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Abstract: Domperidone and Itopride are pro-kinetic agents, regulating the gastric motility and are commonly prescribed as anti emetic drugs. In the present study a simple, rapid and sensitive RP-HPLC/UV method was developed for simultaneous determination of Domperidone and Itopride in pharmaceutical samples and human plasma, using Tenofavir as internal standard. Experimental conditions were optimized and method was validated according to the standard guidelines. Combination of water (pH 3.0) and acetonitrile (65:35 v/v) was used as mobile phase, pumped at the flow rate of 1.5ml/min. Detector wavelength was set at 210nm and column oven temperature was 40oC. Unlike conventional liquid-liquid extraction, simple precipitation technique was applied for drug extraction from human plasma using acetonitrile for deprotienation.
The method showed adequate separation of both the analytes and best resolution was achieved using Hypersil BDS C8 column (150mm x 4.6mm, 5µm). The method was quite linear in the range of 20 - 600ng/ml. Recovery of the method was 92.31% and 89.82% for Domperidone and Itopride, respectively. Retention time of both the analytes and internal standard was below 15min. The lower limit of detection (LLOD) and lower limit of quantification (LLOQ) for Domperidone were 5 and 10ng/ml while for Itopride was 12 and 15ng/ml, respectively.
The developed method was successfully applied for in-vivo analysis of fast dispersible tablets of Domperidone in healthy human volunteer. The proposed method was a part of formulation development study and was efficiently applied for determination of the two drugs in various pharmaceutical products and human plasma.

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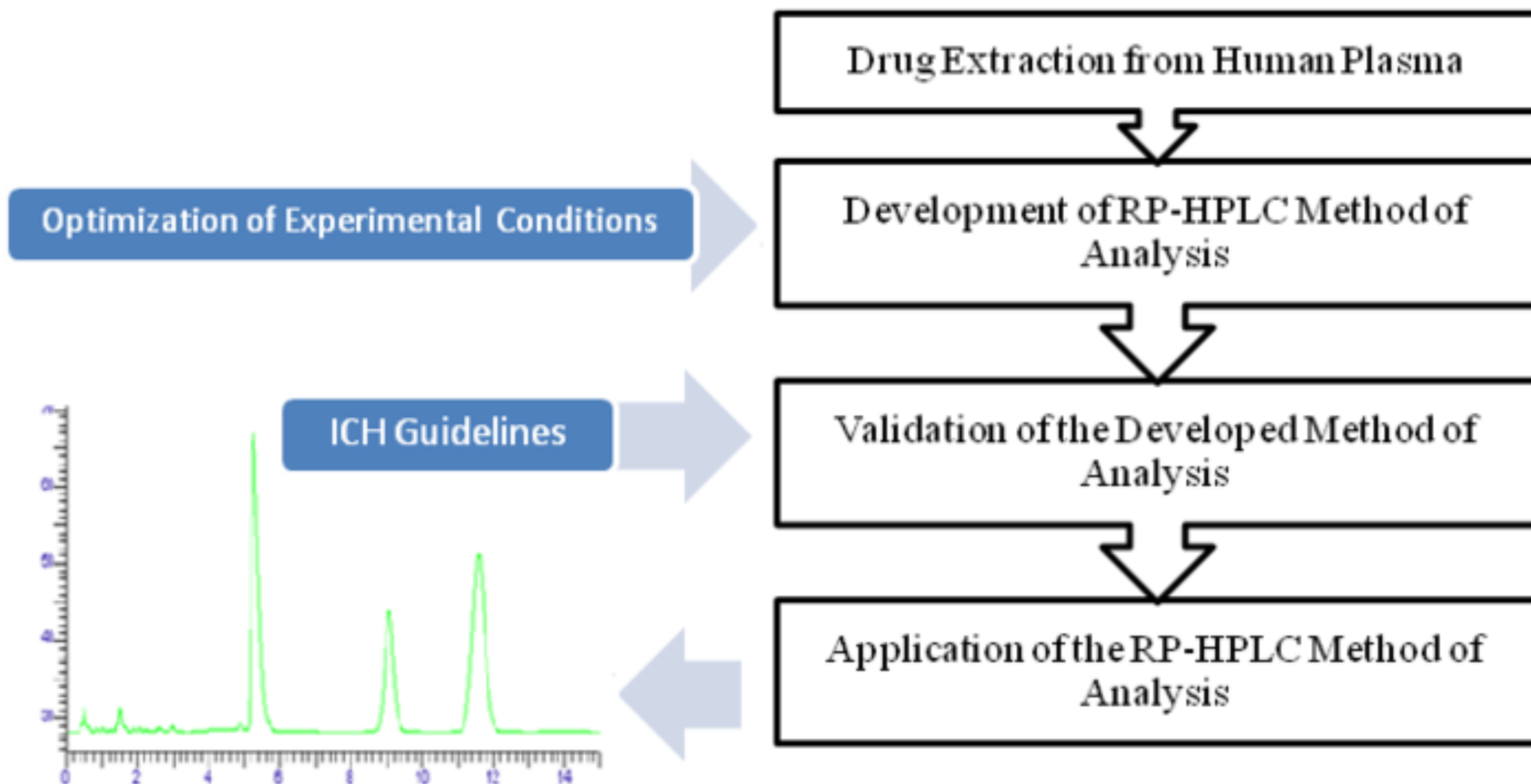
*Graphical Abstract



*Highlights (for review)

Simultaneous Determination of Domperidone and Itopride in Pharmaceuticals and Human plasma using RP-HPLC/UV Detection: Method Development, Validation and Application of the Method in *In-vivo* Evaluation of Fast Dispersible Tablets

Amjad Khan, Zafar Iqbal, Ibrahim Khadra, Lateef Ahmad, Muhammad Imran Khan, Abad Khan, Zia Ullah, Ismail.

Highlights:

- Development and validation of method for simultaneous determination of Domperidone and Itopride in pharmaceuticals and human plasma
- Application of the developed method for *in-vivo* evaluation of fast dispersible tablets of Domperidone
- The developed method has simple mobile phase composition, shorter analysis time and was successfully applied for analysis of Domperidone and Itopride in pharmaceuticals and biological fluids (human plasma)

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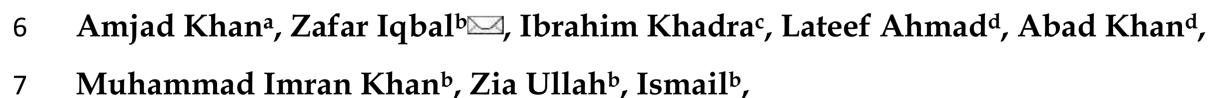
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1 **1. Introduction**

2 Domperidone (6-chloro-3-[1-[3-(2-oxo-3H-benzimidazol-1-yl) propyl] piperidin-4-yl]-1H
3 benzimidazol-2-one) is structurally related to butyrophenones having molecular formula of
4 $C_{22}H_{24}ClN_5O_2$ and molecular weight of 425.91g/mol **[1, 2]**. It is a class-2 drug having pH
5 dependent solubility. Experimental water solubility of Domperidone is 0.986mg/L and its Pka
6 value is 7.9 **[3]**. Domperidone is commonly prescribed as anti emetic with daily recommended
7 dose of 10 – 40 mg in divided doses.

8 Itopride (N-[[4-(2-dimethylaminoethoxy) phenyl] methyl]-3, 4-dimethoxy-benzamide HCl) is a
9 prokinetic agent. Chemically it is a substituted benzamide with molecular formula of
10 $C_{20}H_{26}N_2O_4.HCl$ and molecular weight of 394.89g/mol having pKa value of 8.72 **[4, 5]**.
11 Physically it is a crystalline solid and freely water soluble. Recommended dose of Iopride for
12 adult patients is 150mg/day in divided doses **[6]**. Structural formulae of Domperidone and
13 Itopride are presented in **Fig-1**.

14 Both Domperidone and Itopride are prescribed for treatment of gastric esophageal reflux
15 disorders (GERD) and are commonly used anti emetic **[7]**. Domperidone and Itopride regulate
16 the gastric motility **[8, 9]** by;

17 - Antagonizing inhibitory effect of dopamine at D_2 receptors
18 - Increasing stimulatory effect of acetylcholine by blocking its metabolizing enzyme
19 acetylcholine esterase

20 To date several reversed phase high performance liquid chromatography (RP-HPLC) methods
21 have been reported for analysis of Domperidone and Itopride. Most of the methods are
22 applicable only for analysis of the drugs in various pharmaceutical dosage forms **[10 – 19]**. Due
23 to lack of sensitivity, these methods cannot be applied for analysis of the drugs in biological
24 samples. Few methods exhibited better sensitivity but involved derivatization technique and post
25 column photochemical reactions **[12]**. In these methods drug was extracted from biological
26 matrix using tedious and costly technique of solid phase extraction (SPE). In most of the reported
27 methods, mobile phase composition is very complex, using buffer solutions as aqueous phase
28 **[13, 14, 16]**.

29 It was required to develop a rapid, sensitive and simple method involving no derivatization or
30 other advanced technique. The aim of the presented work was to develop a sensitive, rapid and
31 simple RP-HPLC/UV method for simultaneous determination of Domperidone and Itopride in

pharmaceutical and biological samples (human plasma). The suggested method is economical, involves simple extraction procedure, requires lesser sample volume and possesses a shorter chromatographic analysis run time as compared to other reported methods. The developed method was successfully applied for *in-vivo* evaluation of fast dispersible tablets of Domperidone in human volunteers.

2. Experimental

2.1 Chemicals and reagents

Domperidone (purity 99.9%) was provided by Medicraft Pharmaceuticals, Peshawar, Pakistan, Itopride (purity 99.75%) and Tenofovir were gifted by Ferozsons Laboratories Ltd. Nowshera, Pakistan. HPLC grade acetonitrile (purity 99.9%), methanol (purity 99.9%) and analytical grade ortho-phosphoric acid (OPA; purity 85%) were purchased from Sigma–Aldrich (Oslo, Norway). Ultra pure HPLC grade water was prepared by Milli-Q® system (Millipore, Milford, MA, USA).

2.2. Instrumentation

Chromatographic analysis was performed using HPLC system equipped with an auto sampler, vacuum degasser, peltier column oven, pump, and UV–visible detector (Perkin Elmer Series 200 system, Norwalk, USA). The chromatographic data was analyzed on Perkin Elmer Total-chrom workstation software (version 6.3.1, Bridgeport, USA) linked with the LC-system through network chromatography interface (NCI) 900. Separation of the analytes was carried out using Hypersil BDS C8 column (150mm x 4.6mm, 5µm) protected by a Perkin Elmer pre-column guard cartridge C18 (30mm × 4.6mm, 10µm; Norwalk, USA).

2.3 Optimization of chromatographic conditions

Different mobile phases, consisting of acetonitrile, acidified water (pH adjusted to 3.0 with OPA; 35:65, v/v), methanol, acetonitrile and acidified water (pH adjusted to 3.0 with OPA; 25:25:50 v/v) and methanol, acidified water (25:75, v/v), pumped with different flow rates in the range of 0.5 – 2.0 ml/min in an isocratic mode were evaluated. Chromatographic analyses were performed at different column oven temperatures in the range of 25 – 50 ^{o}C. The injection volume was studied in the range of 10 – 50 µl.

2.4 Preparation of standard solutions

Stock solutions of Domperidone and Itopride and Tenofovir (concentration = 1.0mg/ml) were prepared in methanol and water, separately, on the basis of their respective solubility and stored

1 at –4°C. Stock solutions were diluted with the mobile phase (acetonitrile: water, 35:65, v/v) to
2 obtain various dilutions in the range of 10 – 600ng/ml (i.e., 10, 50, 100, 200, 400, 500 and
3 600ng/ml).
4 **2.5 Sample preparation**
5 Stock solution of both analytes and internal standard were prepared separately in water and
6 organic solvent (methanol), on the basis of their respective solubility. Stock solutions of
7 Domperidone and internal standard (Tenofovir) were prepared in methanol while purified water
8 was used for preparation of Itopride solution. Concentration of the stock solutions of all the
9 compounds was 1mg/ml. Working solutions (10ml) of both analytes were prepared by dilution of
10 stock solution with mobile phase on daily basis.
11 Blood samples (about 3ml) collected from healthy human volunteers in the ethylene diamine
12 tetraacetic acid (EDTA) tubes were centrifuged at 4000 × g for 10min at 4°C to separate plasma.
13 The isolated plasma was deproteinated with acetonitrile. Aliquot of plasma (200µl) was taken in
14 Eppendorf tube (capacity = 1ml), acetonitrile (600µl) was added and vertexed for 2min. Sample
15 was centrifuged at 4000 x g for 10min and supernatant was collected using posture pipette. The
16 separated plasma was stored at –20°C till further use.
17 On the day of analysis, plasma samples were thawed at room temperature and spiked with
18 appropriate concentrations of working solutions to obtain calibration standards {in the range 10 –
19 600ng/ml (Domperidone) and 15 – 600ng/ml (Itopride)}. The calibration curves covered the
20 maximum therapeutic concentrations achieved by both the drug *in-vivo*. The amount of stock
21 solution in all spiked samples was kept below 2.0% of the total sample volume to minimize any
22 systematic errors between real samples and standards.
23 **2.6 Sample collection and extraction**
24 Plasma samples (200µl) were spiked with known amount of Domperidone, and Itopride, internal
25 standard solution (10µl) was added and mixed for 10s. Unlike traditional liquid-liquid extraction,
26 simple precipitation technique was applied for drug extraction from biological sample. That is
27 the sample was deproteinated with acetonitrile and diluted with mobile phase. Schematic
28 presentation of extraction procedure is shown in **Fig-2**.
29
30
31

2.7 Validation of the developed method

The suggested method was validated according to standard guidelines **[20 – 22]** with emphasis on linearity, precision, specificity/selectivity, accuracy, sensitivity (lower limit of detection and lower limit of quantification), sample stability, and robustness.

2.7.1 Linearity

Linearity of the method was evaluated from the calibration curves constructed at seven different concentration points of both the analytes in spiked plasma and mobile phase (standard solutions). The concentration range for calibration curve was 10 – 600ng/ml. The ratio of the peak area of each analyte to the area of internal standard was plotted against their corresponding concentration **[23, 24]** and slope (a), intercept (b) correlation coefficient (r) and covariance (%RSD) were determined from the regression analysis using the least squares method.

2.7.2 Precision

Precision of the method was determined in terms of injection repeatability and analysis repeatability of spiked plasma samples. In order to determine injection repeatability, plasma samples spiked with 250ng/ml each of Domperidone and Itopride were injected (n = 5) into HPLC. Peak area and retention time of both the drugs were determined and results were expressed as mean and covariance (mean; %RSD). Analysis repeatability was verified by determining the recovered amount from plasma samples spiked with known concentration of both drugs (Domperidone and Itopride). Results were presented as mean; %RSD (n = 5).

To determine intermediate precision (intra-day and inter day reproducibility), plasma samples spiked with different concentrations of Domperidone and Itopride were analyzed three times a day (at 8 hours interval) on day-1 (intraday). Inter day study was carried out by analyzing the same sample (spiked samples used in intraday study) on daily basis for next three days. The results were presented as mean ± standard deviation; covariance (n = 5).

2.7.3 Selectivity/specificity

The specificity of the method was evaluated by verifying the complete separation of the peaks of both the analytes and internal standard in standard solutions (solutions of both analytes prepared in mobile phase) and spiked plasma samples **[23]**.

2.7.4 Accuracy

Percent recovery was used for determination of accuracy of the proposed method **[24]**. Percent recovery was calculated by spiking the biological sample at three appropriately nominated

concentrations (100, 250 and 500ng/ml) of both the analytes. Concentration of internal standard (Tenofovir) was kept constant and extraction was made using mobile phase. Sample (20µl) was injected into the HPLC system in triplicate (n = 3) and percent recovery was calculated using following equation;

$$\%\ \text{Recovery} = [A / B] \times 100 \text{ --------------- Eq-1}$$

Where

A = Peak area response ratios of the analytes with reference to IS in the plasma samples

B = Peak area response ratios of the analytes with reference to IS mobile phase

Amount recovered from each sample was calculated using the following equation;

$$C = (X/Y) \times (A/B) \times Cs \times F_D \text{ ----------------- Eq-2}$$

Where

X and Y = Peak areas of the analyte in plasma samples and 1:1 mixture, respectively

A and B = Peak areas of the internal standard in 1:1 mixture and plasma samples,

Cs = Concentration of analyte in the 1:1 mixture

FD = Dilution factor

Standard deviation (SD) and covariance (%RSD) were calculated for each sample

2.7.5 Sensitivity

Sensitivity of the method was evaluated on the basis of determination of the lower limit of detection (LLOD) and lower limit of quantification (LLOQ) for both the analytes. Signal to noise ratio (S/N) was determined for each analyte using HPLC software. Lower limit of detection was the concentration at which signal to noise ratio was 3 (S/N ≈ 3) and lower limit of quantification (LLOQ) was the concentration at which S/N ratio was 10 (S/N ≈ 10).

2.7.6 Sample stability

1 Stability of both the analytes was evaluated by keeping samples at various temperatures. Samples
2 of both the analytes were stored at room temperature and –20 °C for one week and analyzed.
3 Percent stability and percent loss were calculated using the following equations **[24, 25]**;

4 $\% \, Stability = S_t / S_0 \times 100$ ------------------------ Eq-3

5 $\% \, Loss = (S_0 - S_t) \, / \, S_0 \times 100$ ------------------ Eq-4

6 Where

7 S_t = Stability of analyte at time t

8 S_0 = Stability at initial time (stability of fresh solution)

9

10 **2.7.7 Robustness**

11 Robustness of the method was estimated by changing various experimental conditions and
12 evaluating their effect on peak area, peak height and retention time of both the analytes. Small
13 deliberate changes were made in different chromatographic condition like column oven
14 temperature (±5°C), mobile phase composition, mobile phase flow rate (± 0.2 ml/min), detector
15 wave length (± 2nm) and detector voltage (± 50 mV).

16 **2.8 Statistical interpretation and correlation of data**

17 Various statistical tools such as mean (X), standard deviation (SD) and relative standard
18 deviation (%RSD) were calculated using MS Excel and applied for quantification of both the
19 analytes.

20 **2.9 Application of the method in pharmacokinetic evaluation of fast dispersible tablets of**
21 **Domperidone**

22 The method was transferred to the Biopharmaceutical Laboratory of the Department of
23 Pharmacy, University of Peshawar, Pakistan. The method transfer was controlled by a procedure
24 including development of a transfer plane, definition of transfer tests and acceptance criteria
25 (sample analysis: sample time and replication, validation experiments), training of laboratory
26 staff and full method validation. The method parameters were identical with regard to mobile
27 phase composition, HPLC column, detector wavelength and extraction procedure. The HPLC
28 system was different and minor modifications to the laboratory procedures had to be made in
29 order to adapt it to the local conditions. The clinical application of the method was assessed by
30 measuring Domperidone in plasma samples of healthy human volunteers (Pharmacy Graduates
31 in the Department of Pharmacy, University of Peshawar). The study was conducted according to

the guidelines of "World Medical Associations, Declaration of Helsinki-ethical principles for medical research involving human subjects" and was approved by the institutional "Committee for Research Ethics", Department of Pharmacy, University of Peshawar, under license number "UOP/PHARM-EC/387".

Fast dispersible tablet of Domperidone was dispersed in a glass of water (200ml) and taken by each volunteer. Blood samples (3ml) were obtained at specified time intervals (0, 5, 15, 30, 60, 120, 180, 240, 300 and 360min) following tablet administration. Blood samples were collected in EDTA tubes and centrifuged at 4000 x g for 10min at 4 ^{o}C to separate the plasma.

Deproeination of plasma samples was carried out using acetonitrile. An aliquot of plasma (200µl), together with internal standard was taken in eppendorf tube, acetonitrile (3 times volume of plasma sample) was added to it, vertexed for 2min and centrifuged at 4000 x g for 5min. Supernatant was isolated in a separate eppendorf tube (1ml capacity), volume was made up with mobile phase and vertexed for 1min. The plasma samples, isolated in separate eppendorf tubes, were stored at –20 ^{o}C till further analysis. On the day of analysis, the frozen samples were thawed at room temperature. The samples were analyzed in triplicate to ensure satisfactory method performance in accordance with the standard guidelines.

The plasma concentration of Domperidone in human plasma samples was quantified at various time intervals following oral administration of fast dispersible tablets. The data was fitted in the compartmental models to calculate various pharmacokinetic parameters such as t_{max}, C_{max}, Half life (t ½) and area under cure (AUC). The pharmacokinetic data was assessed using Microsoft Excel 2007 and PK-Summit®, a pharmacokinetics software.

3. Results and Discussion

Domperidone and Itopride were analyzed for first time simultaneously using Tenofavir as internal standard. Various experimental conditions were optimized and method was validated according to the ICH guidelines **[22]**. The proposed method was a rapid one and all the analytes were separated within 15min. Of the various analytical conditions studied, best results were achieved with the Hypersil BDS C8 Column (150mm x 4.6mm, 5µm) column, flushed with combination of acetonitrile and acidified water (35:65, v/v) as the mobile phase, pumped at a flow rate of 1.5ml/min at 40 ^{o}C.

3.1 Sample preparation

Methanol, acetonitrile and 1:1 mixture of methanol and acetonitrile were evaluated for protein precipitation; however better protein precipitation was achieved with acetonitrile. Volume of acetonitrile at least 3 times the volume of plasma was used for protein precipetation.

3.2 Optimization of analytical conditions and experimental parameters

Hypersil BDS C8 Column (150 x 4.6mm, 5µm) column was used for separation of Domperidone and Itopride. Other columns like Discovery HS C18 column (150mm × 4.6mm, 5µm), Symmetry C8 column (150mm × 3.9mm, 5µm) and Symmetry C8 (250mm × 4.6mm, 5µm) were also tried. With these peaks overlapping and broadening were observed for both analytes. Hypersil BDS C8 (150mm x 4.6mm, 5µm) column was selected as the best on the basis of excellent peak parameters (retention, separation, tailing, asymmetry, and resolution) and shorter run time.

Different combinations of organic solvents (methanol and acetonitrile) and water were used as mobile phase. Water used in preparation of mobile phase was acidified (pH adjusted to 3) with ortho phosphoric acid (85%) avoiding use of buffer solution. Different isocratic solvent systems, consisting of acetonitrile–acidified water (25:75, 35:65 and 50:50, v/v), methanol–acidified water (25:75 and 50: 50, v/v) and methanol–acetonitrile-acidified water (25:25: 50, v/v) were assessed as the mobile phases. In comparison to methanol, acetonitrile resulted in better peak area and peak shape so acetonitrile was included as organic solvent in mobile phase.

It was noticed that increase in ratio of acetonitrile in mobile phase decreased retention time of all the compounds resulting in shorter analysis time but peak area and peak height were significantly affected. Domperidone is water insoluble compound and by increasing ratio of acetonitrile in mobile phase resulted in increased sensitivity of the method for Domperidone. Similarly area of Itopride reduced with increasing quantity of acetonitrile in mobile phase. Combination of acetonitrile and acidified water (35: 65, v/v) resulted in the best results in terms of peak shape, area, retention time and resolution and was selected as mobile phase for further analysis.

Mobile phase of different pH was prepared by changing pH of water in the range of 2 – 5 with *O*-phosphoric acid. Sensitivity and retention time of all the compounds were significantly affected by pH of the mobile phase. At higher pH sensitivity of the method was reduced and retention time was increased for all the compounds. Effect of changes in pH was more prominent with Domperidone as its retention time was increased up to 20min with very low sensitivity **(Fig-3)**. At lower pH retention time decreased but separation between Domperidone and Itopride

was also reduced significantly. Best peak shape with better resolution and sensitivity were obtained at pH 3 and was selected as optimum pH of mobile phase for further analysis.

Retention time, peak shape and peak area were significantly affected by flow rate of the mobile phase. Best results were obtained at flow rate of 1.5ml/min and selected as optimum flow rate for further analysis.

Column oven temperature was studied in the range of 25 – 50 ^{o}C and best results were obtained at 40 ^{o}C. At higher column oven temperature, sensitivity of the method was increased with improved peak height. Better peak shape and resolution were obtained at 40 ^{o}C and selected as column oven temperature for simultaneous determination of the two drugs.

Internal standard was selected on the basis of its specificity, sensitivity, recovery and compatibility with Domperidone and Itopride. Various compounds like Ciprofloxacin, Naproxin Sodium, Tenofovir and Atenolol were tried as internal standard. Among them Tenofovir exhibited better recovery and peak separation and was used as internal standard.

3.3 Method validation

The proposed method of analysis was validated according to standard guidelines in terms of selectivity, sensitivity, recovery, precision and robustness.

3.3.1 Specificity and selectivity

The proposed method of analysis was highly selective and specific as peaks of both analytes and internal standard were well resolved. Separation of peaks was confirmed by using blank plasma, plasma spiked with both analytes and internal standard. The method is well suitable for analysis of Domperidone and Itopride in pharmaceutical solutions as well as biological samples.

3.3.2 Linearity

Linearity of the method was determined from calibration curve of the standard mixtures and spiked plasma samples. Calibration curves were constructed at seven concentration levels in the range of 10 – 600ng/ml for Domperidone and 15 – 600ng/ml for Itopride, separately for standard mixture and spiked plasma samples. Regression analysis of calibration curves of both drugs showed that the method is quite linear within the said concentration range. Regression equation and correlation coefficient values of calibration curves of both the drugs are presented in **Table-1**.

1 **3.3.3 Accuracy of the method (percent recovery)**
2 Percent recovery from plasma was used for determination of accuracy of the proposed method.
3 Percent recovery was determined at three concentration levels (100ng/ml, 250ng/ml and
4 500ng/ml) of both drugs and was found above 91% and 89% for Domperidone and Itopride,
5 respectively **(Table-2)**.
6 **3.3.4 Precision of the method**
7 Precision of the method was evaluated on the basis of repeatability and intermediate precision.
8 Intermediate precision was determined in terms of intra and inter day reproducibility. Results of
9 analysis repeatability, inter day precision and intraday precision are presented in **Table-2**. There
10 is complete harmony among repeated injections, repeated analysis, inter day and intraday study.
11 **3.3.5 Stability of Solutions**
12 Stability study was conducted at room temperature (18 – 23 ^{o}C) and freezer temperature (-20 ^{o}C)
13 for one week. Both Domperidone and Itopride were stable during the freeze/thaw cycles and for
14 at least 24hr in each step of the analysis. All the samples were stable at –20 ^{o}C for the specified
15 period. At room temperature Domperidone and internal standard (Tenofavir) remained stable
16 while Itopride degraded to a significant level. The percent degradation of drugs in both spiked
17 plasma samples and standard solutions stored at room temperature was higher as compared to the
18 samples stored at –20 ^{o}C. It revealed that all the samples containing Itopride should be stored at –
19 20 ^{o}C to avoid stability issue.
20 **3.3.6 Sensitivity (Limit of Detection and Lower Limit of Quantification)**
21 Lower limit of detection (LLOD) and lower limit of quantification (LLOQ) of the two drugs are
22 quite low as presented in **Table-1**. Values of LLOD and LLOQ proved that the proposed method
23 was more sensitive than the previously reported method for the drugs. Respective
24 chromatograms of LLOD and LLOQ of both drugs are given in **Fig-4**.
25 **3.3.7 Robustness**
26 Minor deliberate changes in analytical parameters did not significantly affect retention times,
27 percent recovery and peak area of both the drugs. Thus, the method was robust for minor
28 changes in column oven temperature, i.e., 40 ± 2 ^{o}C, flow rate, i.e., 1.5 ± 0.2ml/min and pH of
29 the mobile phase (±0.1 units).
30
31

4. Applicability of the method

This method was a part of an extensive formulation development process involving *in-vitro* and *in-vivo* characterization of novel formulations of fast dispersible tablet of Domperidone and Itopride. The developed method was successfully applied for in-vitro evaluation (drug excipients compatibility study, determination of drug content and dissolution rate of compressed tablets; *data not shown*) and determination of pharmacokinetics of Domperidone in healthy human volunteers, following oral administration of fast dispersible tablets. The pharmacokinetics study of fast dispersible tablets of Domperidone (10mg) was carried out in healthy human volunteers (age range = 24 – 29 years). The study was approved by the "Committee for Research Ethics" of Department of Pharmacy, University of Peshawar and written consent form was signed by each volunteer. Drug content of each sample was determined and pharmacokinetic parameters were calculated using pharmacokinetics software PK Summit.

Applicability of the method is illustrated in **Fig-5** showing mean plasma concentration versus time curve for Domperidone. Various pharmacokinetics parameters determined in healthy human volunteers (n = 6) are summarized in **Table-3**.

5. Conclusion

Using isocratic RP-HPLC with UV detection mode, a simple, rapid and reproducible method for simultaneous determination of Domperidone and Itopride in human plasma was developed, optimized and validated. Various experimental parameters such as stationary phase, mobile phase composition, flow rate, pH of the mobile phase, column oven temperature and internal standard were optimized. The proposed method was also validated on the basis of linearity, accuracy, precision, specificity, stability and robustness.

The suggested method offered the advantages of most simple extraction procedure, a short run time and a small sample volume for analysis. The method is robust, precise and easy to automate. Baseline separation of Domperidone, Itopride and internal standard was achieved quickly with no peak interference from endogenous compounds in biological samples. This method could be efficiently applied for the analysis of Domperidone and Itopride in pharmaceuticals and biological sample.

6. Acknowledgement

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Table-1: Calibration range, linearity and sensitivity of the method

Parameters	**Analytes**	
	Domperidone	**Itopride HCl**
Calibration range (ng/ml)	10 – 600	15 – 600
Linearity		
Standard mixture		
Regression equation	y = 0.013x +0.017	y = 0.007x +0.006
Correlation co efficient	0.998	0.999
Spiked plasma samples		
Regression equation	y = 0.011x + 0.021	y = 0.005x +0.001
Correlation co efficient	0.997	0.996
Sensitivity		
Limit of detection (LOD)	5ng/ml	12ng/ml
Lower limit of quantification (LLOQ)	10ng/ml	15ng/ml

Table-2: Recovery and precision of the method

Parameter	Analytes	
	Domperidone	Itopride HCl
Recovery		
Spiked conc. (100ng/ml)	[a](91.26); 0.464	[a](89.82); 0.87
Spiked conc. (250ng/ml)	[a](91.10); 0.958	[a](89.38) ; 1.22
Spiked conc. (500ng/ml) (n = 5)	[a](92.31); 0.502	[a](89.78); 1.98
Precision		
Injection repeatability		
Spiked conc. (250ng/ml)	[b]32670; 1.28	[b]24514; 1.097
Spiked conc. (250ng/ml) (n = 5)	[c]11.39; 1.15	[c]8.37; 0.764
Analysis repeatability		
Spiked conc. (500ng/ml) (n = 5)	[d]0.421; 3.8	[d]0.40; 2.125
Intermediate precision		
Intraday reproducibility		
Spiked conc. (100ng/ml)	[d]91.18; 1.54	[d]83.95; 3.74
Spiked conc. (250ng/ml)	[d]207.50; 2.71	[d]199.12; 3.02
Spiked conc. (500ng/ml) (n = 5)	[d]418.61; 3.19	[d]399.83; 2.76
Inter day reproducibility		
Spiked conc. (100ng/ml)	[d]89.54; 4.13	[d]85.26; 4.58
Spiked conc. (250ng/ml)	[d]209.26; 4.16	[d]189.58; 3.97
Spiked conc. (500ng/ml) (n = 5)	[d]411.59; 1.67	[d]395.30; 4.76

Results are presented as (mean); %RSD

a: percent recovery; b: peak area; c: retention time; d: quantity recovered

Table-3: Various pharmacokinetic parameters in healthy human volunteers following oral administration of fast dispersible tablets of Domperidone (10mg)

Parameter (Unit)	**Results**
C max (ng/ml)	24 ± 6.01
T max (min)	30 ± 5.37
AUC (ng.hr/ml)	168.7 ± 19.33
$t_{1/2}$ (hr)	6.1 ± 1.73

Results are presented as mean ± S.D. (n = 6)

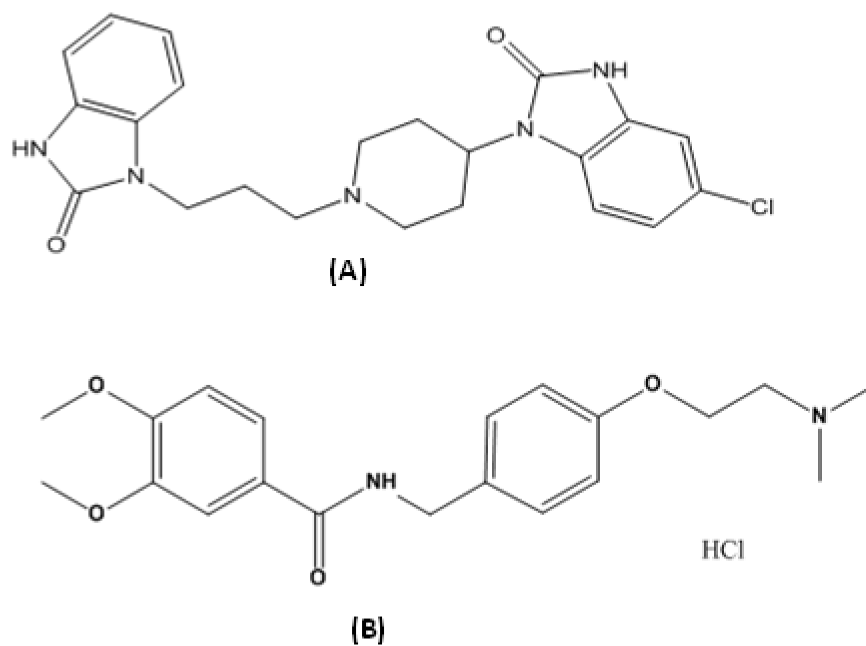

Figure-1: Chemical structures of; A) Domperidone, B) Itopride

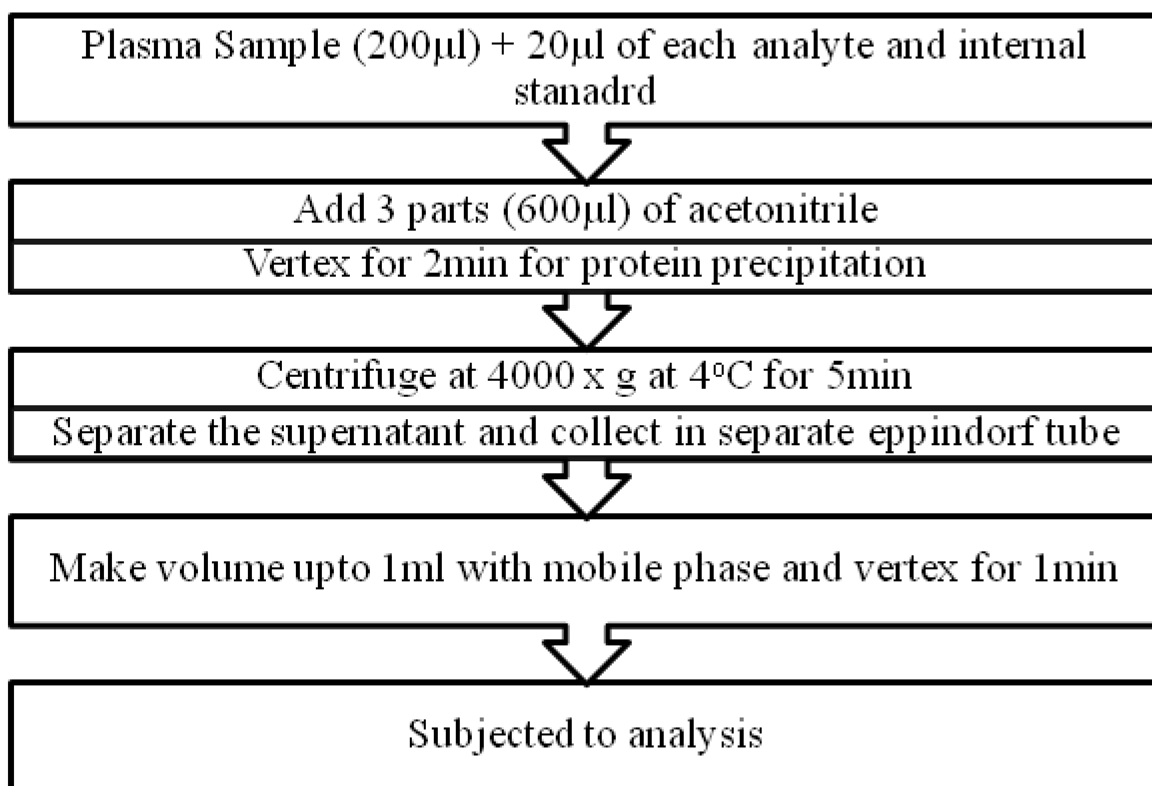

Figure-2: Extraction procedure of Domperidone and Itopride from human plasma

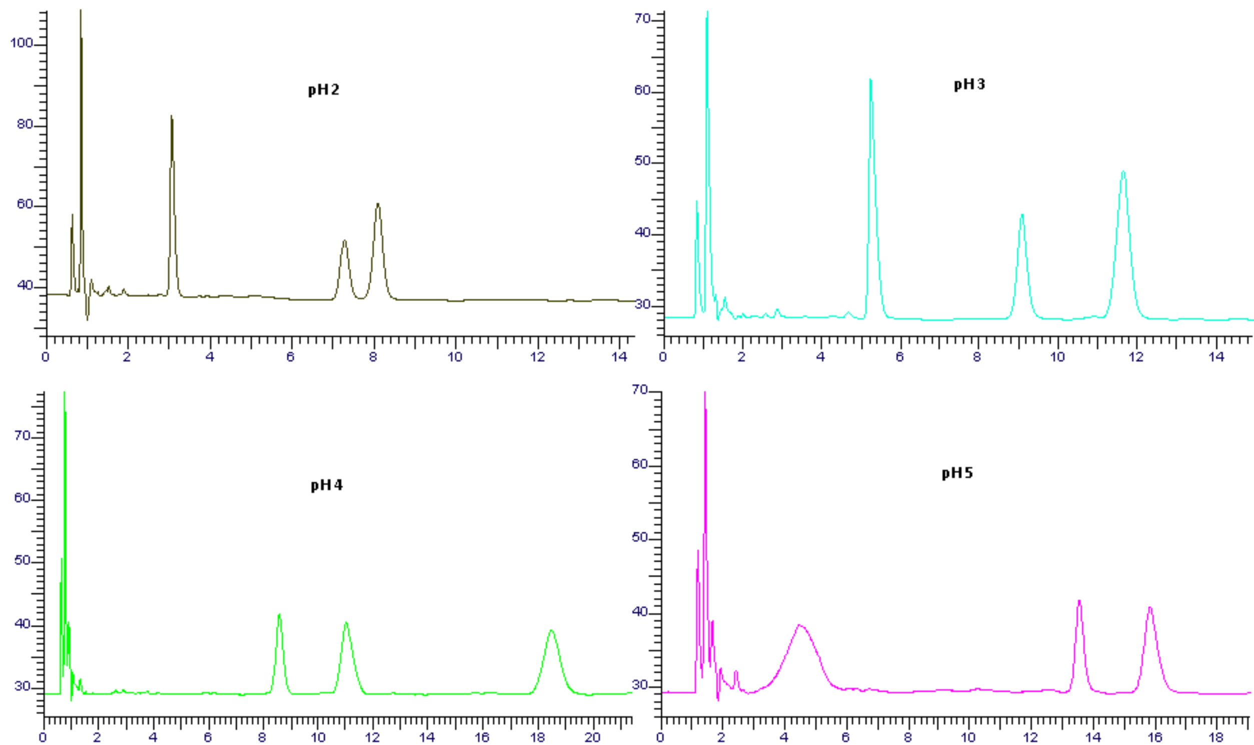

Figure-3: Effect of pH of mobile phase on various peak characteristics,
Where 1) Internal Standard; 2) Itopride; 3) Domperidone

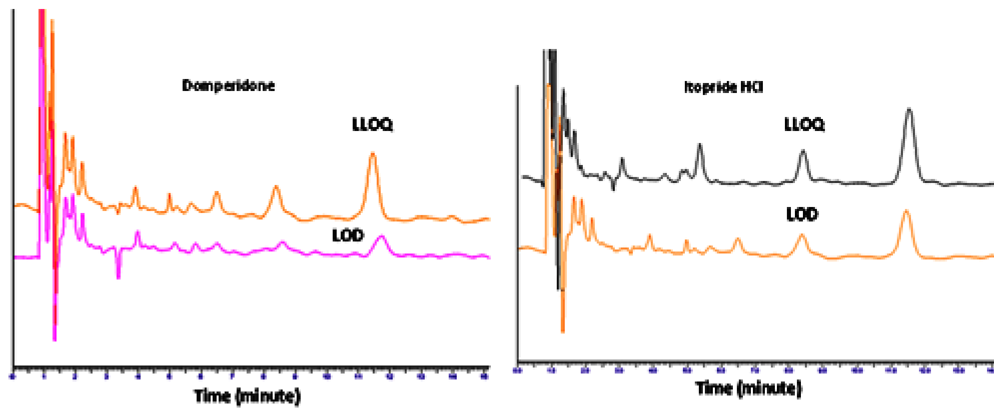

Figure-4: Chromatogram presenting lower limit of detection and lower limit of quantification of Domperidone and Itopride

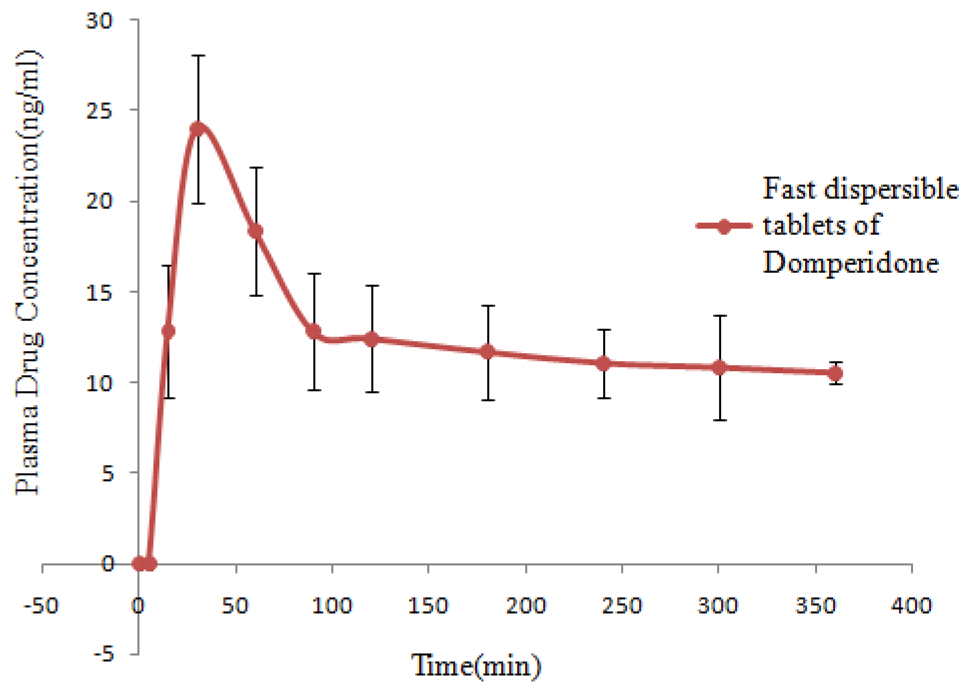

Figure-5: Plasma drug concentrations versus time curve for Domperidone given as fast dispersible tablets (10mg) single oral dose in healthy human volunteers (mean ± SD, n = 6)