast growth factor in the bone microenvironment enhances cell motility and invasion of Ewing's sarcoma family of tumours by activating the FGFR1-PI3K-Rac1 pathway. *British J. Cancer*, 2010, **103**(3), 370-381. doi: 10.1038/sj.bjc.6605775
 68. Dvorak, P.; Dvorakova, D. & Hampl, A. Fibroblast growth factor signaling in embryonic and cancer stem cells. *FEBS letters*, 2006, **580**(12), 2869-2874. doi: 10.1016/j.febslet.2006.01.095
 69. Majka, M.; Janowska-Wieczorek, A.; Ratajczak, J.; Ehrenman, K.; Pietrzakowski, Z.; Kowalska, M.A. & Ratajczak, M.Z. Numerous growth factors, cytokines, and chemokines are secreted by human CD34+ cells, myeloblasts, erythroblasts, and megakaryoblasts and regulate normal hematopoiesis in an autocrine/paracrine manner. *Blood*, 2001, **97**(10), 3075-3085. doi: 10.1182/blood.V97.10.3075
 70. Chesi, M.; Bergsagel, P.L. & Kuehl, W.M. The enigma of ectopic expression of FGFR3 in multiple myeloma: a critical initiating event or just a target for mutational activation during tumor progression. *Current Opinion Hematology*, 2002, **9**(4), 288-293. doi: 10.1097/00062752-200207000-00005
 71. Song, S.; Wientjes, M.G.; Gan, Y. & Au, J.L. S. Fibroblast growth factors: an epigenetic mechanism of broad spectrum resistance to anticancer drugs. *In Proceedings of the National Academy of Sciences*, 2000, **97**(15), 8658-8663. doi: 10.1073/pnas.140210697
 72. Dvorakova, D.; Krejci, P.; Mayer, J.; Fajkus, J.; Hampl, A. & Dvorak, P. Changes in the expression of FGFR3 in patients with chronic myeloid leukaemia receiving transplants of allogeneic peripheral blood stem cells. *British J. Haematology*, 2001, **113**(3), 832-835. doi: 10.1046/j.1365-2141.2001.02829.x
 73. Welm, B.E.; Freeman, K.W.; Chen, M.; Contreras, A.; Spencer, D.M. & Rosen, J. M. Inducible dimerization of FGFR1 development of a mouse model to analyze progressive transformation of the mammary gland. *J. Cell Biology*, 2002, **157**(4), 703-714. doi: 10.1083/jcb.200107119
 74. Jackson, D.; Bresnick, J.; Rosewell, I.; Crafton, T.; Poulosom, R.; Stamp, G. & Dickson, C. Fibroblast growth factor receptor signaling has a role in lobuloalveolar development of the mammary gland. *J. Cell Sci.*, 1997, **110**(11), 1261-1268.
 75. Jain, V. K. & Turner, N. C. Challenges and opportunities in the targeting of fibroblast growth factor receptors in breast cancer. *Breast Cancer Research*, 2012, **14**(3), 1-9. doi: 10.1186/bcr3139
 76. Bai, A.; Meetze, K.; Vo, N.Y.; Kollipara, S.; Mazsa, E. K.; Winston, W.M. & Jiang, J. GP369, an FGFR2-IIIb-Specific Antibody, Exhibits Potent Antitumor Activity against Human Cancers Driven by Activated FGFR2 Signaling. *Cancer Research*, 2010, **70**(19), 7630-7639. doi: 10.1158/0008-5472.CAN-10-1489
 77. Dutt, A.; Salvesen, H. B.; Chen, T.H.; Ramos, A. H.; Onofrio, R.C.; Hatton, C. & Wyhs, N. Drug-sensitive FGFR2 mutations in endometrial carcinoma. *Proceedings National Academy Sci.*, 2008, **105**(25), 8713-8717. doi: 10.1073/pnas.0803379105
 78. Cappellen, D.; De Oliveira, C.; Ricol, D.; de Medina, S.; Bourdin, J.; Sastre-Garau, X. & Radvanyi, F. Frequent activating mutations of FGFR3 in human bladder and cervix carcinomas. *Nature genetics*, 1999, **23**(1), 18-20. doi: 10.1038/12615
 79. Dorkin, T. J.; Robinson, M. C.; Marsh, C.; Bjartell, A.; Neal, D. E. & Leung, H. Y. FGF8 over-expression in prostate cancer is associated with decreased patient survival and persists in androgen independent disease. *Oncogene*, 1999, **18**(17), 2755-2761. doi: 10.1038/sj.onc.1202624
 80. Payson, R.A.; Chotani, M. A. & Chiu, M. Regulation of a promoter of the fibroblast growth factor 1 gene in prostate and breast cancer cells. *The Journal of Steroid Biochemistry Molecular Biology*, 1998, **66**(3), 93-103. doi: 10.1016/S0960-0760(98)00051-X
 81. Song, Z.; Wu, X.; Powell, W. C.; Cardiff, R. D.; Cohen, M.B.; Tin, R.T. & Roy-Burman, P. Fibroblast Growth Factor 8 Isoform b Overexpression in Prostate Epithelium A New Mouse Model for Prostatic Intraepithelial Neoplasia. *Cancer Research*, 2002, **62**(17), 5096-5105.
 82. Sidenius, N. & Blasi, F. The urokinase plasminogen activator system in cancer: recent advances and implication for prognosis and therapy. *Cancer Metastasis Rev.*, 2003, **22**(2-3), 205-222. doi: 10.1023/A:1023099415940
 83. Lee, S. H.; de Menezes, D. L.; Vora, J.; Harris, A.; Ye, H.; Nordahl, L. & Heise, C. In vivo target modulation

- and biological activity of CHIR-258, a multitargeted growth factor receptor kinase inhibitor, in colon cancer models. *Clinical Cancer Res.*, 2005, **11**(10), 3633-3641. doi: 10.1158/1078-0432.CCR-04-2129
84. Hilberg, F.; Roth, G. J.; Krssak, M.; Kautschitsch, S.; Sommergruber, W.; Tontsch-Grunt, U. & Heckel, A. BIBF 1120: triple angiokine inhibitor with sustained receptor blockade and good antitumor efficacy. *Cancer Research*, 2008, **68**(12), 4774-4782. doi: 10.1158/0008-5472.CAN-07-6307
 85. O'Hare, T.; Shakespeare, W. C.; Zhu, X.; Eide, C. A.; Rivera, V. M.; Wang, F. & Metcalf, C. A. AP24534, a pan-BCR-ABL inhibitor for chronic myeloid leukemia, potently inhibits the T315I mutant and overcomes mutation-based resistance. *Cancer Cell*, 2009, **16**(5), 401-412. doi: 10.1016/j.ccr.2009.09.028
 86. Gavine, P. R.; Mooney, L.; Kilgour, E.; Thomas, A. P.; Al-Kadhimi, K.; Beck, S. & Brooks, A. N. AZD4547: an orally bioavailable, potent, and selective inhibitor of the fibroblast growth factor receptor tyrosine kinase family. *Cancer Research*, 2012, **72**(8), 2045-2056. doi: 10.1158/0008-5472.CAN-11-3034
 87. Dieci, M.V.; Arnedos, M.; Andre, F. & Soria, J.C. Fibroblast growth factor receptor inhibitors as a cancer treatment: from a biologic rationale to medical perspectives. *Cancer Discovery*, 2013, **3**(3), 264-279. doi: 10.1158/2159-8290.CD-12-0362
 88. Li, M.O.; Wan, Y.Y.; Sanjabi, S.; Robertson, A.K.L. & Flavell, R.A. Transforming growth factor- β regulation of immune responses. *Annu. Rev. Immunol.*, 2006, **24**, 99-146. doi: 10.1111/j.0105-2896.2006.00405.x
 89. Kingsley, D.M. The TGF-beta superfamily: new members, new receptors, and new genetic tests of function in different organisms. *Genes Development*, 1994, **8**(2), 133-146. doi: 10.1101/gad.8.2.133
 90. Grady, William M. Transforming growth factor- β , Smads, and cancer. *Clinical Cancer Res.*, 2005, **11**(9), 3151-3154. doi: 10.1158/1078-0432.CCR-05-0414
 91. Millet, C. & Zhang, Y.E. Roles of Smad3 in TGF- β signaling during carcinogenesis. *Critical Reviews™ in Eukaryotic Gene Expression*, 2007, **17**(4),
 92. Bierie, B. & Moses, H. L. TGF- β and cancer. *Cytokine Growth Factor Reviews*, 2006, **17**(1), 29-40. doi: 10.1016/j.cytogfr.2005.09.006
 93. Levy, L. & Hill, C.S. Alterations in components of the TGF- β superfamily signaling pathways in human cancer. *Cytokine Growth Factor Rev.*, 2006, **17**(1), 41-58. doi: 10.1016/j.cytogfr.2005.09.009
 94. Rodgarkia-Dara, C.; Vejda, S.; Erlach, N.; Losert, A.; Bursch, W.; Berger, W. & Grusch, M. The activin axis in liver biology and disease. *Mutation Research/Reviews Mutation Res.*, 2006, **613**(2), 123-137. doi: 10.1016/j.mrrev.2006.07.002
 95. Kang, Y.; Chen, C.R. & Massagué, J. A self-enabling TGF β response coupled to stress signaling: Smad engages stress response factor ATF3 for Id1 repression in epithelial cells. *Molecular cell*, 2003, **11**(4), 915-926. doi: 10.1016/S1097-2765(03)00109-6
 96. Sneddon, J.B.; Zhen, H.H.; Montgomery, K.; van de Rijn, M.; Tward, A.D.; West, R. & Brown, P.O. Bone morphogenetic protein antagonist gremlin 1 is widely expressed by cancer-associated stromal cells and can promote tumor cell proliferation. *Proceedings National Academy Sci.*, 2006, **103**(40), 14842-14847. doi: 10.1073/pnas.0606857103
 97. Sjöblom, T.; Jones, S.; Wood, L. D.; Parsons, D.W.; Lin, J.; Barber, T.D. & Szabo, S. The consensus coding sequences of human breast and colorectal cancers. *Science*, 2006, **314**(5797), 268-274. doi: 10.1126/science.1133427
 98. Brodin, G.; ten Dijke, P.; Funa, K.; Heldin, C. H. & Landström, M. Increased smad expression and activation are associated with apoptosis in normal and malignant prostate after castration. *Cancer Research*, 1999, **59**(11), 2731-2738.
 99. Kleeff, J.; Maruyama, H.; Friess, H.; Büchler, M. W.; Falb, D. & Korc, M. Smad6 suppresses TGF- β -induced growth inhibition in COLO-357 pancreatic cancer cells and is overexpressed in pancreatic cancer. *Biochem. Biophys. Res. Comm.*, 1999, **255**(2), 268-273. doi: 10.1006/bbrc.1999.0171
 100. Nakae, J.; Kido, Y. & Accili, D. Distinct and overlapping functions of insulin and IGF-I receptors. *Endocrine Reviews*, 2001, **22**(6), 818-835. doi: 10.1210/edrv.22.6.0452
 101. Siddle, K. Signaling by insulin and IGF receptors: supporting acts and new players. *J. Molecular Endocrinology*, 2011, **47**(1), R1-R10. doi: 10.1530/JME-11-0022
 102. Yu, H. & Rohan, T. Role of the insulin-like growth factor family in cancer development and progression. *J. National Cancer Institute*, 2000, **92**(18), 1472-1489. doi: 10.1093/jnci/92.18.1472
 103. Jones, J. I. & Clemmons, D. R. Insulin-Like Growth Factors and Their Binding Proteins: Biological Actions. *Endocrine Reviews*, 1995, **16**(1), 3-34. doi: 10.1210/er.16.1.3
 104. Minshall, C.; Arkins, S.; Straza, J.; Conners, J.; Dantzer, R.; Freund, G. G. & Kelley, K. W. IL-4 and insulin-like growth factor-I inhibit the decline in Bcl-2 and promote the survival of IL-3-deprived myeloid progenitors. *Journal Immunology*, 1997, **159**(3), 1225-1232.
 105. O'Dell, S.D. & Day, I.N. Molecules in focus Insulin-like growth factor II (IGF-II). *Int. J. Biochem. Cell Bio.*, 1998, **30**(7), 767-771. doi: 10.1016/S1357-2725(98)00048-X
 106. Stewart, C.E. & Rotwein, P. Growth, differentiation, and survival: Multiple physiological functions for insulin-like growth factors. *Physiological Reviews*, 1996, **76**(4), 1005-1026.
 107. Levine, A.J.; Feng, Z.; Mak, T.W.; You, H. & Jin, S. Coordination and communication between the p53 and IGF-1-AKT-TOR signal transduction pathways. *Genes Development*, 2006, **20**(3), 267-275. doi: 10.1101/gad.1363206
 108. Jiang, Y.; Rom, W. N.; Yie, T. A.; Chi, C. X. & Tchou-Wong, K. M. Induction of tumor suppression and glandular differentiation of A549 lung carcinoma cells by dominant-negative IGF-I receptor. *Oncogene*, 1999, **18**(44), doi: 10.1038/sj.onc.1202984
 109. Byrd, J.C.; Devi, G.R.; De Souza, A.T.; Jirtle, R.L. & MacDonald, R.G. Disruption of ligand binding to the insulin-like growth factor II/mannose 6-phosphate receptor by cancer-associated missense mutations. *J. Biol. Chem.*, 1999, **274**(34), 24408-24416. doi: 10.1074/jbc.274.34.24408
 110. Yee, D.; Favoni, R. E.; Lippman, M. E. & Powell, D. R.

- Identification of insulin-like growth factor binding proteins in breast cancer cells. *Breast Cancer Research Treatment*, 1991, **18**(1), 3-10. doi: 10.1007/BF01975437
111. Oh, Y.O.U.N.G.M.A.N.; Müller, H.L.; Lamson, G. & Rosenfeld, R. G. Insulin-like growth factor (IGF)-independent action of IGF-binding protein-3 in Hs578T human breast cancer cells. Cell surface binding and growth inhibition. *J. Biol. Chem.*, 1993, **268**(20), 14964-14971.
 112. Gill, Z.P.; Perks, C.M.; Newcomb, P.V. & Holly, J. M. Insulin-like growth factor-binding protein (IGFBP-3) predisposes breast cancer cells to programmed cell death in a non-IGF-dependent manner. *J. Biol. Chem.*, 1997, **272**(41), 25602-25607. doi: 10.1074/jbc.272.41.25602
 113. Rollison, D.E.; Giuliano, A.R.; Risendal, B. C.; Sweeney, C.; Boulware, D.; Laronga, C. & Slattery, M.L. Serum insulin-like growth factor (IGF)-1 and IGF binding protein-3 in relation to breast cancer among Hispanic and white, non-Hispanic women in the US Southwest. *Breast Cancer Res. Treatment*, 2010, **121**(3), 661-669. doi: 10.1007/s10549-009-0609-5
 114. Wu, Y.; Yakar, S.; Zhao, L.; Hennighausen, L. & LeRoith, D. Circulating insulin-like growth factor-I levels regulate colon cancer growth and metastasis. *Cancer Research*, 2002, **62**(4), 1030-1035.
 115. Dong, Z.Z.; Yao, D.F.; Yao, D.B.; Wu, X.H.; Wu, W.; Qiu, L.W. & Meng, X.Y. Expression and alteration of insulin-like growth factor II-messenger RNA in hepatoma tissues and peripheral blood of patients with hepatocellular carcinoma. *World J. Gastroenterology: WJG*, 2005, **11**(30), 4655-4660. doi: 10.3748/wjg.v11.i30.4655
 116. Rozengurt, E.; Sinnott-Smith, J. & Kisfalvi, K. Crosstalk between insulin/insulin-like growth factor-1 receptors and G protein-coupled receptor signaling systems: a novel target for the antidiabetic drug metformin in pancreatic cancer. *Clinical Cancer Research*, 2010, **16**(9), 2505-2511. doi: 10.1158/1078-0432.CCR-09-2229
 117. Peretz, S.; Kim, C.; Rockwell, S.; Baserga, R. & Glazer, P. M. IGF1 receptor expression protects against microenvironmental stress found in the solid tumor. *Radiation Research*, 2002, **158**(2), 174-180. doi: 10.1667/0033-7587(2002)158[0174:IREPAM]2.0.CO;2
 118. Menu, E.; Jernberg-Wiklund, H.; Stromberg, T.; De Raeve, H.; Girnita, L.; Larsson, O. & Vanderkerken, K. Inhibiting the IGF-1 receptor tyrosine kinase with the cyclolignan PPP: an in vitro and in vivo study in the 5T33MM mouse model. *Blood*, 2006, **107**(2), 655-660. doi: 10.1182/blood-2005-01-0293
 119. Kolb, E. A.; Gorlick, R.; Houghton, P.J.; Morton, C.L.; Lock, R.; Carol, H. & Smith, M. A. Initial testing (stage 1) of a monoclonal antibody (SCH 717454) against the IGF-1 receptor by the pediatric preclinical testing program. *Pediatric Blood Cancer*, 2008, **50**(6), 1190-1197. doi: 10.1002/pbc.21368
 120. Mulvihill, M.J.; Ji, Q.S.; Coate, H.R.; Cooke, A.; Dong, H.; Feng, L. & Nigro, A. I. Novel 2-phenylquinolin-7-yl-derived imidazo [1, 5-a] pyrazines as potent insulin-like growth factor-I receptor (IGF-IR) inhibitors. *Bioorganic & medicinal chemistry*, 2008, **16**(3), 1359-1375. doi: 10.1016/j.bmc.2007.10.061
 121. López-Calderero, I.; Chávez, E.S. & García-Carbonero, R. The insulin-like growth factor pathway as a target for cancer therapy. *Clinical Translational Oncology*, 2010, **12**(5), 326-338. doi: 10.1007/s12094-010-0514-8
 122. Li, M.; He, Z.; Ermakova, S.; Zheng, D.; Tang, F.; Cho, Y. Y. & Bode, A. M. Direct inhibition of insulin-like growth factor-I receptor kinase activity by (-)- epigallocatechin-3-gallate regulates cell transformation. *Cancer Epidemiology Biomarkers Prevention*, 2007, **16**(3), 598-605. doi: 10.1158/1055-9965.EPI-06-0892
 123. Tap, W. D.; Demetri, G.; Barnette, P.; Desai, J.; Kavan, P.; Tozer, R. & Leitch, I. Phase II study of Ganitumab, a fully human anti-type-1 insulin-like growth factor receptor antibody, in patients with metastatic ewing family tumors or desmoplastic small round cell tumors. *J. Clinical Oncology*, 2012, **30**(15), 1849-1856. doi: 10.1200/JCO.2011.37.2359
 124. Dallas, N. A.; Xia, L.; Fan, F.; Gray, M. J.; Gaur, P.; Van Buren, G. & Ellis, L. M. Chemoresistant colorectal cancer cells, the cancer stem cell phenotype, and increased sensitivity to insulin-like growth factor-I receptor inhibition. *Cancer Research*, 2009, **69**(5), 1951-1957. doi: 10.1158/0008-5472.CAN-08-2023
 125. Ma, C. X.; Suman, V. J.; Goetz, M.; Haluska, P.; Moynihan, T.; Nanda, R. & Erlichman, C. A phase I trial of the IGF-1R antibody Cixutumumab in combination with temsirolimus in patients with metastatic breast cancer. *Breast Cancer Research Treatment*, 2013, **139**(1), 145-153. doi: 10.1007/s10549-013-2528-8
 126. Humphrey, P. A.; Zhu, X.; Zarnegar, R.; Swanson, P. E.; Ratliff, T. L.; Vollmer, R. T. & Day, M. L. Hepatocyte growth factor and its receptor (c-MET) in prostatic carcinoma. *American J. Pathology*, 1995, **147**(2), 386.
 127. Falletto, D.L.; Kaplan, D.R.; Halverson, D.O.; Rosen, E.M. & Woude, G.V. Hepatocyte Growth Factor-Scatter Factor and the c-met Receptor. *IE Goldberg and EM Rosen, editors*, 1993, 107-130.
 128. Miyazawa, K.; Shimomura, T. & Kitamura, N. Activation of hepatocyte growth factor in the injured tissues is mediated by hepatocyte growth factor activator. *J. Biol. Chem.*, 1996, **271**(7), 3615-3618. doi: 10.1074/jbc.271.7.3615
 129. Lin, C. Y.; Anders, J.; Johnson, M.; Sang, Q. A. & Dickson, R. B. Molecular cloning of cDNA for matriptase, a matrix-degrading serine protease with trypsin-like activity. *J. Biol. Chem.*, 1999, **274**(26), 18231-18236. doi: 10.1074/jbc.274.26.18231
 130. Jiang, W. G.; Martin, T. A.; Parr, C.; Davies, G.; Matsumoto, K. & Nakamura, T. Hepatocyte growth factor, its receptor, and their potential value in cancer therapies. *Critical Reviews Oncology/Hematology*, 2005, **53**(1), 35-69. doi: 10.1016/j.critrevonc.2004.09.004
 131. Bardelli, A.; Longati, P.; Albero, D.; Goruppi, S.; Schneider, C.; Ponzetto, C. & Comoglio, P. M. HGF receptor associates with the anti-apoptotic protein BAG-1 and prevents cell death. *The EMBO Journal*, 1996, **15**(22), 6205.
 132. Boccaccio, C.; Gaudino, G.; Gambarotta, G.; Galimi, F. & Comoglio, P. M. Hepatocyte growth factor (HGF) receptor expression is inducible and is part of the

- delayed-early response to HGF. *J. Biol. Chem.*, 1994, **269**(17), 12846-12851.
133. Montesano, R. K. T. L.; Matsumoto, K.; Nakamura, T. & Orci, L. Identification of a fibroblast-derived epithelial morphogen as hepatocyte growth factor. *Cell*, 1991, **67**(5), 901-908. doi: 10.1016/0092-8674(91)90363-4
 134. Matsumoto, K. & Nakamura, T. HGF: its organotrophic role and therapeutic potential. In *Ciba Symp.*, 1997, **212**, pp. 198-211,
 135. Birchmeier, W.; Brinkmann, V.; Niemann, C.; Meiners, S.; DiCesare, S.; Naundorf, H. & Sachs, M. Role of HGF/SF and c-Met in Morphogenesis and Metastasis of Epithelial Cells. In *Ciba Foundation Symposium 212-Plasminogen-Related Growth Factors*. John Wiley & Sons, Ltd. 1997, January, pp. 230-251.
 136. Gao, C.F. & Woude, G.F.V. HGF/SF-Met signaling in tumor progression. *Cell Research*, 2005, **15**(1), 49-51. doi: 10.1038/sj.cr.7290264
 137. Birchmeier, C.; Birchmeier, W.; Gherardi, E. & Woude, G.F. V. Met, metastasis, motility and more. *Nature Reviews Molecular Cell Biology*, 2003, **4**(12), 915-925. doi: 10.1038/nrm1261
 138. Shinomiya, N.; Gao, C. F.; Xie, Q.; Gustafson, M.; Waters, D.J.; Zhang, Y.W. & Woude, G.F.V. RNA interference reveals that ligand-independent met activity is required for tumor cell signaling and survival. *Cancer Research*, 2004, **64**(21), 7962-7970. doi: 10.1158/0008-5472.CAN-04-1043
 139. Lee, J. H.; Han, S. U.; Cho, H.; Jennings, B.; Gerrard, B.; Dean, M. & Vande Woude, G. F. A novel germ line juxtamembrane Met mutation in human gastric cancer. *Oncogene*, 2000, **19**(43), 4947-4953. doi: 10.1038/sj.onc.1203874
 140. Ma, P. C.; Kijima, T.; Maulik, G.; Fox, E. A.; Sattler, M.; Griffin, J. D. & Salgia, R. c-MET Mutational Analysis in Small Cell Lung Cancer Novel Juxtamembrane Domain Mutations Regulating Cytoskeletal Functions. *Cancer Research*, 2003, **63**(19), 6272-6281.
 141. Lai, A. Z.; Abella, J.V. & Park, M. (2009, Crosstalk in Met receptor oncogenesis. *Trends Cell Biology*, **19**(10), 542-551. doi: 10.1016/j.tcb.2009.07.002
 142. Jo, M.; Stolz, D. B.; Esplen, J. E.; Dorko, K.; Michalopoulos, G. K. & Strom, S. C. Cross-talk between epidermal growth factor receptor and c-Met signal pathways in transformed cells. *J. Biol. Chem.*, 2000, **275**(12), 8806-8811. doi: 10.1074/jbc.275.12.8806
 143. Organ, S.L. & Tsao, M.S. An overview of the c-MET signaling pathway. *Therapeutic Advances Medical Oncology*, 2011, **3**(1 suppl), S7-S19. doi: 10.1177/1758834011422556
 144. Forbes, S.; Bhamra, G.; Bamford, S.; Dawson, E.; Kok, C.; Clements, J. & Stratton, M. R. The catalogue of somatic mutations in cancer (COSMIC). *Current Protocols Human Genetics*, 2008, 10-11. doi: 10.1002/0471142905.hg1011s57
 145. Kitajima, Y.; Ide, T.; Ohtsuka, T. & Miyazaki, K. Induction of hepatocyte growth factor activator gene expression under hypoxia activates the hepatocyte growth factor/c-Met system via hypoxia inducible factor-1 in pancreatic cancer. *Cancer Science*, 2008, **99**(7), 1341-1347. doi: 10.1111/j.1349-7006.2008.00828.x
 146. Cecchi, F.; Rabe, D.C. & Bottaro, D. P. Targeting the HGF/Met signaling pathway in cancer. *European J. Cancer*, 2010, **46**(7), 1260-1270. doi: 10.1016/j.ejca.2010.02.028
 147. Comoglio, P. M.; Giordano, S. & Trusolino, L. Drug development of MET inhibitors: targeting oncogene addiction and expedience. *Nature Reviews Drug Discovery*, 2008, **7**(6), 504-516. doi: 10.1038/nrd2530
 148. Parr, C. & Jiang, W.G. Hepatocyte growth factor activation inhibitors (HAI-1 and HAI-2) regulate HGF-induced invasion of human breast cancer cells. *International J. Cancer*, 2006, **119**(5), 1176-1183. doi: 10.1002/ijc.21881
 149. Cantiani, L.; Manara, M.C.; Zucchini, C.; De Sanctis, P.; Zuntini, M.; Valvassori, L. & Picci, P. Caveolin-1 reduces osteosarcoma metastases by inhibiting c-Src activity and met signaling. *Cancer Research*, 2007, **67**(16), 7675-7685. doi: 10.1158/0008-5472.CAN-06-4697
 150. Koga, F.; Tsutsumi, S. & Neckers, L.M. Low Dose Geldanamycin Inhibits Hepatocyte Growth Factor- and Hypoxia-Stimulated Invasion of Cancer Cells. *Cell Cycle*, 2007, **6**(11), 1393-1402. doi: 10.4161/cc.6.11.4296
 151. Barde, Y.A. The nerve growth factor family. *Progress Growth Factor Research*, 1990, **2**(4), 237-248. doi: 10.1016/0955-2235(90)90021-B
 152. Levi-Montalcini, R. The nerve growth factor: its mode of action on sensory and sympathetic nerve cells. *Harvey Lectures*, 1965, **60**, 217-259.
 153. Kaplan, D. R.; Hempstead, B. L.; Martin-Zanca, D.; Chao, M. V. & Parada, L. F. The trk proto-oncogene product: a signal transducing receptor for nerve growth factor. *Science*, 1991, **252**(5005), 554-558. doi: 10.1126/science.1850549
 154. Barbacid, M. The Trk family of neurotrophin receptors. *Journal of neurobiology*, 1994, **25**(11), 1386-1403. doi: 10.1002/neu.480251107
 155. Verdi, J.M.; Birren, S.J.; Ibáñez, C.F.; Persson, H.; Kaplan, D. R.; Benedetti, M. & Anderson, D.J. p75^{LNGFR} regulates Trk signal transduction and NGF-induced neuronal differentiation in MAH cells. *Neuron*, 1994, **12**(4), 733-745. doi: 10.1016/0896-6273(94)90327-1
 156. Dobrowsky, R.T.; Jenkins, G.M. & Hannun, Y.A. Neurotrophins induce sphingomyelin hydrolysis modulation by co-expression of p75^{NTR} with Trk receptors. *J. Biol. Chem.*, 1995, **270**(38), 22135-22142. doi: 10.1074/jbc.270.38.22135
 157. Blöchl, A. & Thoenen, H. Characterisation of nerve growth factor (NGF) release from hippocampal neurons: Evidence for a constitutive and an unconventional sodium-dependent regulated pathway. *European J. Neuroscience*, 1995, **7**(6), 1220-1228. doi: 10.1111/j.1460-9568.1995.tb01112.x
 158. Bilderback, T.R.; Grigsby, R.J. & Dobrowsky, R.T. Association of p75^{NTR} with caveolin and localization of neurotrophin-induced sphingomyelin hydrolysis to caveolae. *J. Biol. Chem.*, 1997, **272**(16), 10922-10927. doi: 10.1074/jbc.272.16.10922
 159. Molloy, N. H.; Read, D. E. & Gorman, A. M. Nerve growth factor in cancer cell death and survival. *Cancers*, 2011, **3**(1), 510-530. doi: 10.3390/cancers3010510
 160. Romon, R.; Adriaenssens, E.; Lagadec, C.; Germain, E.; Hondermarck, H. & Le Bourhis, X. Nerve growth factor promotes breast cancer angiogenesis by activating

- multiple pathways. *Molecular Cancer*, 2010, **9**(1), 1. doi: 10.1186/1476-4598-9-157
161. Truzzi, F.; Marconi, A.; Lotti, R.; Dallaglio, K.; French, L. E.; Hempstead, B. L. & Pincelli, C. Neurotrophins and their receptors stimulate melanoma cell proliferation and migration. *J. Investigative Dermatology*, 2008, **128**(8), 2031-2040. doi: 10.1038/jid.2008.21
162. Rende, M.; Rambotti, M.G.; Stabile, A.M.; Pistilli, A.; Montagnoli, C.; Chiarelli, M.T. & Mearini, E. Novel localization of low affinity NGF receptor (p75) in the stroma of prostate cancer and possible implication in neoplastic invasion: an immunohistochemical and ultracytochemical study. *The Prostate*, 2010, **70**(5), 555-561.
163. Zhu, Z.; Kleeff, J.; Kaye, H.; Wang, L.; Korc, M.; Büchler, M.W. & Friess, H. Nerve growth factor and enhancement of proliferation, invasion, and tumorigenicity of pancreatic cancer cells. *Molecular Carcinogenesis*, 2002, **35**(3), 138-147. doi: 10.1002/mc.10083
164. Kuner, P. & Hertel, C. NGF induces apoptosis in a human neuroblastoma cell line expressing the neurotrophin receptor p75NTR. *J. Neuroscience Res.*, 1998, **54**(4), 465-474. doi: 10.1002/(SICI)1097-4547(19981115)54:4<465::AID-JNR4>3.0.CO;2-T
165. Li, C.; MacDonald, J. I.; Hryciw, T. & Meakin, S. O. Nerve growth factor activation of the TrkA receptor induces cell death, by macropinocytosis, in medulloblastoma Daoy cells. *Journal Neurochemistry*, 2010, **112**(4), 882-899. doi: 10.1111/j.1471-4159.2009.06507.x
166. Harel, L.; Costa, B. & Fainzilber, M. On the death Trk. *Developmental Neurobiology*, 2010, **70**(5), 298-303. doi: 10.1002/dneu.20769
167. Descamps, S.; Toillon, R.A.; Adriaenssens, E.; Pawlowski, V.; Cool, S. M.; Nurcombe, V. & Hondermarck, H. Nerve growth factor stimulates proliferation and survival of human breast cancer cells through two distinct signaling pathways. *J. Biol. Chem.*, 2001, **276**(21), 17864-17870. doi: 10.1074/jbc.M010499200
168. Chiarenza, A.; Lazarovici, P.; Lempereur, L.; Cantarella, G.; Bianchi, A. & Bernardini, R. Tamoxifen inhibits nerve growth factor-induced proliferation of the human breast cancerous cell line MCF-7. *Cancer Research*, 2001, **61**(7), 3002-3008.
169. Marshall, J.L.; Kindler, H.; Deeken, J.; Bhargava, P.; Vogelzang, N.J.; Rizvi, N. & Hawkins, M. J. Phase I trial of orally administered CEP-701, a novel neurotrophin receptor-linked tyrosine kinase inhibitor. *Investigational New Drugs*, 2005, **23**(1), 31-37. doi: 10.1023/B:DRUG.0000047103.64335.b0
170. Zage, P. E.; Graham, T. C.; Zeng, L.; Fang, W.; Pien, C.; Thress, K. & Zweidler-McKay, P. A. The selective Trk inhibitor AZ623 inhibits brain-derived neurotrophic factor-mediated neuroblastoma cell proliferation and signaling and is synergistic with topotecan. *Cancer*, 2011, **117**(6), 1321-1391. doi: 10.1002/cncr.25674
171. Miknyoczki, S. J.; Chang, H.; Klein-Szanto, A.; Dionne, C. A. & Ruggeri, B. A. The Trk tyrosine kinase inhibitor CEP-701 (KT-5555) exhibits significant antitumor efficacy in preclinical xenograft models of human pancreatic ductal adenocarcinoma. *Clinical Cancer Res.*, 1999, **5**(8), 2205-2212.
172. Iyer, R.; Evans, A. E.; Qi, X.; Ho, R.; Minturn, J. E.; Zhao, H. & Brodeur, G. M. Lestaurtinib enhances the antitumor efficacy of chemotherapy in murine xenograft models of neuroblastoma. *Clinical Cancer Res.*, 2010, **16**(5), 1478-1485. doi: 10.1158/1078-0432.CCR-09-1531
173. Pasquale, E. B. Eph receptor signaling casts a wide net on cell behaviour. *Nature Rev. Mol. Cell Bio.*, 2005, **6**(6), 462-475. doi: 10.1038/nrm1662
174. Xi, H.Q.; Wu, X.S.; Wei, B. & Chen, L. Eph receptors and ephrins as targets for cancer therapy. *J. Cellular Molecular Medicine*, 2012, **16**(12), 2894-2909. doi: 10.1111/j.1582-4934.2012.01612.x
175. Genander, M. & Frisén, J. Ephrins and Eph receptors in stem cells and cancer. *Current Opinion Cell Bio.*, 2010, **22**(5), 611-616. doi: 10.1016/j.ceb.2010.08.005
176. Brantley-Sieders, D. M.; Fang, W. B.; Hicks, D. J.; Zhuang, G.; Shyr, Y. & Chen, J. Impaired tumor microenvironment in EphA2-deficient mice inhibits tumor angiogenesis and metastatic progression. *FASEB journal*, 2005, **19**(13), 1884-1886. doi: 10.1096/fj.05-4038fje
177. Kumar, S.R.; Singh, J.; Xia, G.; Krasnoperov, V.; Hassanieh, L.; Ley, E.J. & Weaver, F. A. Receptor tyrosine kinase EphB4 is a survival factor in breast cancer. *The American J. Pathology*, 2006, **169**(1), 279-293. doi: 10.2353/ajpath.2006.050889
178. Herath, N.I.; Doecke, J.; Spanevello, M.D.; Leggett, B.A. & Boyd, A. W. Epigenetic silencing of EphA1 expression in colorectal cancer is correlated with poor survival. *British J. Cancer*, 2009, **100**(7), 1095-1102. doi: 10.1038/sj.bjc.6604970
179. Xi, H. Q. & Zhao, P. Clinicopathological significance and prognostic value of EphA3 and CD133 expression in colorectal carcinoma. *J. Clinical Pathology*, 2011, **64**(6), 498-503. doi: 10.1136/jcp.2010.087213
180. Walker-Daniels, J.; Coffman, K.; Azimi, M. E.; Rhim, J.S.; Bostwick, D. G.; Snyder, P. & Kinch, M.S. Overexpression of the EphA2 tyrosine kinase in prostate cancer. *The Prostate*, 1999, **41**(4), 275-280. doi: 10.1002/(SICI)1097-0045(19991201)41:4<275::AID-PROS8>3.0.CO;2-T
181. Wykosky, J.; Gibo, D. M.; Stanton, C. & Debinski, W. EphA2 as a novel molecular marker and target in glioblastomamultiforme. *Molecular Cancer Res.*, 2005, **3**(10), 541-551. doi: 10.1158/1541-7786.MCR-05-0056
182. Stephenson, S. A.; Slomka, S.; Douglas, E. L.; Hewett, P. J. & Hardingham, J. E. Receptor protein tyrosine kinase EphB4 is up-regulated in colon cancer. *BMC Mol. Bio.*, 2001, **2**(1), 1. doi: 10.1186/1471-2199-2-15
183. Nakada, M.; Niska, J.A.; Miyamori, H.; McDonough, W. S.; Wu, J.; Sato, H. & Berens, M. E. The phosphorylation of EphB2 receptor regulates migration and invasion of human glioma cells. *Cancer Research*, 2004, **64**(9), 3179-3185. doi: 10.1158/0008-5472.CAN-03-3667
184. Kinch, M. S. & Carles-Kinch, K. Overexpression and functional alterations of the EphA2 tyrosine kinase in cancer. *Clinical Experimental Metastasis*, 2003, **20**(1), 59-68. doi: 10.1023/A:1022546620495
185. Easty, D. J.; Herlyn, M. & Bennett, D.C. Abnormal protein tyrosine kinase gene expression during melanoma progression and metastasis. *Int. J. Cancer*, 1995, **60**(1), 129-136. doi: 10.1002/ijc.2910600119
186. Shao, Z.; Zhang, W. F.; Chen, X. M. & Shang, Z. J. Expression of EphA2 and VEGF in squamous cell carcinoma of the tongue: correlation with the angiogenesis and clinical

- outcome. *Oral Oncology*, 2008, **44**(12), 1110-1117. doi: 10.1016/j.oraloncology.2008.01.018
187. Coffman, K.T.; Hu, M.; Carles-Kinch, K.; Tice, D.; Donacki, N.; Munyon, K. & Kinch, M.S. Differential EphA2 epitope display on normal versus malignant cells. *Cancer Research*, 2003, **63**(22), 7907-7912.
188. Wang, X.D.; Reeves, K.; Luo, F.R.; Xu, L.A.; Lee, F.; Clark, E. & Huang, F. Identification of candidate predictive and surrogate molecular markers for dasatinib in prostate cancer: Rationale for patient selection and efficacy monitoring. *Genome Bio.*, 2007, **8**(11), R255 doi: 10.1186/gb-2007-8-11-r255
189. Landen, C.N.; Chavez-Reyes, A.; Bucana, C.; Schmandt, R.; Deavers, M.T.; Lopez-Berestein, G. & Sood, A. K. Therapeutic EphA2 gene targeting in vivo using neutral liposomal small interfering RNA delivery. *Cancer Research*, 2005, **65**(15), 6910-6918. doi: 10.1158/0008-5472.CAN-05-0530
190. Maruyama, E.; Sakamoto, T.; Azuma, H.; Ito, Y.; Katsuoka, Y. & Otsuki, Y. Involvement of angiopoietins in cancer progression in association with cancer cell-fibroblast interaction. *Anticancer Research*, 2005, **25**(1A), 171-177.
191. Davis, S.; Aldrich, T. H.; Jones, P. F.; Acheson, A.; Compton, D. L.; Jain, V. & Yancopoulos, G. D. Isolation of angiopoietin-1, a ligand for the TIE2 receptor, by secretion-trap expression cloning. *Cell*, 1996, **87**(7), 1161-1169. doi: 10.1016/S0092-8674(00)81812-7
192. Suri, C.; Jones, P.F.; Patan, S.; Bartunkova, S.; Maisonpierre, P.C.; Davis, S. & Yancopoulos, G.D. Requisite role of angiopoietin-1, a ligand for the TIE2 receptor, during embryonic angiogenesis. *Cell*, 1996, **87**(7), 1171-1180. doi: 10.1016/S0092-8674(00)81813-9
193. Maisonpierre, P.C.; Suri, C.; Jones, P.F.; Bartunkova, S.; Wiegand, S.J.; Radziejewski, C. & Daly, T.J. Angiopoietin-2, a natural antagonist for Tie2 that disrupts in vivo angiogenesis. *Science*, 1997, **277**(5322), 55-60. doi: 10.1126/science.277.5322.55
194. Schlingemann, R.O. Role of growth factors and the wound healing response in age-related macular degeneration. *Graefe's Archive Clinical Experimental Ophthalmology*, 2004, **242**(1), 91-101. doi: 10.1007/s00417-003-0828-0
195. Ashraf, S. & Walsh, D. A. Angiogenesis in osteoarthritis. *Current Opinion Rheumatology*, 2008, **20**(5), 573-580. doi: 10.1097/BOR.0b013e3283103d12
196. Sfiligoi, C.; de Luca, A.; Cascone, I.; Sorbello, V.; Fuso, L.; Ponzzone, R. & Sismondi, P. Angiopoietin-2 expression in breast cancer correlates with lymph node invasion and short survival. *Int. J. Cancer*, 2003, **103**(4), 466-474. doi: 10.1002/ijc.10851
197. Kobayashi, H. & Lin, P. C. Angiopoietin/Tie2 signaling, tumor angiogenesis and inflammatory diseases. *Front Biosci*, 2005, **10**, 666-674. doi: 10.2741/1561
198. Creamer, D.; Sullivan, D.; Bicknell, R. & Barker, J. N. W. N. Angiogenesis in psoriasis. *Angiogenesis*, 2002, **5**(4), 231-236. doi: 10.1023/A:1024515517623
199. Rini, B.; Szczylik, C.; Tannir, N.M.; Koralewski, P.; Tomczak, P.; Deptala, A. & Rogowski, W. AMG 386 in combination with sorafenib in patients with metastatic clear cell carcinoma of the kidney. *Cancer*, 2012, **118**(24), 6152-6161. doi: 10.1002/cncr.27632
200. Gerald, D.; Chintharlapalli, S.; Augustin, H.G. & Benjamin, L.E. Angiopoietin-2: an attractive target for improved antiangiogenic tumor therapy. *Cancer Research*, 2013, **73**(6), 1649-1657. doi: 10.1158/0008-5472.CAN-12-4697
201. Ruggeri, B.; Underiner, T.; Gingrich, D.; Hudkins, R.; Angeles, T.; Albom, M. & Jones-Bolin, S. CEP-11981: A potent TIE-2/Pan-VEGF-R inhibitor with broad kinase inhibitory activity exhibits significant antitumor and antiangiogenic efficacy in preclinical tumor models. *Molecular Cancer Therapeutics*, 2007, **6**(11 Supplement), B264-B264.
202. Lowy, I.; Thurston, G. & Daly, C. Washington, DC: U.S. Patent and Trademark Office. 2015, U.S. Patent No. 8,980,268.